Mechanisms of human drug-induced anaphylaxis

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Drug-induced anaphylaxis is a hyperacute reaction affecting multiple organs that can be of fatal consequence. Its incidence is increasing, consistent with a global increased sensitization to various allergens and drugs in the population. Few risk factors and mechanisms have been identified from human studies due to the rarity of anaphylactic events and their unpredictability. This systemic reaction is caused by the rapid release of a large range of functionally diverse mediators, including histamine and platelet-activating factor as the main drivers identified. Mechanisms defined from models of experimental anaphylaxis identify drug-specific antibodies of the IgE and IgG class that link the drug to antibody receptors on multiple cell types, causing their activation and mediator release. In the case of drugs with peculiar chemical structures, antibodies may not be necessary because drug-binding receptors, such as Mas-related G protein–coupled receptor member X2, have been identified. This review describes the complex reaction leading to drug-induced anaphylaxis that can involve various antibody classes, various cell types—including mast cells, neutrophils, platelets, basophils, macrophages, and monocytes—and their mediators and receptors that, importantly, can be activated alone or in association to participate in the severity of the reaction. (J Allergy Clin Immunol 2021;147:1133-42.)

Key words: *Anaphylaxis, drugs, IgE, IgG, MRGPRX2, platelet-activating factor, histamine, serotonin, mast cells, basophils, neutrophils, platelets*

Abbreviations used

CD32A:	Human-activating IgG receptor FcγRIIA
FcεRI:	High-affinity receptor for the Fc portion of IgE
FcγR:	Receptor for the Fc portion of IgG
Mrgprb2:	Mas-related G protein–coupled receptor member b2
MRGPRX2:	Mas-related G protein–coupled receptor member X2
NMBA:	Neuromuscular-blocking agent
PAF:	Platelet-activating factor
PAF-R:	PAF-receptor
QA:	Quaternary ammonium

Anaphylaxis is a hyperacute reaction that can be of fatal consequence. It is a systemic reaction caused by the rapid and systemic release of a large range of functionally diverse mediators affecting multiple organs. These mediators typically induce urticaria, vasodilatation, increased vascular permeability and vascular leakage, edema, and bronchoconstriction, leading to a drop in arterial pressure, tachycardia, bronchospasm, and digestive troubles. Death can be caused by the resulting cardiac failure and/or asphyxia or pulmonary edema following major bronchospasm. Anaphylactic reactions cannot, in general, be foreseen. Because of their life-threatening nature, they represent an emergency situation for the medical staff.

The more recent publications describe a world incidence of anaphylaxis in humans at between 50 and 112 episodes per 100,000 person-years, and drug allergy mortality is estimated at 0.05 to 0.51 per million people/y.[1] Interestingly, drug-induced anaphylaxis incidence is increasing, consistent with a global increased sensitization to various allergens in the population, including drugs.[2] Almost 60% of fatal anaphylaxis cases have been attributed to drugs.[3,4] Because of their increasing availability, the anaphylaxis to mAbs jumped at an average rate of 0.77% of total anaphylaxis reports per year in the United States, from 2.00% in 1999 to 17.37% in 2019; it was the fastest increase observed among all the drugs responsible for anaphylaxis.[5] Surprisingly, very different drugs—whether considering chemical nature or structure, size, target, mode of action, or biodistribution—lead to anaphylactic events with similar symptoms and consequences. The most frequent culprit drugs are antibiotics (mostly penicillin and cephalosporins), nonsteroidal anti-inflammatory drugs, injected radiocontrast agents (iodinated contrast media and gadolinium), antineoplastic drugs, therapeutic antibodies, and neuromuscular-blocking agents (NMBAs) used during surgery.[3,4,6] Even more surprisingly, the size of most of these compounds is 100 to 1000 times smaller than that of "classical" allergens—linked to allergic reactions to pollens, house dust mite, food allergens—and due to this minimal size, these drugs would rather qualify as ***haptens*** (Fig 1): antibiotics, for example, penicillin, 334 Da; ciprofloxacin, 331 Da; nonsteroidal anti-inflammatory drugs, for example, ibuprofen, 206 Da, and

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Terms in boldface and italics are detailed in the glossary on page 1134.

dicolfenac, 296 Da; radiocontrast agents, for example, diatrizoate, 613 Da; and NMBA, for example, suxamethonium, 361 Da, and rocuronium, 530 Da. Their small size, allowing them to passively diffuse systemically, could be interpreted as a common feature of drugs with anaphylactic potential. Nevertheless, drugs of radically larger sizes, proteins of 20 to 180 kDa including therapeutic antibodies, for example, infliximab, 149 kDa, and cetuximab, 152 kDa, or enzymes used for enzyme replacement therapy, for example, glucocerebrosidase, 60 kDa, and polymers, 200 to 35,000 kDa, contained in drug preparations such as polyethylene glycol[7,8] (Fig 1) that do not diffuse passively are also reported to cause drug anaphylaxis with similar kinetics. Adding to the complexity, the route of administration of the drug responsible for anaphylaxis can be multiple: oral, infused, injected (intravenous, intradermal, subcutaneous, intramuscular). Concerning the inhalation route, some cases have been reported in asthmatic children using inhaled corticosteroids, probably related to milk protein traces.[9] This variability in chemical nature, size, and biodistribution of culprit drugs for anaphylactic events makes it difficult to envision a single mechanism responsible for anaphylaxis induction.

Evidence of the mechanisms responsible for anaphylaxis from human studies is scarce due to the rarity of anaphylaxis and its unpredictability, and thus of the very few prospective clinical studies performed so far. Similarities between local allergic reactions (eg, skin rashes and edema) and low-grade systemic anaphylaxis has led to proposing mechanisms of allergic reactions as the basis of severe anaphylactic reactions also without solid evidence to support them. Thus, clinical research in anaphylaxis has mainly focused on accumulating evidence of an "allergic" mechanism, including the presence of certain mediators (eg, histamine), enzymes (eg, tryptase), and antibodies (eg, IgE) classically involved in local allergic reactions. Whether histamine, tryptase, and allergen-specific IgE are rather biomarkers than actual triggers of the anaphylactic reaction, which may be induced by other mechanisms entirely, will be discussed herein. As an example, histamine is found at elevated levels during anaphylactic reactions and proposed as the main mediator of anaphylaxis, but antihistamines do not demonstrate efficacy on severe anaphylaxis symptoms. Below are summarized risk factors and evidence from human studies to propose that anaphylaxis is an integration of diverse mechanisms leading to systemic organ failure rather than, simply put, an extreme allergic reaction.

RISK FACTORS AND EVIDENCE FROM HUMAN STUDIES

Few risk factors have been identified that increase the risk of developing a drug-induced anaphylactic event. Sex remains a matter of debate with controversies on higher rates of drug-induced anaphylaxis in women,[3,10] whereas old age has been linked to both an increased risk of severe reactions and a higher incidence,[11] with preexisting cardiovascular morbidity being an important cofactor.[4] Surprisingly, ***atopy*** and allergic status of the patients do not appear to be convincingly related to a higher risk of drug-induced anaphylaxis,[6] suggestive that different or additional mechanisms may be at play in "systemic" anaphylaxis compared with more "local" allergic reactions. Nevertheless, patients with mastocytosis, a disease characterized by the presence of high numbers of mast cells in various organs, have a high occurrence of anaphylaxis,[12] suggesting a role of mast cells—the crucial effector cell of allergic reactions and inflammation[13]—in anaphylaxis.

Mast cells are notorious for their ability to quickly release histamine, the major mediator recognized in hypersensitivity reactions. Although antihistamines have not proven efficacious to prevent or treat severe anaphylaxis, intravenous administration of histamine in volunteers has been shown to reproduce most signs and symptoms of anaphylaxis, including cutaneous flushing, headache, airway obstruction, and transient hemodynamic changes, mainly evidenced by systemic hypotension and tachycardia.[14,15] Thus, histamine has the capacity to mediate the symptoms of anaphylaxis, but is clearly not the sole mediator involved. Vadas et al[16] indeed reported in their landmark study in 2008 that

GLOSSARY

ALLERGIC DESENSITIZATION: A method to develop tolerance to a particular allergen through the administration of progressively larger doses of the allergen to decrease IgE-mediated responses.

ATOPY: A genetic-related tendency to produce heightened allergic reactions against otherwise harmless non–self-antigens. It is associated with increased tendency to develop allergic rhinitis, asthma, and atopic dermatitis.

BASOPHIL ACTIVATION TEST: A flow cytometry–based test in which some basophil surface expression of activation markers is quantified following stimulation with different allergens. A positive result is considered to be an *in vitro* surrogate to an immediate allergic reaction *in vivo*. Basophil activation test is a good diagnostic tool in addition to skin tests.

ENDOTYPES: A subtype of a disease condition that is caused by a distinct functional or pathophysiological mechanism.

EPITOPE: The part of the antigen molecule to which an antibody binds.

