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Magnetic resonance elastography of slow and fast shear waves illuminates differences in shear and tensile moduli in anisotropic tissue

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1 **Abstract**

2
3 Mechanical anisotropy is an important property of fibrous tissues; for example, the anisotropic
4 mechanical properties of brain white matter may play a key role in the mechanics of traumatic brain
5 injury (TBI). The simplest anisotropic material model for small deformations of soft tissue is a nearly
6 incompressible, transversely isotropic (ITI) material characterized by three parameters: minimum shear
7 modulus (μ), shear anisotropy ($\phi = \mu_1/\mu - 1$) and tensile anisotropy ($\zeta = E_1/E_2 - 1$). These
8 parameters can be determined using magnetic resonance elastography (MRE) to visualize shear
9 waves, if the angle between the shear-wave propagation direction and fiber direction is known. Most
10 MRE studies assume isotropic material models with a single shear (μ) or tensile (E) modulus. In this
11 study, two types of shear waves, “fast” and “slow”, were analyzed for a given propagation direction to
12 estimate anisotropic parameters μ , ϕ , and ζ in two fibrous soft materials: turkey breast *ex vivo* and
13 aligned fibrin gels. As expected, the speed of slow shear waves depended on the angle between fiber
14 direction and propagation direction. Fast shear waves were observed when the deformations due to
15 wave motion induced stretch in the fiber direction. Finally, MRE estimates of anisotropic mechanical
16 properties in turkey breast were compared to estimates from direct mechanical tests.

17 1. Introduction

18 Accurate characterization of soft tissue material properties is important to medical clinicians and
19 researchers. The identification of parameters for soft tissue is especially relevant to traumatic brain
20 injury (TBI) research, as it will enable more accurate mechanical modeling and simulation of TBI.
21 Magnetic resonance elastography (MRE) is a technique for non-invasive estimation of material
22 parameters in soft tissues. In MRE, shear waves are excited in the soft tissue; the resulting shear wave
23 speed is measured and used to estimate material parameters.

24 MRE was originally developed using isotropic, elastic material models (Muthupillai and Ehman,
25 1996; Muthupillai et al., 1995) and similar isotropic, elastic or viscoelastic models have been used to
26 characterize tissues such as liver (Asbach et al., 2008; Klatt et al., 2010a; Mariappan et al., 2009),
27 breast (Sinkus et al., 2005), and brain (Atay et al., 2008; Clayton et al., 2011a; Feng et al., 2013a;
28 Green et al., 2008; Johnson et al., 2013; Murphy et al., 2013; Sack et al., 2008). However, since
29 biological tissue is often anisotropic (Feng et al., 2013b), techniques used in MRE should be extended
30 to account for directionally-dependent material properties. Work has recently been published on
31 anisotropic material models in three general categories: theoretical studies, ultrasound elastography
32 studies (involving estimation of two or three parameters), and MRE studies involving estimation of two,
33 three, or five or more parameters. Rouze et al. (2013) showed agreement between predictions of a
34 three-parameter, incompressible, transversely isotropic (ITI) theory and a finite element model. Royer
35 et al. (2011) outlined an incompressible, transversely isotropic (ITI) model and approach for ultrasound
36 elastography. Gennisson et al. (2003) also using ultrasound elastography, studied transversely
37 isotropic phantoms and measured shear moduli parallel and perpendicular to the fibers. Other recent
38 ultrasound studies (Aristizabal et al., 2014; Wang et al., 2013) describe two different shear-wave
39 speeds in transversely isotropic phantoms.

40 Anisotropic MRE has most commonly been used to estimate two elastic parameters: the shear
41 moduli governing shear in planes parallel and perpendicular to the fiber direction. Such studies have
42 been performed by Sinkus et al. (2005) (breast tissue); Green et al. (2013), Klatt et al. (2010b),

43 Papazoglou et al. (2006), Qin et al. (2014, 2013) (muscle tissue); Qin et al. (2013) (anisotropic
44 phantoms); and Namani et al. (2009) (aligned fibrin gels). MRE can also be used to estimate three
45 parameters for ITI material models (Feng et al., 2013b; Guo et al., 2015) and five parameters for
46 general TI material models, or more for general orthotropic models (Romano et al., 2012). Papazoglou
47 et al. (2006) derived a three parameter model for skeletal muscle tissue and an approach for
48 estimation, though only two shear moduli were reported. Feng et al. (2013b) demonstrated that at least
49 three elastic parameters are required to describe white matter in the brain because of anisotropy in
50 shear and tensile moduli. Romano et al. (2012) identified five parameters in brain white matter
51 (corticospinal tracts) with the application of spatial-spectral filters and Helmholtz decomposition to
52 separate shear and pressure waves in a waveguide. These authors also applied their technique to
53 patients with amyotrophic lateral sclerosis (Romano et al., 2014). Guo et al. (2015) recently measured
54 three parameters in skeletal muscle by inverting the curl field measured by MRE. Tweten et al. (2015)
55 showed by simulation that two types of shear waves must exist, with propagation of both waves in
56 different directions, in order to estimate accurately the three material parameters.

57 While these recent studies have illustrated the plausibility of different approaches to estimate
58 parameters for TI material models, several key issues remain to be addressed. In most studies, the
59 performance of the anisotropic inversion algorithm has not been validated, either by using data from
60 simulations or from phantoms with known anisotropic properties. The notable exception is the study of
61 (Qin et al., 2013), which found close agreement between MRE and direct measurements of shear
62 anisotropy. Most importantly, in prior experimental work, the criterion that both slow (pure transverse, or
63 PT) and fast shear (quasi-transverse, or QT) waves must be present (at significant amplitudes, with
64 multiple directions) in the estimation region in order to estimate accurately all three ITI parameters
65 (Tweten et al., 2015) was not explicitly met. Many studies did not take into account the effects of tensile
66 moduli on the fast shear-wave speed. In this study, we explicitly separate wave fields into fast and slow
67 components in multiple propagation directions. We estimate the two separate wave speeds occurring in

68 transversely isotropic soft tissues and show how these wave speeds can be used to estimate the three
 69 elastic material parameters for a linear ITI model.

70

71 2. Methods

72 2.1 Theory: fast and slow shear-wave speeds in elastic, incompressible, transversely isotropic materials

73 In a linear ITI material model, there are three independent material parameters: shear modulus
 74 μ , shear anisotropy $\phi = \mu_1/\mu - 1$, and tensile anisotropy $\zeta = E_1/E_2 - 1$ (other parameters, which are
 75 linear combinations of these, can also be chosen; see Appendix A). “Slow” (PT) shear waves in ITI
 76 materials exhibit displacements perpendicular to both the wave propagation direction, \mathbf{n} , and the
 77 normal to the plane of isotropy (nominally the fiber direction), \mathbf{a} . The unit vector in the direction of
 78 displacement, or polarization direction, of the slow wave can be shown to be (Appendix A; also see
 79 Rouze et al., 2013; Tweten et al., 2015):

$$80 \quad \mathbf{m}_s = \mathbf{n} \times \mathbf{a} / |\mathbf{n} \times \mathbf{a}|. \quad [1]$$

81 (The special case when \mathbf{n} and \mathbf{a} are parallel, is discussed below). Since the slow shear-wave
 82 polarization is always perpendicular to the fiber direction, it does not stretch the fibers and wave speed
 83 depends only on the baseline shear modulus, μ , shear anisotropy, ϕ , and the angle, θ , between \mathbf{n}
 84 and \mathbf{a} (Appendix A):

$$85 \quad c_s^2 = (\mu/\rho)(1 + \phi \cos^2 \theta). \quad [2]$$

86 The polarization direction of a “fast” (QT) shear wave is perpendicular to both the propagation direction
 87 and the slow shear wave polarization direction and lies in the plane defined by \mathbf{n} and \mathbf{a} (Appendix A;
 88 also Rouze et al., 2013; Tweten et al., 2015):

$$89 \quad \mathbf{m}_f = \mathbf{n} \times \mathbf{m}_s \quad [3]$$

90 Fig.1 shows the relationship between \mathbf{m}_s , \mathbf{m}_f , \mathbf{n} and \mathbf{a} . Deformations associated with fast shear waves
 91 induce stretch in the fiber direction, and the speed of the fast shear wave thus depends on the tensile
 92 anisotropy, ζ (Appendix A; also Tweten et al., 2015):

$$c_f^2 = (\mu/\rho)(1 + \phi \cos^2 2\theta + \zeta \sin^2 2\theta). \quad [4]$$

93
94 Note that when the propagation direction \mathbf{n} is parallel to the fiber direction \mathbf{a} , Eq. 1 and Eq. 3 are
95 undefined, $\theta = 0$, and there is only one wave speed.

