

Deciphering the complexities of atopic dermatitis: Shifting paradigms in treatment approaches

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1. To appreciate the heterogeneity of atopic dermatitis (AD) phenotypes.
2. To review the role of cytokines in patients with AD.
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Atopic dermatitis (AD) is the most common chronic inflammatory skin disease. It often precedes the development of food allergy and asthma. Recent insights into AD reveal abnormalities in terminal differentiation of the epidermal epithelium leading to a defective stratum corneum, which allows enhanced allergen penetration and systemic IgE sensitization. Atopic skin is also predisposed to colonization or infection by pathogenic microbes, most notably *Staphylococcus aureus* and herpes simplex virus. Causes of this abnormal skin barrier are complex and driven by a combination of genetic, environmental, and immunologic factors. These factors likely account for the

heterogeneity of AD onset and the severity and natural history of this skin disease. Recent studies suggest prevention of AD can be achieved through early interventions to protect the skin barrier. Onset of lesional AD requires effective control of local and systemic immune activation for optimal management. Early intervention might improve long-term outcomes for AD and reduce the systemic allergen sensitization that leads to associated allergic diseases in the gastrointestinal and respiratory tract. (*J Allergy Clin Immunol* 2014;134:769-79.)

Key words: Atopic dermatitis, eczema, skin epithelium, immune, infection, filaggrin

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Atopic dermatitis (AD) is the most common chronic inflammatory skin disease.^{1,2} Recent studies reveal strong associations between mental health disorders and AD, suggesting the need to effectively manage this disease for patient's general well-being and their family's quality of life.³ AD is often associated with food allergy and asthma.^{4,5} The abnormal skin barrier in patients with AD might allow epicutaneous absorption of environmental allergens through the skin and promote systemic allergen sensitization, which predisposes to the development of food allergy and asthma. In this month's issue of the *Journal*, researchers report

Abbreviations used

AD: Atopic dermatitis

ADEH+: Atopic dermatitis with a history of eczema herpeticum

ADEH−: Atopic dermatitis without a history of eczema herpeticum

AMP: Antimicrobial peptide

DC: Dendritic cell

FLG: Filaggrin

TLR: Toll-like receptor

TMEM79: Transmembrane protein 79

that AD increases the effect of environmental peanut exposure in children with AD.⁶⁻⁸ Because there are currently no cures for food allergy and asthma, the development of effective treatments for AD might be an important strategy for prevention of the atopic march. Therefore elucidation of the underlying mechanisms of AD provides a critical opportunity for early intervention.

AD is a complex disease with a genetic predisposition strongly influenced by innate and adaptive immune responses, as well as environmental factors, including allergen exposure, irritants, microbes, diet, stress, and air quality.⁹⁻¹³ Although it is commonly referred to as a single disease,¹⁴ recent studies suggest that the time has come to distinguish various AD phenotypes and endotypes^{15,16} in much the same manner in which attempts have been made to categorize asthma and rhinosinusitis into different subtypes based on a constellation of the onset, biomarkers, immune polarization, gene variants, and natural history of the disease.¹⁷⁻¹⁹ Identification of immune pathway polarity will be of particular importance as biologic agents become more readily available to target specific immune pathways, such as the T_H2 and T_H22 pathways, as well as various inflammatory cytokines and mediators.^{20,21}

This month's issue of the *Journal* focuses particularly on the importance of both genetic and acquired causes of epithelial skin barrier dysfunction in driving the natural history of AD.²²⁻²⁴ Two original articles this month report that early emollient use to protect the skin barrier might prevent AD.^{25,26} Although dermatologists and allergists often debate the relative importance of genetic defects in the skin barrier giving rise to a leaky skin epithelial barrier that allows penetration of allergens and microbes into the skin of patients with AD (ie, the so-called "outside-in hypothesis" as opposed to the "inside-out hypothesis," a polarized immune response giving rise to a defective skin barrier), this argument is moot in patients with established AD because both processes are equally important (Fig 1). The majority of patients with AD constitute admixtures of genetic defects in the skin barrier and immune responses strongly influenced by environmental factors. This review will highlight recent insights into the crosstalk between the skin barrier and immune dysfunction leading to AD. Effective prevention and treatment of AD requires a multipronged approach involving the maintenance of skin barrier integrity, control of skin inflammation, nutrition, and identification and management of allergenic and microbial triggers.²⁷

COMPLEX CAUSES OF EPITHELIAL SKIN BARRIER DYSFUNCTION IN PATIENTS WITH AD

Multifunctional role of filaggrin

The robust association of loss-of-function mutations in the skin barrier gene encoding filaggrin (*FLG*) with risk of AD has focused attention on the important role of epithelial barrier dysfunction in

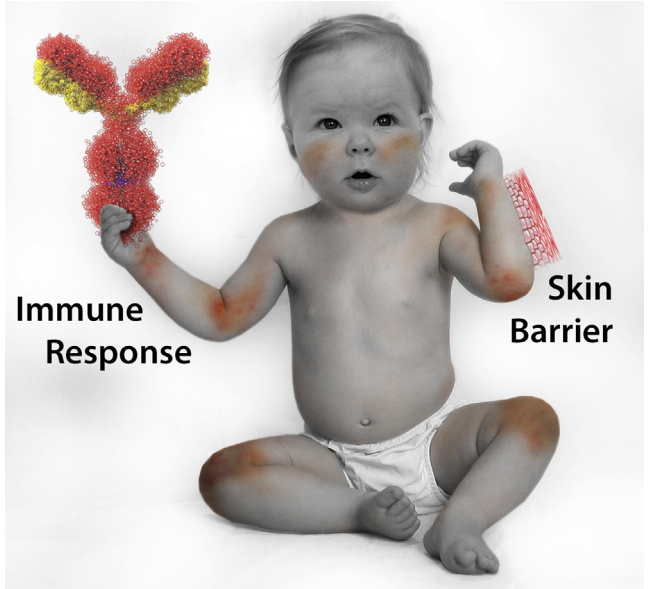


FIG 1. Is it clinically relevant whether skin barrier dysfunction or an immune response occurred first? Once AD is established, the physician needs to address both aspects of AD pathophysiology. However, prevention of AD might require identification of patients with primary defects in barrier versus immune dysfunction. Figure courtesy of Boyd Jacobson, National Jewish Health, Denver, Colorado.

TABLE I. Comparison of clinical and biophysical features of patients with AD with (AD_{FLG}) and without (AD_{NON-FLG}) filaggrin mutations*

	Clinical features	Biophysical features
AD _{FLG}	Palmar hyperlinearity	Severe decrease in NMF
	More persistent	pH
	↑ Allergic sensitization	IL-1β
	↑ Risk of asthma	Type 1 interferon-mediated stress response
	↑ Severity of AD	
AD _{NON-FLG}	↑ Eczema herpeticum	
	No palmar hyperlinearity	Mild decrease in NMF
	Less persistent	pH lower compared with patients with AD _{FLG}
	Less allergic sensitization	IL-1β low compared with patients with AD _{FLG}
	Lower risk of asthma	Dysregulation of lipid metabolic processes

NMF, Natural moisturizing factor.
*Modified with permission from McAleer and Irvine.²⁸

patients with this skin disease.^{28,29} Patients with filaggrin mutations have been found to have dry skin and early-onset AD that is more persistent and often associated with asthma, food allergy, and microbial infection.³⁰⁻³² Recent studies suggest that stratification of patients with versus without filaggrin mutations identifies patients with different mechanistic pathways of inflammation (Table I).²⁸ Patients with filaggrin loss-of-function mutations are associated with enhanced expression of IL-1 cytokines in their stratum corneum and type 1 interferon-mediated stress response.^{33,34} Children with AD with normal filaggrin genes have been reported to have dysregulation of lipid metabolic processes.³⁴ Filaggrin-dependent secretion of sphingomyelinase has also been found to protect against staphylococcal

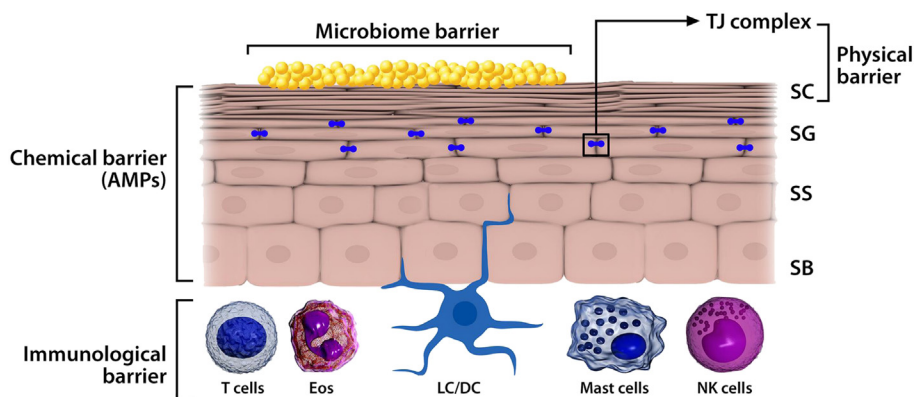


FIG 2. The skin as a multitiered barrier. The stratum corneum (SC) is the first physical barrier protecting the skin from the environment. Gene mutations (eg, filaggrin-null mutations) or cytokines (eg, IL-4, IL-13, IL-25, and IL-33) downregulating epidermal proteins, including filaggrin, leads to allergen or microbial penetration through this barrier. Tight junctions (TJs) found at the level of the stratum granulosum (SG) provide an additional barrier. Disruption of both physical barriers enables the uptake of allergens, irritants, and microbes by Langerhans cells (LCs)/DCs. Keratinocytes produce AMPs as a chemical barrier in response to pathogen colonization/infection. The skin surface is colonized by a diverse array of microorganisms (microbiome barrier) that dysregulate local immune responses and inhibit pathogenic microbes. There is also infiltration of a number of cells into the AD skin lesion, including T cells, eosinophils (Eos), DCs, natural killer (NK) cells, and mast cells/basophils. Collectively, these cells constitute the cutaneous immunologic barrier. Pattern recognition receptors regulate the function of all of these barriers (physical, chemical, microbiome, and immunologic). SB, Stratum basale; SS, stratum spinosum. This figure is modified from Kuo et al.⁵¹

α -toxin–induced keratinocyte death.³⁵ This strongly suggests that patients with filaggrin mutations have a distinct endotype of AD with different mechanistic outcomes (Table I), which could be used to identify one subset of AD, particularly for the development of new therapies targeting skin barrier function.