HAPTENS: Small molecules that elicit immune responses only when attached to a larger carrier molecule, such as a protein.

IgG RECYCLING RECEPTOR: Cell surface protein that allows for transcytosis of IgG and albumin and increases their half-life in the serum by bypassing lysosomal degradation.

NETosis: Physiological process by which neutrophils release neutrophil extracellular traps (NETs), which are web-like complexes of chromosomal DNA, histones, and granule and cytoplasmic proteins, to ensnare extracellular pathogens and participate in their destruction/killing. NETosis involves plasma membrane disruption and is a form of active cell death in some situations. They also participate in tissue damage in acute or chronic inflammation, as well as in autoimmunity.

SKIN PRICK TEST: A test that screens for hypersensitivity to different allergens. It involves placing small drops of allergen in various dilutions onto different areas at the skin surface, introducing the allergens into the top layer of the skin through pricks, and then assessing for wheal and flare local reactions after 15 minutes.

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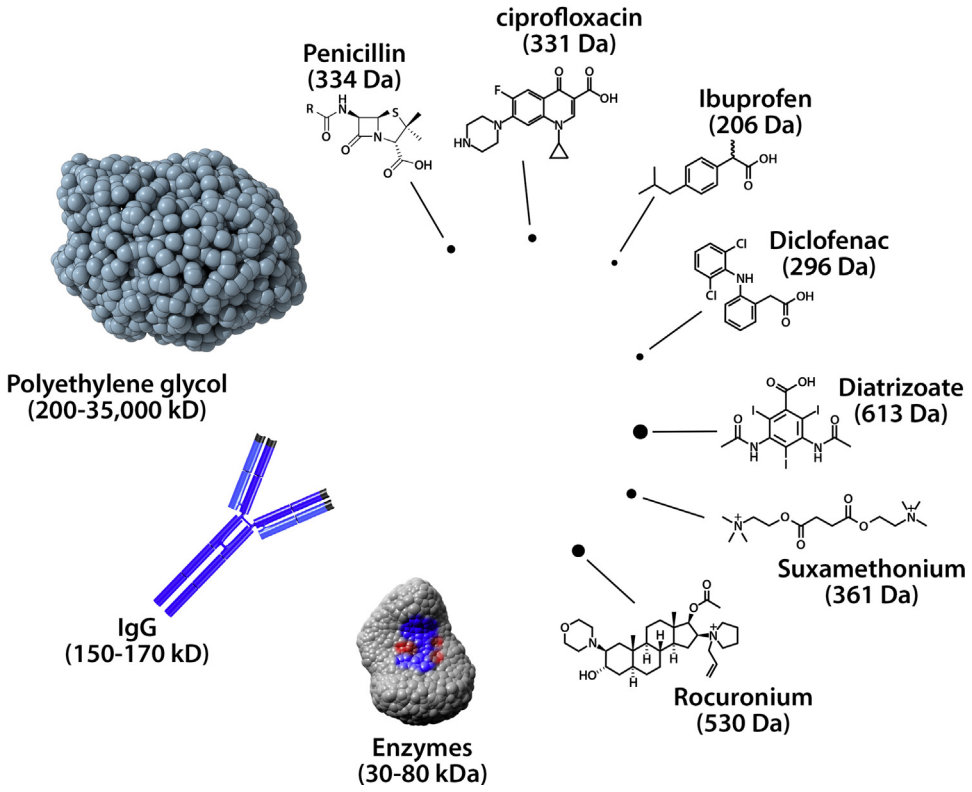

FIG 1. Relative drug sizes implicated in drug-induced anaphylaxis: size does not matter. Schematic representations of polyethylene glycol (Image Credit: StudioMolekuul/Shutterstock.com), IgG, enzyme (Image credit; PDB 9LYZ), and chemical structures of indicated drugs and contrast agents. Dots represent the relative size of the depicted small molecules (<1 kDa) compared with those of polyethylene glycol, IgG, and enzymes.

platelet-activating factor (PAF) levels in serum were directly correlated, and the activity of its degrading enzyme, PAF acetylhydrolase, inversely correlated with the severity of anaphylaxis. Their follow-up work[17] reported that histamine, PAF, and tryptase, the major enzyme of mast cell secretory granules already identified as a biomarker for anaphylaxis,[18] were all detected in serum of patients who underwent anaphylaxis of low (grade 1), mild (grade 2), and severe (grade 3) severity. However, serum concentrations of PAF and tryptase, but not histamine, correlated with anaphylaxis severity.[17] PAF is an extremely potent lipid mediator that can activate various cells that express the PAF-receptor (PAF-R), including endothelium, smooth muscle, and myeloid cells including mast cells. Thus, PAF could directly elicit the circulatory and respiratory symptoms of anaphylaxis while also eliciting the generation of other mediators involved in anaphylaxis propagation and severity. Intracutaneous injection or inhalation of PAF elicit symptoms resembling grade 1 anaphylaxis and bronchoconstriction, respectively, in human subjects.[19,20] Even though deficiencies in the enzyme degrading PAF, PAF acetylhydrolase (ie, leading to high levels of PAF), have been correlated with respiratory deficiencies in asthmatic children,[21] no study has linked it yet to anaphylaxis. Nevertheless, PAF-acetylhydrolase activity inversely correlates with anaphylaxis severity, and can be used as its marker.[16,22] In contrast to PAF, mast cell tryptase is not thought to elicit rapid responses that contribute to immediate manifestation of anaphylaxis, although its effects are only partially described.[23] Mast cells in the vicinity of an activated mast cell releasing tryptase have been reported to become activated in turn and to release histamine.[24] Tryptase is considered mainly a "practical" marker of mast cell activation because it can be easily detected in serum.[18] Altogether, these data suggest that among identified immediate mediators with potency to be anaphylaxis inducers, PAF, rather than histamine, is the contributor of the more severe forms of anaphylaxis (grade 3) and potentially of lethality (grade 4) when highly abundant systemically. More explorations of anaphylactogenic mediators (eg, leukotrienes and prostaglandins) in human drug-induced anaphylaxis remain to be performed to understand fully the mechanisms leading to moderate and severe symptoms, or even to lethality.

Until recently, only 1 pathway had been universally accepted as the mechanistic explanation of anaphylaxis induction: the IgE antibody pathway. Antibodies of the IgE class are generated in small quantities by B lymphocytes. Once produced, IgE antibodies have a very short half-life in circulation because they cannot be recycled by the neonatal ***IgG recycling receptor*** FcRn. Total IgE levels are thus only 50 to 200 ng/mL in healthy individuals but generally several fold higher in patients with allergy, with patients having allergen-specific IgE levels up to 200 ng/mL, particularly those who experienced anaphylactic events.[25] Direct evidence of the role of IgE in human anaphylaxis is based on transfer of purified human IgE in the skin of human volunteers that transferred allergen reactivity[26] and several clinical trials using the anti-IgE therapeutic antibody omalizumab, with one suggesting less spontaneous episodes of anaphylaxis in patients with mastocytosis,[27] and others proposing anti-IgE therapy as an adjunct therapy for ***allergic desensitization***, leading to fewer anaphylactic episodes.[28-30]

Although the precise conditions for human B cells to start producing an IgE remain speculative and extrapolated from data

obtained in animal models,[31] recent human studies found evidence that allergen-specific IgE B cells arise from mature B cells, producing initially an allergen-specific IgG.[32] Although elusive, human circulating nonsecreting IgE B cells, that is, IgE memory B cells, as well as noncirculating IgE-secreting B cells, that is, IgE plasma cells, have recently been identified (in extremely low numbers) in the blood and bone marrow, respectively, of patients with allergy.[33,34] Indirectly supporting a role for these IgE-secreting cells located in the bone marrow in human anaphylaxis, bone marrow transplantation from allergic donors to nonallergic recipients has been reported in a few cases to transfer drug hypersensitivity, penicillin hypersensitivity for example (reviewed in Khan et al[35]). Although several donor cell types in the allograft may contribute to the transfer of hypersensitivity, specific penicillin IgE could be detected 3 months after transplant, supporting the likely importance of graft-associated IgE-producing B cells.[36] This hypothesis is further supported by the observation that transplantation of livers from fatal anaphylaxis cases transferred food hypersensitivity (nuts or peanuts) to recipients with either detectable specific IgE or positive ***skin prick test*** result (reviewed in Khan et al[35]). Of note, IgE-producing B cells related to food allergy have been identified in the gut, and arise most probably from mature B cells, producing initially a food allergen-specific IgA.[37] Wherever IgE is anatomically secreted—bone marrow, liver, gut—it has the unique ability to "sensitize" not only human mast cells in tissue but also human basophils in blood, empowering them with the ability to react to various specific targets, including allergens and drugs. This phenomenon, unique among antibody classes, relies on the IgE receptor FcεRI (high-affinity receptor for the Fc portion of IgE) that these cells express constitutively. FcεRI is of such high affinity that once bound an IgE remains on a mast cell for weeks.[38] Upon penetration of a drug/allergen in the body, it will bind to IgE-sensitized cells, and provoke FcεRI aggregation on their surface, leading to cell activation, degranulation, and mediator release, including histamine, tryptase, and PAF.