96 To extend the theory above to viscoelastic materials, the correspondence principle (Flügge,
97 1975) may be invoked, in which the complex shear modulus, $\mu^* = \mu' + i\mu''$, describes the relationship
98 between harmonic stress and strain in the plane of isotropy. If dissipative effects are due to fluid
99 motion, and thus approximately isotropic, the loss factor $\eta = \mu''/\mu'$ would govern waves in all directions.
100 In this study, each material is studied at a single frequency, so that μ represents the magnitude of the
101 complex modulus $|\mu^*|$, and ϕ and ζ the anisotropy in moduli, at that frequency.

102

103 2.2 Experiments: slow and fast shear waves in cylindrical and cube specimens

104 Detailed descriptions of materials and methods are included in Appendix B. Briefly, two different
105 fibrous, biological materials, turkey breast and magnetically-aligned fibrin gel, were studied to assess
106 wave speed differences in tissue based on fiber orientation. Two sample geometries, cylindrical and
107 cube, were used for both materials (Fig. 2, Appendix B). Cylindrical (45 mm diameter) samples of both
108 turkey breast and fibrin gel were embedded in a gelatin mixture (Okamoto et al., 2011) in a cylindrical
109 container (Fig. 2 (a, d, f) and excited by vertical vibrations of a central axial rod. This setup produced
110 shear waves with approximately radial propagation ($\mathbf{n} \approx \mathbf{e}_r$). Cube-shaped specimens of turkey breast
111 and aligned fibrin with fibers oriented $\sim 45^\circ$ downward from the top surface were also imaged. Waves in
112 cube specimens were produced by lateral vibration of the top surface (Fig. 2 (b, c, e)). Two excitation
113 cases (Fig. 2 (b, c)) were applied to each cube sample. In one case (Fig. 2 (b)) the actuation
114 direction, \mathbf{m} , was aligned with $\mathbf{m}_s = \mathbf{n} \times \mathbf{a}$ to excite slow shear waves and in the other case (Fig. 2 (c))
115 the actuation was aligned with $\mathbf{m}_f = \mathbf{n} \times \mathbf{m}_s$ to excite fast shear waves. Frequencies of actuation were
116 chosen to produce multiple wavelengths in the specimen, and multiple voxels per wavelength.

117 Images of shear-wave propagation in cylindrical and cube samples of turkey breast and aligned
118 fibrin gel were acquired using previously-described spin-echo MRE sequences (Clayton et al., 2011a;
119 Appendix B). To verify the average fiber orientation in turkey breast, diffusion tensor imaging (DTI) was
120 performed in the same imaging session as MRE. Figure 3 shows a three-dimensional diffusion tensor
121 field to indicate the fiber orientation inside a representative cube turkey breast sample. The relatively
122 sparse networks of aligned fibrils in fibrin gels do not constrain water diffusion and thus DTI was not
123 performed on the fibrin gels. Instead the direction of the magnetic field during the magnetic aligning
124 process was physically marked on the sample container and was noted during all tests.

125 For comparison with MRE, estimates of viscoelastic shear modulus from dynamic shear testing
126 (DST) were obtained in separate circular samples of turkey breast following techniques in (Feng et al.,
127 2013b; Namani et al., 2012) (see Appendix B).

128

129 2.3 Image analysis: characterization of anisotropic wave propagation

130 First, for a simple measure of anisotropy, radially-propagating shear waves in anisotropic
131 cylindrical specimens were fitted to ellipses (Fig. 4). Peaks of the elliptical wavefront in multiple
132 directions were manually picked on the 2D image of axial (w) displacement in the xy plane, in five
133 contiguous slices from each sample. Ellipses were fitted using an algorithm that minimized the squared-
134 error between the curves and the picked points (Fitzgibbon et al., 1999).

135 Second, directional filtering (Appendix C) was used to isolate slow and fast shear waves in
136 specific propagation and polarization directions. Figs. 5 (b, c) and 6 (c, d) show examples of
137 directionally filtered waves in cylindrical and cube samples, respectively. From directionally filtered
138 wave fields, peaks and valleys were selected manually to capture the wavelength. The wavelength in
139 each of the 16 directions was averaged over 5 representative slices. Values of average wave speed as
140 a function of angle, θ , were fitted to Eq. 2 using a weighted, least-squares fitting algorithm to estimate
141 the slow wave-speed parameters, μ and ϕ .

142 To characterize dissipative (viscous) effects, attenuation per wavelength was found from the
143 ratios of amplitudes of successive peaks in directionally filtered waves (Appendix C; Figure C1).

144

145 3. Results

146 3.1 Imaging experiments

147 Wave patterns consistent with a transversely isotropic (TI) material model were observed in
148 both cylindrical and cube samples. Axially-excited cylindrical samples (actuation in the m_s direction)
149 exhibited the slow shear-wave patterns predicted by mathematical models (Tweten et al., 2015).
150 Elliptical waves were observed in specimens with a dominant fiber orientation (Fig. 4 (a, b, c)), and
151 circular waves were observed in isotropic gel (Fig. 4 (d)), in planes perpendicular to the cylinder axis
152 (Fig. 2 (a)). For ellipses fitted to radially-propagating wavefronts in axially excited cylindrical specimens,
153 the ratio of major semi-axes and minor semi-axes was used to describe shear anisotropy. The average
154 ratio of semi-axes was found to be 1.65 ± 0.24 for turkey breast (800 Hz, $n=4$), 1.37 ± 0.14 for aligned
155 fibrin (200 Hz, $n = 3$), and 1.03 ± 0.01 for isotropic gelatin (200 Hz, $n = 3$).

156 Analysis of the directionally-filtered wave speeds in cylindrical specimens (Fig. 9(a, c)) revealed
157 dependence of slow shear-wave speed, c_s , on the angle between fiber orientation and propagation
158 direction, θ (Fig. 5). The observed dependence is consistent with theoretical predictions based on Eq.
159 2 (red curves) with fitted parameters. The parameters estimated for turkey breast were found to be
160 $\mu = 33.1 \pm 11.4$ kPa and $\phi = 1.3 \pm 0.7$ for turkey breast (800 Hz, $n = 4$). For aligned fibrin gel,
161 $\mu = 1.1 \pm 0.5$ kPa and $\phi = 1.1 \pm 0.2$ (200 Hz, $n = 3$). Close agreement between average fiber direction
162 estimated from DTI (turkey breast) or magnetically induced alignment direction (fibrin) and the direction
163 of fastest shear-wave propagation confirms that aligned fibers produce mechanical anisotropy.

164 In cube specimens, strong evidence for the importance of both slow and fast shear waves is
165 provided by the dependence of propagation speed on polarization direction. Polarization direction was
166 controlled by actuation direction. When the actuation direction was in the plane containing the fibers

167 and at an angle of $\sim 45^\circ$ from the fiber axis in this plane, displacements were induced in the m_f
 168 direction. In this case, a substantial component of stretch is in the direction parallel to the fibers, the
 169 cubes exhibited primarily fast shear waves (Figs. 6(a) and 8(a,c)). In contrast, when the polarization
 170 direction was perpendicular to the fibers, in the m_s direction, the cube exhibited slow shear waves
 171 (Figs. 6(b) and 8(b,d)). The difference between fast and slow shear wave speed in this sample is
 172 largely due to tensile anisotropy, characterized by the parameter ζ . This parameter was estimated from
 173 the ratio between slow and fast shear-wave speeds (Fig 9(b,d)), using Eqs. 2 and 4. Because Eqs. 2
 174 and 4 contain three unknown parameters (after measuring the slow and fast wave speed and $\theta=45^\circ$),
 175 the average value of ϕ estimated from cylindrical specimens was used to solve for both μ and ζ . For
 176 turkey breast cubes, the estimated parameters are $\mu = 32.2 \pm 16.8$ kPa and $\zeta = 9.2 \pm 4.9$ (800 Hz, $n =$
 177 5) and for aligned fibrin gel the estimated parameters are $\mu = 4.5$ kPa and $\zeta = 2.7$ (200 Hz, $n = 1$).