Clinical expression of AD is dependent on gene-environment and gene-gene interactions. Gene-environment interactions are best illustrated in filaggrin-deficient mice, in which exposure of the skin to allergens or microbes predictably leads to the development of eczema.³⁶ Three recent articles in the *Journal* demonstrate that environmental peanut might drive sensitization to peanut allergy in patients with AD, particularly those with filaggrin mutations, which is a clinically relevant example of the importance that environmental exposures in house dust might contribute to allergen sensitization in patients with AD.⁶⁻⁸

The importance of gene-gene interactions is illustrated in flaky tail mice, an animal model of AD with spontaneous eczema under pathogen-free conditions. These mice carry a double mutation involving the matted (*ma*) gene, giving them a matted hair phenotype, as well as a deletion in *Flg*. It was originally thought that the filaggrin deficiency in flaky tail mice explained the propensity of these mice to have AD. Surprisingly, the derivation of genetically engineered filaggrin-deficient mice that were free of the *ma* gene mutation were found to display impaired barrier function but to lack the propensity for spontaneous eczema.^{37,38} The matted phenotype in flaky tail mice was found to be due to a loss-of-function mutation in the transmembrane protein 79 (*Tmem79*) gene. Unexpectedly, the *Tmem79* mutation, rather than the deletion in *FLG*, was found to be associated with the development of dermatitis in mice. Interestingly, *Tmem79* encodes lamellar granules that are required for processing of filaggrin, lipids, proteases, and antimicrobial peptides (AMPs).²² Saunders et al³⁷ have also found that a single nucleotide polymorphism in the human *TMEM79* gene confers a significant risk for AD in human subjects, even when controlling for the effect of *FLG* mutations, suggesting both genes are involved in AD and the need for gene-gene interactions.

Depending on the population, *FLG* mutations are found in up to 40% of patients with severe AD, but less than 20% of these patients with severe disease are homozygous or compound heterozygous for *FLG* mutations.³⁹ Furthermore, only a minority of European American and Asian patients and none of the African American patients with AD have *FLG* mutations.^{28,29,40,41} Reductions in filaggrin expression are also pronounced in the skin of patients with AD who have no detectable *FLG*-null mutations but are most profound when combined with *FLG* mutations.⁴² Thus there are multiple causes for low *FLG* expression in the skin. The most common reason is likely to be immune activation.⁴²⁻⁴⁴ Intragenic copy number variation within the filaggrin gene also contributes to the risk of AD with a dose-dependent effect.⁴⁵ The expression of *FLG* gene expression can also be reduced by means of epigenetic modification.⁴⁶

Skin barrier dysfunction: Beyond filaggrin

Aside from *FLG*, AD has been associated with variants in other genes that encode a cluster of proteins in the epidermal differentiation complex located on chromosome 1q21.⁴⁷ These include filaggrin 2,⁴⁸ hornerin,⁴⁹ and the cornified envelope precursor SPRR3.⁵⁰ However, it is noteworthy that unlike *FLG*, the biologic function of these epidermal differentiation complex gene variants as it relates to AD is not well understood. However, a substantial amount of information indicates that loss-of-function mutations in serine protease inhibitors (eg, SPINK5) augment protease-activated pathways that enhance T_H2 responses, supporting the argument that epidermal barrier dysfunction can induce allergic skin diseases.²³ This complexity in epidermal gene variants is further modified by variants in genes that control innate and adaptive immune responses, as reviewed by Barnes⁹ and Kuo et al.⁵¹

The normal skin can be viewed as containing a series of interrelated barriers the function of which is retention of moisture and repelling penetration of the skin by allergens and microbial invasion. Once the stratum corneum is breached, such as through a

deficiency of structural proteins (eg, filaggrin, involucrin, and loricrin), lipids (eg, ceramides), or both, other barrier structures are engaged (Fig 2). These include tight junction proteins, such as the claudins, which are found on opposing membranes of stratum granulosum keratinocytes directly below the stratum corneum and thereby form a second physical barrier in the epidermis.⁵¹ Gene profiling in the epidermis of patients with AD has revealed downregulation of claudin protein and function in these patients. Once these 2 physical barriers (filaggrin and tight junctions) are breached, a rapid innate immune response must be initiated to prevent further microbial invasion. Keratinocytes and antigen-presenting cells in the skin express innate pattern recognition receptors, such as Toll-like receptors (TLRs). Stimulation of TLRs by microbes or tissue injury leads to the release of AMPs and enhanced strength of tight junctions to limit penetration of allergens and microbes.⁵¹ Patients with AD have reduced TLR function.

Loss of skin barrier function and increased severity of AD predisposes to microbial colonization and chronic skin inflammation. This is due to increased expression of tissue receptors for *Staphylococcus aureus*, which leads to colonization of *S aureus* in atopic skin.^{52,53} Keratinocytes from the skin of patients with AD have also been found to be deficient in their ability to produce the AMPs that are needed to control *S aureus* and viral replication.^{54,55} Interestingly, commensal bacteria also produce AMPs capable of controlling *S aureus* growth.⁵⁶ *S aureus* produces high levels of serine proteases that can degrade the skin barrier.⁵⁷ Therefore an overabundance of *S aureus* in patients with poorly controlled AD can reduce barrier function through multiple mechanistic pathways.

Immune-mediated barrier dysfunction

Although there are strong arguments for the “outside-in” hypothesis suggesting that AD is fundamentally a disease of fixed (genetic) epidermal barrier defects,^{22,23} there are equally compelling arguments that some forms of AD are primarily driven by polarized immune pathways that downregulate keratinocyte terminal differentiation, thereby creating a secondary skin barrier defect. The arguments against a primary role of the barrier defect in triggering keratinocyte hyperplasia and secondary immune activation include the following.

1. The *FLG* mutation is absent in most patients with AD.^{28,29,58}
2. The majority of children with AD outgrow their disease, even in the presence of an *FLG* mutation.⁵⁹
3. Unlike ichthyosis vulgaris, in which the entire skin is affected at birth, in the same genetic background patients with AD with *FLG* mutations have both lesional and non-lesional skin and the disease develops at some later time point and does not start at birth.
4. Both lesional and nonlesional AD skin exhibit a broad range of differentiation abnormalities beyond filaggrin (eg, loricrin, involucrin, corneodesmosin, and claudins), suggesting reactive epidermal differentiation/cornification alterations.^{60,61}
5. Treatment of keratinocytes with IL-4, IL-13, IL-22, IL-25, and IL-31 directly downregulates filaggrin expression and increases kallikrein function, which can directly cause barrier dysfunction.^{21,23,42-44,62,63} IL-22 directly induces keratinocyte hyperplasia and downregulation of filaggrin expression.^{64,65}

6. Mice that are genetically engineered to overexpress T_H2 cytokines in their skin spontaneously have AD and *in vivo* skin barrier defects.⁶⁶⁻⁶⁹
7. Filaggrin expression is restored by using anti-inflammatory regimens with either topical calcineurin inhibitors or topical corticosteroids.⁷⁰
8. The strongest argument is the resolution of clinical AD disease activity in patients with moderate-to-severe disease with broad-based immunosuppressive therapies, such as cyclosporine or narrow-band UV phototherapy,^{71,72} and immune-targeted therapeutics (dupilumab), which is coupled with resolution of the abnormal epidermal responses.²⁰

It is noteworthy that AD skin lesions are always associated with underlying immune activation.^{61,73,74} In patients with chronic AD, several underlying features are invariably present: (1) increased skin infiltration by T cells (approximately 10-fold increase over background T-cell levels in normal skin); (2) increased skin infiltration by myeloid (CD11c⁺) dendritic cells (DCs; also approximately 10-fold increase over normal skin levels), with most DCs having an “inflammatory phenotype” (BDCA1[−]/CD11c⁺)⁷⁵; (3) increased production of cytokines and chemokines by activated T cells and DCs within skin lesions, as measured by means of quantitative mRNA measures for individual molecules and immunohistochemical detection of associated protein products in skin lesions; and (4) reactive epidermal hyperplasia or “regenerative maturation,” showing an unusual hyperplasia response in which mRNAs and proteins of epidermal cornification are highly suppressed in the associated epidermis.^{21,76-78}

Immune pathways driving AD skin lesions

In patients with AD with increased IgE levels, nonlesional AD is associated with a selective expansion of T_H2 cells in a dermal perivascular distribution.⁷³ The acute initiation of AD skin lesions is associated with T_H2, T_H22, and also T_H17 cytokine activation (Fig 3).²¹ In parallel, there are epidermal (S100) responses marked by an extremely high increase in the expression of the proinflammatory epidermal differentiation complex cluster-encoded S100A genes (S100A7-9), which are known to be regulated by IL-22 and IL-17 cytokines.^{65,79} In patients with chronic AD, intensification of T_H2 and T_H22 activation occurs with the appearance of a significant T_H1 component but not a complete switch to T_H1.²¹ Although T_H2 cytokine effects remain during chronic AD, the increase in IFN-γ levels contributes to the inflammatory response and causes keratinocyte apoptosis.⁸⁰

