

Accepted Manuscript

Metal adsorption on mosses: towards a universal adsorption model

A.G. González, O.S. Pokrovsky

PII: S0021-9797(13)00928-4

DOI: <http://dx.doi.org/10.1016/j.jcis.2013.10.028>

Reference: YJCIS 19149

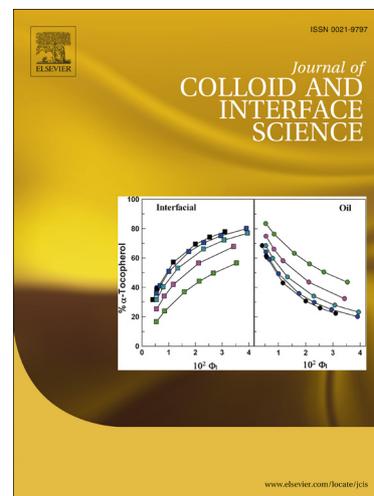
To appear in: *Journal of Colloid and Interface Science*

Received Date: 21 July 2013

Accepted Date: 16 October 2013

Please cite this article as: A.G. González, O.S. Pokrovsky, Metal adsorption on mosses: towards a universal adsorption model, *Journal of Colloid and Interface Science* (2013), doi: <http://dx.doi.org/10.1016/j.jcis.2013.10.028>

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.



Metal adsorption on mosses: towards a universal adsorption model

A.G. González^{1#}, O.S. Pokrovsky^{1,2*}

¹*Geosciences Environment Toulouse (GET), CNRS, UMR 5563, Observatoire Midi-Pyrénées, 14 Avenue Edouard Belin, 31400 Toulouse, France*

²*Institute of Ecological Problems of the Northern Regions, URoRAS, 23 Naberezhnaja Sev. Dviny, Arkhangelsk, Russia*

#First corresponding author. Email: aridaneglez@gmail.com

*Second corresponding author: Tel: +33 561 332 625; Fax: +33 561 332 650

Email: oleg@get.obs-mip.fr

Highlights

- We studied the adsorption of 5 metals on 4 common mosses.
- pH-dependent adsorption edge and constant pH isotherms are similar among mosses.
- *Sphagnum sp.* is the most inert and efficient metal and proton adsorbent.
- A universal adsorption edge of metals on mosses is recommended.
- Mosses are among the most efficient organic adsorbents for heavy metals.

Abstract

This study quantifies the adsorption of heavy metals on 4 typical moss species used for environmental monitoring in the moss bag technique. The adsorption of Cu^{2+} , Cd^{2+} , Ni^{2+} , Pb^{2+} and Zn^{2+} onto *Hypnum sp.*, *Sphagnum sp.*, *P. purum* and *B. rutabulum* has been investigated using a batch reactor in a wide range of pH (1.3-11.0) and metal concentrations in solution (1.6 μM – 3.8 mM). A Linear Programming Model (LPM) was applied for the experimental data to derive equilibrium constants and the number of surface binding sites. The surface acid-base titration performed for 4 mosses at a pH range of 3 to 10 in 0.1 M NaNO_3 demonstrated that *Sphagnum sp.* is the most efficient adsorbent as it has the maximal number of proton-binding sites on the surface (0.65 mmol g^{-1}). The pK_a computed for all the moss species suggested the presence of 5 major functional groups: phosphodiester, carboxyl, phosphoryl, amine and polyphenols. The results of pH edge experiments demonstrated that *B. rutabulum* exhibits the highest percentage of metal adsorption and has the highest number of available sites for most of the metals studied. However, according to the results of the constant-pH “Langmuirian” isotherm, *Sphagnum sp.* can be considered as the strongest adsorbent, although the relative difference from other mosses is within 20%. The LPM was found to satisfactorily fit the experimental data in the full range of the studied solution parameters. The results of this study demonstrate a rather similar pattern of five metal

adsorptions on mosses, both as a function of pH and metal concentration, which is further corroborated by similar values of adsorption constants. Therefore, despite the species and geographic differences between the mosses, a universal adsorption edge and constant pH adsorption isotherm can be recommended for 4 studied mosses. The quantitative comparison of metal adsorption with other common natural organic and inorganic materials demonstrates that mosses are among the most efficient natural adsorbents of heavy metals.

Keywords: Adsorption, metal, moss, pH edge, Langmuirian isotherm

1. INTRODUCTION

Atmospheric pollution constitutes one of the most important environmental problems of human health [1-3]. This is especially true for heavy metal pollutions that enter the food chain via plant uptake and subsequent amplification [4]. To assess the degree of atmospheric contamination by metals, bioindicators have been widely used both in urban and industrial areas. Among various bioindicators, mosses were among the first ones for tracing pollution in Europe [5-6], notably in the industrial areas [7-13]. Their capacity to reflect the chemical composition of the surrounding atmosphere is due to the fact that mosses do not have either cuticle or root and owing to their ectohydric nature, they obtain most elements and nutrients directly from atmospheric deposition [14]. There are several other sorbents that have been tested as pollution monitors but the cost of moss production is low and they have the possibility of reutilization which, together with their high adsorption capacity, gives the moss an extra value [15]. Despite several studies on heavy metal adsorption on mosses [16-17], the detailed physico-chemical mechanism of these important biosorbents operations remains rather limited in contrast to comprehensive models and experimental data on other organic surfaces such as bacteria [18-19], fungi [20], diatoms [21-22] and organic-rich soils [23].

Moss is a phylum of small, soft plants with around 12000 species classified as Bryophyta [24] which inhabit most of the earth. Mosses are unique in the sense that they 1) are able to store water up to 16-26 times dry weight and 2) the phenolic compounds embedded in the mosses' cell walls readily avoid moss decay [25]. Peat moss can also acidify its surroundings by taking up cations such as Ca^{2+} and Mg^{2+} , and releasing H^+ . These characteristics determine the very important role of moss as the interface between atmosphere and hydrosphere/biosphere in metal biogeochemical cycles.

This work presents a concerted study of chemical characterization of four species of common European mosses, *Hypnum sp.*, *Sphagnum sp.*, *Pseudoscleropodium purum* and *Brachytecium rutabulum*, comprising acid/base characterization of the moss surfaces and adsorption of five toxic metals (Cu^{2+} , Cd^{2+} , Ni^{2+} , Pb^{2+} and Zn^{2+}) as a function of pH and metal concentration in solution. Nickel, cadmium and lead are especially hazardous for human health [26], causing cancer and mutations in living organisms [27-32] notably when transported as atmospheric aerosols. Copper has been responsible for neurological disorders in humans [33] and behavioral changes in animals [34]. Over the past decade, significant progress has been achieved in the application of passive biomonitors to assess the level of atmospheric contamination by heavy metals using the moss bag technique [35-36]. Despite the apparent success in using a moss bag for tracing air integral pollution [37-38], fundamental mechanisms controlling heavy metal interaction with the main sorbent, green moss, still remain poorly known and the degree of heavy metal retention by moss biomass cannot be easily predicted.

The present study is therefore aimed at quantifying the first-order physico-chemical parameters of divalent metal adsorption on moss surfaces trying to address the following specific questions:

- 1) What is the most efficient metal adsorbent among 4 selected species that can be recommended for the moss bag biomonitoring procedure?
- 2) What is the chemical nature of the main metal-binding group at the moss surface and how does it vary depending on moss species, the identity and aqueous concentration of metals?
- 3) Can we suggest “universal” thermodynamic adsorption parameters for the prediction of metal adsorption on mosses under a wide range of solution parameters?

Via providing straightforward and quantitative answers to the above-listed questions we create a comprehensive model of chemical reactions between heavy metals and the moss-aqueous solution interface, suitable for a number of environmental applications.

2. EXPERIMENTAL

2.1. *Moss species*

The mosses examined in this research study were the dominant European species: *Hypnum sp.*, *Sphagnum sp.*, *Pseudoscleropodium purum* and *Brachytecium rutabulum*.

They were harvested in June 2012 in NW Spain in non-urban areas. Before the experiments, the whole moss was cleaned three times with Milli-Q water (18 MQ) and inactivated at 120°C following the standard procedure of moss bag preparation [36].

Intact whole mosses without grinding or disaggregation were used throughout the study because the physical and biological status of mosses under investigation should be as

close as possible to that of moss bags envisaged in the environmental exposure conditions. The biomass concentration in the experiments was kept constant at $1 \text{ g}_{\text{dry}}\text{L}^{-1}$.

2.2. Chemicals

The adsorption experiments were carried out at 20°C individually for each metal for Cu^{2+} , Cd^{2+} , Ni^{2+} , Pb^{2+} and Zn^{2+} . All metals were used as nitrate salts (Sigma-Aldrich). The electrolyte solution was NaNO_3 0.01M for all the experiments. All the solutions were prepared with Milli-Q water (18 MQ). The experiments were carried out at constant pH and were buffered by 2.5 mM MES (Merck) for pH 5.5, or HEPES for pH 6.5 (Sigma-Aldrich).

2.3. Surface acid-base titration

The acid-base titration of moss surfaces was carried out in 0.01 M NaNO_3 at room temperature ($20 \pm 1^{\circ}\text{C}$). Solutions were conditioned for 1 h before titration and were also pre-saturated with nitrogen. The titration was performed in two steps, acid titration by adding aliquots of 0.07 M HCl and basic titration by adding small amounts of 0.09 M NaOH. The acid-base titration experiments were done by triplicate for each moss in a whole range of pH between 3 and 11. The reference solution was the supernatant solution after the conditioning time and removing the moss biomass. The pH was measured by a combined electrode (Mettler Toledo^R) in a pH-meter ion analyzer (PHM250-MeterlabTM) with an uncertainty of ± 0.002 units. The excess of charge was computed as the difference of the acid/base concentration in the suspension and in the reference solution according to usual procedures of biomass titration [39-40].

2.4. Adsorption of metals onto moss

The metal adsorption experiments were designed to provide a quantitative physico-chemical characterization of metal binding by moss species as a function of pH (pH-dependent adsorption edge) and as a function of aqueous metal concentration (adsorption isotherm). All the experiments were performed in the solution undersaturated with respect to any metal oxide, hydroxide or carbonate as verified by speciation calculations with the MINTEQA2 computer code and corresponding database [41-42]. Experiments were performed in polypropylene beakers continuously agitated with a suspended Teflon coated magnet stirrer and N₂ bubbling.

In the pH-edge experiments, the initial metal concentration was set at 52 μM, 29 μM, 56 μM, 16 μM and 50 μM for Cu²⁺, Cd²⁺, Ni²⁺, Pb²⁺ and Zn²⁺ respectively, while the pH ranged from 1.28 to 11.02, depending on each metal. The pH was adjusted by adding aliquots of NaOH (0.1-0.01 M) or HNO₃ (0.1-0.01 M). In the second series of experiments, at constant pH (Langmuirian adsorption isotherm), the metal ion concentration ranged as follows: 1.6 μM - 3.8 mM M for Cu²⁺, 2.3 μM - 1.5 mM M for Cd²⁺, 9.1 μM - 3.2 mM M for Ni²⁺, 1.9 μM - 1.0 mM M for Pb²⁺ and 7.0 μM - 2.9 mM M for Zn²⁺. In this case, the pH was kept constant by adding MES (pH ≈ 5.5) for Cu²⁺ and Pb²⁺, or HEPES (pH ≈ 6.5) for Cd²⁺, Ni²⁺ and Zn²⁺.