178 The attenuation per wavelength (ratio between successive peaks) for slow shear waves in
 179 turkey breast was $r = 0.27 \pm 0.18$, corresponding to a loss factor of $\eta = \mu''/\mu' = 0.43 \pm 0.17$ (Auld,
 180 1990). In fibrin the attenuation per wavelength was $r = 0.50 \pm 0.21$ ($\eta = 0.22 \pm 0.11$) and in gelatin the
 181 attenuation ratio was $r = 0.67 \pm 0.13$ ($\eta = 0.13 \pm 0.06$). No statistically significant differences in
 182 attenuation per wavelength were found for different directions of propagation in MRE.

183 Parameter estimates are summarized in Table 1. The amplitudes of slow and fast shear waves
 184 turkey breast and fibrin are summarized in Table D2 in Appendix D.

185

186 3.2 Direct mechanical characterization

187 Parallel and perpendicular shear moduli for turkey breast ($N = 33$) were averaged over the
 188 frequencies from 30-40 Hz using DST. The storage modulus was found to be $\mu'_{\parallel} = 4.8 \pm 1.6$ kPa when
 189 fibers were aligned parallel to the direction of imposed shear displacement and $\mu'_{\perp} = 3.2 \pm 1.1$ kPa
 190 when fibers were aligned perpendicular to the shear displacement. The loss modulus was $\mu''_{\parallel} = 2.2 \pm$
 191 0.7 kPa for the parallel orientation and $\mu''_{\perp} = 1.1 \pm 0.3$ kPa for the perpendicular orientation. The ratio

192 between parallel and perpendicular moduli $\mu'_{\parallel}/\mu'_{\perp} = 1.5 \pm 0.3$ for storage modulus, $\mu''_{\parallel}/\mu''_{\perp} = 2.0 \pm 0.3$ for
193 loss modulus, and $\mu_{\parallel}/\mu_{\perp} = 1.6 \pm 0.3$ ($\phi = 0.6 \pm 0.3$) for the magnitude. Fig. 10 summarizes these data.

194

195 4. Discussion

196 In this experimental study, shear waves were imaged using MR elastography procedures in
197 both muscle tissue *ex vivo* (turkey breast) and aligned fibrin gel. In axially-excited cylindrical samples,
198 slow shear waves were found to propagate with elliptical wave fronts through both transversely
199 isotropic materials, and with circular wave fronts in an isotropic medium (gelatin). In cube samples
200 excited by tangential vibration on one face, measurements of slow and fast shear waves with differing
201 polarization direction showed the effects of tensile anisotropy on wave speed. These results confirm
202 that MRE can detect anisotropic shear moduli and tensile moduli in these two prototypical soft, fibrous,
203 materials.

204 Direct mechanical tests (DST) in the current study confirmed mechanical anisotropy in turkey
205 breast. Values of the shear anisotropy ratio from DST (turkey breast $\phi \sim 0.6$, or $\mu_{\parallel}/\mu_{\perp} \sim 1.6$ at 30-40 Hz)
206 were comparable to values estimated from MRE ($\phi \sim 1.3$, or $\mu_{\parallel}/\mu_{\perp} \sim 2.3$, at 800 Hz). While both tests
207 give ratios of shear modulus near 2, the fact that MRE estimates of anisotropy at 800 Hz are about 40%
208 higher than DST at 30-40 Hz must be acknowledged. This difference may reflect limitations of each
209 method. Accuracy and precision of MRE estimates are limited by the practical challenges of wavelength
210 estimation from discretely sampled data with limited resolution. Results from DST may be affected by
211 slip, nonlinearity or non-affine deformation. Alternatively, shear anisotropy may truly depend on
212 frequency; the frequency ranges of the instruments in this study precluded a direct test of this
213 possibility, but it is a topic for future investigation. Although DST of turkey breast was done at much
214 lower frequencies (30-40 Hz) than in MRE (800 Hz), the values of the baseline storage modulus ($\mu \sim 4$
215 kPa at 30 Hz and $\mu \sim 33$ kPa at 800 Hz) are consistent with the expected increase in modulus with
216 frequency in viscoelastic muscle tissue. Both DST and MRE estimates of μ are consistent with a

217 previous MRE study of the viscoelastic properties of bovine muscle *ex vivo*, in which estimates of
218 (isotropic) shear modulus increase from $\mu \sim 12$ kPa at 200 Hz to $\mu \sim 35$ kPa at 800 Hz (Riek et al., 2011).

219 In a previous study of aligned fibrin (Namani et al., 2012), DST also detected anisotropy in
220 shear ($\phi \sim 0.9$, at 20-40 Hz). This result is similar to estimates of shear anisotropy of aligned fibrin from
221 MRE in the current study ($\phi \sim 0.6$ from measured wavelengths at 200 Hz). Also, the tensile anisotropy
222 parameter for aligned fibrin estimated from MRE ($\zeta \sim 2.1$ at 600 Hz) in this study is comparable to the
223 tensile anisotropy estimated from asymmetric indentation of aligned fibrin ($\zeta \sim 3.5$, quasi-static) in
224 previous work (Namani et al., 2012).

225 The current results demonstrate the ability to both (1) characterize two distinct shear-wave
226 types in soft transversely isotropic materials using MRE, and (2) use these shear wave measurements
227 to estimate three elastic parameters. Notably, tensile anisotropy can cause large differences between
228 slow and fast shear-wave speeds. Both fast and slow shear waves exhibit direction-dependent
229 propagation speed. For slow shear waves, only shear anisotropy causes this directional dependence;
230 tensile anisotropy plays no role in slow shear wave speed. In contrast, the directional dependence of
231 fast shear wave speed is due to both shear anisotropy and tensile anisotropy. The estimation of
232 mechanical parameters by directional filtering and isolation of separate shear-wave components is
233 robust to noise (Tweten et al., 2015), since it does not rely on multiple numerical derivatives.

234 Romano et al. (2012, 2014) also used directional filters to isolate shear wave components.
235 These authors analyzed wave speeds for a set of propagation and polarization directions aligned with a
236 specific fiber tract, and estimated five to nine components of the elasticity tensor. For nearly-
237 incompressible materials, elements of the elasticity tensor will have widely varying magnitude, as some
238 elements approach infinity due to the contribution of the bulk modulus (see Appendix A). In the current
239 approach, only three parameters (μ , ϕ , and ζ) are sought, which for incompressible materials,
240 completely specifies the compliance tensor. An advantage of the current approach is that components
241 of the compliance tensor converge to finite values as the bulk modulus approaches infinity.

242 The current material model is constrained by the assumptions of linear elasticity and
243 incompressibility. In applications *in vivo*, because tissue is slightly compressible and excitation may
244 have a longitudinal component, longitudinal waves must be considered. The current method can be
245 applied even in the presence of longitudinal waves, because fast and slow shear waves are isolated by
246 a combination of directional filtering and projection onto the corresponding polarization directions. In the
247 isotropic case, taking the curl of the displacement field eliminates the contributions of longitudinal
248 waves. In the anisotropic case, the curl of the displacement field does not uncouple the equations.
249 However, Guo et al. (2015) were able to estimate transversely isotropic material parameters from the
250 curl field; the assumption of incompressibility was used to obtain the equations used in the inversion.