In the last decade, T_H2 and T_H22 cytokines have been reported to modulate the epidermal barrier, including suppression of keratinocyte differentiation, hyperplasia, keratinocyte apoptosis, and AMP production.^{42-44,54,81-84} The cytokine effects include (1) suppression of terminal differentiation genes, such as filaggrin (*FLG*), loricrin, and involucrin by T_H2 cytokines (IL-4, IL-13, and IL-31) and T_H22/IL-22 cytokines; (2) inhibition of AMP production by T_H2 cytokines (IL-4 and IL-13); (3) upregulation of S100As by IL-17 and IL-22^{21,83-85}; and (4) induction of epidermal hyperplasia by the T_H22 IL-22 cytokine.⁷⁸

AD disease activity (quantified by using the SCORAD score) has also been shown to positively correlate with lesional and nonlesional skin expression of T_H2 and T_H22 mediators (ie, IL-13 and IL-22) and negatively with expression of terminal differentiation markers.^{71,81-84} Although immune activation is significantly higher in lesional than nonlesional skin, impressive reductions in

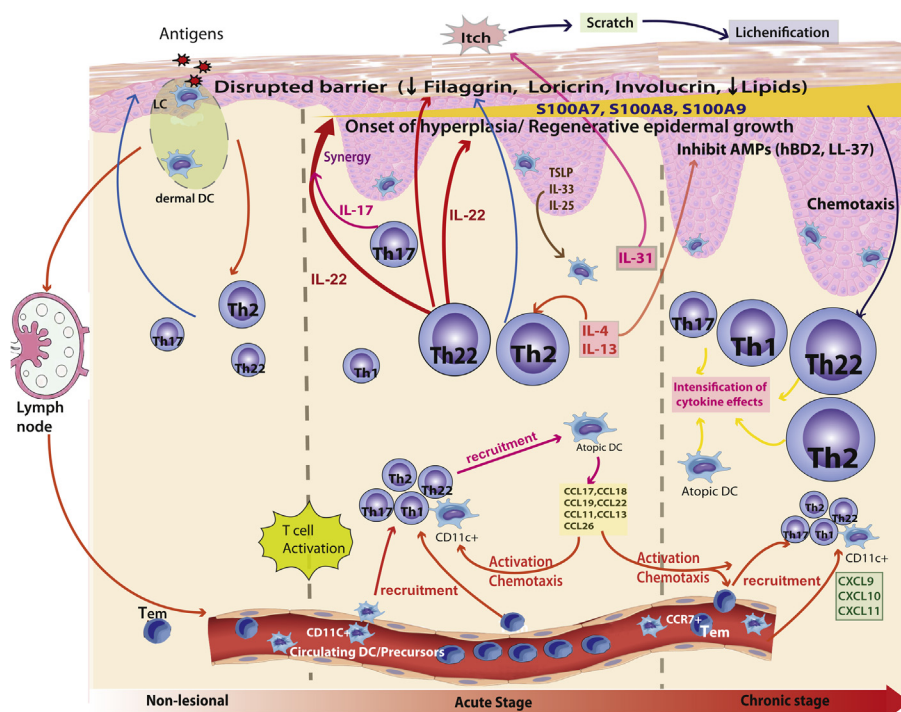


FIG 3. Immunologic pathways involved in different phases of AD. Nonlesional AD skin lesions contain immune infiltrates that produce cytokines, such as IL-4 and IL-13, which contribute to a defective epidermal barrier. Barrier defects lead to penetration by epicutaneous allergens that encounter Langerhans cells in the epidermis and dermal DCs in the dermis to activate T_H2 and T_H22 cells involved in acute disease onset. Smaller increases in T_H1 and T_H17 immune axes are also found in acute lesions. A progressive activation of T_H2 and T_H22 , as well as T_H1 , pathways is characteristic of patients with chronic AD. IL-22 induces epidermal hyperplasia and, synergistically with the T_H17 cytokine IL-17, drives an abrupt increase in a subset of terminal differentiation genes, specifically S100A7, S100A8, and S100A9 proteins. The increases in levels of these barrier proteins contrast with the uniformly disrupted epidermal differentiation gene products (eg, filaggrin, loricrin, and corneodesmosin) throughout nonlesional, acute, and chronic AD skin. The T_H2 and T_H22 cytokines contribute to inhibition of the terminal differentiation proteins. IL-31 is thought to contribute to the itch in patients with acute AD. TSLP, Thymic stromal lymphopoietin. Updated with permission from Gittler et al.²¹

expression of a broad array of epidermal differentiation genes (ie, loricrin, periplakin, and involucrin in addition to filaggrin) characterize both lesional and nonlesional AD skin. Because T_H2 (IL-4/IL-13) and T_H22 (IL-22) cytokines have been shown to inhibit expression of terminal differentiation products in keratinocytes, increased circulating levels of these cytokines might cause this global suppression of barrier proteins, as well as increases in levels of S100As detected at the onset of acute lesions.²¹

IMPLICATIONS OF AD PATHOBIOLOGY FOR GENERAL MANAGEMENT APPROACHES

Patients with established AD have a combination of skin barrier dysfunction and skin inflammation driving their skin disease. Therefore keys to the successful management of AD should include skin hydration and skin barrier repair, topical anti-inflammatory medications (topical corticosteroids or calcineurin inhibitors), control of infection, and elimination of factors (including allergens, irritants, and emotional triggers) that might exacerbate the scratch-itch cycle. Treatment should use a step-wise approach that is dependent on the severity of skin disease. The reader is referred to several recent excellent reviews on the management of AD.^{27,85-87}

In managing patients with chronic AD, it is important to recognize that gene profiling and immunohistologic studies reveal

subclinical inflammation and downregulation of terminal epithelial differentiation with reduced skin barrier protein levels and increased transepidermal water loss, even in nonlesional AD skin.^{47,61,74,87,88} Thus it is important to maintain skin barrier therapy in the form of emollient therapy, even during periods of remission. In patients with AD who are prone to frequent relapses, the subclinical inflammation can be managed with intermittent (2 times per week) corticosteroids or alternate-day topical calcineurin inhibitors as maintenance therapy.⁸⁹ For acute AD exacerbations, medium- and high-potency corticosteroids can be used for short periods of time to control the disease. Oral or systemic corticosteroids should be avoided because of rebound flares when patients are being weaned from oral corticosteroids.

In patients with AD who are refractory or do not clinically respond to conventional treatment approaches, a number of alternative strategies can be used, including cyclosporine, methotrexate, azathioprine, IL-6 blockade, dust mite immunotherapy (when indicated), wet wrap therapy, and UV light.⁹⁰⁻⁹⁴ Recent studies with broad-based targeting therapeutics^{71,72} have used disease-related cellular and molecular biomarkers to (1) show that the chronic AD phenotype can be reversed to a nonlesional state, as has been shown for patients with psoriasis treated with effective therapeutics, and (2) map inflammatory disease-related pathways. The narrow-band UVB phototherapy and cyclosporine trials showed elimination of the pathologic epidermal hyperplasia

(suprabasal K16 expression immunohistochemically and increased K16 and Ki67 mRNA expression) after 12 weeks of treatment. The improvement in disease activity as identified by using the SCORAD score and epidermal pathology were highly linked to clearance of excess T-cell and DC infiltrates, as well as decreased expression of inflammatory markers.^{71,72,95,96}

PREVENTION OF AD BY EARLY INTERVENTION

Because current treatment approaches are not curative, there is considerable interest in studying approaches to prevent AD.⁹⁷ The use of probiotic therapy or bacterial lysates early in the course of illness to prevent AD remains an area of active investigation,⁹⁸⁻¹⁰⁰ but results have been inconsistent. This might be due to lack of standardization of the bacterial preparations or lack of biomarkers to identify which AD phenotype would benefit from this approach.

The potential contribution of vitamin D deficiency to allergic inflammation, corticosteroid insensitivity, and downregulation of innate immune responses has also been an active area of study.¹⁰¹⁻¹⁰⁶ Reproducible well-controlled studies of oral vitamin D supplementation are lacking, and when they have been done, they have yielded confusing results. The greatest benefits are likely in populations who have extremely low vitamin D levels, such as persons living in upper latitudes during the winter or darkly pigmented persons. In the current issue of the *Journal*, Camargo et al¹³ present results from a randomized, placebo-controlled trial demonstrating that winter-related AD can be improved with vitamin D oral supplementation.

The importance of AD skin barrier dysfunction in driving allergen sensitization is highlighted by 3 articles in the current issue suggesting that severe AD drives sensitization with environmental peanut.⁶⁻⁸ In 2 of these articles, filaggrin mutations predicted increased association of AD with peanut allergy. These articles suggest the possibility that controlling environmental peanut levels in the household might reduce peanut allergen sensitization. Alternatively, studies are needed to determine whether effective control of AD with barrier therapy and anti-inflammatory treatment to reduce AD skin severity will reduce absorption of environmental allergens and decrease onset of food allergies or respiratory allergy.

Considering the important role that skin barrier dysfunction plays in the initiation of AD, the current issue of the *Journal* contains 2 different international investigations assessing early intervention with skin emollient therapy to prevent AD and allergic sensitization during infancy.^{25,26} Simpson et al²⁵ performed a randomized controlled trial of 124 neonates at high risk for AD. Parents in the intervention arm were instructed to apply full-body emollient therapy at least once per day starting within 3 weeks of birth. Parents in the control arm were asked to use no emollients. The primary outcome was the cumulative incidence of AD at 6 months. Their results demonstrated a statistically significant protective effect with the use of daily emollient on the cumulative incidence of AD, with a relative risk reduction of 50%.