The adsorption of Cu²⁺ and Zn²⁺ was also studied in a series of kinetics experiments conducted for *Sphagnum sp.*, at constant pH and various metal concentrations in solution, via ranging the exposure time from 5 min to 28 days. These experiments demonstrated the lack of any measurable effect of the exposure time between 5 min and

28 days on the adsorbed metal concentration (see below). As such, the time of contact was 5 min for most of the adsorption experiments.

All sampled solutions were filtered (0.45 μm) and acidified with bidistilled HNO_3 and analyzed for aqueous metal concentration using flame atomic adsorption spectroscopy (Perkin Elmer AAnalyst 400) with an uncertainty of $\pm 2\%$ and a detection limit of 0.05 mg L^{-1} . The concentration of metal for the initial moss biomass was measured by ICP-MS (Agilent 7500 series) with a detection limit of 0.001 $\mu\text{g L}^{-1}$ and precision of $\pm 5\%$.

The Dissolved Organic Carbon (DOC) concentration in solution was monitored for most of the experiments and was analyzed by using a Carbon Total Analyzer (Shimadzu TOC-V_{CSN}) with an uncertainty of 3% and a detection limit of 0.1 mg L^{-1} . Altogether, 170 individual experiments for 5 metals and 4 mosses were performed in this study.

2.5. *Lineal Programming Model (LPM)*

The LPM model was applied for the acid-base surface titration, pH-edge and fixed pH experiments in order to compute the apparent equilibrium constants and the site densities for each individual experiment following the approaches elaborated for bacteria [39-40, 43]. This model is convenient for describing complex 3-D multi-layer systems having both organic components and rigid cell walls [44-46]. The details of the model description are presented in the Electronic Supplementary Material (ESM-1).

3. RESULTS

In order to define the optimal experimental conditions for adsorption experiments, notably the minimal metal concentration in solution and at the moss surface, two types of preliminary experiments were conducted: (1) analysis of bulk metal concentration in non-contaminated mosses, prior to the adsorption experiments, and (2) metal release

from non-contaminated mosses into aqueous solution at the typical condition of adsorption experiments.

3.1. Metal concentration in mosses

The total elementary composition of the studied mosses is listed in the Electronic Supplementary Material (Table ESM-1). It can be seen that Zn^{2+} was the most abundant metal for each moss (27-40 mg kg⁻¹), except for *Sphagnum sp.*, where Pb^{2+} showed the highest concentration (42.9 mg kg⁻¹). According to their elementary composition, the studied mosses can be ranked in the following order: for Cu^{2+} , *B. rutabulum* > *Hypnum sp.* > *P. purum* > *Sphagnum sp.*, for Cd^{2+} , *Sphagnum sp.* > *Hypnum sp.* ≈ *P. purum* ≥ *B. rutabulum*, for Ni^{2+} , *B. rutabulum* > *Hypnum sp.* > *Sphagnum sp.* > *P. purum*, for Pb^{2+} , *Sphagnum sp.* > *Hypnum sp.* > *B. rutabulum* > *P. purum*. Finally, for Zn^{2+} , *P. purum* > *B. rutabulum* > *Sphagnum sp.* > *Hypnum sp.* Based on these results, we defined the minimal charge of moss surface by adsorbed metals as a factor of 2 higher than the metal concentration in intact biomass.

3.2. Metal release by moss

The metal released by the intact biomass into aqueous solution reflects the degree of moss degradability under given experimental conditions. The concentration of released metal therefore defined the minimal threshold of aqueous metal loading in moss adsorption experiments. Below this threshold value, specific for each metal and each moss, the addition of metal in solution for adsorption on selected biomass was considered unwarranted. For the purpose of quantifying this threshold value, the released metal concentration of Cu^{2+} , Cd^{2+} , Ni^{2+} , Pb^{2+} and Zn^{2+} was measured in 0.01 M $NaNO_3$ for a 1 g_{dry}L⁻¹ biomass of each moss, after 1 min, 12 min, 1 hr, 5 hrs and 8.5 hrs of exposure (Table ESM-2). Typically, the released metal concentrations are 2 orders of

magnitude lower than the minimal (starting) metal concentration in our adsorption experiments. As such, the released metals do not interfere with adsorption constant measurements in this study.

According to the released metal ion concentrations, the studied moss species can be ranked as follows: for Cu^{2+} and Pb^{2+} , *B. rutabulum* > *P. purum* > *Sphagnum sp.* \geq *Hypnum sp.*. For Cd^{2+} , *P. purum* > *B. rutabulum* > *Hypnum sp.* \geq *Sphagnum sp.*. For Ni^{2+} , *B. rutabulum* > *Hypnum sp.* > *Sphagnum sp.* \geq *P. purum*. For Zn^{2+} , *P. purum* > *B. rutabulum* > *Sphagnum sp.* \approx *Hypnum sp.*.

Table ESM-2 also shows that the DOC concentration in solution and contacting with mosses increased with time during the first 1.5 hours of experiments, and then stabilized around 38 mg L^{-1} , 21 mg L^{-1} , 60 mg L^{-1} and 61 mg L^{-1} for *Hypnum sp.*, *Sphagnum sp.*, *P. purum* and *B. rutabulum*, respectively. In this regard, *Sphagnum sp.* and *Hypnum sp.* seem to be the most inert and stable moss species in neutral aqueous solutions excreting the lowest amount of DOC.

3.3. Surface acid-base titration

Surface acid-base titration allows to quantify the proton and hydroxyl buffer capacity of mosses in a wide range of pH and thus determine the concentration of amphoteric surface functional groups. These acid-base titrations were performed for a pH range from 3 to 11 as illustrated in Fig. 1. The pH values of the zero net proton adsorption (pH_{PZC}) were equal to 5.01 ± 0.13 (*Hypnum sp.*), 4.64 ± 0.10 (*Sphagnum sp.*), 4.96 ± 0.14 (*P. purum*) and 6.23 ± 0.25 (*B. rutabulum*). These differences in pH_{PZC} can be understood in terms of the different concentrations of surface functional groups of each moss species. *B. rutabulum* showed the highest excess of adsorbed protons (0.21 mmol L^{-1}), whereas *Sphagnum sp.* exhibits the highest negative surface charge. Consequently

Sphagnum sp. may be the most efficient cation adsorbent, as it has the highest number of negatively charged moieties on the surface.

The results of the LPM application for the surface titration experiments are summarized in Table 1. The moss species can be ranked according to the total number of binding sites available on the surface as: *Sphagnum sp.* (0.65 mmol g^{-1}) > *P. purum* (0.55 mmol g^{-1}) > *Hypnum sp.* (0.49 mmol g^{-1}) \geq *B. rutabulum* (0.48 mmol g^{-1}). The values of pK_a obtained from the LPM fit can be tentatively linked to several possible functional groups: phosphodiester ($\text{pK}_a = 3.6\text{-}3.7$), carboxyl ($\text{pK}_a = 4.7\text{-}5.7$), phosphoryl ($\text{pK}_a = 5.9\text{-}7.4$), amine ($\text{pK}_a = 7.7\text{-}9.2$) and polyphenols ($\text{pK}_a = 10.1\text{-}10.4$), present in all of the four moss species.

3.4. Long-term adsorption of metals

The adsorption of Cu^{2+} and Zn^{2+} was studied as a function of the metal ion concentration in solution ($0.04\text{-}2.6 \text{ mM}$) over different exposure periods, from 5 min. to 28 days (Fig. 2). The result of these experiments allows us to choose the optimal exposure time for adsorption of metals on moss. It can be seen from Fig. 2 that the adsorption of Cu^{2+} and Zn^{2+} achieved the maximum during the first 5 min. The adsorption of Cu^{2+} and Zn^{2+} was similar at different exposure times, thus strongly suggesting the achievement of an adsorption equilibrium during the first several minutes of reaction.

3.5. Adsorption of metals as a function of pH (pH-edge)

The adsorption of Cu^{2+} , Cd^{2+} , Ni^{2+} , Pb^{2+} and Zn^{2+} on moss was studied as a function of pH that ranged from 1.8 to 6.5, 1.3 to 11.0, 1.9 to 10.5, 1.3 to 7.4, and 1.8 to 10.1, for Cu^{2+} , Cd^{2+} , Ni^{2+} , Pb^{2+} and Zn^{2+} , respectively (Fig. 3). The pH-dependent adsorption edge is rather similar among all mosses, in the first order agreement with the principal

of “universal adsorption edge” developed earlier for heterotrophic bacteria and their consortia [19, 39, 47-48].

The adsorption of metals typically starts at pH around 2 and the maximum adsorption percentage is achieved at pH > 6 depending on the identity of the metal. For Cu²⁺, the adsorption was 82% at pH 6.1 for *Sphagnum sp.* whereas the highest percentage of adsorption was reached for *P. purum* (92%) at pH 5.2. For Cd²⁺, the maximum adsorption was reached at pH 8.7 (91%) for *Sphagnum sp.* The lowest adsorption of Cd²⁺ percentage, 79%, was at pH = 8.8 for *P. purum*. For Ni²⁺, *Sphagnum sp.* also had the highest adsorption capacity with 70% at pH = 7.2. The adsorption of Pb²⁺ was quite similar among different species (around 97% at pH = 5.5). Finally, Zn²⁺ exhibited the lowest maximal adsorption of 73% at pH = 7.8 on *Sphagnum sp.*.

Based on the results of the pH-dependent adsorption edge, the mosses investigated in this study can be ranked as following: for Cu²⁺, *P. purum* > *B. rutabulum* ≥ *Hypnum sp.* > *Sphagnum sp.*, for Cd²⁺, *B. rutabulum* > *Sphagnum sp.* ≥ *P. purum* > *Hypnum sp.*, for Ni²⁺, *B. rutabulum* > *P. purum* ≈ *Hypnum sp.* ≈ *Sphagnum sp.*, for Pb²⁺, *B. rutabulum* ≈ *Sphagnum sp.* ≈ *Hypnum sp.* ≈ *P. purum* and finally for Zn²⁺, *B. rutabulum* ≥ *Sphagnum sp.* ≥ *Hypnum sp.* ≥ *P. purum*.

A plot of DOC concentration as a function of pH during metal adsorption experiments demonstrated a slight increase of [DOC] with pH (Fig. 4). Among 4 studied mosses, *P. purum* and *B. rutabulum* are the most reactive species, excreting ≥ 2 times more DOC compared to *Sphagnum sp.* and *Hypnum sp.* Therefore, *Sphagnum sp.* and *Hypnum sp.* are the most inert species in terms of biomass degradation and organic carbon leaching. This conclusion is consistent with results of metal release from the moss biomass (see section 3.2).

The LPM model was applied for the experimental data on the pH-dependent adsorption edge of divalent metals examined in this study (Table 2). The smallest pK_s corresponding to the strongest binding were found for *Sphagnum sp.*, -3.15 and -4.40 for Cu^{2+} and Cd^{2+} respectively. For Ni^{2+} , *Hypnum sp.* yielded pK_s of -3.50, and for Pb^{2+} and Zn^{2+} , *B. rutabulum* showed the strongest binding pK_s , -3.20 and -0.65, respectively. The number of surface binding sites capable of adsorbing cationic metals was computed to be the highest for *P. purum* ($4.0 \cdot 10^{-2} \text{ mmol g}^{-1}$) in the presence of Cu^{2+} . *Sphagnum sp.* contained $1.8 \cdot 10^{-2} \text{ mmol g}^{-1}$ for Cd^{2+} adsorption experiments, whereas *B. rutabulum* exhibited the highest number of sites for Ni^{2+} , Pb^{2+} and Zn^{2+} with $3.2 \cdot 10^{-2}$, $1.3 \cdot 10^{-2}$ and $2.0 \cdot 10^{-2} \text{ mmol g}^{-1}$, respectively.