251 Neither dissipation (viscosity, or complex shear modulus) nor nonlinearity is addressed
252 comprehensively. The deformations in the current study are small (<1% strain) and thus linear theory is
253 applicable. Though the focus of the current study is on elastic anisotropy, viscoelastic effects are clearly
254 important for describing the complete response of tissue (Clayton et al., 2011b; Green et al., 2008; Qin
255 et al., 2013). Dissipative effects are often attributed to fluid viscosity and approximated as isotropic, in
256 which case the directional dependence of shear wave speeds can be attributed to anisotropy of elastic
257 moduli. While this model was not rigorously tested in these materials, our observations of wave
258 attenuation in MRE are consistent with an approximately isotropic loss factor (Appendix C, Figure C1).
259 Current estimates of loss factor from attenuation of waves in MRE in turkey breast ($\eta = 0.43$) are similar
260 to those observed in prior studies of muscle ($\eta \approx 0.4$, Riek et al., 2011), as well as to the ratios of loss
261 and storage moduli estimated by DST (Figure 10). Current estimates from MRE in fibrin ($\eta = 0.22$)
262 approximate those from a prior study of fibrin ($\eta \approx 0.2$; Namani et al., 2012). The estimated loss factor in
263 gelatin ($\eta = 0.13$) also agrees with prior observations ($\eta \approx 0.1$; Okamoto et al., 2011). The elastic
264 constants estimated here may be interpreted as effective moduli at the given experimental frequency.
265 In future work, complex moduli could be determined as functions of frequency.

266 The current study focused on the physics of wave propagation in ITI materials, and hence
267 simple, manual methods were used to estimate wavelength and wave speed. This approach was

268 chosen to separate the characterization of physical phenomena from the performance of automated
269 algorithms. Future studies and related work (Tweten et al., 2015, e.g.) will address the development of
270 robust, automated methods for estimating wavelengths of slow and fast shear waves in TI materials.

271 Samples of roughly uniform material were used in this study. The current approach, in which the
272 wavelength is estimated and averaged over five slices and then the average substituted into the
273 equations to estimate the shear moduli, is adequate for globally homogenous materials, such as ex vivo
274 specimens and phantoms. In actual tissue measurements in vivo, tissue homogeneity may only be
275 assumed very locally. More work is needed to address parameter estimation in heterogeneous
276 materials such as brain tissue *in vivo*. In such materials, it is critically important that both slow and fast
277 shear waves propagating in multiple directions are present in a given volume, in order to obtain valid
278 estimates of transversely anisotropic material properties in that region.

279

280 5. Conclusion

281 Both slow and fast shear waves propagate in soft, fibrous, materials and can be imaged by
282 MRE. Shear-wave speed depends on the angles between propagation direction, polarization direction,
283 and fiber direction. Three elastic parameters may be estimated from these data, allowing for concise
284 characterization of nearly-incompressible, transversely isotropic materials. Such material
285 characterization can lead to improved modeling of white matter in the brain and a greater
286 understanding of TBI.

287

288 **Conflict of Interest Statement**

289 None of the authors have a conflict of interest that could influence the work described in this
290 manuscript.

291

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294 **References**

- 295 Aristizabal, S., Amador, C., Qiang, B., Kinnick, R.R., Nenadic, I.Z., Greenleaf, J.F., Urban, M.W., 2014.
296 Shear wave vibrometry evaluation in transverse isotropic tissue mimicking phantoms and skeletal
297 muscle. *Phys. Med. Biol.* 59, 7735–7752.
- 298 Asbach, P., Klatt, D., Hamhaber, U., Braun, J., Somasundaram, R., Hamm, B., Sack, I., 2008.
299 Assessment of liver viscoelasticity using multifrequency MR elastography. *Magn. Reson. Med.* 60,
300 373–379.
- 301 Atay, S.M., Kroenke, C.D., Sabet, A., Bayly, P. V, 2008. Measurement of the dynamic shear modulus of
302 mouse brain tissue in vivo by magnetic resonance elastography. *J. Biomech. Eng.* 130.
- 303 Auld, B.A., 1990. *Acoustic fields and waves in solids*, 2nd ed. Krieger Publishing Company, New York,
304 NY.
- 305 Clayton, E.H., Garbow, J.R., Bayly, P. V, 2011a. Frequency-dependent viscoelastic parameters of
306 mouse brain tissue estimated by MR elastography. *Phys. Med. Biol.* 56, 2391–2406.
- 307 Clayton, E.H., Garbow, J.R., Bayly, P. V, 2011b. Frequency-dependent viscoelastic parameters of
308 mouse brain tissue estimated by MR elastography. *Phys Med Biol* 56, 2391–2406.
- 309 Feng, Y., Clayton, E.H., Chang, Y., Okamoto, R.J., Bayly, P. V, 2013a. Viscoelastic properties of the
310 ferret brain measured in vivo at multiple frequencies by magnetic resonance elastography. *J.*
311 *Biomech.* 46, 863–870.
- 312 Feng, Y., Okamoto, R.J., Namani, R., Genin, G.M., Bayly, P. V., 2013b. Measurements of mechanical
313 anisotropy in brain tissue and implications for transversely isotropic material models of white
314 matter. *J. Mech. Behav. Biomed. Mater.* 23, 117–132.
- 315 Fitzgibbon, A., Pilu, M., Fisher, R.B., 1999. Direct least square fitting of ellipses. *IEEE Trans. Pattern*
316 *Anal. Mach. Intell.* 21, 476–480.
- 317 Flügge, W., 1975. *Viscoelasticity*, 2d rev. ed. Springer-Verlag, New York, NY.
- 318 Gennisson, J.L., Catheline, S., Chaffai, S., Fink, M., 2003. Transient elastography in anisotropic
319 medium: Application to the measurement of slow and fast shear wave speeds in muscles. *J.*
320 *Acoust. Soc. Am.* 114, 536–541.
- 321 Green, M. a., Geng, G., Qin, E., Sinkus, R., Gandevia, S.C., Bilston, L.E., 2013. Measuring anisotropic
322 muscle stiffness properties using elastography. *NMR Biomed.* 26, 1387–1394.
- 323 Green, M.A., Bilston, L.E., Sinkus, R., 2008. In vivo brain viscoelastic properties measured by magnetic
324 resonance elastography. *NMR Biomed.* 21, 755–764.
- 325 Guo, J., Hirsch, S., Scheel, M., Braun, J., Sack, I., 2015. Three-parameter shear wave inversion in MR
326 elastography of incompressible transverse isotropic media: Application to in vivo lower leg
327 muscles. *Magn. Reson. Med.* In press. Available online doi: 10.1002/mrm.25740.
- 328 Holzapfel, G.A., 2000. *Nonlinear Solid Mechanics: A Continuum Approach for Engineering*. John Wiley

- 329 & Sons, Ltd.
- 330 Johnson, C.L., McGarry, M.D.J., Van Houten, E.E.W., Weaver, J.B., Paulsen, K.D., Sutton, B.P.,
331 Georgiadis, J.G., 2013. Magnetic resonance elastography of the brain using multishot spiral
332 readouts with self-navigated motion correction. *Magn. Reson. Med.* 70, 404–412.
- 333 Klatt, D., Friedrich, C., Korth, Y., Vogt, R., Braun, J., Sack, I., 2010a. Viscoelastic properties of liver
334 measured by oscillatory rheometry and multifrequency magnetic resonance elastography.
335 *Biorheology* 47, 133–141.
- 336 Klatt, D., Papazoglou, S., Braun, J., Sack, I., 2010b. Viscoelasticity-based MR elastography of skeletal
337 muscle. *Phys. Med. Biol.* 55, 6445–6459.
- 338 Knutsson, H., Westin, C.-F., Granlund, G., 1994. Local multiscale frequency and bandwidth estimation.
339 In: *Proceedings of 1st International Conference on Image Processing*. IEEE Comput. Soc. Press,
340 pp. 36–40.
- 341 Manduca, A., Lake, D.S., Kruse, S.A., Ehman, R.L., 2003. Spatio-temporal directional filtering for
342 improved inversion of MR elastography images. *Med. Image Anal.* 7, 465–473.
- 343 Mariappan, Y.K., Glaser, K.J., Manduca, A., Romano, A.J., Venkatesh, S.K., Yin, M., Ehman, R.L.,
344 2009. High-frequency mode conversion technique for stiff lesion detection with magnetic
345 resonance elastography (MRE). *Magn. Reson. Med.* 62, 1457–1465.
- 346 Murphy, M.C., Huston, J., Jack, C.R., Glaser, K.J., Senjem, M.L., Chen, J., Manduca, A., Felmlee, J.P.,
347 Ehman, R.L., 2013. Measuring the characteristic topography of brain stiffness with magnetic
348 resonance elastography. *PLoS One* 8, 1–14.
- 349 Muthupillai, R., Ehman, R.L., 1996. Magnetic resonance elastography. *Nat. Med.* 2, 601–603.
- 350 Muthupillai, R., Lomas, D.J., Rossman, P.J., Greenleaf, J.F., Manduca, A., Ehman, R.L., 1995.
351 Magnetic resonance elastography by direct visualization of propagating acoustic strain waves.
352 *Science* 269, 1854–7.
- 353 Namani, R., Feng, Y., Okamoto, R.J., Jesuraj, N., Sakiyama-Elbert, S.E., Genin, G.M., Bayly, P. V.,
354 2012. Elastic characterization of transversely isotropic soft materials by dynamic shear and
355 asymmetric indentation. *J. Biomech. Eng.* 134, 061004.
- 356 Namani, R., Wood, M.D., Sakiyama-Elbert, S.E., Bayly, P. V., 2009. Anisotropic mechanical properties
357 of magnetically aligned fibrin gels measured by magnetic resonance elastography. *J. Biomech.* 42,
358 2047–2053.
- 359 Okamoto, R.J., Clayton, E.H., Bayly, P. V., 2011. Viscoelastic properties of soft gels: comparison of
360 magnetic resonance elastography and dynamic shear testing in the shear wave regime. *Phys.*
361 *Med. Biol.* 56, 6379–400.
- 362 Papazoglou, S., Rump, J., Braun, J., Sack, I., 2006. Shear wave group velocity inversion in MR
363 elastography of human skeletal muscle. *Magn. Reson. Med.* 56, 489–497.
- 364 Qin, E.C., Lauriane, J., Lambert, S.A., Paradis, V., Sinkus, R., Bilston, L.E., 2014. In Vivo Anisotropic