Horimukai et al²⁶ performed a randomized controlled trial with early moisturizer intervention conducted in 116 neonatal participants at high familial risk for AD. The primary outcome was the cumulative incidence of AD as of week 32, as evaluated by a dermatologist. The intervention with the moisturizer significantly decreased (by approximately 40%) the risk of AD compared with that seen in the control subjects ($P = .002$) as of week 32. The 2 groups showed similar rates of allergic sensitization. However,

TABLE II. Clinical phenotypes of AD

<ul style="list-style-type: none"> • Onset in infancy, outgrown in childhood • Onset in infancy, persistent severe eczema • Adolescent-adult onset, mild-to-moderate eczema • Adolescent-adult onset, persistent severe eczema • Increased IgE levels with food or aeroallergen sensitization (extrinsic) • Non-IgE mediated (intrinsic) • AD with <i>S aureus</i> infection/colonization • AD with history of disseminated viral infections (eg, eczema herpeticum)
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the rate of allergic sensitization of infants with AD was significantly higher than that seen in the rest of the infants.

These 2 studies suggest that early intervention with emollient therapy from birth represents a safe and effective approach for AD prevention. If confirmed to be effective in future studies, emollient therapy from birth would be a simple and low-cost intervention that could reduce the global burden of allergic diseases. Whether this form of intervention can prevent the atopic march is unresolved and might require combination with intermittent anti-inflammatory therapy and environmental control.

CLINICAL PHENOTYPES OF AD

AD is primarily defined by clinical criteria.¹⁰⁷ However, there is increasing recognition that AD is a complex syndrome with multiple causes and mechanistic pathways that clinically can be distinguished by age of onset, severity of illness, racial modifiers, response to therapy, and triggers (including infections, allergens, stress, and irritant threshold). Table II lists some of the major clinical phenotypes of AD.^{15,16} These phenotypes often have overlapping features but contain dominant characteristics that distinguish them from each other (Fig 4).¹⁰⁸ The majority of infants who present with mild AD will outgrow their skin disease in later childhood. However, a group of difficult-to-manage patients exist who have early-onset eczema with severe lifelong AD. Adult-onset AD has also been increasingly reported, although it is unclear whether these might be patients who had eczema during infancy and then went into a prolonged remission only to have relapse of eczema later in life because recall history can be unreliable. Up to 50% but certainly not all patients with AD have associated asthma, allergic rhinitis, or food allergy. Identification of genetic markers and biomarkers of patients likely to undergo the atopic march would allow early intervention for prevention of mucosal allergy, including food allergy and asthma.

Approximately 80% of patients with AD have increased serum IgE levels and/or immediate skin test reactivity to allergens, but 20% of patients with AD have no IgE to food or inhalant allergens. However, it is possible that such intrinsic or nonatopic patients might have IgE or autoreactive T cells to autoallergens or microbial antigens, which are not routinely measured.¹⁰⁹⁻¹¹¹ Other AD subsets exist, including those who are prone to skin infection, such as *S aureus* skin infections or eczema herpeticum.¹¹²⁻¹¹⁶ Although up to 90% of patients with AD might have problems with *S aureus* skin colonization, actual overt skin infections requiring systemic antibiotic treatment affect less than 50%. Less than 3% of patients with AD are predisposed to eczema herpeticum. These different phenotypes likely arise from a complex combination of mutations and epigenetic effects on genes controlling protein expression in the skin barrier and the innate and adaptive immune response controlled by environmental influences.^{9,117}

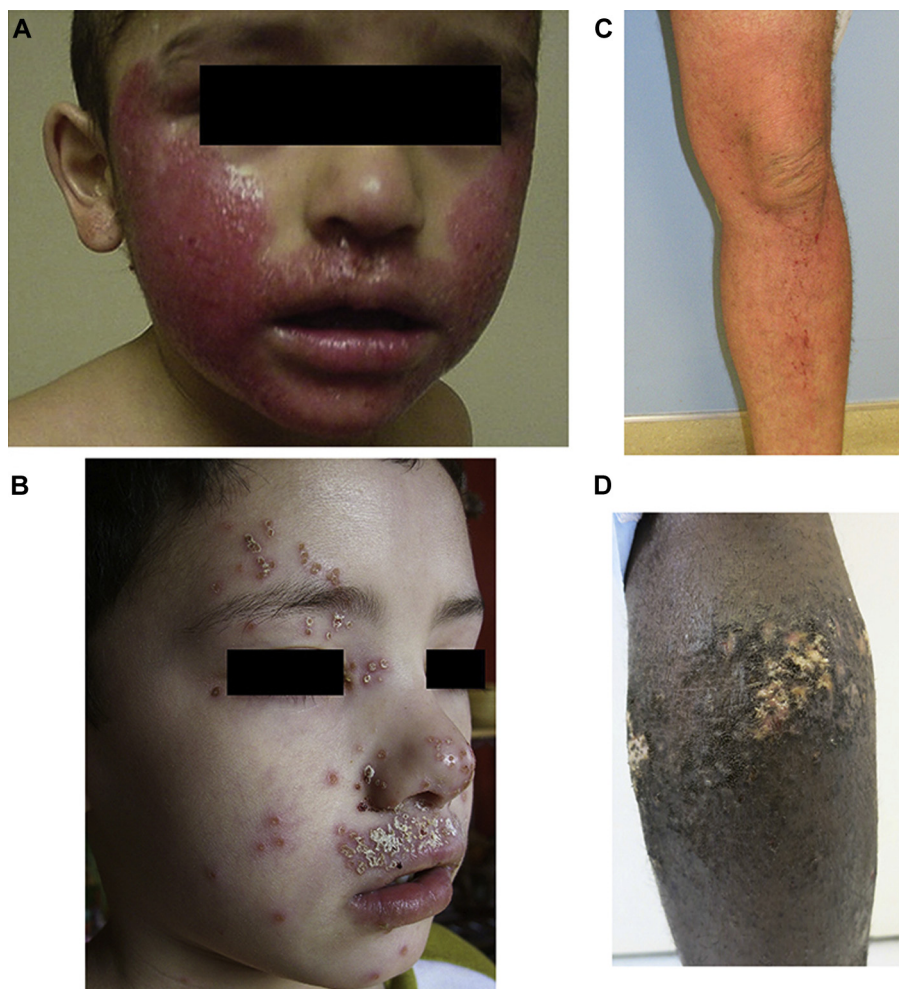


FIG 4. Clinical phenotypes in patients with AD: eczema herpeticum (**A**), *S aureus*-colonized AD (**B**), mild AD (**C**), and severe AD (**D**). Fig 4, **A** and **B**, are from Boguniewicz and Leung.¹⁰⁸ Fig 4, **C** and **D**, were contributed by Dr Emma Guttman-Yassky at the Icahn School of Medicine at Mt Sinai, New York.

TABLE III. Summary of cytokine effects on the epidermis in patients with AD

- Induce epidermal hyperplasia (IL-22)
- Induce spongiosis (T_H2 cytokines IL-4/IL-13 and TNF)
- Inhibit keratinocyte terminal differentiation (IL-4, IL-13, IL-31, IL-25/T_H2, IL-22/T_H22, and TNF) with potential for feedback hyperplasia
- Inhibit synthesis of AMPs (T_H2 cytokines IL-4, IL-13, and IL-33)
- Inhibit lipid synthesis (T_H2 cytokines IL-4/IL-13, IL-31, and TNF)
- Increase expression of S100A7, S100A8, and S100A9 (IL-22 plus IL-17)
- Induce TSLP production in KCs (IL-4/IL-13 and TNF)
- Promote itch (IL-31 and TSLP)
- Promote antiviral responses (IFN- γ , IFN- α , and IL-29)

KCs, Keratinocytes; TSLP, thymic stromal lymphopoietin.

DEFINING ENDOTYPES IN PATIENTS WITH AD

The importance of eventually defining endotypes in patients with AD is that these new subtypes can be used in clinical study design and drug development to target therapies to patients most likely to benefit from a mechanism-based treatment. In the future, AD might be stratified by genotype and biomarkers reflecting immune polarization to complement their clinical phenotype. As

noted in Table I, filaggrin genotyping defines AD subsets with different mechanistic pathways. Importantly, the severity of AD is related to filaggrin expression with a dose-dependent effect.⁴⁵

Patients with AD with homozygous filaggrin-null mutations or compound heterozygotes compared with patients with normal filaggrin gene expression have early onset of skin disease and more persistent severe eczema. They often have palmar hyperlinearity, greater risk of allergen sensitization, and a history of food allergy and asthma. In addition, they have increased pH values in their stratum corneum, which might predispose them to *S aureus* colonization. Patients with heterozygous *FLG* mutations have an intermediate phenotype and can outgrow their AD in adolescence, whereas homozygotes or compound heterozygotes can have lifelong disease.⁵⁹ These patients might also serve as a target for filaggrin therapy.¹¹⁸

Given the complex genetic milieu of AD, the development of biomarkers is important to assess the final immune-polarized pathways that might exist in various AD subsets. The best biomarkers for AD currently define patients who are T_H2 polarized versus those who are not. In the future, a combination of epidermal proteomics, genomics, gene transcriptomes, and blood biomarkers in combination with the clinical phenotype will offer more precision in defining endotypes of AD.¹¹⁹⁻¹²¹