3.6. Adsorption of metals as a function of metal concentration in solution (“Langmuirian” isotherm)

The adsorption of Cu^{2+} , Cd^{2+} , Ni^{2+} , Pb^{2+} and Zn^{2+} on *Hypnum sp.*, *Sphagnum sp.*, *P. purum* and *B. rutabulum* was studied at constant pH 5.5, 6.5, 5.6, 6.5 and 6.8 respectively, in the range of 1.6 μM -3.8 mM (Cu^{2+}), 2.3 μM -1.5 mM (Cd^{2+}), 9.1 μM -3.2 mM (Ni^{2+}), 1.9 μM -1.0 mM (Pb^{2+}) and 7.0 μM -2.9 mM (Zn^{2+}) as shown in Table 3 and Fig. 5. The adsorption curve is rather similar among all four mosses and depends significantly on the identity of the metal. The concentration of adsorbed metal increased linearly with $[\text{Me}^{2+}]_{\text{aq}}$ until 0.5 mM. Above this concentration, Cu^{2+} , Cd^{2+} , Ni^{2+} , and, in a lesser degree, Pb^{2+} , demonstrated the beginning of surface sites saturation.

The Langmuirian adsorption isotherm describes a large number of adsorption experiments on biosorbents [49]. It was used to rationalize the adsorption data according to:

$$\frac{[Me^{2+}]_{aq}}{[Me^{2+}]_{ads}} = \frac{1}{K_L q_{max}} + \frac{[Me^{2+}]_{aq}}{q_{max}} \quad (1)$$

where K_L is the Langmuir equilibrium (g mmol^{-1}) constant and q_{max} is the maximum adsorption capacity (mmol g^{-1}). This equation provided an adequate fit to the data with $R^2 > 0.98$; obtained Langmuirian parameters are listed in Table 4.

The highest value of K_L was obtained for Ni^{2+} adsorption ($7.1\text{-}11.4 \text{ g mmol}^{-1}$) while the K_L for adsorption of Cu^{2+} , Pb^{2+} and Zn^{2+} ranged between 0.3 and 1.0 g mmol^{-1} . The maximum adsorption capacity (q_{max}) was reached for Pb^{2+} on *B. rutabulum* (2.6 mmol g^{-1}). An integral parameter of metal adsorption on mosses is defined as the sum of all 4 metals q_{max} value which follows the order: *B. rutabulum* (4.9 mmol g^{-1}) > *Sphagnum sp.* (4.2 mmol g^{-1}) > *Hypnum sp.* (3.6 mmol g^{-1}) > *P. purum* (3.5 mmol g^{-1}).

Individually for each metal, the moss adsorption capacity can be ranked as following: For Cu^{2+} , *P. purum* \geq *Sphagnum sp.* > *Hypnum sp.* \geq *B. rutabulum*; for Cd^{2+} , *Sphagnum sp.* > *B. rutabulum* \geq *Hypnum sp.* > *P. purum*; for Ni^{2+} , *Sphagnum sp.* \geq *Hypnum sp.* \geq *B. rutabulum* \geq *P. purum*; for Pb^{2+} , *B. rutabulum* > *Hypnum sp.* > *Sphagnum sp.* > *P. purum*; for Zn^{2+} , *B. rutabulum* \geq *Sphagnum sp.* \approx *P. purum* \geq *Hypnum sp.*

Overall, *Sphagnum sp.* can be considered to be among the strongest adsorbents, although the relative differences to other mosses are within 20%.

All the data collected for Langmuir isotherm experiments were used to apply the LPM model and the results are shown in Table 3. The LPM revealed small but statistically significant differences in metal binding by different species. In particular, *Hypnum* exhibited the highest amount of binding sites for Cd^{2+} (40 mmol g^{-1}), Pb^{2+} (50 mmol g^{-1}) and Ni^{2+} exposure (8 mmol g^{-1}), and *Sphagnum sp.* exhibited the maximal amount of binding sites for Zn^{2+} (29 mmol g^{-1}) among 4 moss species.

4. DISCUSSION

4.1. Moss chemical composition and degradation in aqueous solution

The total chemical composition of the four moss species examined in this research study is in general agreement with results reported for other moss species [50-52]. In particular, Castello [52] reported the chemical composition of *H. cupressiforme* and *P. purum*. while for both *P. purum* the concentration of Al, Cu and Zn are in the same order; As, Cd, Cr, Fe, Pb concentrations are lower in the present study and Mn and Ti concentrations are higher in this study compared to Castello [52]. Overall, all studied species are enriched in Al, As, Fe, Mn, and Ti and *Sphagnum sp.* and *B. rutabulum* are enriched in Pb and Cr, respectively, compared to the literature data.

The differences in composition between mosses could be understood in terms of morphologic specificity and growth rates. The smaller species are able to form compact communities and more crowded leaves allow them to reach high efficiency to bind metals [50]. In terms of growth rate, the species with lower rates has a longer time to bind metals and increases the concentration of several metals inside the cells or cell walls [53]. Eventually, the difference in composition can also be explained by a different capacity to cation exchange, because of the differences in the chemical composition of the membranes and cell walls [54].

The concentrations of released metals during moss interaction with aqueous solution were significantly lower than those used in adsorption experiments. During the 9 hours of solution exposure experiments, *Sphagnum sp.* and *Hypnum sp.* proved to be the most inert species in terms of both DOC and metal release, which is certainly linked to the specificity of their cell wall chemical composition as described below.

4.2. Acid-base properties of mosses

The amphoteric properties of the moss stem from acid/base dissociation of protonated organic moieties on the surface of the cell wall. The acid-base titration showed that *Sphagnum sp.* exhibits the highest excess of negative charges corresponding to its highest capacity for metal adsorption. The acid-base titration of 4 mosses demonstrated a certain variability of pK_a among mosses likely linked to the different composition of their cell walls. In this study, *Hypnum sp.*, *Sphagnum sp.*, *P. purum* and *B. rutabulum* showed pK_a values ~ 4 , 4.5-5.75, ~ 6 -7.35, 8-9.15 and ~ 10 . These pK_a can be tentatively related with carboxyl/phosphodiester, carboxyl, phosphoryl, amine and polyphenol functional groups. *Sphagnum sp.* contains the highest amount of total binding sites, followed by *P. purum* (15% smaller), *Hypnum sp.* (25% smaller) and *B. rutabulum* (25% smaller). It is important to note that *Sphagnum sp.* exhibits the dominance of carboxyl, phosphoryl, amine functional groups, the main metal-binding moieties on the biological surfaces [19, 48]. As such, *Sphagnum sp.* is the most efficient metal adsorbent given the carboxyl and phosphoryl groups are the primary metal-binding groups at a high concentration of metals [55-56] whereas the sulphhydryl and amine groups can be determinant especially under extreme pH conditions and low metal concentrations [57]. The pK_a computed reported for different microorganisms are around ~ 3 , 4-5, 6-7 and 9-10 with total binding sites around 0.044-0.113 mmol g^{-1} of bacteria [39, 48, 57-58]. The relative percentage of functional groups for the different microorganisms inferred from surface titration (Fig. 6) showed that mosses possess a relatively higher percentage of carboxyl/phosphodiester, amine/polyphenol groups compared to bacteria [48, 59] and cyanobacteria [43, 60], whereas the number of carboxyl groups on mosses is smaller compared to bacteria.

Lignin and cellulose represent the main organic composition of the mosses cell walls [61]. These polysaccharides contain alcohols, aldehydes, ketones, acids, phenolic and hydroxides as the main functional groups. Accordingly, the carboxylic and phenolic groups have been suggested to be responsible for the adsorption of metals on peat moss [62], similar to humic and fulvic acids [63].

4.3. *Metal adsorption on mosses*

All 4 studied mosses demonstrated very fast adsorption kinetics as the equilibrium or steady-state metal concentration in solution in contact with devitalized biomass is achieved within several minutes of reaction and remains constant over almost a month of exposure. This corroborated numerous previous observations on other biological surfaces on the fast equilibrium adsorption of divalent metals [64-66] and on organic-rich abiotic surfaces such as soils [67].

Heavy metals can be bound to most surface layers of cell wall through cation exchange, assimilated within the cells for cellular metabolism or distributed within the porous matrix of the surface layer. Regardless of the nature of final, biologically-active metal compartment in the cells, reversible adsorption on the cell surface represents the first and often limiting step of metal uptake by the microorganisms. Increasing the pH in solution leads to deprotonation of available surface sites that become therefore available to complex metals. The adsorption of metal on moss as a function of pH allowed us to rank the mosses according to their adsorption capacity for each metal. Considering the pH-edge adsorption, *B. rutabulum* seems to be the most efficient species because it reaches the highest percentage of adsorption and has the highest number of available

sites for almost all the metals studied. However, high DOC concentration released by *B. rutabulum* during its interaction with aqueous solution suggests its high instability in water and precludes its use as a biomonitor. In contrast, *Sphagnum sp.* seems to be the most promising species as a potential bioindicator, because it releases a relatively small amount of DOC and it is capable of efficiently binding metals in the full range of pH investigated as follows from its maximal adsorption capacity (q_{\max}).

The number of major binding sites determined for mosses is equal to 34.8 mmol g⁻¹ for Cu²⁺, 40.3 mmol g⁻¹ for Cd²⁺, 7.7 mmol g⁻¹ for Ni²⁺, 50.4 mmol g⁻¹ for Pb²⁺ and 28.6 mmol g⁻¹ for Zn²⁺. These values are significantly higher than those reported for other microorganisms which typically rank between 0.1 and 4 mmol g⁻¹ of dry biomass, for aerobic soil bacteria *Pseudomonas aureofaciens* [39], heterotrophic bacteria [46, 68], bacterial consortia [48], marine diatoms and freshwater species [69] and cyanobacteria [43].

The q_{\max} obtained for 4 mosses was compared with that for aquatic plants [70-73], yeast [74], herbaceous peat [26, 75], Sphagnum peat [26], fungus [20, 76], bacteria [77-78], plants [79], algae [80-81] and soil [82]. The comparison is presented in Fig. 7. It can be clearly seen that the q_{\max} value is the highest for mosses compared to all other studied organic surfaces, especially for Cu²⁺ and Pb²⁺ adsorption. Aquatic plants were the second group with highest adsorption capacities, reaching the maximum adsorption capacity in *P. luteus* with q_{\max} = 0.496 mmol g⁻¹ for Zn²⁺ and 0.642 mmol g⁻¹ for Cu²⁺ [70]. Herbaceous peat also demonstrated significant adsorption capacities, although much lower than the mosses. Overall, the mosses examined in this study exhibit one of the highest adsorption capacity among all known biological sorbents and as such can be efficient candidates for the environmental biomonitoring application.

4.4. Universal adsorption parameters

The adsorption of metals on mosses as a function of pH follows a universal adsorption pattern that is well comparable with other organic materials. In Fig. 3 we presented a pH-dependent adsorption edge for bacteria consortia and individual bacteria species obtained at experimental conditions very similar to those used in the present study. In full accord with numerous previous observations [40, 43, 47-48, 64, 83], the existence of “universal metal (Cu^{2+} , Cd^{2+} , Ni^{2+} , Pb^{2+} , Zn^{2+}) adsorption edge” both for bacteria and bryophyte can be concluded. In contrast to former studies, dealing only with Cd^{2+} for establishing this “universal edge”, this study extends to 4 other important divalent metals, all of them with a similar universal dependence on the percentage of adsorption on pH. This finding should certainly facilitate quantitative modeling of metal interaction with biosorbents under various environmental conditions.