Although IgE may be responsible for many cases of anaphylaxis to drug or allergen, it may be undetectable in others.[39] In such cases, the terms "anaphylactoid reactions" (ie, anaphylaxis-like reaction) or "idiopathic anaphylaxis" (ie, anaphylaxis of unclear trigger) may be applied. IgG antibodies are proposed as causative agents, and can be detected in patients who react to NMBAs,[40] polyethylene glycol,[41] therapeutic antibodies, and other drugs (reviewed in Finkelman et al[42]). IgG antibodies could trigger activation of neutrophils and other cells bearing activating IgG receptors (FcγR), either directly inducting anaphylaxis or acting in concert with IgE having the same specificity (refer to section NMBA-induced anaphylaxis exemplflies multiple mechanisms at play). In addition, anaphylactic reactions to certain drugs may be caused by direct interaction with Mas-related G protein–coupled receptor member X2 (MRGPRX2),[43,44] which is expressed at high level in primary human skin and synovial mast cells, but not in primary lung mast cells.[45] Many drugs capable of directly inducing histamine release can bind and activate MRGPRX2[46] (Fig 2).

MECHANISMS IDENTIFED IN EXPERIMENTAL ANAPHYLAXIS

Most animal models of systemic anaphylaxis are directly relevant to drug-induced anaphylaxis because they are based on the injection of a bolus of allergen/antigen/drug into a sensitized animal. Two main types of models are used. In passive systemic anaphylaxis, naive animals are directly injected with an anaphylactogenic mediator (eg, histamine and PAF) or with antibodies thought to be responsible for anaphylaxis induction followed by a challenge with an allergen/antigen/drug in the next hours or days. In active systemic anaphylaxis, animals are exposed to low doses of an allergen/antigen/drug to induce an antibody response against that molecule followed by a challenge several weeks later. In the latter case, initial exposures can be performed in the presence or absence of an adjuvant. Surprisingly, models of active sensitization reveal that the presence or absence of adjuvant influences the principal mechanisms leading to anaphylaxis induction.[47-49] Thus, each animal model of anaphylaxis, even each experimental protocol thereof, will draw a different picture of what pathways of anaphylaxis in humans may be (discussed in Finkelman et al[42]). Even though data may be considered conflicting between studies, animal models have provided an enlightened understanding of the multiple mechanisms at play, which are, in our view, the current basis of human anaphylaxis exploration and dogma: antibodies of the IgE and/or IgG class to the culprit drug triggering multiple cell types through activating antibody receptors, or mast cell– activating receptors directly triggered by some drugs, notably Mas-related G protein–coupled receptor member b2 (Mrgprb2), the mouse ortholog of human MRGPRX2.

Passive systemic anaphylaxis

Injection of histamine or PAF in mice leads to symptoms resembling systemic anaphylaxis that are dependent on the presence of histamine receptors or PAF-R, respectively.[50] Injection of allergen-specific IgE or allergen-specific IgG followed by allergen challenge (generally intravenous injection) hours to days later provokes systemic, sometimes fatal, anaphylaxis that requires expression of FcεRI or IgG receptors (FcγR [receptor for the Fc portion of IgG]), respectively (reviewed in Finkelman[47] and Gillis et al[48]). Both IgE and IgG receptors require cross-linking to trigger cell activation, implying that multiple IgE or IgG molecules need to bind the same drug molecule, or that the drug has been haptenized onto a carrier molecule to allow multimeric interactions. Among mouse IgG subclasses, allergen-specific IgG_{2a} and IgG_{2b} are potent inducers, whereas IgG_1 is weak,[51] in line with its preferential binding to inhibitory mouse FcγR[52] and its rather anti-inflammatory role in mice.[53] Ciprofloxacin, an antibiotic of the fluoroquinolone family, can induce antibody-independent anaphylaxis in mice. Mice lacking Mrgprb2 (the mouse ortholog of MRGPRX2) are protected from ciprofloxacin-induced anaphylaxis.[44] As is the case for MRGPRX2 in humans, Mrgprb2 in mice is expressed almost exclusively on mast cells and is therefore considered a direct target of several drugs belonging to fluoroquinolones, NMBA (eg, atracurium and rocuronium) chemical classes, and cationic peptides.[43] A novel mouse model allowing for the development of human mast cells expressing MRGPRX2 reported local mast cell degranulation after exposure to contrast agents, but did not investigate systemic reactions.[54] Altogether passive models of anaphylaxis validate antibody classes IgE and IgG and their receptors, Mrgprb2, and mediators histamine and PAF as potential inducers of anaphylaxis in simplified models (Fig 2), but are not able to rank them or discriminate among them for their relevance in human anaphylaxis.

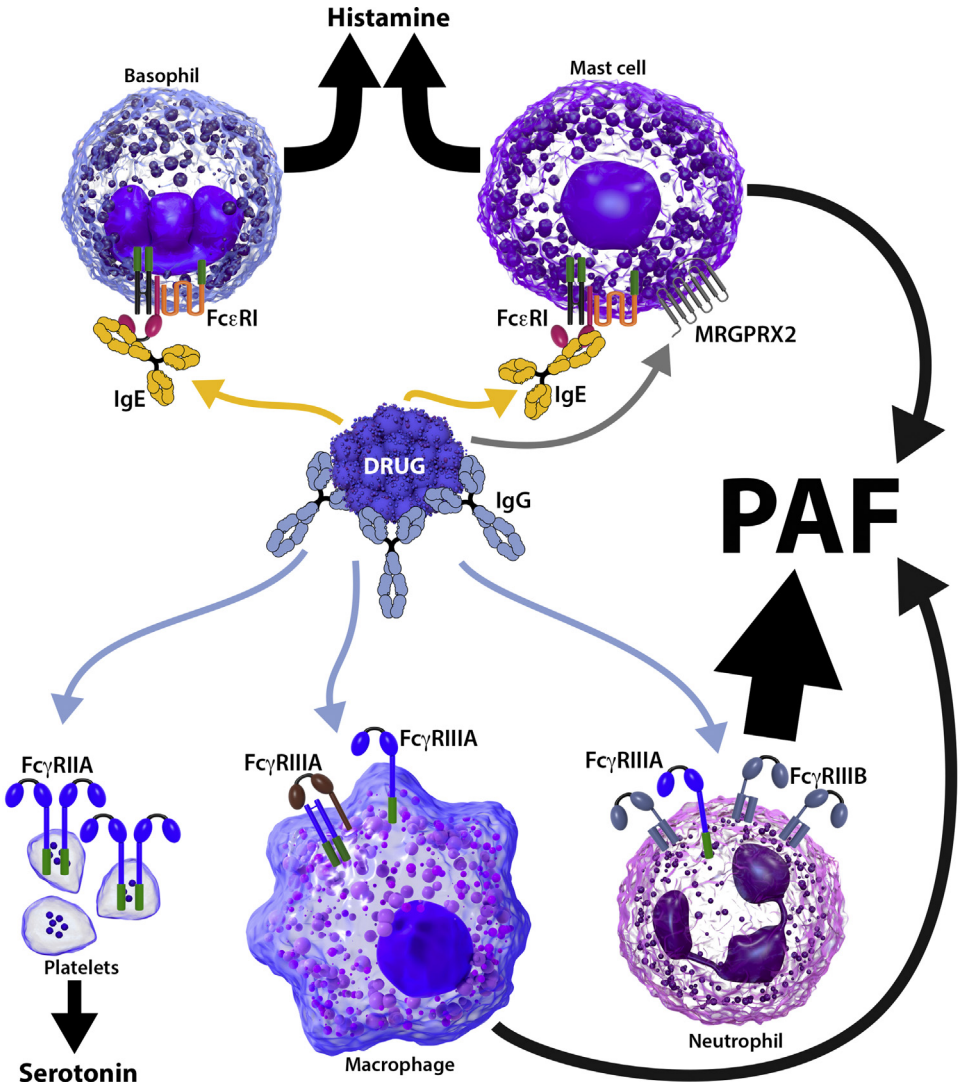

FIG 2. Potential pathways in drug-induced anaphylaxis. Once administrated a drug can be bound by (a) drug-specific IgE antibodies, prebound on their high-affinity IgE receptor (FcεRI)-expressing mast cells and basophils, leading to their release of anaphylactogenic mediators, histamine, and to some extent PAF (*Note*: human mast cells are thought to make little or no serotonin); (b) drug-specific IgG antibodies, forming drug-IgG immune complexes that can bind to their low-affinity IgG receptor (FcγR)-expressing neutrophils (eg, FcγRIIA and FcγRIIIB) and monocyte/macrophages (eg, FcγRIIA and FcγRIIIA), leading to their release of PAF, and to FcγRIIA-platelets, leading to their release of serotonin; (c) mast cell–expressed MRGPRX2 if that drug has affinity for this receptor, leading to mast cell degranulation and histamine and PAF release. The thickness of the black arrows represents their contribution to the indicated mediator release.