- 365 Mechanical Properties of Dystrophic Skeletal Muscles Measured by Anisotropic MR Elastographic
366 Imaging : The mdx Mouse Model of Muscular Dystrophy. *Radiology* 273, 726 – 735.
- 367 Qin, E.C., Sinkus, R., Geng, G., Cheng, S., Green, M., Rae, C.D., Bilston, L.E., 2013. Combining MR
368 elastography and diffusion tensor imaging for the assessment of anisotropic mechanical
369 properties: a phantom study. *J. Magn. Reson. Imaging* 37, 217–226.
- 370 Riek, K., Klatt, D., Nuzha, H., Mueller, S., Neumann, U., Sack, I., Braun, J., 2011. Wide-range dynamic
371 magnetic resonance elastography. *J. Biomech.* 44, 1380–1386.
- 372 Romano, A., Guo, J., Prokscha, T., Meyer, T., Hirsch, S., Braun, J., Sack, I., Scheel, M., 2014. In vivo
373 waveguide elastography: Effects of neurodegeneration in patients with amyotrophic lateral
374 sclerosis. *Magn. Reson. Med.* 1761, 1755–1761.
- 375 Romano, A., Scheel, M., Hirsch, S., Braun, J., Sack, I., 2012. In vivo waveguide elastography of white
376 matter tracts in the human brain. *Magn. Reson. Med.* 68, 1410–1422.
- 377 Rouze, N.C., Wang, M.H., Palmeri, M.L., Nightingale, K.R., 2013. Finite element modeling of impulsive
378 excitation and shear wave propagation in an incompressible, transversely isotropic medium. *J.*
379 *Biomech.* 46, 2761–2768.
- 380 Royer, D., Gennisson, J.L., Deffieux, T., Tanter, M., 2011. On the elasticity of transverse isotropic soft
381 tissues. *J. Acoust. Soc. Am.* 129, 2757–2760.
- 382 Sack, I., Beierbach, B., Hamhaber, U., Klatt, D., Braun, J., 2008. Non-invasive measurement of brain
383 viscoelasticity using magnetic resonance elastography. *NMR Biomed.* 21, 265–271.
- 384 Sinkus, R., Tanter, M., Catheline, S., Lorenzen, J., Kuhl, C., Sondermann, E., Fink, M., 2005. Imaging
385 anisotropic and viscous properties of breast tissue by magnetic resonance-elastography. *Magn.*
386 *Reson. Med.* 53, 372–387.
- 387 Tweten, D.J., Okamoto, R.J., Schmidt, J.L., Garbow, J.R., Bayly, P. V, 2015. Estimation of material
388 parameters from slow and fast shear waves in an incompressible, transversely isotropic material.
389 *J. Biomech.* 48, 4002–4009.
- 390 Wang, M., Byram, B., Palmeri, M., Rouze, N., Nightingale, K., 2013. Imaging transverse isotropic
391 properties of muscle by monitoring acoustic radiation force induced shear waves using a 2-D
392 matrix ultrasound array. *IEEE Trans. Med. Imaging* 32, 1671–1684.

393

394 **Table 1**

	Turkey breast		Aligned fibrin gel	
	Cylinder (800 Hz, N=4)	Cube (800 Hz, N=5)	Cylinder (200 Hz, N=3)	Cube (600 Hz, N=1)
μ kPa	33.1 ± 11.4	32.2 ± 16.8	1.1 ± 0.5	4.7
ϕ	1.3 ± 0.7	NA*	1.1 ± 0.2	NA*
ζ	NA	9.2 ± 4.9	NA	2.7
η	0.43 ± 0.17		0.22 ± 0.11	

395

396 **Table 1:** Summary of incompressible, transversely isotropic (ITI) material parameter estimates from
 397 MRE of turkey breast muscle tissue and aligned fibrin gel. Parameters are: baseline shear modulus, μ ;
 398 shear anisotropy, ϕ ; tensile anisotropy, ζ ; loss factor, η .

399 (* To estimate ζ in cube specimens, ϕ was set to the value measured in cylindrical specimens.)

400 **Figures and captions**

401

402 **Figure 1.** The propagation direction (denoted by unit vector \mathbf{n}) and polarization directions (unit vectors
403 \mathbf{m}_s and \mathbf{m}_f) of slow and fast shear waves, respectively, in an incompressible, transversely isotropic,
404 elastic material. The unit vector \mathbf{a} denotes the normal to the plane of isotropy.

405

406 **Figure 2.** Schematic diagrams of: (a) cylindrical specimen with axial excitation; (b) cube specimen with
407 tangential excitation in a plane parallel to the fiber direction to induce “fast” shear waves. (c) cube
408 specimen with tangential excitation perpendicular to the dominant fiber direction to induce “slow” shear
409 waves. (d) Photograph of cylindrical turkey breast specimen embedded in gelatin (corresponding to
410 panel a). (e) Photograph of experimental setup for cube turkey breast (corresponding to panel b;
411 actuator on left). (f) Photograph of a cylindrical sample placed in RF coil with actuator on right.

412

413 **Figure 3.** Fiber orientation estimated by DTI in (a) cylindrical and (b) cube specimens of turkey breast.
414 Maximum principal diffusion direction vectors (cyan) are superimposed on fractional anisotropy maps
415 (FA, grey) for each voxel.

416

417 **Figure 4.** Wave propagation in axially-excited, cylindrical specimens. (a-c) Representative images of
418 elliptical waves exhibiting direction-dependent propagation with different wave speeds in different
419 directions. (a) Representative sample #1 of turkey breast, 800 Hz; (b) Representative sample #2 of
420 turkey breast, 800 Hz; (c) aligned fibrin gel, 200 Hz. (d) Circular waves in (isotropic) gelatin, 200 Hz. (e)
421 Ellipses were fitted to the wave images (white and black lines in b-d) and the average ratios of their
422 semi-axes are shown for the different materials.

423

424 **Figure 5.** Wave propagation in a cylindrically aligned fibrin gel (200 Hz actuation) specimen, illustrating
425 analysis by directional filtering. (a) Elliptical waves exhibiting direction dependent propagation with
426 different wave speeds in different directions. (b-c) Displacement field after directional filtering in each of
427 two propagation directions specified by angle, θ , from the dominant fiber direction. (b) $\theta = 0^\circ$ and (c) $\theta =$
428 90° .

429

430 **Figure 6.** Wave propagation in a cube specimen of aligned fibrin with dominant fiber direction at 45°
431 from horizontal (Figures 1(b,c)), illustrating analysis by directional filtering. (a) Excitation (600 Hz) in
432 the m_f direction (with a component along the fibers, as in Figure 1(b)) leads to predominantly
433 downward-propagating fast shear waves. (b) Excitation (600 Hz) in the m_s direction, perpendicular to
434 the fibers, as in Figure 1(c), leads to predominantly downward-propagating slow shear waves. Panels
435 (c,d): Directionally filtered waves in the [0 -1 0] direction corresponding to panels (a,b) respectively.