The universal adsorption edge, consisting in similarity of the pH-dependent adsorption edge on various biosorbents for each metal, likely stem from the dominance of carboxylates and phosphorylates as the main binding sites for metal complexation at the moss surface. The same binding sites are most frequently reported on bacteria [39-40, 43, 48, 64], periphytic biofilms [67] and diatoms [22]. Consequently, regardless of the biological nature of the sample and the bacteria/plankton/plant kingdom, the adsorption curve remains rather similar reflecting the dominance of main metal-binding moieties. However, given their 1) large surface area, 2) high stability of devitalized mosses in aqueous solution, notably *Sphagnum sp.*, and 3) low cost of natural or artificially grown (notably cloned) moss species, devitalized moss should remain by far the best biomonitor that can be used for passive adsorption of divalent metals.

5. CONCLUSIONS

The interaction of 5 heavy metals (Cu^{2+} , Cd^{2+} , Ni^{2+} , Pb^{2+} and Zn^{2+}) with 4 mosses (*Hypnum sp.*, *Sphagnum sp.*, *P. purum* and *B. rutabulum*) demonstrated their potential use as bioindicator of atmospheric pollution. Via combining thorough solid and solution analyses, including surface acid/base titration, pH-dependent adsorption edge and “Langmuirian” adsorption at constant pH and variable metal concentration, we conclude that *Sphagnum sp.* exhibits the highest proton and metal adsorption capacity while being most stable in aqueous solution in terms of DOC release and biomass degradation. Compared with other biosorbents, mosses possess significantly higher concentration of surface groups capable to bind divalent metals at the cell surface.

Acknowledgments

This study received financial support from the MOSSclone project by the European Union in the Seventh Framework Programme (FP7) for Research and Technological Development. Additional support from the Russian Ministry of Science and Education and Tomsk State University (Mega-grant BIO-GEO-CLIM No 14.B25.31.0001) is also acknowledged. We also thank Katrin Meier for the English revision of the manuscript.

REFERENCES

- [1] N. Bernard, M. Saintot, C. Astre, M. Gerber, Personal exposure to nitrogen dioxide pollution and effect on plasma antioxidants, *Archives of Environmental Health: An International Journal* 53 (2) (1998) 122–128.
- [2] M. Saintot, N. Bernard, C. Astre, M. Gerber, Inserm-crlc, Ozone exposure and blood antioxidants: a study in a periurban area in Southern France, *Archives of Environmental Health: An International Journal* 54 (1) (1999) 34–39.
- [3] M. Saintot, N. Bernard, C. Astre, P. Galan, S. Hercberg, M. Gerber, Nitrogen dioxide and ozone exposures in a population sample from ile-de-france], *Revue d'épidémiologie et de santé publique* 48 (2000) 2S54.

- [4] M. J. McLaughlin, D. Parker, J. Clarke, Metals and micronutrients–food safety issues, *Field crops research* 60 (1) (1999) 143–163.
- [5] A. Brinkman, Hepatics and sites: A short study in the ecology of hepatics, *Bryol.* 32 (1929) 29–31.
- [6] O. Gilbert, AirPollution-Proc.-1st Eur. Cong.- Inflow Air Pollution Plants and Animals., Wageningen, 1969, Ch. The effects of SO₂ on lichens and bryophytes around Newcastle-upon-Tyne, pp. 223–235.
- [7] B. Whitton, P. Say, J. Wehr, Use of plants to monitor heavy metals in rivers, *Heavy Metals in Northern England: Environmental and Biological Aspects*. Department of Botany, University of Durham, England (1981) 135–145.
- [8] C. Mouvet, Accumulation of chromium and copper by the aquatic moss *Fontinalis antipyretica* L. ex Hedw transplanted in a metal-contaminated river, *Environ. Technol.* 5 (12) (1984) 541–548.
- [9] C. Mouvet, Use of aquatic bryophytes to monitor heavy metals pollution of freshwaters as illustrated by case studies, *Verhandlung Internationale Vereinigung Limnologie* 22 (4).
- [10] M. Kelly, C. Girton, B. Whitton, Use of moss-bags for monitoring heavy metals in rivers, *Water Res.* 21 (11) (1987) 1429–1435.
- [11] E. P. Gonçalves, R. A. Boaventura, C. Mouvet, Sediments and aquatic mosses as pollution indicators for heavy metals in the Ave river basin (Portugal), *Science of the total environment* 114 (1992) 7–24.
- [12] E. P. Gonçalves, H. M. Soares, R. A. Boaventura, A. A. Machado, J. C. Esteves da Silva, Seasonal variations of heavy metals in sediments and aquatic mosses from the Cávado river basin (Portugal), *Science of the total environment* 142 (3) (1994) 143–156.
- [13] J. M. Glime, Economic and ethnic uses of bryophytes, *Flora of North America* 27 (2007) 14–41.
- [14] J. Fabure, C. Meyer, F. Denayer, A. Gaudry, D. Gilbert, N. Bernard, Accumulation capacities of particulate matter in an acrocarpous and a pleurocarpous moss exposed at three differently polluted sites (industrial, urban and rural), *Water, Air, & Soil Pollution* 212 (1-4) (2010) 205–217.
- [15] N. Abdel-Jabbar, S. Al-Asheh, B. Hader, Modeling, parametric estimation, and sensitivity analysis for copper adsorption with moss packed-bed, *Separ. Sci. Technol.* 36 (13) (2001) 2811–2833.
- [16] Y. Ho, D. J. Wase, C. Forster, Batch nickel removal from aqueous solution by sphagnum moss peat, *Water Res.* 29 (5) (1995) 1327 – 1332.
- [17] S. Al-Asheh, Z. Duvnjak, Adsorption of metal ions by moss, *Adv. Environ. Res.* 2 (1997) 194–212.
- [18] J. B. Fein, C. J. Daughney, N. Yee, T. Davis, A chemical equilibrium model of metal adsorption onto bacterial surfaces, *Geochim. Cosmochim. Acta* 61 (1997) 3319–3328.

- [19] J. B. Fein, A. M. Martin, P. G. Wightman, Metal adsorption onto bacterial surfaces: development of a predictive approach, *Geochim. Cosmochim. Acta.* 65 (23) (2001) 4267–4273.
- [20] E. Fourest, J.-C. Roux, Heavy metal biosorption by fungal mycelial by-products: mechanisms and influence of pH, *Appl. Microbiol. Biot.* 37 (3) (1992) 399–403.
- [21] M. Gonzalez-Davila, J. M. Santana-Casiano, J. Perez-Pena, F. J. Millero, Binding of Cu (II) to the surface and exudates of the alga *Dunaliella tertiolecta* in seawater, *Environ. Sci. Technol.* 29 (2) (1995) 289–301.
- [22] A. Gélabert, O. Pokrovsky, J. Schott, A. Boudou, A. Feurtet-Mazel, J. Mielczarski, E. Mielczarski, N. Mesmer-Dudons, O. Spalla, Study of diatoms/aqueous solution interface. I. acid-base equilibria and spectroscopic observation of freshwater and marine species, *Geochim. Cosmochim. Acta.* 68 (20) (2004) 4039–4058.
- [23] O. Pokrovsky, G. Pokrovski, L. Shirokova, A. Gonzalez, E. Emnova, A. Feurtet Mazel, Chemical and structural status of copper associated with oxygenic and anoxygenic phototrophs and heterotrophs: possible evolutionary consequences, *Geobiology* 10 (2012) 130–149.
- [24] B. Goffinet, W. Buck, Systematics of the Bryophyta (Mosses): From molecules to a revised classification, *Monographs in Systematic Botany. Molecular Systematic of Bryophytes* 98 (2004) 205–239.
- [25] M. Hübers, H. Kerp, Oldest known mosses discovered in Mississippian (late Viséan) strata of Germany, *Geology* 40 (8) (2012) 755–758.
- [26] R. Gündogan, B. Acemioglu, M. H. Alma, Copper (II) adsorption from aqueous solution by herbaceous peat, *J. Colloid. Interf. Sci.* 269 (2) (2004) 303–309.
- [27] E. Rojas, L. A. Herrera, L. A. Poirier, P. Ostrosky-Wegman, Are metals dietary carcinogens?, *Mutation Research/Genetic Toxicology and Environmental Mutagenesis* 443 (1–2) (1999) 157 – 181.
- [28] F. Calevro, S. Campani, C. Filippi, R. Batistoni, P. Deri, S. Bucci, M. Raghianti, G. Mancino, Bioassays for testing effects of Al, Cr and Cd using development in the amphibian *Pleurodeles waltl* and regeneration in the planarian *Dugesia etrusca*, *Aquatic Ecosystem Health and Management* 2 (3) (1999) 281 – 288.
- [29] J. Blasiak, J. Kowalik, A comparison of the in vitro genotoxicity of tri- and hexavalent chromium, *Mutation Research/Genetic Toxicology and Environmental Mutagenesis* 469 (1) (2000) 135 – 145.
- [30] L. Amor, C. Kennes, M. Veiga, Kinetics of inhibition in the biodegradation of monoaromatic hydrocarbons in presence of heavy metals, *Bioresour. Technol.* 78 (2) (2001) 181 – 185.
- [31] S. Kawanishi, S. Inoue, S. Oikawa, N. Yamashita, S. Toyokuni, M. Kawanishi, K. Nishino, Oxidative {DNA} damage in cultured cells and rat lungs by carcinogenic nickel compounds, *Free Radical Biology and Medicine* 31 (1) (2001) 108 – 116.
- [32] S. Monni, C. Uhlig, E. Hansen, E. Magel, Ecophysiological responses of *Empetrum nigrum* to heavy metal pollution, *Environ. Pollut.* 112 (2) (2001) 121 – 129.

- [33] D. Strausak, J. F. Mercer, H. H. Dieter, W. Stremmel, G. Multhaupt, Copper in disorders with neurological symptoms: Alzheimer's, menkes, and wilson diseases, *Brain Research Bulletin* 55 (2) (2001) 175 – 185.
- [34] H. Lefcort, E. Ammann, S. Eiger, Antipredatory behavior as an index of heavy-metal pollution? A test using snails and caddisflies, *Archives of Environmental Contamination and Toxicology* 38 (3) (2000) 311–316.
- [35] F. De Nicola, F. Murena, M. A. Costagliola, A. Alfani, D. Baldantoni, M. V. Prati, L. Sessa, V. Spagnuolo, S. Giordano, A multi-approach monitoring of particulate matter, metals and PAHs in an urban street canyon, *Environ. Sci. Pollut. Res.* 20 (7) (2013) 1–11.
- [36] S. Giordano, P. Adamo, V. Spagnuolo, M. Tretiach, R. Bargagli, Accumulation of airborne trace elements in mosses, lichens and synthetic materials exposed at urban monitoring stations: Towards a harmonisation of the moss-bag technique, *Chemosphere*. 90 (2) (2013) 292-299.
- [37] P. Adamo, S. Giordano, S. Vingiani, R. Castaldo Cobianchi, P. Violante, Trace element accumulation by moss and lichen exposed in bags in the city of Naples (Italy), *Environ. Pollut.* 122 (1) (2003) 91–103.
- [38] P. Adamo, P. Crisafulli, S. Giordano, V. Minganti, P. Modenesi, F. Monaci, E. Pittao, M. Tretiach, R. Bargagli, Lichen and moss bags as monitoring devices in urban areas. Part II: Trace element content in living and dead biomonitors and comparison with synthetic materials, *Environ. Pollut.* 146 (2) (2007) 392–399.
- [39] A. González, L. Shirokova, O. Pokrovsky, E. Emnova, R. Martínez, J. Santana Casiano, M. González-Dávila, G. Pokrovski, Adsorption of copper on *Pseudomonas aureofaciens*: Protective role of surface exopolysaccharides, *J. Colloid. Interf. Sci.* 350 (1) (2010) 305–314.
- [40] O. S. Pokrovsky, R. E. Martínez, E. I. Kompantseva, L. S. Shirokova, Interaction of metals and protons with anoxygenic phototrophic bacteria *Rhodobacter blasticus*, *Chemical Geology* 335 (2013) 75–86.
- [41] J. D. Allison, D. S. Brown, J. Kevin, MINTEQA2/PRODEFA2, a geochemical assessment model for environmental systems: Version 3.0 user's manual, Environmental Research Laboratory, Office of Research and Development, US Environmental Protection Agency Athens, Georgia, USA, 1991.
- [42] A. Martell, R. Smith, R. Motekaitis, Critically selected stability constants of metal complexes database, NIST Standard Reference Database 46.
- [43] O. S. Pokrovsky, R. E. Martínez, S. V. Golubev, E. I. Kompantseva, L. S. Shirokova, Adsorption of metals and protons on *Gloeocapsa sp.* cyanobacteria: A surface speciation approach, *Appl. Geochem.* 23 (9) (2008) 2574–2588.
- [44] J. S. Cox, S. Smith, L. A. Warren, F. G. Ferris, Characterizing heterogeneous bacterial surface functional groups using discrete affinity spectra for proton binding, *Environ. Sci. Technol.* 33 (1999) 4514–4521.
- [45] I. Sokolov, D. Smith, G. Henderson, Y. Gorby, F. Ferris, Cell surface electrochemical heterogeneity of the Fe (III)-reducing bacteria *Shewanella putrefaciens*, *Environ. Sci. Technol.* 35 (2) (2001) 341–347.