Active systemic anaphylaxis

Mice sensitized with allergen in the presence of adjuvants show detectable IgE and IgG specific for the allergen, and develop anaphylaxis upon allergen challenge (intravenous, gavage), with severity increasing with higher doses of allergen. Surprisingly, IgE-deficient or FcεRI-deficient mice were protected from some active anaphylaxis models, but not others, demonstrating that the "IgE pathway" is not necessary in some models of active systemic anaphylaxis (reviewed in Gillis et al,[48] Finkelman et al,[42] and Reber et al[55]). In contrast, mice lacking all activating IgG and IgE receptors (mice deficient for the FcRγ chain, lacking all activating IgG and IgE receptors) were resistant to anaphylaxis, as well as mice lacking only IgG receptors (mice deficient for FcγRI, FcγRIIB, FcγRIII, and FcγRIV).[56-58] Transgenic expression in mice deficient for FcγRI, FcγRIIB, FcγRIII, and FcγRIV of a single[59] or of multiple human FcγR[57,58] restored anaphylaxis, demonstrating the requirement of the "IgG pathway" in severe active systemic anaphylaxis. Even though convincing animal studies on the potential contribution of the complement system to anaphylaxis are still lacking (discussed in Finkelman et al[42]), some compounds trigger complement component C3a production, leading to myeloid cell activation through their complement receptors and, thus, to PAF and histamine release.[60] Mice deficient in either PAF-R or cytosolic phospholipase A2, which is required for PAF generation (and, in the case of cytosolic phospholipase A2, leukotriene and

prostaglandin generation), had markedly reduced anaphylaxis symptoms.[50,56] PAF-R antagonists consistently strongly inhibited anaphylaxis symptoms and protected from lethality in different mouse models of active anaphylaxis, whereas antihistamines had moderate to negligible effects, unless in concert with PAF-R antagonists.[49,51,56,61,62] Depending on the active systemic anaphylaxis model and/or the mouse strain used, tissue-resident mast cells and macrophages, and circulating basophils, monocytes, and neutrophils have all been convincingly reported to be main actors of the anaphylactic reaction (reviewed in Gillis et al,[48] Finkelman et al,[42] and Reber et al[55]). More recently, platelets were added to this list through 2 independent reports using human-activating IgG receptor FcγRIIA (CD32A) transgenic mice, proposing that platelets release pathogenic serotonin in response to FcγRIIA triggering on their surface by circulating IgG-antigen/allergen immune complexes (Fig 2)[58,63] that form following exposure to high amounts of antigen/allergen as is mostly the case in drug anaphylaxis. The abundance and systemic distribution of platelets suggests a plausible role in anaphylaxis. Because humans and nonhuman primates, but not rodents, express FcγRIIA,[52] the potential contribution of platelets in anaphylaxis models may be missed in mice lacking transgenic expression of FcγRIIA.

Although animal studies suggest a diverse range of initiating pathways for anaphylaxis, demonstrating the contribution of each pathway in severe human anaphylaxis is challenging, because sampling of blood is typically undertaken after the reaction has occurred. Nevertheless, markers have been proposed to confirm the engagement of the IgE pathway (increase in IL-4 and soluble IL-4 receptor levels) and/or the IgG pathway (decrease in FcγR expression),[64] which have been reported to occur simultaneously in a mouse model of fatal anaphylaxis.[61] Our group demonstrated that the reduced FcγR expression was a marker of the IgG pathway in passive and active mouse models of anaphylaxis[51,57] and in our recent clinical study on NMBA-induced anaphylaxis (described in the next section).[40] The diverse range of candidate effector cell types identified in animal studies makes their relative contribution difficult to comprehend in humans. The contributions from different cell types are likely influenced by cell numbers, their capacities for activation, and the abundance of mediators generated per cell. Among circulating cells, platelets (150,000-450,000/μL) are approximately 70, approximately 700, and approximately 100,000-fold more abundant than neutrophils (2,500-6,000/μL), monocytes (200-600/μL), and basophils (1-3/μL). How these compare to mast cell and macrophage numbers is unclear because both cell types reside in various human tissues in which they differentiate into different subpopulations expressing different enzymes, receptors (including different IgG receptors and levels of MRGPRX2 for mast cells), and mediators.[65,66] Skin mast cell densities in humans vary depending on the anatomical location.[67] Considering numbers only, platelets and neutrophils would probably largely dominate over mast cells and macrophages, whereas mast cells and basophils would likely be the dominant cell types activated through IgE and MRGPRX2-dependent pathways. Considering the high levels of specific IgG necessary for generating immune complexes with their target drug to trigger FcγRs, compared with few specific IgE-bound FcεRI on sensitized mast cells and basophils necessary to trigger their activation (Fig 2), IgE would largely dominate over IgG.[68] These considerations apply even more in anaphylaxis to ingested drugs/antigens that require the compound to reach circulation, because only a small fraction of the ingested compound is absorbed.[69,70] Solving this equation is next to impossible, but informs clinicians of the possibility that 1, 2, or even 3 different pathways, involving different antibody classes (or none), different receptors, and different cells types in circulation and tissue resident, may be at play simultaneously in a drug-induced severe anaphylactic reaction. This next section will propose a clinician's view on drug-induced anaphylaxis, taking NMBA hypersensitivity as an example.

NMBA-induced anaphylaxis exemplflies multiple mechanisms at play

As emphasized above with the animal models, the possible mechanisms leading to anaphylaxis in human begin to be better understood, as more and more actors are evidenced at the cellular or soluble levels that can interact and define complex ***endotypes***. The example we chose to describe in this section in detail is NMBA-induced anaphylaxis during the perioperative period because it may be representative of drug-induced severe anaphylaxis implicating several pathways that synergize to increase severity.

The incidence of perioperative anaphylaxis varies between the geographical locations, with rates of 1 in 10,000 to 20,000 anesthesia procedures. For the 2011-2012 period, in 714 patients who experienced perioperative anaphylaxis in France, the most common cause was NMBA administration (60%).[71] The classical IgE-dependent mechanism that involves basophil and mast cell degranulation has been clearly documented in various studies for several years, mainly using the morphine quaternary ammonium (QA) as a surrogate ***epitope*** of antibody responses to NMBA, even if one can be critical on this laboratory reagent, the only one commercially available so far.[72] NMBA are small molecules that may be considered as haptens and it remains unclear if 1 NMBA molecule can be bound by 2 different antibodies to trigger antibody receptor activation. Interestingly, the historical hapten concept has been recently revisited by its author, Dr Werner J. Pichler, who postulates now that in the minutes following the reexposure to a drug, a massive mast cell degranulation occurs in response to IgE cross-linking by noncovalent drug-carrier complexes called "fake antigens."[73] However, concerning NMBAs, the absence of any sign of IgE-dependent immune activation despite evident clinical anaphylaxis in 10% to 20% of patients led us to test the hypothesis of an IgG-induced neutrophil activation, as we previously described in mouse models.[56]

We prospectively conducted a multicenter study of 86 patients with suspected anaphylaxis to NMBAs during general anesthesia and 86 matched controls (age, sex, drug, type of surgery).[40] We found that circulating anti-QA IgE was undetectable in a large percentage of the patients, whereas anti-QA IgG levels were significantly increased as compared with matched controls. Moreover, both anti-QA IgE and IgG levels correlated with anaphylaxis severity. We then found that downregulation of FcγRs (CD32A, CD16) at the neutrophil surface was also associated with reaction severity, suggesting that anti-QA specific IgG formed immune complexes with NMBA to rapidly activate circulating neutrophils. This was further supported by increased neutrophil expression of CD11b and CD66b, elevated circulating levels of degranulated elastase, and decreased PAF-acetyl hydrolase activity related to PAF secretion. Moreover, high levels of neutrophil extracellular