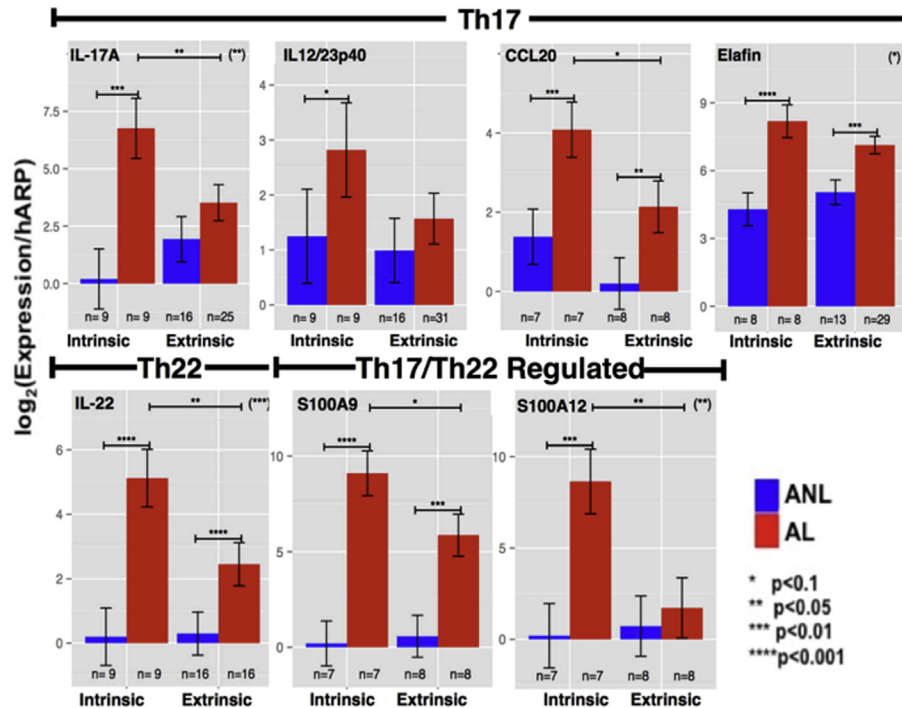


FIG 5. Measures of mRNA levels (normalized to hARP mRNA) for specific T_H17 /IL-23 and T_H22 cytokines and inflammatory products in nonlesional (ANL) and lesional (AL) AD skin from patients with intrinsic versus extrinsic disease. From Suarez-Farinas et al.¹²⁴

Approximately 80% of patients with AD have increased serum IgE levels, often with increased eosinophilia and serum levels of the T_H2 chemokine thymus and activation-regulated chemokine. Additional markers are needed to better monitor patients with so-called intrinsic AD. However, it is noteworthy that studies of patients with so-called intrinsic AD who lacked IgE to conventional inhalant and food allergens had detectable serum IgE to autoantigens in the skin and microbial antigens from bacterial and fungi that colonize the skin.^{110,111} Therefore a wider range of IgE screens to various exogenous and endogenous antigens is warranted to determine potential triggers of AD because it might have an important effect on pathways triggering allergic skin inflammation.

Elucidation of the mechanisms underlying *S aureus* and disseminated viral infections in patients with AD is an active area of investigation. These patients are generally very atopic, with increased serum IgE levels and eosinophilia. This might reflect high-level T_H2 cytokine pathway activation, which is known to reduce skin barrier function, enhance *S aureus* skin colonization, reduce AMP production, and impair innate immune responses. However, because eczema herpeticum is extremely rare and herpes simplex virus exposure is very common, it is likely that additional immunologic and genetic factors contribute to atopic dermatitis with a history of eczema herpeticum (ADEH+). To identify novel gene signatures, in the current issue of the *Journal*, Bin et al¹²² used an RNA-sequencing approach to evaluate global transcriptional changes in PBMCs from patients with ADEH+ and those with atopic dermatitis without a history of eczema herpeticum. They found that PBMCs from patients with ADEH+ stimulated with herpes simplex virus 1 were deficient in their antiviral immune response involving interferon regulatory factor 3 and 7 innate immune pathways. This likely

contributes to the reduced interferon response in ADEH+ that predisposes to increased susceptibility to disseminated viral infection. Interestingly, dedicator of cytokinesis 8 deficiency, which presents with eczema and recurrent herpetic skin infections, has recently been found to respond well to treatment with IFN- α 2b.¹²³

Disease severity appears to be related to the magnitude and polarity of immune activation, as well as effects of immune cytokines on epidermal responses (Table III).^{21,61,95,96,124} Severe AD might thus be associated with systemic immune activation, including significant pathology in nonlesional skin,⁶¹ explaining the frequent need for systemic immune suppressants and the inadequacy of treating only lesional skin with topical agents. In contrast, because of minimal systemic involvement, it might be most appropriate to treat mild disease with topical agents directed only to lesional sites.

Immune studies suggest that different AD phenotypes are associated with distinct patterns of activation (or suppression) of polar immune axes and corresponding tissue responses. There are distinct differences in cytokine production in patients with intrinsic versus extrinsic AD.¹²⁴ In patients with intrinsic AD, who have normal IgE levels, there is significantly increased expression of IL-17, IL-23, IL-22, and their respective keratinocyte-induced products (ie, S100As, elafin/PI3, and CCL20), suggesting a subset with potential for greater responsiveness to suppression of the IL-17/IL-23/IL-22 axes (Fig 5).¹²⁴ Despite high IgE levels in patients with extrinsic AD, similar expression levels of T_H2 -related products were observed in both patients with extrinsic and those with intrinsic AD, suggesting that these phenotypes might have similar responses to IL-4 receptor antagonism. Indeed, Beck et al²⁰ showed that responses to dupilumab treatment were similar in patients with intrinsic and those with extrinsic AD.

TABLE IV. Phenotypes of severity and treatment response

- Preclinical: Use skin barrier cream for prevention of AD
- Chronic AD in remission: Use barrier cream in combination with maintenance topical corticosteroid or calcineurin inhibitors to prevent relapse
- Relapse of mild-to-moderate AD: Use topical corticosteroids or calcineurin inhibitors for control of inflammation, identify and avoid triggers (irritants, allergens, and infection), immunotherapy for allergen-driven AD
- Persistent moderate-to-severe AD not controlled with topical corticosteroids or calcineurin inhibitors: Wet wrap therapy, allergen immunotherapy, and nonspecific immunosuppressives (cyclosporine, methotrexate, narrow-band UV phototherapy, and mycophenolate)
- Future targeted therapies for moderate-to-severe AD (anti-IL-4 receptor α , anti-IL-22, anti-IL-23/IL-17, or other biologic agents that interrupt polarized immune pathways)

Strong support for the changing pathogenic and therapeutic AD paradigm comes from a recent report on the use of IL-4 receptor α antagonist/dupilumab conducted in patients with moderate-to-severe AD.²⁰ This study showed major improvement (approximately 70%) in disease activity at the higher dose (compared with only approximately 20% in the placebo group) coupled with large improvement in the AD genomic phenotype and reversal of the epidermal hyperplasia (as quantified by larger reductions in K16 expression).²⁰ In fact, the reduction in hyperplasia was much higher in the 4-week trial than with 5 mg/kg cyclosporine given for 12 weeks to patients with similar disease activity.⁹⁵

CONCLUDING COMMENTS: THE TRANSLATIONAL REVOLUTION IN AD

AD presents a large unmet need for more effective topical and systemic therapeutics. In addition to T_H2 antagonists (ie, anti-IL-4 receptor/dupilumab), the key role of thymic stromal lymphopoietin receptor signaling^{125,126} and IL-22,¹²⁷ as studied in clinical trials with agents targeting thymic stromal lymphopoietin, T_H22, and T_H17/IL-23, will be of interest. Selection of immune-targeted therapeutics for patients with different degrees of disease severity or recognized AD phenotypes should not be done by serendipity but should be guided by defining the extent of activation of polar immune circuits in skin and blood (Table IV). For example, anti-IL-23/IL-17 might provide beneficial responses in patients with AD, particularly patients with intrinsic AD. The individual contributions of the T_H22, T_H17, and T_H2 immune pathways to the disease phenotype will be clarified through clinical trials coupled with mechanistic studies that are currently in progress.