- [46] R. E. Martinez, D. S. Smith, E. Kulczycki, F. G. Ferris, Determination of intrinsic bacterial surface acidity constants using a Donnan shell model and a continuous pK_a distribution method, *J. Colloid. Interf. Sci.* 253 (1) (2002) 130–139.
- [47] N. Yee, J. Fein, Cd adsorption onto bacterial surfaces: A universal adsorption edge?, *Geochim. Cosmochim. Acta.* 65 (13) (2001) 2037–2042.
- [48] D. Borrok, J. B. Fein, Distribution of protons and Cd between bacterial surfaces and dissolved humic substances determined through chemical equilibrium modelling, *Geochim. Cosmochim. Acta* 68 (2004) 3043–3052.
- [49] J. Febrianto, A. Kosasih, J. Sunarso, J. Yi-Hsu, Equilibrium and kinetic studies in adsorption of heavy metals using biosorbent, *J. Hazard. Mater.* 162 (2009) 616–645.
- [50] R. Bargagli, E. Battisti, E. Cardaioli, P. Formichi, L. Nelli, La deposizione atmosferica di elementi in tracce in Italia. prime rilevazioni mediante i muschi., *Inquinamento* 36 (2) 48-58.
- [51] R. Bargagli, D. Brown, L. Nelli, Metal biomonitoring with mosses: Procedures for correcting for soil contamination, *Environ. Pollut.* 89 (2) (1995) 169 – 175.
- [52] M. Castello, A comparison between two moss species used as transplants for airborne trace element biomonitoring in ne Italy, *Environ. Monit. Asses.* 133 (1-3) (2007) 267–276.
- [53] H. Zechmeister, Growth rates of five pleurocarpous moss species under various climatic conditions, *J. Bryol.* 18 (3) (1995) 455–468.
- [54] D. Brown, Mineral nutrition, in: *Bryophyte ecology*, Springer, 1982, pp. 383–444.
- [55] M. Boyanov, S. Kelly, K. Kemner, B. Bunker, J. Fein, D. Fowle, Adsorption of cadmium to *Bacillus subtilis* bacterial cell walls: a ph-dependent X-ray absorption fine structure spectroscopy study, *Geochim. Cosmochim. Acta.* 67 (18) (2003) 3299–3311.
- [56] O. S. Pokrovsky, G. S. Pokrovski, A. Gãclabert, J. Schott, A. Boudou, Speciation of Zn associated with diatoms using X-ray absorption spectroscopy, *Environ. Sci. Technol.* 39 (12) (2005) 4490– 4498.
- [57] B. Mishra, M. Boyanov, B. A. Bunker, S. D. Kelly, K. M. Kemner, J. B. Fein, High and low-affinity binding sites for Cd on the bacterial cell walls of *Bacillus subtilis* and *Shewanella oneidensis*, *Geochim. Cosmochim. Acta.* 74 (15) (2010) 4219–4233.
- [58] J. B. Fein, J.-F. Boily, N. Yee, D. Gorman-Lewis, B. F. Turner, Potentiometric titrations of *Bacillus subtilis* cells to low ph and a comparison of modeling approaches, *Geochim. Cosmochim. Acta.* 69 (5) (2005) 1123–1132.
- [59] M. Ueshima, B. R. Ginn, E. A. Haack, J. E. S. Szymanowski, J. B. Fein, Cd adsorption onto *Pseudomonas putida* in the presence and absence of extracellular polymeric substances, *Geochim. Cosmochim. Acta.* 72 (24) (2008) 5885–5895.
- [60] R. E. Martinez, O. S. Pokrovsky, J. Schott, E. H. Oelkers, Surface charge and zeta potential of metabolically active and dead cyanobacteria, *J. Colloid. Interf. Sci.* 323 (2) (2008) 317–325.

- [61] B. S. Gupta, M. Curran, S. Hasan, T. Ghosh, Adsorption characteristics of Cu and Ni on Irish peat moss, *J. Environ. Manage.* 90 (2) (2009) 954 – 960.
- [62] L. Ringqvist, I. Öborn, Copper and zinc adsorption onto poorly humified sphagnum and carex peat, *Water Res.* 36 (9) (2002) 2233 – 2242
- [63] T. Karlsson, Complexation of cadmium, copper and methyl mercury to functional groups in natural organic matter, PhD Thesis (2005).
- [64] O. S. Pokrovsky, J. Viers, E. E. Emnova, E. I. Kompantseva, R. Freydisier, Copper isotope fractionation during its interaction with soil and aquatic microorganisms and metal oxy(hydr)oxides: Possible structural control, *Geochim. Cosmochim. Acta.* 72 (7) (2008) 1742–1757.
- [65] A. Gélabert, O. Pokrovsky, J. Schott, A. Boudou, A. Feurtet-Mazel, Cadmium and lead interaction with diatom surfaces: a combined thermodynamic and kinetic approach, *Geochim. Cosmochim. Acta.* 71 (15) (2007) 3698–3716.
- [66] J. Ha, A. Gélabert, A.M. Spormann, G.E. Brown Jr., role of extracellular polymeric substances in metal ion complexation on *Shewanella oneidensis*: Batch uptake, thermodynamic modeling, ATR-FTIR, and EXAFS study. *Geochim. Cosmochim. Acta.* 74 (1) (2010) 1-15.
- [67] O. Pokrovsky, A. Feurtet-Mazel, R. Martinez, S. Morin, M. Baudrimont, T. Duong, M. Coste, Experimental study of cadmium interaction with periphytic biofilms, *Appl. Geochem.* 25 (3) (2010) 418–427.
- [68] C. Lamelas, M. Benedetti, K. J. Wilkinson, V. I. Slaveykova, Characterization of H⁺ and Cd²⁺ binding properties of the bacterial exopolysaccharides, *Chemosphere* 1362 (2006) 1370.
- [69] A. Gélabert, O. Pokrovsky, J. Viers, J. Schott, A. Boudou, A. Feurtet-Mazel, Interaction between zinc and freshwater and marine diatom species: surface complexation and Zn isotope fractionation, *Geochim. Cosmochim. Acta.* 70 (4) (2006) 839–857.
- [70] O. Keskinan, M. Goksu, A. Yuceer, M. Basibuyuk, C. Forster, Heavy metal adsorption characteristics of a submerged aquatic plant (*Myriophyllum spicatum*), *Process Biochemistry* 39 (2) (2003) 179–183.
- [71] T. Wang, J. Weissman, G. Ramesh, R. Varadarajan, J. Benemann, Parameters for removal of toxic heavy metals by water milfoil (*Myriophyllum spicatum*), *Bulletin of environmental contamination and toxicology* 57 (5) (1996) 779–786.
- [72] I. A. H. Schneider, J. Rubio, Sorption of heavy metal ions by the nonliving biomass of freshwater macrophytes, *Environ. Sci. Technol.* 33 (13) (1999) 2213–2217.
- [73] P. Pavasant, R. Apiratikul, V. Sungkhum, P. Suthiparinyanont, S. Wattanachira, T. F. Marhaba, Biosorption of Cu²⁺, Cd²⁺, Pb²⁺, and Zn²⁺ using dried marine green macroalga *Caulerpa lentillifera*, *Bioresour. Technol.* 97 (18) (2006) 2321–2329.
- [74] A. Öztürk, T. Artan, A. Ayar, Biosorption of nickel (II) and copper (II) ions from aqueous solution by *Streptomyces coelicolor* a3 (2), *Colloid. Surface. B.* 34 (2) (2004) 105–111.

- [75] T. Gosset, J.-L. Trancart, D. R. Thévenot, Batch metal removal by peat. Kinetics and thermodynamics, *Water Res.* 20 (1) (1986) 21–26.
- [76] G. Yan, T. Viraraghavan, Heavy-metal removal from aqueous solution by fungus *Mucor rouxii*, *Water Research* 37 (18) (2003) 4486–4496.
- [77] A. Öztürk, Removal of nickel from aqueous solution by the bacterium *Bacillus thuringiensis*, *J. Hazard. Mater.* 147 (1) (2007) 518–523.
- [78] R. J. Martins, R. Pardo, R. A. Boaventura, Cadmium (II) and zinc (II) adsorption by the aquatic moss *Fontinalis antipyretica*: effect of temperature, pH and water hardness, *Water Res.* 38 (3) (2004) 693–699.
- [79] R. Rakhshae, M. Khosravi, M. T. Ganji, Kinetic modeling and thermodynamic study to remove Pb (II), Cd (II), Ni (II) and Zn (II) from aqueous solution using dead and living *Azolla filiculoides*, *J. Hazard. Mater.* 134 (1) (2006) 120–129.
- [80] D. Roy, P. N. Greenlaw, B. S. Shane, Adsorption of heavy metals by green algae and ground rice hulls, *Journal of Environmental Science & Health Part A* 28 (1) (1993) 37–50.
- [81] A. Sarı, M. Tuzen, Biosorption of Pb (II) and Cd (II) from aqueous solution using green alga *Ulva lactuca* biomass, *J. Hazard. Mater.* 152 (1) (2008) 302–308.
- [82] O. S. Pokrovsky, A. Probst, E. Leviel, B.-h. Liao, Interactions between cadmium and lead with acidic soils: Experimental evidence of similar adsorption patterns for a wide range of metal concentrations and the implications of metal migration, *J. Hazard. Mater.* 199 (2012) 358–366.
- [83] B. R. Ginn, J. B. Fein, The effect of species diversity on metal adsorption onto bacteria, *Geochim. Cosmochim. Acta.* 72 (16) (2008) 3939–3948.

LIST OF FIGURES

Fig. 1. Surface acid-base titration of each moss species in 0.01M NaNO₃ and 1.0 g_{dry}L⁻¹ biomass. Each experiment was carried out by triplicate. The solutions were conditioned during 1 hour. Lines represent the LPM model results.