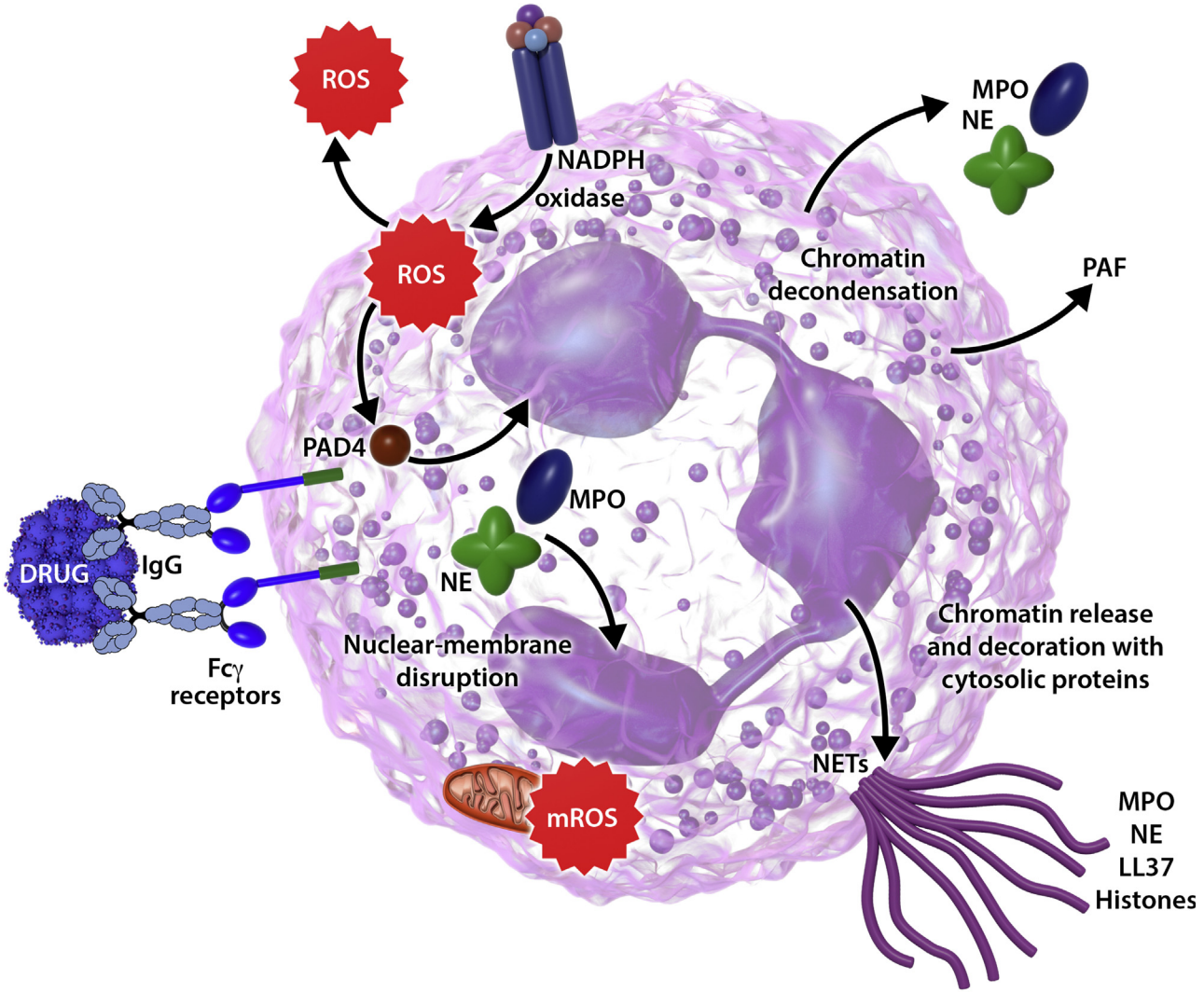

FIG 3. Mechanisms of IgG-induced neutrophil activation during drug anaphylaxis. The classical and historical pathway of anaphylaxis is based on mediator release by mast cells and basophils activated by the engagement of FcεRI after their interaction with a drug/antidrug IgE immune complex (IC). A second pathway was recently demonstrated both in mice and in human. The drug can react with specific IgG and form an IC that binds to several FcγRs at the neutrophil surface and activate the cell. In addition to nicotinamide adenine dinucleotide phosphate oxidase (NADPH oxidase)-induced reactive oxygen species (ROS) and proteases release such as neutrophil elastase (NE) and myeloperoxidase (MPO), neutrophils release PAF and neutrophil extracellular traps (NETs), also involved in anaphylaxis clinical manifestations. The release of NETs is the consequence of ROS production, in particular due to mitochondrial-derived ROS (mROS) production and peptidyl arginase deiminase 4 (PAD4) activation leading to chromatin decondensation, nuclear membrane disruption, and chromatin extracellular release. *LL37*, Active form of cathelicidin antimicrobial peptide.

traps, detected as DNA-myeloperoxidase complexes, were found in severe patients as compared with mild patients and controls. Altogether, using a large panel of neutrophil activation markers, we could confirm that, in humans, an IgG-dependent neutrophil activation occurs during NMBA anaphylaxis with, or independently of, IgE-dependent mast cell/basophil activation (Fig 3).

Supporting anti-NMBA IgG contribution to anaphylaxis and our finding that IgG receptor CD32A-expressing platelets can induce anaphylaxis in animal models, platelet activation in the same patient cohort suffering from NMBA anaphylaxis was associated with anaphylaxis severity and was accompanied by a reduction in circulating platelet numbers.[58] To better document IgG-mediated mechanisms in anaphylaxis, we isolated rocuronium-specific IgG from 1 patient and found that they could form immune complexes with rocuronium that could in turn activate neutrophils isolated from healthy controls, as evidenced by the activation of oxidative burst and neutrophil extracellular trap release (Fig 3). These results reconcile clinical and experimental data on the role of IgE and IgG during anaphylaxis and can modify our biological diagnostic approaches to NMBA-induced anaphylaxis, even if skin tests remain major tools for IgE-mediated reactions. Indeed, we can now suggest to implement the classical biological evaluation of suspected NMBA anaphylaxis[74,75] by exploring both IgE-basophil and IgG-neutrophil pathways.

Exploration of NMBA-induced anaphylaxis for the determination of specific IgEs against QA (as a surrogate epitope of NMBAs) and against each independent NMBA (rocuronium,

atracurium, suxamethonium) can be made using commercial (ImmunoCAP; ThermoFisher, Uppsala, Sweden) or home-made techniques, but the specificity is not optimal and false positivity and numerous cross-reactivities are observed.[76] The calculation of specific to total IgE ratio did not improve the biological diagnosis of rocuronium allergy.[77] Sensitization to NMBAs might originate from exposure to other drugs or compounds that contain also a QA epitope, such as pholcodine (a morphine derivative contained in anticough medications): indeed, antipholcodine IgE can be detected in NMBA-hypersensitive patients.[78] Altogether, these recent studies emphasize the difficulties of correctly quantifying the circulating anti-NMBA IgEs. New methods are needed, and we can assume that the recent luciferase-linked immunosorbent assay that demonstrated 10- to 100-fold better sensitivity than ImmunoCAP for peanut allergen-specific IgE could be adapted to other allergens and drugs including NMBA-specific IgE detection, providing perhaps enhanced sensitivity and specificity.[79] In addition to anti-NMBA specific IgE, we now recommend, using similar techniques, to assay anti-NMBA specific IgG to increase the understanding of our clinical findings reported in 2019.[40]

Assays to detect soluble mediators can be used to invoke mechanisms involved in anaphylaxis. Histamine and tryptase measurements, routinely used to confirm an anaphylactic reaction, reflect the activation of mast cells and, in the case of histamine, basophils. An elevated level of serum tryptase remains one of the very best markers of anaphylaxis. However, the level of tryptase at baseline (after resolution of the anaphylactic reaction) is required to calculate the acute tryptase levels using the following algorithm: tryptase levels are acute if more than [1.2 × baseline tryptase] + 2 μg/L.[80] Baseline levels of tryptase are elevated in patients with mastocytosis (reflecting increased mast cell burden), and are associated with an increased risk of recurrent perioperative anaphylaxis.[81] As noted previously, human mast cells strongly express MRGPRX2 (which is also expressed more weakly by basophils in humans[82]). MRGPRX2 can directly bind a number of drugs leading to mast cell activation and mediator release such as tryptase,[44] meaning that elevated circulating tryptase levels at the initial phase of anaphylaxis can thus be generated by IgE- or MRGPRX2-dependent mast cell activation, or both. Initially, the list of drugs activating MRGPRX2 included NMBAs atracurium and rocuronium among others, but conflicting results have placed this assumption under debate.[83,84] The ability and importance of rocuronium-induced MRGPRX2 activation is under evaluation, investigating effects of MRGPRX2 mutations.[85,86]

Both blood basophils and neutrophils can be studied *ex vivo* in the patients to improve diagnosis. The ***basophil activation test*** is a useful tool to document NMBA anaphylaxis[87] that needs to be performed 4 to 6 weeks after the episode. This flow cytometry–based *ex vivo* assay can be adapted to other NMBAs.[88] The versatility of basophil activation test may make it an increasingly used tool in the diagnosis of NMBA-induced anaphylaxis. In addition, elastase levels (using ELISA) and DNA-MPO levels (markers of increased ***netosis***) may be useful for detecting neutrophil activation during anaphylaxis (Fig 3).[89] We can propose that a simple phenotypic study such as CD11b and CD66b expression, monitored over the course of a clinical reaction in parallel with tryptase, can document a potential specific anti-NMBA IgG-induced neutrophil activation at the time of the reaction in case of the presence of specific IgG.