REFERENCES

- Shaw TE, Currie GP, Koudelka CW, Simpson EL. Eczema prevalence in the United States: data from the 2003 National Survey of Children's Health. *J Invest Dermatol* 2011;131:67-73.
- Schmitt J, Langan S, Deckert S, Svensson A, von Kobyletzki L, Thomas K, et al. Assessment of clinical signs of atopic dermatitis: a systematic review and recommendation. *J Allergy Clin Immunol* 2013;132:1337-47.
- Yaghmaie P, Koudelka CW, Simpson EL. Mental health comorbidity in patients with atopic dermatitis. *J Allergy Clin Immunol* 2013;131:428-33.
- Boguniewicz M, Leung DYM. Atopic dermatitis: a disease of altered skin barrier and immune dysregulation. *Immunol Rev* 2011;242:233-46.
- McLean WH, Palmer CN, Henderson J, Kabesch M, Weidinger S, Irvine AD. Filaggrin variants confer susceptibility to asthma. *J Allergy Clin Immunol* 2008;121:1294-5.
- Venkataraman D, Soto-Ramirez N, Kurukulaaratchy RJ, Holloway JW, Karmaus W, Ewart SL, et al. Filaggrin loss-of-function mutations are associated with food allergy in childhood and adolescence. *J Allergy Clin Immunol* 2014;134:876-82.
- Brough HA, Simpson A, Makinson K, Hankinson J, Brown S, Douiri A, et al. Peanut allergy: effect of environmental peanut exposure in children with filaggrin loss-of-function mutations. *J Allergy Clin Immunol* 2014;134:867-75.
- Brough HA, Liu AH, Sicherer S, Makinson K, Douiri A, Brown S, et al. Atopic dermatitis increases the impact of exposure to peanut antigen in dust on peanut sensitization and likely peanut allergy. *J Allergy Clin Immunol* 2014; In press.
- Barnes KC. An update on the genetics of atopic dermatitis: scratching the surface in 2009. *J Allergy Clin Immunol* 2010;125:16-29.
- Roduit C, Frei R, Loss G, Büchele G, Weber J, Depner M, et al. Development of atopic dermatitis according to age of onset and association with early-life exposures. *J Allergy Clin Immunol* 2012;130:130-6.
- Illi S, Depner M, Genuneit J, Horak E, Loss G, Strunz-Lehner C, et al. Protection from childhood asthma and allergy in Alpine farm environments-the GABRIEL Advanced Studies. *J Allergy Clin Immunol* 2012;129:1470-7.
- Kim J, Kim EH, Ohm I, Jung K, Han Y, Cheong HK, et al. Symptoms of atopic dermatitis are influenced by outdoor air pollution. *J Allergy Clin Immunol* 2013;132:495-8.
- Camargo CA Jr, Ganma D, Sidbury R, Erdenedelger KH, Radnaakhand N, Khandsuren B. Randomized trial of vitamin D supplementation for winter-related atopic dermatitis in children. *J Allergy Clin Immunol* 2014;134:831-5.
- Schmitt J, Spuls PI, Thomas KS, Simpson E, Furue M, Deckert S, et al. The Harmonising Outcome Measures for Eczema (HOME) statement to assess clinical signs of atopic eczema in trials. *J Allergy Clin Immunol* 2014;134:800-7.
- Garmhausen D, Hagemann T, Bieber T, Dimitriou I, Fimmers R, Diepgen T, et al. Characterization of different courses of atopic dermatitis in adolescent and adult patients. *Allergy* 2013;68:498-506.
- Bieber T. Atopic dermatitis 2.0: from the clinical phenotype to the molecular taxonomy and stratified medicine. *Allergy* 2012;67:1475-82.
- Ingram JL, Kraft M. IL-13 in asthma and allergic disease: asthma phenotypes and targeted therapies. *J Allergy Clin Immunol* 2012;130:829-42.
- Carolan BJ, Sutherland ER. Clinical phenotypes of chronic obstructive pulmonary disease and asthma: recent advances. *J Allergy Clin Immunol* 2013;131:627-34.
- Akdis CA, Bachert C, Cingi C, Dykewicz MS, Hellings PW, Naclerio RM, et al. Endotypes and phenotypes of chronic rhinosinusitis: a PRACTALL document of the European Academy of Allergy and Clinical Immunology and the American Academy of Allergy, Asthma & Immunology. *J Allergy Clin Immunol* 2013;131:1479-90.
- Beck LA, Thaci D, Hamilton JD, Graham NM, Bieber T, Rocklin R, et al. Dupilumab treatment in adults with moderate-to-severe atopic dermatitis. *N Engl J Med* 2014;371:130-9.
- Gittler J, Shemer A, Suárez-Fariñas M, Fuentes-Duculan J, Gulewicz KJ, Wang CQF, et al. Progressive activation of T(H)2/T(H)22 cytokines and selective epidermal proteins characterizes acute and chronic atopic dermatitis. *J Allergy Clin Immunol* 2012;130:1344-54.
- Elias P, Wakefield JS. Mechanisms of abnormal lamellar body secretion and the dysfunctional skin barrier in patients with atopic dermatitis. *J Allergy Clin Immunol* 2014;134:781-91.
- Samuelov L, Sprecher E. Peeling off the genetics of atopic dermatitis-like congenital disorders. *J Allergy Clin Immunol* 2014;134:808-15.
- Thyssen JP, Kezic S. Causes of epidermal filaggrin reduction and their role in the pathogenesis of atopic dermatitis. *J Allergy Clin Immunol* 2014;134:792-9.
- Simpson EL, Chalmers JR, Hanifin JM, Thomas KS, Cork MJ, McLean WHI, et al. Emollient enhancement of the skin barrier from birth offers effective atopic dermatitis prevention. *J Allergy Clin Immunol* 2014;134:818-23.
- Horimukai K, Morita K, Narita M, Kondo M, Kitazawa H, Nozaki M, et al. Application of moisturizer to neonates prevents development of atopic dermatitis. *J Allergy Clin Immunol* 2014;134:824-30.
- Schneider L, Lio P, Boguniewicz M, Beck L, LeBovidge J, Novak N, et al. Atopic dermatitis: a practice parameter update 2012. *J Allergy Clin Immunol* 2013;131:295-9.
- McAleer MA, Irvine AD. The multifunctional role of filaggrin in allergic skin disease. *J Allergy Clin Immunol* 2013;131:280-91.
- Irvine AD, McLean WHI, Leung DYM. Filaggrin mutations associated with skin and allergic diseases. *N Engl J Med* 2011;365:1315-27.
- Margolis DV, Apter AJ, Gupta J, Hoffstad O, Papadopoulos M, Campbell LE, et al. The persistence of atopic dermatitis and filaggrin (FLG) mutations in a US longitudinal cohort. *J Allergy Clin Immunol* 2012;130:912-7.
- Böhme M, Söderhöl C, Kull I, Bergström A, van Hage M, Wahlgren C. Filaggrin mutations increase the risk for persistent dry skin and eczema independent of sensitization. *J Allergy Clin Immunol* 2012;129:1153-5.