Fig. 2. Long-term adsorption of Copper (A) at pH = 5.30 ± 0.01 and Zinc (B) at pH = 6.20 ± 0.05. [Cu²⁺] = 0.05-2.42 mmol g⁻¹ and [Zn²⁺] = 0.04-2.54 mmol g⁻¹. Biomass was always kept constant as 1.0 g_{dry}L⁻¹.

Fig. 3. Percentage of metal adsorbed onto moss surface as a function of pH, in 0.01M NaNO₃ and 1.0 g_{dry}L⁻¹ biomass. Initial metal concentration 0.052 mM. Lines represent the LPM model results. Dashed line was recalculated from Pokrovsky et

al. [64] at 4 $\text{g}_{\text{wet}}\text{L}^{-1}$ (A), Borrok et al. [48] at 10 $\text{g}_{\text{dry}}\text{L}^{-1}$ (B), Pokrovsky et al. [40] at 10 $\text{g}_{\text{wet}}\text{L}^{-1}$ (C), Ginn and Fein [83] at 1 $\text{g}_{\text{dry}}\text{L}^{-1}$ and Pokrovsky et al. [43] at 4 $\text{g}_{\text{wet}}\text{L}^{-1}$ (D), Pokrovsky et al. [43] (E) at 1.01 $\text{g}_{\text{dry}}\text{L}^{-1}$.

Fig. 4. Dissolved Organic Carbon measured during metal adsorption experiments as a function of pH, in 0.01M NaNO_3 and 1.0 $\text{g}_{\text{dry}}\text{L}^{-1}$ biomass.

Fig. 5. Metal adsorbed onto moss surface as a function of metal concentration in solution (Langmuir-isotherm), in 0.01M NaNO_3 and 1.0 $\text{g}_{\text{dry}}\text{L}^{-1}$ biomass at constant pH (see Table 3). Lines represent the LPM model results.

Fig. 6. Relative percentage of binding functional groups obtained from the acid-base titration for Bacteria consortia [48], Soil bacteria [39, 59], Cyanobacteria [43, 60] and Moss (this study).

Fig. 7. The value of q_{max} for (A) Copper and Zinc and (B) Cadmium and Lead. These data were collected from the literature for aquatic plants [70-73], yeast [74], herbaceous peat [26, 75], Sphagnum peat [26], fungus [20, 76], bacteria [77-78], plants [79], algae [80-81] and soil [82].