CONCLUSION AND THERAPEUTIC AVENUES

Drug-induced anaphylaxis is (1) a very severe clinical reaction that needs to be rapidly and extensively documented and diagnosed by adequate biological tools and (2) a complex reaction that can involve various cell types, mediators, receptors, and intracellular pathways that can be activated alone or in association (Fig 2), with NMBA-induced anaphylaxis being an ideal example.

Recent human clinical data proposed even further possible mechanisms in addition to MRGPRX2, exemplified by the role of the contact system via factor XII activation and bradykinin release in some penicillin-induced anaphylaxis,[90] or after heparin injection and severe hypotension due to oversulfated chondroitin sulfate contamination.[91] Explorations outside of basophil activation, that is, neutrophil, platelet, and monocyte activation, and of antidrug IgE, that is, presence of antidrug IgG,[40,41,92] should increase in clinical research to improve our understanding of anaphylaxis and define markers for its endotypes. Because many anaphylactic reactions to drugs happen at first exposure, identifying potential cross-reactivities is of major importance to discourage the use of some drugs in potentially susceptible patients; hypersensitivity to the oligosaccharide alpha-gal as a consequence of tick bites leading to cetuximab anaphylaxis is a good example.[93] In contrast, sensitization to some cereal and peach allergens (lipid transfer protein) is a high-risk factor to nonsteroidal anti-inflammatory drugs anaphylaxis, without any cross-reactivity identified.[94]

The first-line treatment of any type of anaphylaxis, whatever the mechanism, is adrenaline (epinephrine). As far as therapeutic tools are concerned, the avoidance of the drug is the only efficient action when possible. If not possible, anaphylaxis might be prevented by pretreating patients with the anti-IgE antibody omalizumab because it is known to be useful in drug desensitization.[95] Antidrug therapy to prevent IgE engagement might also be considered: allergen desensitization by anticat allergen antibody therapy has indeed been reported already,[96] and might be transposed to drugs.[97] Antidrug therapy to capture the drug remains a poorly explored avenue to remove quickly the culprit drug and thereby arrest the ongoing anaphylactic reaction: attempts have been described in NMBA-induced anaphylaxis[98] due to the existence of a rocuronium and vecuronium capture reagent, sugammadex, but remains debated.[99-101] Unfortunately, a significant number of (IgE-mediated) sugammadex-induced anaphylactic reactions have been described,[102-104] making this particular therapeutic compound nonideal to explore drug capture as a therapy for drug-induced anaphylaxis. Novel antidrug therapies need to be developed to understand the potential of drug capture to reduce anaphylaxis severity or even to stop an ongoing anaphylactic reaction.

REFERENCES

1. Cardona V, Ansotegui IJ, Ebisawa M, El-Gamal Y, Fernandez Rivas M, Fineman S, et al. World Allergy Organization anaphylaxis guidance 2020. World Allergy Organ J 2020;13:100472.
2. Tanno LK, Demoly P. Epidemiology of anaphylaxis. Curr Opin Allergy Clin Immunol 2021;21:168-74.
3. Jerschow E, Lin RY, Scaperotti MM, McGinn AP. Fatal anaphylaxis in the United States, 1999-2010: temporal patterns and demographic associations. J Allergy Clin Immunol 2014;134:1318-28.e7.
4. Turner PJ, Jerschow E, Umasunthar T, Lin R, Campbell DE, Boyle RJ. Fatal anaphylaxis: mortality rate and risk factors. J Allergy Clin Immunol Pract 2017;5:1169-78.