32. Kawasaki H, Nagao K, Kubo A, Hata T, Shimizu A, Mizuno H, et al. Altered stratum corneum barrier and enhanced percutaneous immune responses in filaggrin-null mice. *J Allergy Clin Immunol* 2012;129:1538-46.
33. Kezic S, O'Regan GM, Lutter R, Jakasa I, Koster ES, Saunders S, et al. Filaggrin loss-of-function mutations are associated with enhanced expression of IL-1 cytokines in the stratum corneum of patients with atopic dermatitis and in a murine model of filaggrin deficiency. *J Allergy Clin Immunol* 2012;129:1031-9.
34. Cole C, Kroboth K, Schurch NJ, Sandilands A, Sherstnev A, O'Regan GM, et al. Filaggrin-stratified transcriptome analysis of paediatric skin identifies mechanistic pathways in atopic dermatitis. *J Allergy Clin Immunol* 2014;134:82-91.
35. Brauweiler AM, Bin L, Kim BE, Oyoshi MK, Geha RS, Goleva E, et al. Filaggrin dependent secretion of sphingomyelinase protects against Staphylococcal alpha-toxin-induced keratinocyte death. *J Allergy Clin Immunol* 2013;131:421-7.
36. Oyoshi MK, Murphy GF, Geha RS. Filaggrin-deficient mice exhibit Th17-dominated skin inflammation and permissiveness to epicutaneous sensitization with protein antigen. *J Allergy Clin Immunol* 2009;124:485-93.
37. Saunders SP, Goh CS, Brown SJ, Palmer CN, Porter RM, Cole C, et al. Tmem79/Matt is the matted mouse gene and is a predisposing gene for atopic dermatitis in human subjects. *J Allergy Clin Immunol* 2013;132:1121-9.
38. Sasaki T, Shiohama A, Kubo A, Kawasaki H, Ishida-Yamamoto A, Yamada T, et al. A homozygous nonsense mutation in the gene for Tmem79, a component for lamellar granule secretory system, produces spontaneous eczema in an experimental model of atopic dermatitis. *J Allergy Clin Immunol* 2013;132:1111-20.
39. Mohiuddin M, Ramamoorthy P, Reynolds PR, Curran-Everett D, Leung DYM. Increased compound heterozygous filaggrin mutations in severe atopic dermatitis in the United States. *J Allergy Clin Immunol Pract* 2013;1:534-6.
40. Thawer-Esmail F, Jakasa I, Todd G, Wen YR, Brown SJ, Kroboth K, et al. South African amaXhosa patients with atopic dermatitis have decreased levels of filaggrin breakdown products but no loss-of-function mutations in filaggrin. *J Allergy Clin Immunol* 2014;133:280-2.
41. Garrett JPD, Hoffstad O, Apter AJ, Margolis DJ. Racial comparison of filaggrin null mutations in asthmatic patients with atopic dermatitis in a US population. *J Allergy Clin Immunol* 2013;132:1232-4.
42. Howell MD, Kim BE, Gao P, Grant AV, Boguniewicz M, DeBenedetto A, et al. Cytokine modulation of atopic dermatitis filaggrin skin expression. *J Allergy Clin Immunol* 2007;120:150-5.
43. Kim BE, Bin L, Ye YM, Ramamoorthy P, Leung DYM. IL-25 Enhances HSV-1 replication by inhibiting filaggrin expression, and acts synergistically with TH2 cytokines to enhance HSV-1 replication. *J Invest Dermatol* 2013;133:2678-85.
44. Cornelissen C, Marquardt Y, Czaja K, Wenzel J, Lüscher-Firzlaff J, Lüscher B, et al. IL-31 regulates differentiation and filaggrin expression in human organotypic skin models. *J Allergy Clin Immunol* 2012;129:426-33.
45. Brown SJ, Kroboth K, Sandilands A, Campbell LE, Pohler E, Kezic S, et al. Intragenic copy number variation within filaggrin contributes to the risk of atopic dermatitis with a dose-dependent effect. *J Invest Dermatol* 2012;132:98-104.
46. Rodriguez E, Baurecht H, Wahn AF, Kretschmer A, Hotze M, Zeilinger S, et al. An integrated epigenetic and transcriptomic analysis reveals distinct tissue-specific patterns of DNA methylation associated with atopic dermatitis. *J Invest Dermatol* 2014;134:1873-83.
47. Pellerin L, Henry J, Hsu CY, Balica S, Jean-Decoster C, Mechin MC, et al. Defects of filaggrin-like proteins in both lesional and nonlesional atopic skin. *J Allergy Clin Immunol* 2013;131:1094-102.
48. Margolis DJ, Gupta J, Apter AJ, Ganguly T, Hoffstad O, Papadopoulos M, et al. Filaggrin-2 variation is associated with more persistent atopic dermatitis in African American subjects. *J Allergy Clin Immunol* 2014;133:784-9.
49. Henry J, Hsu CY, Haftek M, Nachat R, de Koning HD, Gardinal-Galera I, et al. Hornerin is a component of the epidermal cornified cell envelopes. *FASEB J* 2011;25:1567-76.
50. Marenholz I, Rivera VA, Esparza-Gordillo J, Bauerfeind A, Lee-Kirsch MA, Ciechanowicz A, et al. Association screening in the Epidermal Differentiation Complex (EDC) identifies an SPRR3 repeat number variant as a risk factor for eczema. *J Invest Dermatol* 2011;131:1644-9.
51. Kuo I, Yoshida T, De Benedetto A, Beck LA. The cutaneous innate immune response in patients with atopic dermatitis. *J Allergy Clin Immunol* 2013;131:266-78.
52. Cho S-H, Strickland I, Tomkinson A, Gelfand EW, Leung DYM. Preferential binding of *Staphylococcus aureus* to skin sites of Th2-mediated inflammation in a murine model. *J Invest Dermatol* 2001;116:658-63.
53. Cho SH, Strickland I, Boguniewicz M, Leung DYM. Fibronectin and fibrinogen contribute to the enhanced binding of *Staphylococcus aureus* to atopic skin. *J Allergy Clin Immunol* 2001;108:269-74.
54. Ong PY, Ohtake T, Brandt C, Strickland I, Boguniewicz M, Ganz T, et al. Endogenous antimicrobial peptides and skin infections in atopic dermatitis. *N Engl J Med* 2002;347:1151-60.
55. Howell MD, Wollenberg A, Gallo RL, Flaig M, Streib JE, Wong C, et al. Cathelicidin deficiency predisposes to eczema herpeticum. *J Allergy Clin Immunol* 2006;117:836-41.
56. Lai Y, Cogen AL, Radek KA, Park HJ, Macleod DT, Leichte A, et al. Activation of TLR2 by a small molecule produced by *Staphylococcus epidermidis* increases antimicrobial defense against bacterial skin infections. *J Invest Dermatol* 2010;130:2211-21.
57. Schlievert PM, Strandberg KL, Lin Y-C, Peterson ML, Leung DYM. Secreted virulence factor comparison between methicillin-resistant and methicillin-sensitive *Staphylococcus aureus*, and its relevance to atopic dermatitis. *J Allergy Clin Immunol* 2010;125:39-49.
58. Czarnecki T, Krueger JG, Guttman-Yassky E. Skin barrier and immune dysregulation in atopic dermatitis: an evolving story with important clinical implications. *J Allergy Clin Immunol Pract* 2014;2:371-9.
59. Henderson J, Northstone K, Lee SP, Liao H, Zhao Y, Pembrey M, et al. The burden of disease associated with filaggrin mutations: a population-based longitudinal birth cohort study. *J Allergy Clin Immunol* 2008;121:872-7.
60. Guttman-Yassky E, Suarez-Farinas M, Chiricozzi A, Nogales KE, Shemer A, Fuentes-Duculan J, et al. Broad defects in epidermal cornification in atopic dermatitis identified through genomic analysis. *J Allergy Clin Immunol* 2009;124:1235-44.
61. Suarez-Farinas M, Tintle SJ, Shemer A, Chiricozzi A, Nogales K, Cardinale I, et al. Nonlesional atopic dermatitis skin is characterized by broad terminal differentiation defects and variable immune abnormalities. *J Allergy Clin Immunol* 2011;127:954-64.
62. Kim BE, Leung DYM, Boguniewicz M, Howell MD. Loricrin and involucrin expression is down-regulated by Th2 cytokines through STAT-6. *Clin Immunol* 2008;126:332-7.
63. Morizane S, Yamasaki K, Kajita A, Ikeda K, Zhan M, Aoyama Y, et al. TH2 cytokines increase kallikrein 7 expression and function in patients with atopic dermatitis. *J Allergy Clin Immunol* 2012;130:259-61.
64. Zheng Y, Danilenko DM, Valdez P, Kasman I, Eastham-Anderson J, Wu J, et al. Interleukin-22, a T(H)17 cytokine, mediates IL-23-induced dermal inflammation and acanthosis. *Nature* 2007;445:648-51.
65. Nogales KE, Zaba LC, Guttman-Yassky E, Fuentes-Duculan J, Suárez-Fariñas M, Cardinale I, et al. Th17 cytokines interleukin (IL)-17 and IL-22 modulate distinct inflammatory and keratinocyte-response pathways. *Br J Dermatol* 2008;159:1092-102.
66. Chan LS, Robinson N, Xu L. Expression of interleukin-4 in the epidermis of transgenic mice results in a pruritic inflammatory skin disease: an experimental animal model to study atopic dermatitis. *J Invest Dermatol* 2001;117:977-83.
67. Sehra S, Yao Y, Howell MD, Nguyen ET, Kansas GS, Leung DYM, et al. IL-4 regulates skin homeostasis and the predisposition towards allergic skin inflammation. *J Immunol* 2010;184:3186-90.
68. Dillon SR, Sprecher C, Hammond A, Bilsborough J, Rosenfeld-Franklin M, Presnell SR, et al. Interleukin 31, a cytokine produced by activated T cells, induces dermatitis in mice. *Nat Immunol* 2004;5:752-60.
69. Ziegler SF. Thymic stromal lymphopoietin and allergic disease. *J Allergy Clin Immunol* 2012;130:845-52.
70. Jensen JM, Pfeiffer S, Witt M, Bräutigam M, Neumann C, Weichenthal M, et al. Different effects of pimecrolimus and betamethasone on the skin barrier in patients with atopic dermatitis. *J Allergy Clin Immunol* 2009;124:19-28.
71. Tintle S, Shemer A, Suarez-Farinas M, Fujita H, Gilleaudeau P, Sullivan-Whalen M, et al. Reversal of atopic dermatitis with narrow-band UVB phototherapy and biomarkers for therapeutic response. *J Allergy Clin Immunol* 2011;128:583-93.
72. Khattri S, Shemer A, Rozenblit M, Dhingra N, Czarnecki T, Finney R, et al. Cyclosporine in patients with atopic dermatitis modulates activated inflammatory pathways and reverses epidermal pathology. *J Allergy Clin Immunol* 2014;133:1626-34.
73. Leung DYM, Bhan AK, Schneeberger EE, Geha RS. Characterization of the mononuclear cell infiltrate in atopic dermatitis using monoclonal antibodies. *J Allergy Clin Immunol* 1983;71:47-56.
74. Hamid Q, Boguniewicz M, Leung DYM. Differential in situ cytokine gene expression in acute versus chronic atopic dermatitis. *J Clin Invest* 1994;94:870-6.
75. Novak N. An update on the role of human dendritic cells in patients with atopic dermatitis. *J Allergy Clin Immunol* 2012;129:879-86.
76. Dhingra N, Suarez-Farinas M, Fuentes-Duculan J, Gittler JK, Shemer A, Raz A, et al. Attenuated neutrophil axis in atopic dermatitis compared to psoriasis reflects T(H)17 pathway differences between these diseases. *J Allergy Clin Immunol* 2013;132:498-501.
77. Boyman O, Werfel T, Akdis CA. The suppressive role of IL-10 in contact and atopic dermatitis. *J Allergy Clin Immunol* 2012;129:160-1.
78. Gittler JK, Krueger JG, Guttman-Yassky E. Atopic dermatitis results in intrinsic barrier and immune abnormalities: implications for contact dermatitis. *J Allergy Clin Immunol* 2013;131:300-13.