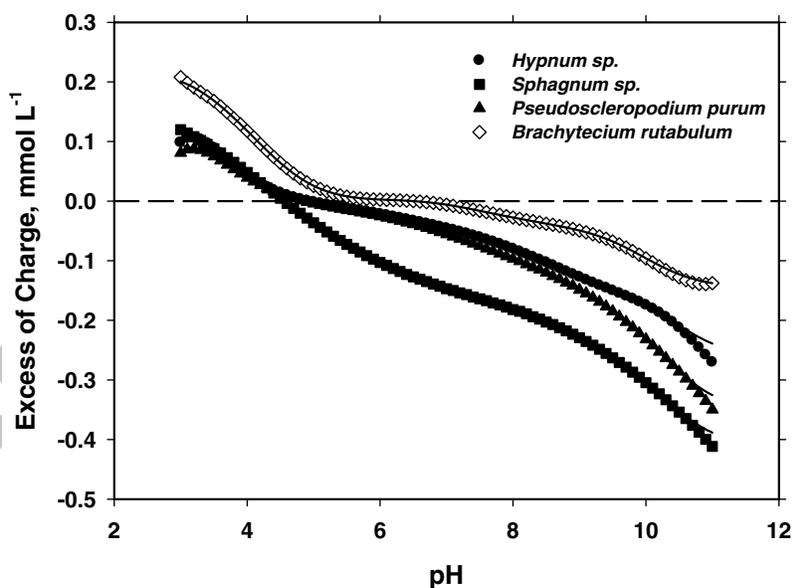


Fig. 1

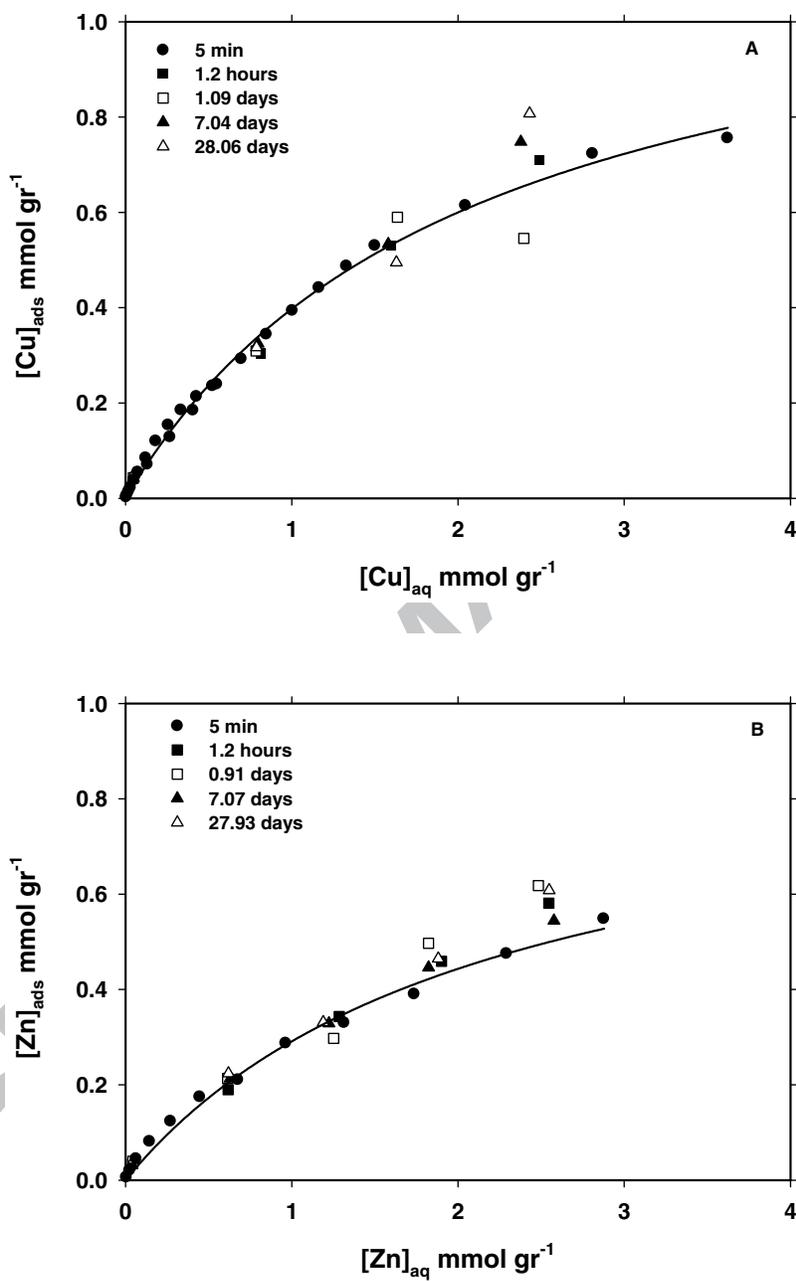


Fig. 2

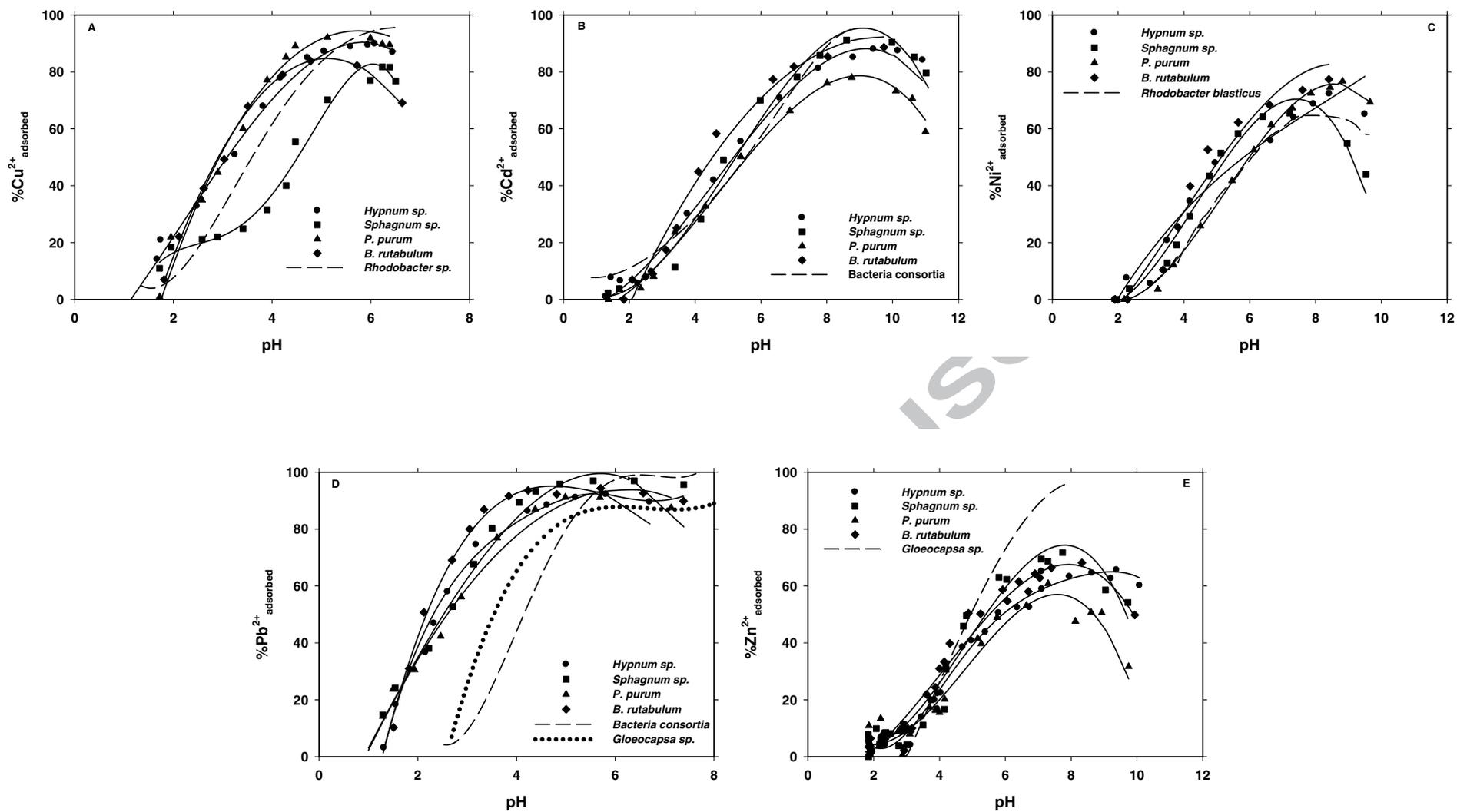


Fig. 3

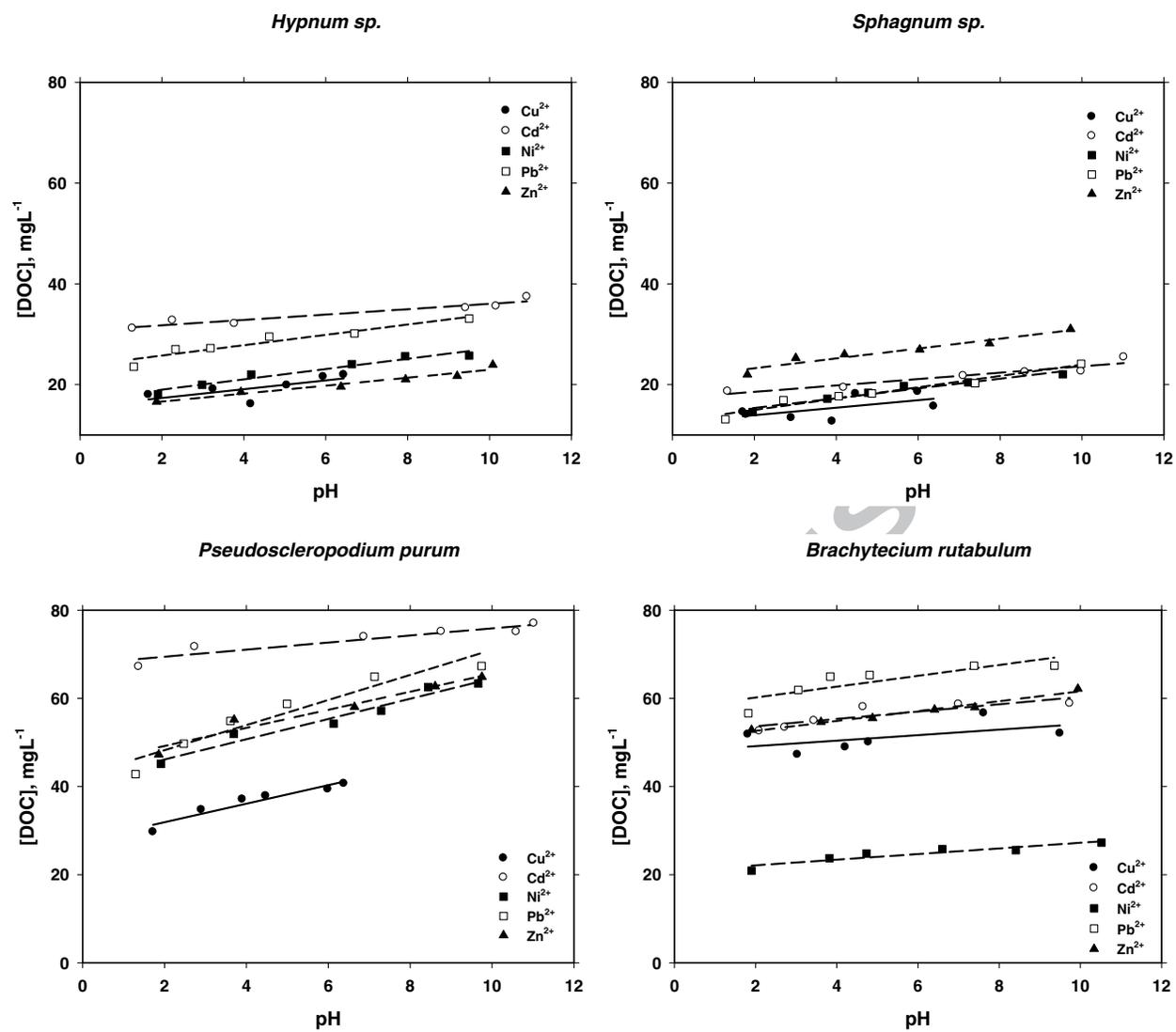


Fig. 4

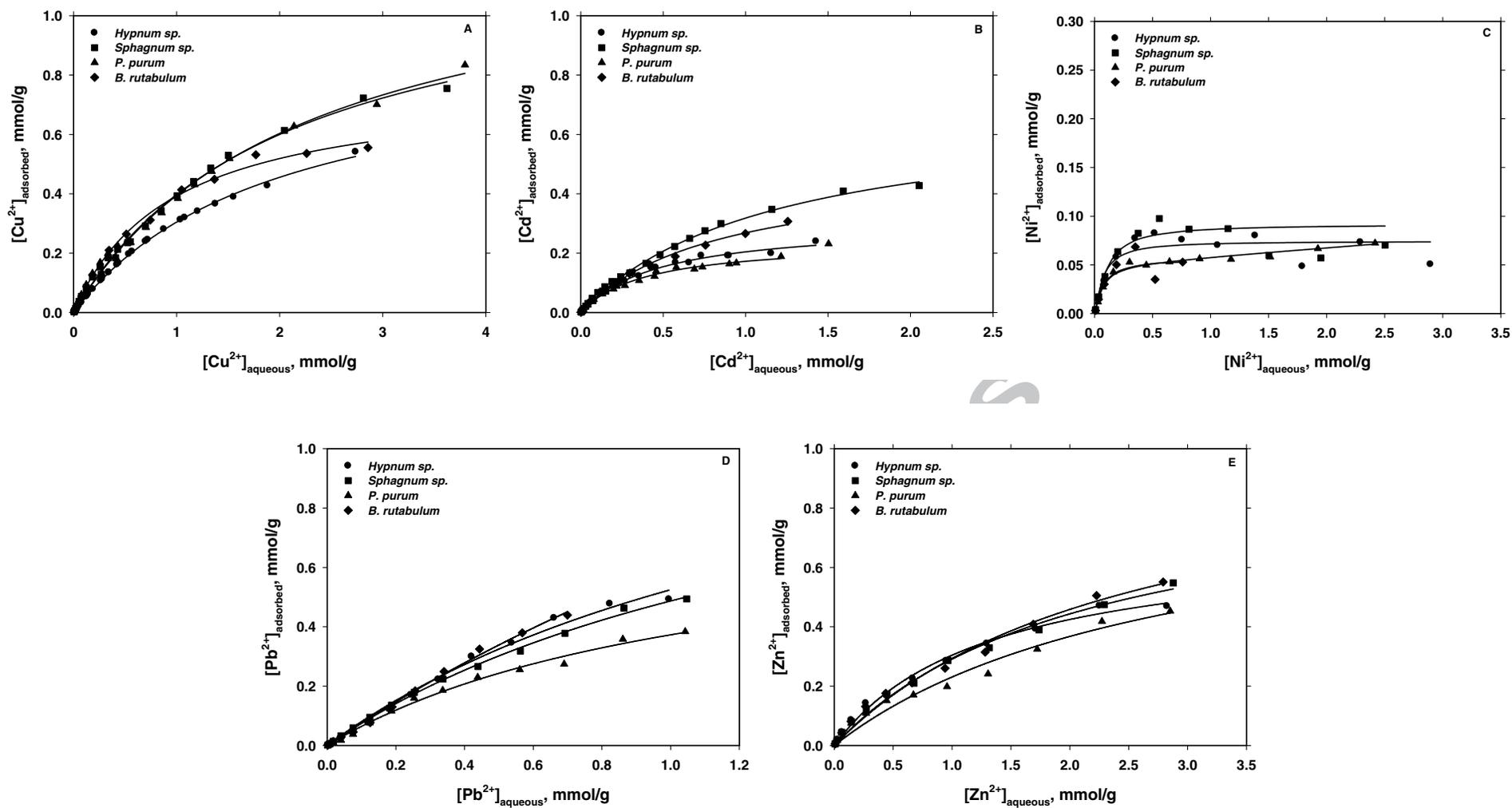


Fig. 5

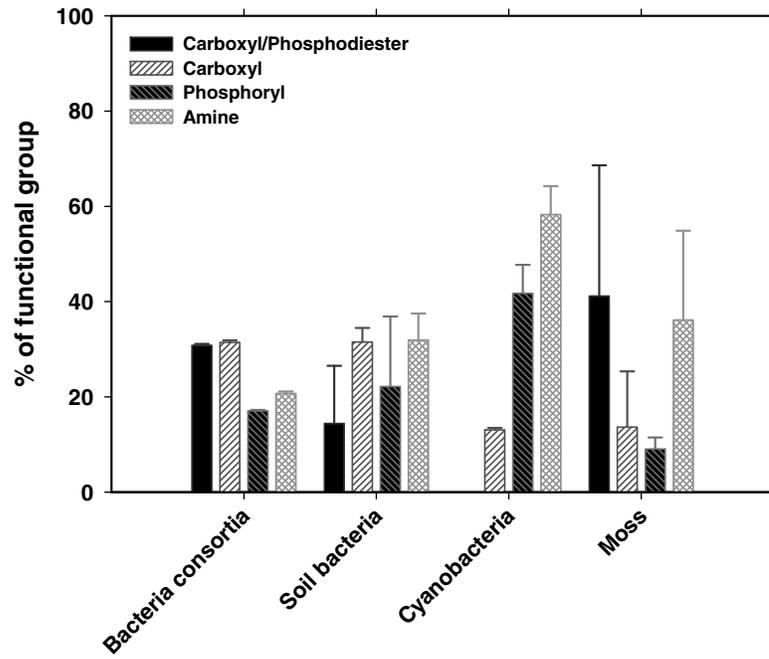


Fig. 6

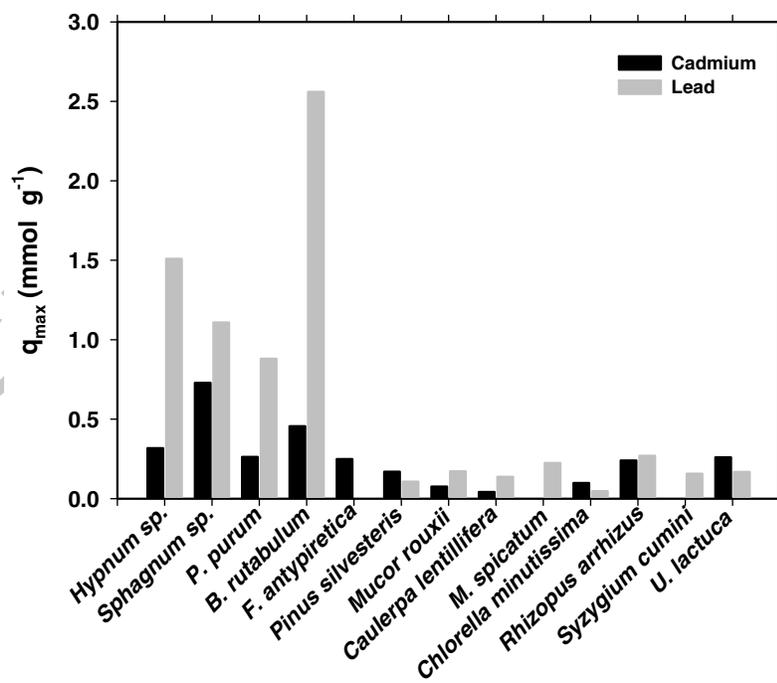
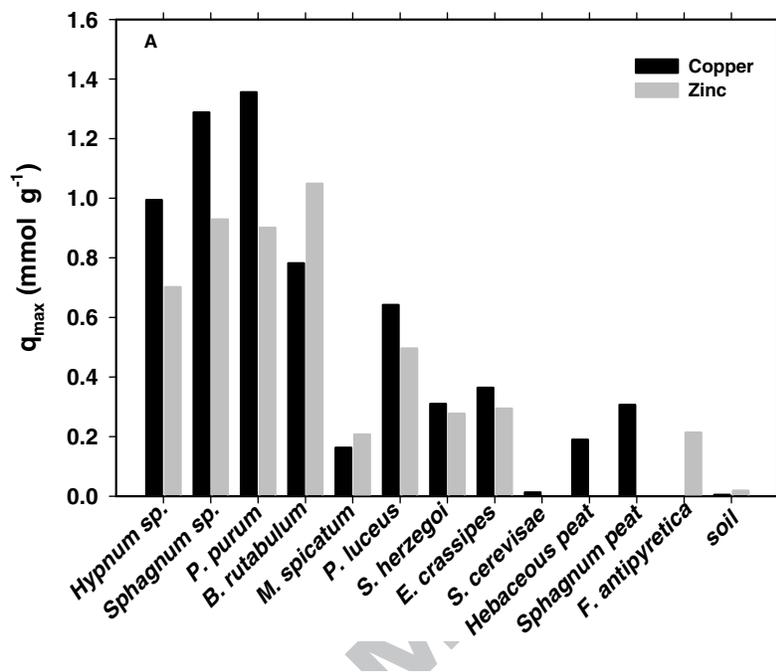