5. Yu RJ, Krantz MS, Phillips EJ, Stone CA Jr. Emerging causes of drug-induced anaphylaxis: a review of anaphylaxis-associated reports in the FDA Adverse Event Reporting System (FAERS). J Allergy Clin Immunol Pract 2021;9:819-29.e2.
6. Kuruvilla M, Khan DA. Anaphylaxis to drugs. Immunol Allergy Clin North Am 2015;35:303-19.
7. Sellaturay P, Nasser S, Ewan P. Polyethylene glycol-induced systemic allergic reactions (anaphylaxis). J Allergy Clin Immunol Pract 2021;9:670-5.
8. Wenande E, Garvey LH. Immediate-type hypersensitivity to polyethylene glycols: a review. Clin Exp Allergy 2016;46:907-22.
9. Savvatianos S, Giavi S, Stefanaki E, Siragakis G, Manousakis E, Papadopoulos NG. Cow's milk allergy as a cause of anaphylaxis to systemic corticosteroids. Allergy 2011;66:983-5.
10. Banerji A, Rudders S, Clark S, Wei W, Long AA, Camargo CA Jr. Retrospective study of drug-induced anaphylaxis treated in the emergency department or hospital: patient characteristics, management, and 1-year follow-up. J Allergy Clin Immunol Pract 2014;2:46-51.
11. Clark S, Wei W, Rudders SA, Camargo CA Jr. Risk factors for severe anaphylaxis in patients receiving anaphylaxis treatment in US emergency departments and hospitals. J Allergy Clin Immunol 2014;134:1125-30.
12. Schuch A, Brockow K. Mastocytosis and anaphylaxis. Immunol Allergy Clin North Am 2017;37:153-64.
13. Bischoff SC. Role of mast cells in allergic and non-allergic immune responses: comparison of human and murine data. Nat Rev Immunol 2007;7:93-104.
14. Vigorito C, Russo P, Picotti GB, Chiariello M, Poto S, Marone G. Cardiovascular effects of histamine infusion in man. J Cardiovasc Pharmacol 1983;5:531-7.
15. Kaliner M, Sigler R, Summers R, Shelhamer JH. Effects of infused histamine: analysis of the effects of H-1 and H-2 histamine receptor antagonists on cardiovascular and pulmonary responses. J Allergy Clin Immunol 1981;68:365-71.
16. Vadas P, Gold M, Perelman B, Liss GM, Lack G, Blyth T, et al. Platelet-activating factor, PAF acetylhydrolase, and severe anaphylaxis. N Engl J Med 2008;358:28-35.
17. Vadas P, Perelman B, Liss G. Platelet-activating factor, histamine, and tryptase levels in human anaphylaxis. J Allergy Clin Immunol 2013;131:144-9.
18. Schwartz LB, Metcalfe DD, Miller JS, Earl H, Sullivan T. Tryptase levels as an indicator of mast-cell activation in systemic anaphylaxis and mastocytosis. N Engl J Med 1987;316:1622-6.
19. Basran GS, Page CP, Paul W, Morley J. Platelet-activating factor: a possible mediator of the dual response to allergen? Clin Allergy 1984;14:75-9.
20. Cuss FM, Dixon CM, Barnes PJ. Effects of inhaled platelet activating factor on pulmonary function and bronchial responsiveness in man. Lancet 1986;2:189-92.
21. Miwa M, Miyake T, Yamanaka T, Sugatani J, Suzuki Y, Sakata S, et al. Characterization of serum platelet-activating factor (PAF) acetylhydrolase. Correlation between deficiency of serum PAF acetylhydrolase and respiratory symptoms in asthmatic children. J Clin Invest 1988;82:1983-91.
22. Pravettoni V, Piantanida M, Primavesi L, Forti S, Pastorello EA. Basal platelet-activating factor acetylhydrolase: prognostic marker of severe Hymenoptera venom anaphylaxis. J Allergy Clin Immunol 2014;133:1218-20.
23. Payne V, Kam PC. Mast cell tryptase: a review of its physiology and clinical significance. Anaesthesia 2004;59:695-703.
24. He S, Gaca MD, Walls AF. A role for tryptase in the activation of human mast cells: modulation of histamine release by tryptase and inhibitors of tryptase. J Pharmacol Exp Ther 1998;286:289-97.
25. Platts-Mills TAE, Schuyler AJ, Erwin EA, Commins SP, Woodfolk JA. IgE in the diagnosis and treatment of allergic disease. J Allergy Clin Immunol 2016;137:1662-70.
26. Ishizaka K, Ishizaka T, Richter M. Effect of reduction and alkylation on allergen-combining properties of reaginic antibody. J Allergy 1966;37:135-44.
27. Carter MC, Robyn JA, Bressler PB, Walker JC, Shapiro GG, Metcalfe DD. Omalizumab for the treatment of unprovoked anaphylaxis in patients with systemic mastocytosis. J Allergy Clin Immunol 2007;119:1550-1.
28. Nadeau KC, Kohli A, Iyengar S, DeKruyff RH, Umetsu DT. Oral immunotherapy and anti-IgE antibody-adjunctive treatment for food allergy. Immunol Allergy Clin North Am 2012;32:111-33.
29. Boni E, Incorvaia C, Mauro M. Dose-dependence of protection from systemic reactions to venom immunotherapy by omalizumab. Clin Mol Allergy 2016;14:14.
30. MacGinnitie AJ, Rachid R, Gragg H, Little SV, Lakin P, Cianferoni A, et al. Omalizumab facilitates rapid oral desensitization for peanut allergy. J Allergy Clin Immunol 2017;139:873-81.e8.
31. Davies JM, Platts-Mills TA, Aalberse RC. The enigma of IgE+ B-cell memory in human subjects. J Allergy Clin Immunol 2013;131:972-6.
32. Looney TJ, Lee JY, Roskin KM, Hoh RA, King J, Glanville J, et al. Human B-cell isotype switching origins of IgE. J Allergy Clin Immunol 2016;137:579-86.e7.
33. Croote D, Darmanis S, Nadeau KC, Quake SR. High-affinity allergen-specific human antibodies cloned from single IgE B cell transcriptomes. Science 2018;362:1306-9.
34. Asrat S, Kaur N, Liu X, Ben LH, Kajimura D, Murphy AJ, et al. Chronic allergen exposure drives accumulation of long-lived IgE plasma cells in the bone marrow, giving rise to serological memory. Sci Immunol 2020;5:eaav8402.
35. Khan F, Hallstrand TS, Geddes MN, Henderson WR Jr, Storek J. Is allergic disease curable or transferable with allogeneic hematopoietic cell transplantation? Blood 2009;113:279-90.
36. Walker SA, Riches PG, Wild G, Ward AM, Shaw PJ, Desai S, et al. Total and allergen-specific IgE in relation to allergic response pattern following bone marrow transplantation. Clin Exp Immunol 1986;66:633-9.
37. Hoh RA, Joshi SA, Lee JY, Martin BA, Varma S, Kwok S, et al. Origins and clonal convergence of gastrointestinal IgE(+) B cells in human peanut allergy. Sci Immunol 2020;5:eaay4209.
38. Kinet JP. The high-affinity IgE receptor (Fc epsilon RI): from physiology to pathology. Annu Rev Immunol 1999;17:931-72.
39. Laroche D, Chollet-Martin S, Leturgie P, Malzac L, Vergnaud MC, Neukirch C, et al. Evaluation of a new routine diagnostic test for immunoglobulin E sensitization to neuromuscular blocking agents. Anesthesiology 2011;114:91-7.
40. Jonsson F, de Chaisemartin L, Granger V, Gouel-Cheron A, Gillis CM, Zhu Q, et al. An IgG-induced neutrophil activation pathway contributes to human drug-induced anaphylaxis. Sci Transl Med 2019;11:eaat1479.
41. Zhou ZH, Stone CA Jr, Jakubovic B, Phillips EJ, Sussman G, Park J, et al. Anti-PEG IgE in anaphylaxis associated with polyethylene glycol [published online ahead of print November 17, 2020]. J Allergy Clin Immunol Pract. https://doi.org/10.1016/j.jaip.2020.11.011.
42. Finkelman FD, Khodoun MV, Strait R. Human IgE-independent systemic anaphylaxis. J Allergy Clin Immunol 2016;137:1674-80.
43. Tatemoto K, Nozaki Y, Tsuda R, Konno S, Tomura K, Furuno M, et al. Immunoglobulin E-independent activation of mast cell is mediated by Mrg receptors. Biochem Biophys Res Commun 2006;349:1322-8.
44. McNeil BD, Pundir P, Meeker S, Han L, Undem BJ, Kulka M, et al. Identification of a mast-cell-specific receptor crucial for pseudo-allergic drug reactions. Nature 2015;519:237-41.
45. Varricchi G, Pecoraro A, Loffredo S, Poto R, Rivellese F, Genovese A, et al. Heterogeneity of human mast cells with respect to MRGPRX2 receptor expression and function. Front Cell Neurosci 2019;13:299.
46. Lagunoff D, Martin TW, Read G. Agents that release histamine from mast cells. Annu Rev Pharmacol Toxicol 1983;23:331-51.
47. Finkelman FD. Anaphylaxis: lessons from mouse models. J Allergy Clin Immunol 2007;120:506-15; quiz 16-7.
48. Gillis CM, Gouel-Chéron A, Bruhns P. Anaphylaxis (immediate hypersensitivity): from old to new mechanisms. In: Compendium of inflammatory diseases. Parnham MJ, ed. Basel: Springer; 2016. Available at: https://doi.org/10.1007/978-3-7643-8550-7_115.
49. Balbino B, Sibilano R, Starkl P, Marichal T, Gaudenzio N, Karasuyama H, et al. Pathways of immediate hypothermia and leukocyte infiltration in an adjuvant-free mouse model of anaphylaxis. J Allergy Clin Immunol 2017;139:584-96.e10.
50. Ishii S, Kuwaki T, Nagase T, Maki K, Tashiro F, Sunaga S, et al. Impaired anaphylactic responses with intact sensitivity to endotoxin in mice lacking a platelet-activating factor receptor. J Exp Med 1998;187:1779-88.
51. Beutier H, Gillis CM, Iannascoli B, Godon O, England P, Sibilano R, et al. IgG subclasses determine pathways of anaphylaxis in mice. J Allergy Clin Immunol 2017;139:269-80.e7.
52. Bruhns P. Properties of mouse and human IgG receptors and their contribution to disease models. Blood 2012;119:5640-9.
53. Strait RT, Posgai MT, Mahler A, Barasa N, Jacob CO, Kohl J, et al. IgG1 protects against renal disease in a mouse model of cryoglobulinaemia. Nature 2015;517:501-4.
54. Mencarelli A, Gunawan M, Yong KSM, Bist P, Tan WWS, Tan SY, et al. A humanized mouse model to study mast cells mediated cutaneous adverse drug reactions. J Leukoc Biol 2020;107:797-807.
55. Reber LL, Hernandez JD, Galli SJ. The pathophysiology of anaphylaxis. J Allergy Clin Immunol 2017;140:335-48.
56. Jönsson F, Mancardi DA, Kita Y, Karasuyama H, Iannascoli B, Van Rooijen N, et al. Mouse and human neutrophils induce anaphylaxis. J Clin Invest 2011;121:1484-96.
57. Gillis CM, Jonsson F, Mancardi DA, Tu N, Beutier H, Van Rooijen N, et al. Mechanisms of anaphylaxis in human low-affinity IgG receptor locus knock-in mice. J Allergy Clin Immunol 2017;139:1253-65.e14.