436

437 **Figure 7.** Average (\pm std. deviation) slow shear-wave speeds (blue *) plotted vs the angle between
438 propagation direction and the horizontal axis of the cylinder, in cylindrical specimens. (a)
439 Representative sample #1 of turkey breast (800 Hz). (b) Representative sample #2 of turkey breast
440 (800 Hz). (c) Aligned fibrin gel (200 Hz). (d) Gelatin (200 Hz). Each plot is for a single sample; average
441 values for each direction are computed over 5 slices. Theoretical curves (red lines) are obtained from
442 Eq. 2 using values of μ and ϕ estimated by weighted, least-squares fitting for each sample.

443

444 **Figure 8.** Wave propagation visualized by MRE in cube samples with different directions of excitation
445 relative to fiber orientation. Fibers are oriented approximately 45° from horizontal as in Figure 2(b,c).
446 Top panels (a,b) show fast and slow wave propagation in turkey breast actuated at 800 Hz and bottom
447 panels (c,d) show aligned fibrin actuated at 600 Hz. Left panels (a,c): Actuation in the m_f direction with
448 a component along the fibers (as in Figure 2(b)) leads to downward-propagating, fast shear waves.

449 Right panels (b,d): Actuation in the m_s direction, perpendicular to the fibers (as in Figure 2(c) leads to
 450 downward-propagating, slow shear waves.

451

452 **Figure 9:** Summary of shear-wave speeds in turkey breast (a, b) and aligned fibrin (c, d) at different
 453 angles θ of propagation direction relative to fiber direction. (a) Slow shear-wave speed in cylindrical
 454 turkey breast specimens (800 Hz, N = 4 samples). Estimated material parameters: $\mu = 33.1 \pm 11.4$
 455 kPa, $\phi = 1.3 \pm 0.7$ (b) Fast and slow shear-wave speeds in cube specimens (800 Hz, N=5). Estimated
 456 parameters: $\mu = 33.2 \pm 16.7$, $\zeta = 9.2 \pm 4.9$. (c) Slow shear-wave speeds in a cylindrical fibrin
 457 specimen (200 Hz, N=3). Estimated material parameters: $\mu = 1.1 \pm 0.5$ kPa, $\phi = 1.1 \pm 0.2$ (d) Average
 458 fast and slow shear-wave speeds in a cube specimen of aligned fibrin (600 Hz, N=1). Estimated
 459 parameters: $\mu = 4.7$ kPa, $\zeta = 2.7$.

460

461 **Figure 10:** Storage (elastic) and loss (viscous) components of the complex shear modulus
 462 $\mu^* = \mu' + i\mu''$ of turkey breast (N=33, 30-40 Hz) measured by direct mechanical testing (DST). The
 463 ratio of the storage moduli was $\mu'_{\parallel}/\mu'_{\perp} = 1.5 \pm 0.3$, the ratio of the loss moduli was $\mu''_{\parallel}/\mu''_{\perp} = 2.0 \pm 0.3$, and
 464 the ratio of the magnitudes was $\mu_{\parallel}/\mu_{\perp} = 1.6 \pm 0.3$ ($\phi = 0.6 \pm 0.3$).