79. Nukui T, Ehama R, Sakaguchi M, Sonogawa H, Katagiri C, Hibino T, et al. S100A8/A9, a key mediator for positive feedback growth stimulation of normal human keratinocytes. *J Cell Biochem* 2008;104:453-64.
80. Goyette J, Geczy CL. Inflammation-associated S100 proteins: new mechanisms that regulate function. *Amino Acids* 2011;41:821-42.
81. Rebane A, Zimmermann M, Aab A, Baurecht H, Koreck A, Karelson M, et al. Mechanisms of IFN-induced apoptosis of human skin keratinocytes in patients with atopic dermatitis. *J Allergy Clin Immunol* 2012;129:1297-306.
82. Howell MD, Fairchild HR, Kim BE, Bin L, Boguniewicz M, Redzic JS, et al. Th2 cytokines act on S100A11 to downregulate keratinocyte differentiation. *J Invest Dermatol* 2008;128:2248-58.
83. Wolk K, Kunz S, Witte E, Friedrich M, Asadullah K, Sabat R. IL-22 increases the innate immunity of tissues. *Immunity* 2004;21:241-54.
84. Wolk K, Witte E, Wallace E, Döcke WD, Kunz S, Asadullah K, et al. IL-22 regulates the expression of genes responsible for antimicrobial defense, cellular differentiation, and mobility in keratinocytes: a potential role in psoriasis. *Eur J Immunol* 2006;36:1309-23.
85. Boniface K, Bernard FX, Garcia M, Gurney AL, Lecron JC, Morel F. IL-22 inhibits epidermal differentiation and induces proinflammatory gene expression and migration of human keratinocytes. *J Immunol* 2005;174:3695-702.
86. Boguniewicz M, Leung DYM. The ABC's of managing patients with severe atopic dermatitis. *J Allergy Clin Immunol* 2013;132:511-2.
87. Arkwright PD, Stafford JC, Sharma Y. Atopic dermatitis in children. *J Allergy Clin Immunol Pract* 2014;2:388-95.
88. Tang TS, Bieber T, Williams HC. Are the concepts of induction of remission and treatment of subclinical inflammation in atopic dermatitis clinically useful? *J Allergy Clin Immunol* 2014;133:1615-25.
89. Schmitt J, von Kobyletzki L, Svensson A, Apfelbacher C. Efficacy and tolerability of proactive treatment with topical corticosteroids and calcineurin inhibitors for atopic eczema: systematic review and meta-analysis of randomized controlled trials. *Br J Dermatol* 2011;164:415-28.
90. Roekevisch E, Spuls PI, Kuester D, Limpens J, Schmitt J. Efficacy and safety of systemic treatments for moderate-to-severe atopic dermatitis: a systematic review. *J Allergy Clin Immunol* 2014;133:429-38.
91. Flohr C, Irvine AD. Systemic therapies for severe atopic dermatitis in children and adults. *J Allergy Clin Immunol* 2013;132:774.
92. Nicol NH, Boguniewicz M, Strand M, Klennert MD. Wet wrap therapy in children with moderate to severe atopic dermatitis in a multidisciplinary treatment program. *J Allergy Clin Immunol Pract* 2014;2:400-6.
93. Novak N, Bieber T, Hoffmann M, Fölster-Holst R, Homey B, Werfel T, et al. Efficacy and safety of subcutaneous allergen-specific immunotherapy with depigmented polymerized mite extract in atopic dermatitis. *J Allergy Clin Immunol* 2012;130:925-31.
94. Bae JM, Choi YY, Park CO, Chung KY, Lee K. Efficacy of allergen-specific immunotherapy for atopic dermatitis: a systematic review and meta-analysis of randomized controlled trials. *J Allergy Clin Immunol* 2013;132:110-7.
95. Rozenblit M, Suárez-Fariñas M, Shemer A, Khattri S, Gilleaudeau P, Sullivan-Whalen M, et al. Residual genomic profile after cyclosporine may offer insights into atopic dermatitis recurrence. *J Allergy Clin Immunol* 2014;134:955-7.
96. Suarez-Farinas M, Gittler J, Shemer A, Cardinale I, Krueger JG, Guttman-Yassky E. Residual genomic signature of atopic dermatitis despite clinical resolution with narrowband UVB. *J Allergy Clin Immunol* 2013;131:577-9.
97. Simpson EL, Keck LE, Chalmers JR, Williams HC. How should an incident case of atopic dermatitis be defined? A systematic review of primary prevention studies. *J Allergy Clin Immunol* 2012;130:137-44.
98. Jensen MP, Meldrum S, Taylor AL, Dunstan JA, Prescott SL. Early probiotic supplementation for allergy prevention: long term outcomes. *J Allergy Clin Immunol* 2012;130:1209-11.
99. Lau S, Gerhold K, Zimmermann K, Ockeloen CW, Rossberg S, Wagner P, et al. Oral application of bacterial lysate in infancy decreases the risk of atopic dermatitis in children with 1 atopic parent in a randomized, placebo-controlled trial. *J Allergy Clin Immunol* 2012;129:1040-7.
100. Penders J, Gerhold K, Stobbering EE, Thijs C, Zimmermann K, Lau S, et al. Establishment of the intestinal microbiota and its role for atopic dermatitis in early childhood. *J Allergy Clin Immunol* 2013;132:601-7.
101. Muehleisen B, Gallo R. Vitamin D in allergic disease: shedding light on a complex problem. *J Allergy Clin Immunol* 2013;131:324-9.
102. Zhang Y, Leung DYM, Goleva E. Anti-inflammatory and corticosteroid-enhancing actions of vitamin D in the monocytes of patients with steroid-resistant and those with steroid-sensitive asthma. *J Allergy Clin Immunol* 2014;133:1744-52.
103. Nanzer AM, Chambers ES, Ryanna K, Richards DF, Black C, Timms CM, et al. Enhanced production of IL-17A in patients with severe asthma is inhibited by 1 α ,25-dihydroxyvitamin D3 in a glucocorticoid-independent fashion. *J Allergy Clin Immunol* 2013;132:297-304.e3.
104. Cheng HM, Kim S, Park GH, Chang SE, Bang S, Won CH, et al. Low vitamin D levels are associated with atopic dermatitis, but not allergic rhinitis, asthma, or IgE sensitization, in the adult Korean population. *J Allergy Clin Immunol* 2014;133:1048-55.
105. Mohiuddin MS, Curran-Everett D, Leung DYM. Vitamin D and food allergy in patients with severe atopic dermatitis. *J Allergy Clin Immunol* 2013;132:1011.
106. Allen KJ, Koplin J, Ponsonby AL, Vuillermin P, Dharmage S. Vitamin D and food allergy in patients with severe atopic dermatitis. *J Allergy Clin Immunol* 2013;132:1011-2.
107. Eichenfield LF, Tom WL, Chamlin SL, Feldman SR, Hanifin JM, Simpson EL, et al. Guidelines of care for the management of atopic dermatitis: section 1. Diagnosis and assessment of atopic dermatitis. *J Am Acad Dermatol* 2014;70:33-51.
108. Boguniewicz M, Leung DY. Recent insights into atopic dermatitis and implications for management of infectious complications. *J Allergy Clin Immunol* 2010;125:4-15.
109. Tang TS, Bieber T, Williams H. Does "autoreactivity" play a role in eczema? *J Allergy Clin Immunol* 2012;129:1209-15.
110. Novak N, Allam J-P, Bieber T. Allergic hyperreactivity to microbial components: a trigger factor of "intrinsic" atopic dermatitis? *J Allergy Clin Immunol* 2003;112:215-6.
111. Hiragun T, Ishii K, Hiragun M, Suzuki H, Kan T, Mihara S, et al. Fungal protein MGL_1304 in sweat is an allergen for atopic dermatitis patients. *J Allergy Clin Immunol* 2013;132:608-15.
112. Kong HH, Oh J, Deming C, Conlan S, Grice EA, Beatson MA, et al. Temporal shifts in the skin microbiome associated with disease flares and treatment in children with atopic dermatitis. *Genome Res* 2012;22:850-9.
113. Bin L, Kim BE, Brauweiler A, Goleva E, Streib J, Ji Y, et al. *Staphylococcus aureus* a-toxin modulates skin host response to viral infection. *J Allergy Clin Immunol* 2012;130:683-91.
114. Leung DY, Gao PS, Grigoryev DN, Rafaels NM, Streib JE, Howell MD, et al. Human atopic dermatitis complicated by eczema herpeticum is associated with abnormalities in IFN- γ response. *J Allergy Clin Immunol* 2011;127:965-73.
115. Beck LA, Boguniewicz M, Hata TR, Schneider LC, Hanifin JM, Gallo RL, et al. Phenotype of atopic dermatitis subjects with a history of eczema herpeticum. *J Allergy Clin Immunol* 2009;124:260-9.
116. Silverberg JI, Silverberg NB. Childhood atopic dermatitis and warts are associated with increased risk of infection: a US population-based study. *J Allergy Clin Immunol* 2014;133:1041-7.
117. Rebane A, Runnel T, Aab A, Maslovskaja J, Rückert B, Zimmermann M, et al. MicroRNA-146a alleviates chronic skin inflammation in atopic dermatitis through suppression of innate immune responses in keratinocytes. *J Allergy Clin Immunol* 2014;134:836-47.
118. Otsuka A, Doi H, Egawa G, Maekawa A, Fujita T, Nakamizo S, et al. Possible new therapeutic strategy to regulate atopic dermatitis through upregulating filaggrin expression. *J Allergy Clin Immunol* 2014;133:139-46.
119. Broccardo CJ, Mahaffey S, Schwarz J, Wruck L, David G, Schlievert PM, et al. Comparative proteomic profiling of patients with atopic dermatitis based on history of eczema herpeticum infection and *Staphylococcus aureus* colonization. *J Allergy Clin Immunol* 2011;127:186-93.
120. Sakabe J, Kamiya K, Yamaguchi H, Ikeya S, Suzuki T, Aoshima M, et al. Proteome analysis of stratum corneum from atopic dermatitis patients by hybrid quadrupole-orbitrap mass spectrometer. *J Allergy Clin Immunol* 2014;134:957-60.
121. Choy DF, Hsu DK, Seshasayee D, Fung MA, Modrusan Z, Martin F, et al. Comparative transcriptomic analyses of atopic dermatitis and psoriasis reveal shared neutrophilic inflammation. *J Allergy Clin Immunol* 2012;130:1335-43.
122. Bin L, Edwards MG, Heiser R, Streib J, Richers B, Hall C, et al. Identification of novel gene signatures in patients with atopic dermatitis complicated by eczema herpeticum. *J Allergy Clin Immunol* 2014;134:848-55.
123. Keles S, Jabara H, Reisli I, McDonald D, Barlan I, Hanna-Wakim R, et al. Plasmacytoid dendritic cell depletion in DOCK8 deficiency: Rescue of severe herpetic infections with IFN- α 2b therapy. *J Allergy Clin Immunol* 2014;133:1753-5.
124. Suarez-Farinas M, Dhingra N, Gittler J, Shemer A, Cardinale I, Strong CD, et al. Intrinsic atopic dermatitis shows similar T(H)2 and higher T(H)17 immune activation compared with extrinsic atopic dermatitis. *J Allergy Clin Immunol* 2013;132:361-70.
125. Nakajima S, Igyártó BZ, Honda T, Egawa G, Otsuka A, Hara-Chikuma M, et al. Langerhans cells are critical in epicutaneous sensitization with protein antigen via thymic stromal lymphopoietin receptor signaling. *J Allergy Clin Immunol* 2012;129:1048-55.
126. Landheer J, Giovannone B, Mattson JD, Tjabringa S, Bruijnzeel-Koomen CAFM, McClanahan T, et al. Epicutaneous application of house dust mite induces thymic stromal lymphopoietin in non-lesional skin of patients with atopic dermatitis. *J Allergy Clin Immunol* 2013;132:1252-4.
127. Teraki Y, Sakurai A, Izaki S. IL-13/IL-22-coproducing T cells, a novel subset, are increased in atopic dermatitis. *J Allergy Clin Immunol* 2013;132:971-4.