58. Beutier H, Hechler B, Godon O, Wang Y, Gillis CM, de Chaisemartin L, et al. Platelets expressing IgG receptor FcgammaRIIA/CD32A determine the severity of experimental anaphylaxis. Sci Immunol 2018;3:eaan5997.
59. Jönsson F, Mancardi DA, Zhao W, Kita Y, Iannascoli B, Khun H, et al. Human FcgammaRIIA induces anaphylactic and allergic reactions. Blood 2012;119:2533-44.
60. Khodoun M, Strait R, Orekov T, Hogan S, Karasuyama H, Herbert DR, et al. Peanuts can contribute to anaphylactic shock by activating complement. J Allergy Clin Immunol 2009;123:342-51.
61. Liu E, Moriyama H, Abiru N, Miao D, Yu L, Taylor RM, et al. Anti-peptide autoantibodies and fatal anaphylaxis in NOD mice in response to insulin self-peptides B:9-23 and B:13-23. J Clin Invest 2002;110:1021-7.
62. Arias K, Baig M, Colangelo M, Chu D, Walker T, Goncharova S, et al. Concurrent blockade of platelet-activating factor and histamine prevents life-threatening peanut-induced anaphylactic reactions. J Allergy Clin Immunol 2009;124:307-14.e1-2.
63. Cloutier N, Allaeys I, Marcoux G, Machlus KR, Mailhot B, Zufferey A, et al. Platelets release pathogenic serotonin and return to circulation after immune complex-mediated sequestration. Proc Natl Acad Sci U S A 2018;115:E1550-9.
64. Khodoun MV, Strait R, Armstrong L, Yanase N, Finkelman FD. Identification of markers that distinguish IgE- from IgG-mediated anaphylaxis. Proc Natl Acad Sci U S A 2011;108:12413-8.
65. Kalesnikoff J, Galli SJ. New developments in mast cell biology. Nat Immunol 2008;9:1215-23.
66. Epelman S, Lavine KJ, Randolph GJ. Origin and functions of tissue macrophages. Immunity 2014;41:21-35.
67. Janssens AS, Heide R, den Hollander JC, Mulder PG, Tank B, Oranje AP. Mast cell distribution in normal adult skin. J Clin Pathol 2005;58:285-9.
68. Strait RT, Morris SC, Finkelman FD. IgG-blocking antibodies inhibit IgE-mediated anaphylaxis in vivo through both antigen interception and Fc gamma RIIb cross-linking. J Clin Invest 2006;116:833-41.
69. Strait RT, Mahler A, Hogan S, Khodoun M, Shibuya A, Finkelman FD. Ingested allergens must be absorbed systemically to induce systemic anaphylaxis. J Allergy Clin Immunol 2011;127:982-9.e1.
70. Kucuk ZY, Strait R, Khodoun MV, Mahler A, Hogan S, Finkelman FD. Induction and suppression of allergic diarrhea and systemic anaphylaxis in a murine model of food allergy. J Allergy Clin Immunol 2012;129:1343-8.
71. Tacquard C, Collange O, Gomis P, Malinovsky JM, Petitpain N, Demoly P, et al. Anaesthetic hypersensitivity reactions in France between 2011 and 2012: the 10th GERAP epidemiologic survey. Acta Anaesthesiol Scand 2017;61:290-9.
72. Baldo BA, Fisher MM. Substituted ammonium ions as allergenic determinants in drug allergy. Nature 1983;306:262-4.
73. Pichler WJ. Anaphylaxis to drugs: overcoming mast cell unresponsiveness by fake antigens [published online ahead of print August 11, 2020]. Allergy. https://doi.org/10.1111/all.14554.
74. Ansotegui IJ, Melioli G, Canonica GW, Caraballo L, Villa E, Ebisawa M, et al. IgE allergy diagnostics and other relevant tests in allergy, a World Allergy Organization position paper. World Allergy Organ J 2020;13:100080.
75. Broyles AD, Banerji A, Barmettler S, Biggs CM, Blumenthal K, Brennan PJ, et al. Practical guidance for the evaluation and management of drug hypersensitivity: specific drugs. J Allergy Clin Immunol Pract 2020;8:S16-116.
76. van der Poorten MM, Van Gasse AL, Hagendorens MM, Faber MA, De Puysseleyr L, Elst J, et al. Serum specific IgE antibodies in immediate drug hypersensitivity. Clin Chim Acta 2020;504:119-24.
77. Van Der Poorten MM, Molina-Molina G, Van Gasse AL, Hagendorens MM, Faber MA, De Puysseleyr L, et al. Application of specific-to-total IgE ratio does not benefit diagnostic performance of serologic testing for rocuronium allergy. Br J Anaesth 2020;125:e443-6.
78. Anderson J, Green S, Capon M, Krupowicz B, Li J, Fulton R, et al. Measurement of pholcodine-specific IgE in addition to morphine-specific IgE improves investigation of neuromuscular blocking agent anaphylaxis. Br J Anaesth 2020;125:e450-2.
79. Goyard S, Balbino B, Chinthrajah RS, Lyu SC, Janin YL, Bruhns P, et al. A highly sensitive bioluminescent method for measuring allergen-specific IgE in microliter samples. Allergy 2020;75:2952-6.
80. Vitte J, Amadei L, Gouitaa M, Mezouar S, Zieleskiewicz L, Albanese J, et al. Paired acute-baseline serum tryptase levels in perioperative anaphylaxis: an observational study. Allergy 2019;74:1157-65.
81. Banerji A, Bhattacharya G, Huebner E, Fu X, Camargo CA Jr, Guyer A, et al. Perioperative allergic reactions: allergy assessment and subsequent anesthesia [published online ahead of print November 26, 2020]. J Allergy Clin Immunol Pract. https://doi.org/10.1016/j.jaip.2020.11.025.
82. Elst J, Sabato V, Hagendorens MM, van Houdt M, Faber MA, Bridts CH, et al. Measurement and functional analysis of the Mas-related G protein-coupled receptor MRGPRX2 on human mast cells and basophils. Methods Mol Biol 2020;2163:219-26.
83. Lansu K, Karpiak J, Liu J, Huang XP, McCorvy JD, Kroeze WK, et al. In silico design of novel probes for the atypical opioid receptor MRGPRX2. Nat Chem Biol 2017;13:529-36.
84. Ebo DG, Van der Poorten ML, Elst J, Van Gasse AL, Mertens C, Bridts C, et al. Immunoglobulin E cross-linking or MRGPRX2 activation: clinical insights from rocuronium hypersensitivity. Br J Anaesth 2021;126:e27-9.
85. Elst J, Sabato V, Mertens C, Garvey LH, Ebo DG. Association between mutated Mas-related G protein-coupled receptor-X2 and rocuronium-induced intraoperative anaphylaxis. Comment on Br J Anaesth 2020;125:e446-e448. Br J Anaesth 2020;125:e448-50.
86. Alkanfari I, Gupta K, Jahan T, Ali H. Naturally occurring missense MRGPRX2 variants display loss of function phenotype for mast cell degranulation in response to substance P, hemokinin-1, human beta-defensin-3, and icatibant. J Immunol 2018;201:343-9.
87. Ebo DG, Bridts CH, Mertens CH, Sabato V. Principles, potential, and limitations of ex vivo basophil activation by flow cytometry in allergology: a narrative review [published online ahead of print November 2, 2020]. J Allergy Clin Immunol. https://doi.org/10.1016/j.jaci.2020.10.027.
88. Dewachter P, Chollet-Martin S, Mouton-Faivre C, de Chaisemartin L, Nicaise-Roland P. Comparison of basophil activation test and skin testing performances in NMBA allergy. J Allergy Clin Immunol Pract 2018;6:1681-9.
89. Granger V, Taille C, Roach D, Letuve S, Dupin C, Hamidi F, et al. Circulating neutrophil and eosinophil extracellular traps are markers of severe asthma. Allergy 2020;75:699-702.
90. Gao Y, Han Y, Zhang X, Fei Q, Qi R, Hou R, et al. Penicillin causes non-allergic anaphylaxis by activating the contact system. Sci Rep 2020;10:14160.
91. Hogwood J, Naggi A, Torri G, Page C, Rigsby P, Mulloy B, et al. The effect of increasing the sulfation level of chondroitin sulfate on anticoagulant specific activity and activation of the kinin system. PLoS One 2018;13:e0193482.
92. Daguet A, Watier H. 2nd Charles Richet et Jules Hericourt workshop: therapeutic antibodies and anaphylaxis; May 31-June 1, 2011, Tours, France. MAbs 2011;3:417-21.
93. Steinke JW, Platts-Mills TA, Commins SP. The alpha-gal story: lessons learned from connecting the dots. J Allergy Clin Immunol 2015;135:589-96; quiz 97.
94. Sanchez-Lopez J, Araujo G, Cardona V, Garcia-Moral A, Casas-Saucedo R, Guilarte M, et al. Food-dependent NSAID-induced hypersensitivity (FDNIH) reactions: unraveling the clinical features and risk factors [published online ahead of print December 8, 2020]. Allergy. https://doi.org/10.1111/all.14689.
95. Fernandez J, Ruano-Zaragoza M, Blanca-Lopez N. Omalizumab and other biologics in drug desensitization. Curr Opin Allergy Clin Immunol 2020;20:333-7.
96. Orengo JM, Radin AR, Kamat V, Badithe A, Ben LH, Bennett BL, et al. Treating cat allergy with monoclonal IgG antibodies that bind allergen and prevent IgE engagement. Nat Commun 2018;9:1421.
97. Vultaggio A, Matucci A, Nencini F, Bormioli S, Vivarelli E, Maggi E. Mechanisms of drug desensitization: not only mast cells. Front Pharmacol 2020;11:590991.
98. McDonnell NJ, Pavy TJ, Green LK, Platt PR. Sugammadex in the management of rocuronium-induced anaphylaxis. Br J Anaesth 2010;106:199-201.
99. Leysen J, Bridts CH, De Clerck LS, Ebo DG. Rocuronium-induced anaphylaxis is probably not mitigated by sugammadex: evidence from an in vitro experiment. Anaesthesia 2011;66:526-7.
100. Raft J, Leclercq M, Longrois D, Meistelman C. Fast recovery of haemodynamic and ventilatory functions after sugammadex bolus following rocuronium-induced anaphylactic shock refractory to conventional treatment [in French]. Ann Fr Anesth Reanim 2012;31:158-61.
101. Spoerl D, D'Incau S, Roux-Lombard P, Harr T, Czarnetzki C. Non-IgE-dependent hypersensitivity to rocuronium reversed by sugammadex: report of three cases and hypothesis on the underlying mechanism. Int Arch Allergy Immunol 2016;169:256-62.
102. Savic L, Savic S, Hopkins PM. Anaphylaxis to sugammadex. Anaesth Intensive Care 2014;42:7-9.
103. Ebo DG, Baldo BA, Van Gasse AL, Mertens C, Elst J, Sermeus L, et al. Anaphylaxis to sugammadex-rocuronium inclusion complex: an IgE-mediated reaction due to allergenic changes at the sugammadex primary rim. J Allergy Clin Immunol Pract 2020;8:1410-5.e3.
104. Orihara M, Takazawa T, Horiuchi T, Sakamoto S, Nagumo K, Tomita Y, et al. Comparison of incidence of anaphylaxis between sugammadex and neostigmine: a retrospective multicentre observational study. Br J Anaesth 2020;124:154-63.