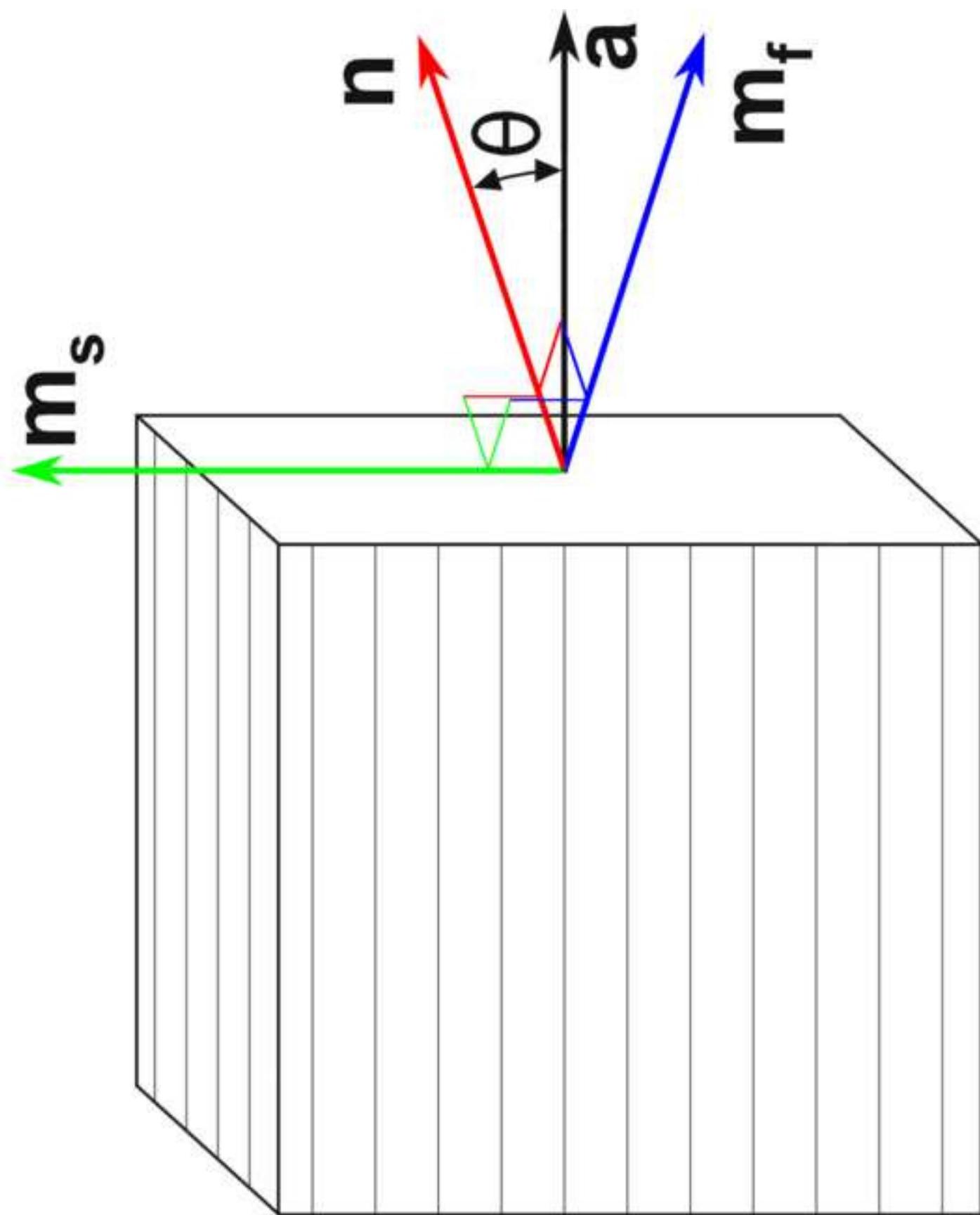
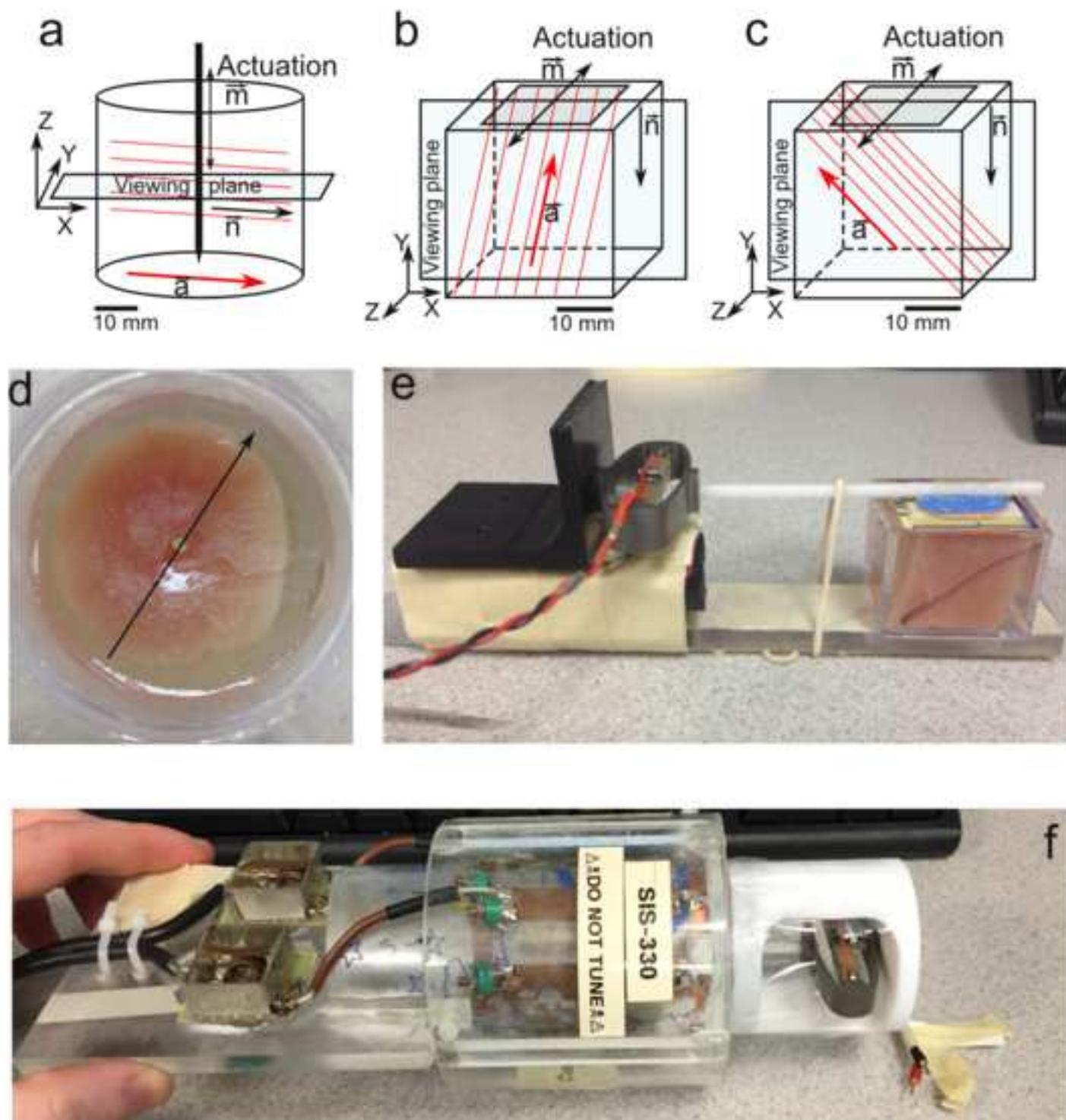
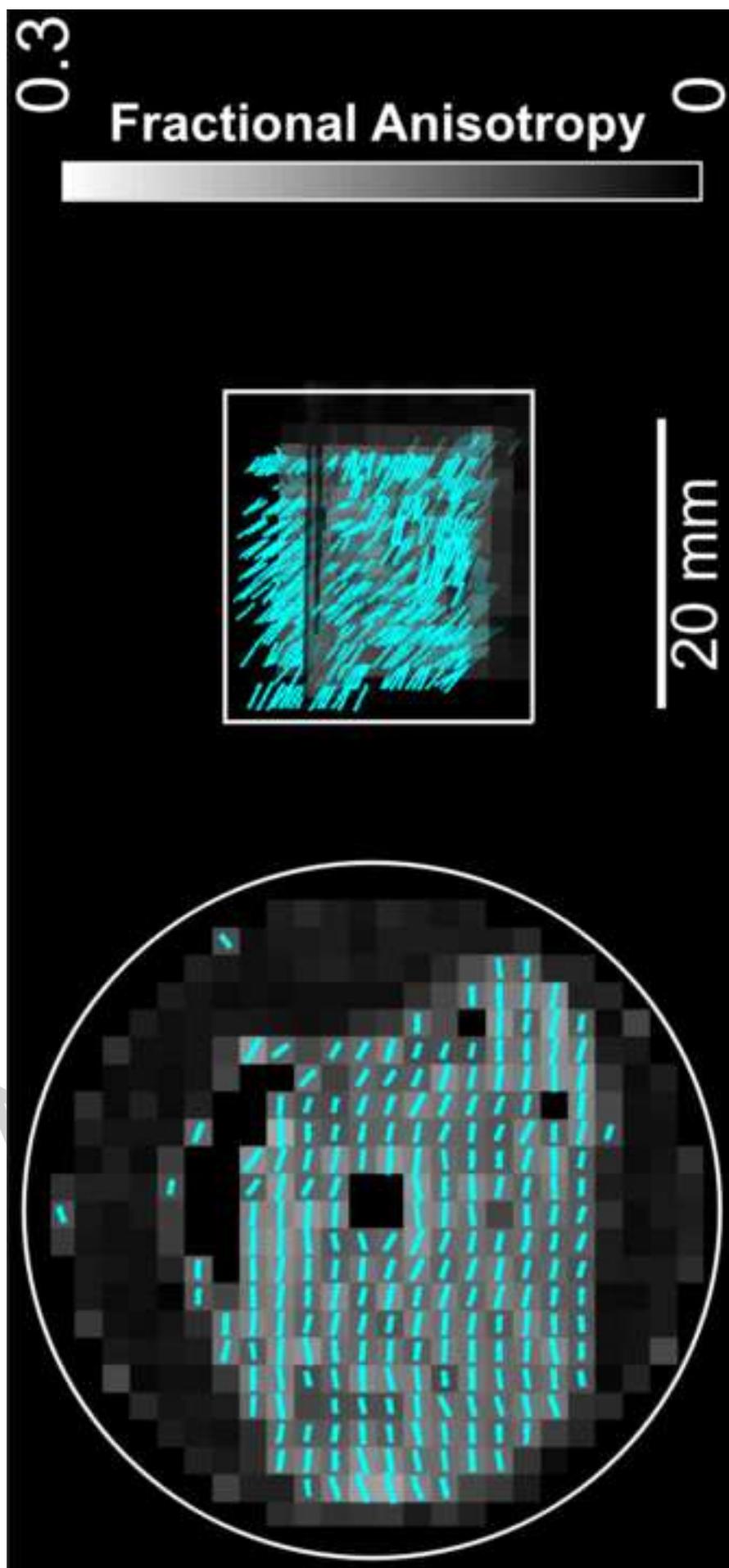


Figure 1





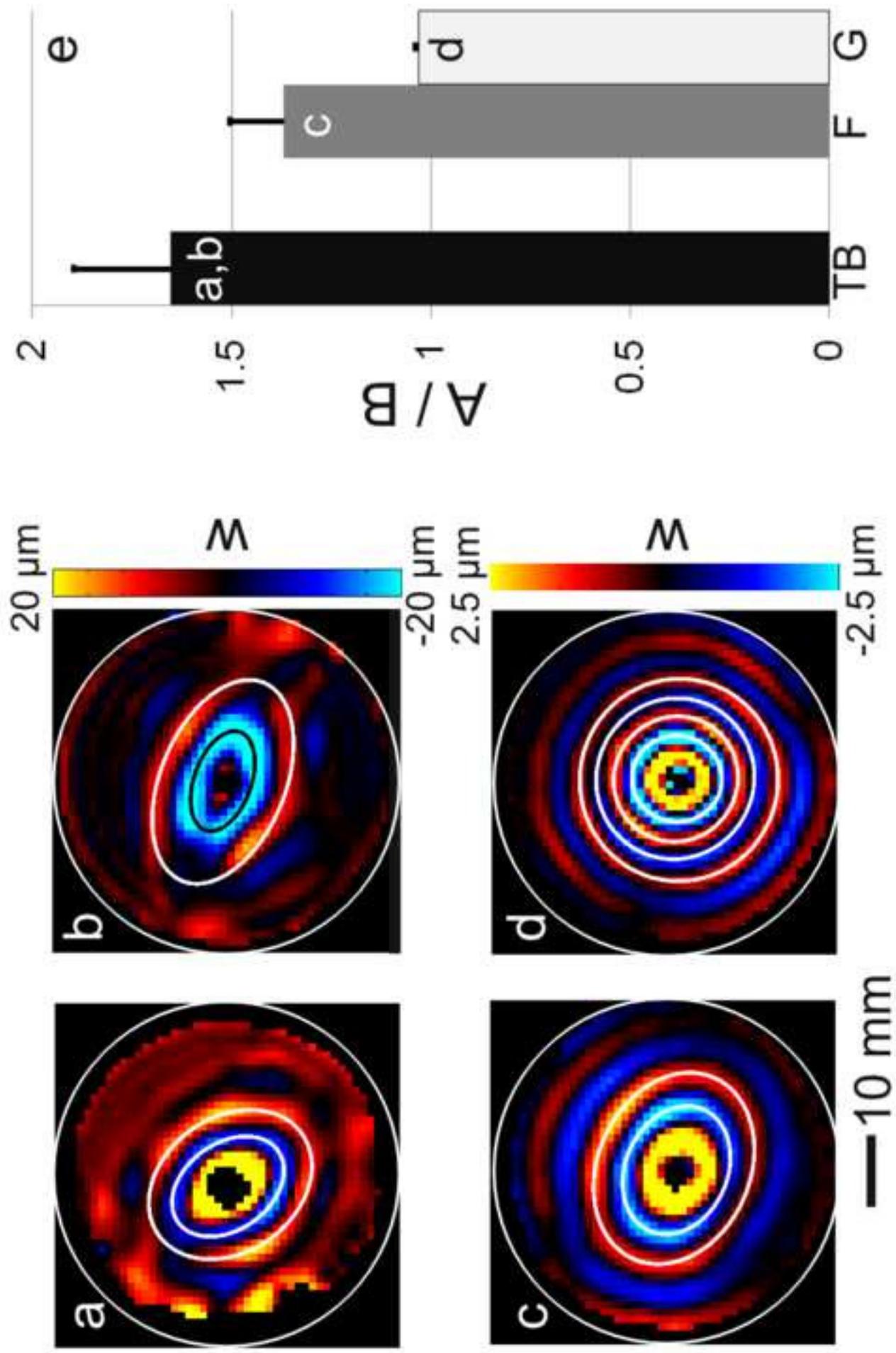


Figure 4

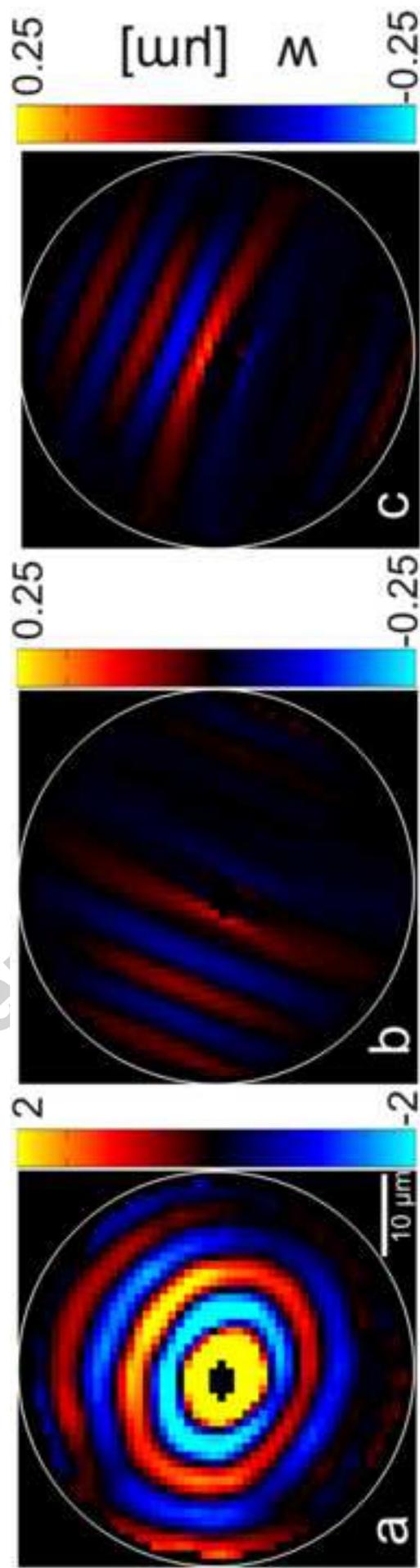


Figure 5

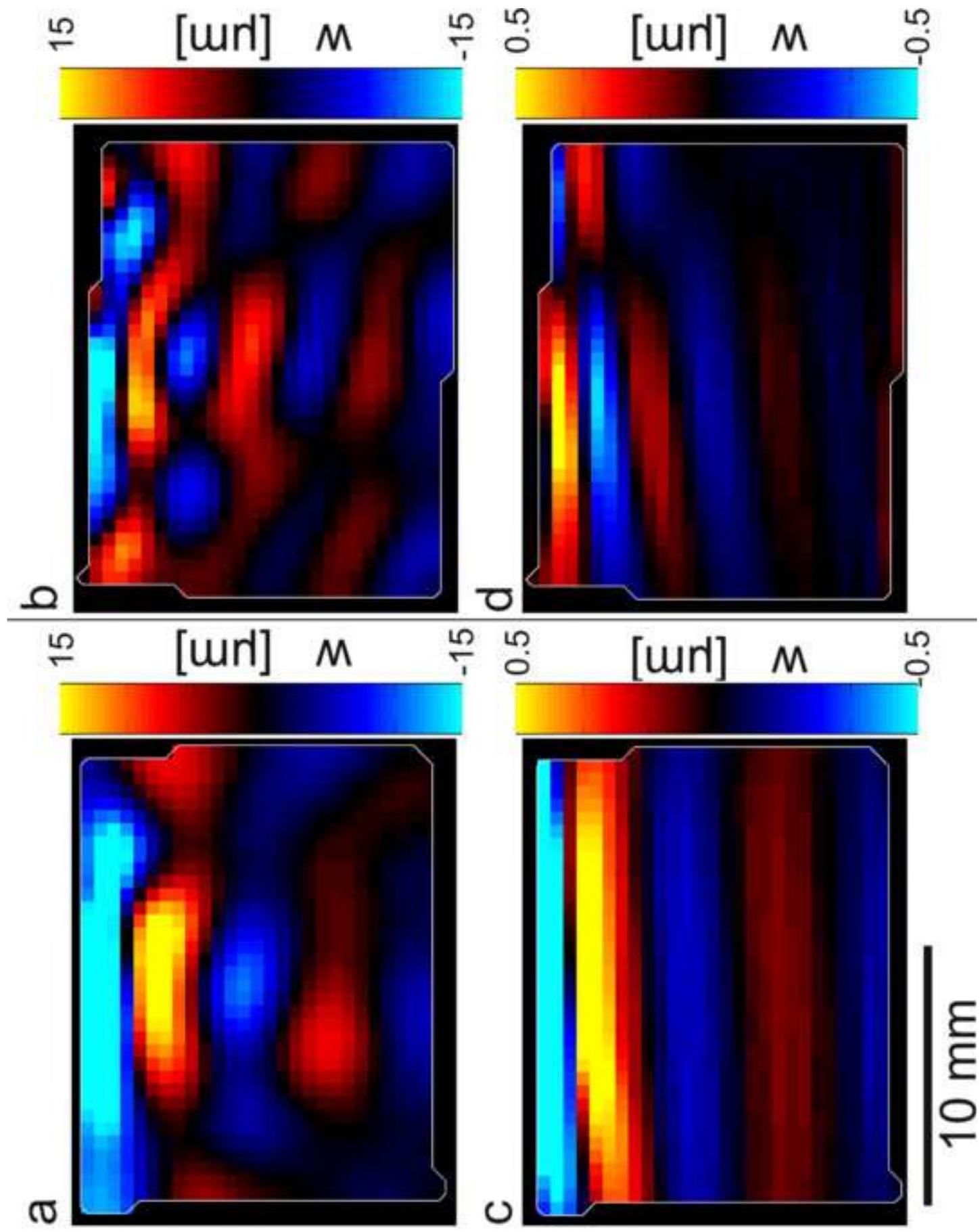


Figure 6