Fig. 7

LIST OF TABLES

Table 1. Surface acid-base titration and LPM parameters for moss in 0.01 M NaNO₃ with 1.0 g_{dry}L⁻¹ of biomass. Conditioning time of 1 hour.

| Specie | pK _a | Binding sites mmol g ⁻¹ | Possible functional group |
|--------------------------------------------------------|-----------------------|---------------------------------------|------------------------------|
| <i>Hypnum sp.</i> (S _T =3) | 3.70 | 1.83·10 ⁻¹ | Carboxyl/Phosphodiester |
| | 5.35 | 1.10·10 ⁻² | Carboxyl |
| | 6.10 | 2.13·10 ⁻² | Phosphoryl |
| | 6.90 | 3.32·10 ⁻² | Phosphoryl |
| | 7.73 | 3.45·10 ⁻² | Amine |
| | 8.68 | 6.66·10 ⁻² | Amine |
| | 10.43 | 1.40·10 ⁻² | Polyphenol |
| <i>Sphagnum sp.</i> (S _T =3) | 3.58 | 1.56·10 ⁻¹ | Carboxyl/Phosphodiester |
| | 4.73 | 9.48·10 ⁻² | Carboxyl |
| | 5.63 | 4.77·10 ⁻² | Carboxyl |
| | 6.45 | 3.43·10 ⁻² | Phosphoryl |
| | 7.05 | 3.10·10 ⁻² | Phosphoryl |
| | 7.85 | 3.36·10 ⁻² | Amine |
| | 8.05 | 2.84·10 ⁻² | Amine |
| | 9.10 | 6.72·10 ⁻² | Amine |
| 10.30 | 1.56·10 ⁻¹ | Polyphenol | |
| <i>Pseudoscleropodium purum</i> (S _T =3) | 3.75 | 1.52·10 ⁻¹ | Carboxyl/Phosphodiester |
| | 4.95 | 1.85·10 ⁻² | Carboxyl |
| | 5.75 | 1.70·10 ⁻² | Carboxyl |
| | 6.55 | 2.42·10 ⁻² | Phosphoryl |
| | 7.30 | 3.46·10 ⁻² | Phosphoryl |
| | 8.00 | 3.40·10 ⁻² | Amine |
| | 9.05 | 6.77·10 ⁻² | Amine |
| | 10.30 | 2.06·10 ⁻¹ | Polyphenol |
| <i>Brachytecium rutabulum</i> (S _T =3) | 3.60 | 1.51·10 ⁻¹ | Carboxyl/Phosphodiester |
| | 4.50 | 1.22·10 ⁻¹ | Carboxyl |
| | 5.93 | 1.52·10 ⁻² | Phosphoryl |
| | 7.35 | 3.38·10 ⁻² | Phosphoryl |
| | 8.18 | 1.21·10 ⁻² | Amine |
| | 9.15 | 1.32·10 ⁻² | Amine |
| | 10.13 | 1.38·10 ⁻¹ | Polyphenol |

Table 2. Experimental conditions and LPM parameters for metal adsorption on moss as a function of pH in 0.01 M NaNO₃ with biomass of 1.0 g_{dry}L⁻¹. K_s corresponds with the equilibrium constant for the reaction between metal in solution and the available sites as a function of pH.

| Specie | Metal studied | pH-range | [Me ²⁺] μM | pK _s | Binding sites mmol g ⁻¹ | |
|---------------------------------|---------------|--------------|---------------------------|-----------------------|---------------------------------------|-----------------------|
| <i>Hypnum sp.</i> | Copper | 1.67 – 6.44 | 51.94 | -1.80 | 3.32·10 ⁻² | |
| <i>Sphagnum sp.</i> | | 1.72 – 6.50 | | -3.15 | 8.69·10 ⁻³ | |
| <i>Pseudoscleropodium purum</i> | | 1.72 – 6.38 | | -0.60 | 1.95·10 ⁻² | |
| <i>Brachytecium rutabulum</i> | | 1.81 – 6.63 | | -1.90 | 3.96·10 ⁻² | |
| <i>Hypnum sp.</i> | Cadmium | 1.28 – 10.21 | 29.36 | -4.15 | 1.28·10 ⁻³ | |
| | | | | | -1.30 | 8.55·10 ⁻³ |
| | | | | | 0.25 | 9.29·10 ⁻³ |
| <i>Sphagnum sp.</i> | | 1.35 – 11.02 | | -4.40 | 8.26·10 ⁻⁴ | |
| <i>Pseudoscleropodium purum</i> | | 1.37 – 11.02 | | -0.35 | 1.78·10 ⁻² | |
| | | | | -1.55 | 7.81·10 ⁻³ | |
| <i>Brachytecium rutabulum</i> | | 1.84 – 9.74 | | 0.00 | 7.85·10 ⁻³ | |
| | | | | -2.50 | 1.31·10 ⁻³ | |
| | -1.05 | | 1.38·10 ⁻² | | | |
| <i>Hypnum sp.</i> | Nickel | 1.90 – 9.50 | 56.23 | -0.15 | 6.97·10 ⁻³ | |
| | | | | -3.50 | 5.71·10 ⁻⁴ | |
| <i>Sphagnum sp.</i> | | 1.96 – 9.53 | | -0.70 | 2.46·10 ⁻² | |
| <i>Pseudoscleropodium purum</i> | | 1.91 – 9.66 | | -0.55 | 2.48·10 ⁻² | |
| | | | | -0.60 | 6.38·10 ⁻³ | |
| <i>Brachytecium rutabulum</i> | 1.90 – 10.52 | 0.15 | 1.95·10 ⁻² | | | |
| <i>Hypnum sp.</i> | Lead | 1.31 – 6.70 | 15.93 | -0.50 | 3.20·10 ⁻² | |
| <i>Sphagnum sp.</i> | | 1.29 – 7.39 | | -2.95 | 1.27·10 ⁻² | |
| <i>Pseudoscleropodium purum</i> | | 1.29 – 7.13 | | -3.00 | 1.30·10 ⁻² | |
| <i>Brachytecium rutabulum</i> | | 1.51 – 7.38 | | -2.95 | 1.11·10 ⁻² | |
| <i>Hypnum sp.</i> | Zinc | 1.87 – 10.08 | 50.48 | -3.20 | 1.32·10 ⁻² | |
| <i>Sphagnum sp.</i> | | 1.83 – 9.72 | | -0.55 | 1.57·10 ⁻² | |
| <i>Pseudoscleropodium purum</i> | | 1.86 – 9.75 | | -0.45 | 1.81·10 ⁻² | |
| <i>Brachytecium rutabulum</i> | | 1.90 – 9.94 | | -0.35 | 1.39·10 ⁻² | |
| | | | -0.65 | 2.00·10 ⁻² | | |

Table 3. Experimental conditions and LPM parameters for metal adsorption on moss as a function of metal concentration in solution (Langmuirian-isotherm) in 0.01 M NaNO₃ with biomass of 1.0 g_{dry}L⁻¹. K_m corresponds with the equilibrium constant for the reaction between metal in solution and the available sites as a function metal aqueous concentration in solution.

| Specie | Metal studied | pH-range | [Me ²⁺] M | pK _m | Binding sites mmol g ⁻¹ | |
|---------------------------------|---------------|-------------|-----------------------------------------------|-----------------------------------------------|---------------------------------------|-----------------------------------------------|
| <i>Hypnum sp.</i> | Copper | 5.50 ± 0.04 | 5.19·10 ⁻⁶ – 3.65·10 ⁻³ | 2.25 | 0.430 | |
| | | | | 5.10 | 34.828 | |
| <i>Sphagnum sp.</i> | | 5.52 ± 0.03 | 1.57·10 ⁻⁶ – 3.62·10 ⁻³ | 0.70 | 0.051 | |
| | | | | 2.65 | 1.038 | |
| <i>Pseudoscleropodium purum</i> | | 5.51 ± 0.04 | 1.57·10 ⁻⁶ – 3.78·10 ⁻³ | 0.75 | 0.104 | |
| | | | | 2.85 | 1.220 | |
| <i>Brachytecium rutabulum</i> | | 5.53 ± 0.04 | 9.13·10 ⁻⁶ – 2.85·10 ⁻³ | 1.45 | 0.181 | |
| | | | | 2.40 | 0.523 | |
| <i>Hypnum sp.</i> | | Cadmium | 6.52 ± 0.04 | 2.31·10 ⁻⁶ – 1.41·10 ⁻³ | 2.55 | 0.172 |
| | | | | | 5.90 | 14.526 |
| <i>Sphagnum sp.</i> | 6.52 ± 0.05 | | 2.31·10 ⁻⁶ – 1.34·10 ⁻³ | 1.45 | 0.029 | |
| | | | | 3.35 | 0.591 | |
| <i>Pseudoscleropodium purum</i> | 6.52 ± 0.05 | | 2.31·10 ⁻⁶ – 1.47·10 ⁻³ | 1.15 | 6·10 ⁻³ | |
| | | | | 2.45 | 0.114 | |
| | | | | 6.20 | 40.266 | |
| <i>Brachytecium rutabulum</i> | 6.62 ± 0.08 | | 3.11·10 ⁻⁶ – 1.23·10 ⁻³ | 2.45 | 0.133 | |
| | | | | 4.60 | 1.329 | |
| | | | | 4.90 | 1.306 | |
| <i>Hypnum sp.</i> | Nickel | | 5.63 ± 0.03 | 9.37·10 ⁻⁶ – 2.89·10 ⁻³ | 1.25 | 0.073 |
| | | | | | 5.65 ± 0.05 | 9.88·10 ⁻⁶ – 3.17·10 ⁻³ |
| <i>Sphagnum sp.</i> | | 5.67 ± 0.03 | 9.08·10 ⁻⁶ – 2.41·10 ⁻³ | 1.35 | 0.053 | |
| | | | | 5.70 | 7.669 | |
| <i>Brachytecium rutabulum</i> | | 5.58 ± 0.04 | 9.37·10 ⁻⁶ – 2.91·10 ⁻³ | 1.20 | 0.057 | |
| <i>Hypnum sp.</i> | | Lead | 6.55 ± 0.07 | 1.93·10 ⁻⁶ – 9.93·10 ⁻⁴ | 2.95 | 0.799 |
| | | | | | <i>Sphagnum sp.</i> | 6.52 ± 0.05 |
| 3.45 | | | 0.961 | | | |
| <i>Pseudoscleropodium purum</i> | 6.55 ± 0.06 | | 1.98·10 ⁻⁶ – 1.03·10 ⁻³ | 2.80 | 0.334 | |
| | | | | 6.00 | 50.400 | |
| <i>Brachytecium rutabulum</i> | 6.53 ± 0.05 | | 2.03·10 ⁻⁶ – 6.97·10 ⁻⁴ | 3.10 | 1.117 | |
| <i>Hypnum sp.</i> | 6.76 ± 0.05 | | 7.19·10 ⁻⁶ – 2.83·10 ⁻³ | 2.15 | 0.084 | |
| | | | | 3.95 | 0.653 | |

| | | | | | |
|---------------------------------|------|-----------------|-------------------------------------------|------|--------|
| <i>Sphagnum sp.</i> | Zinc | 6.79 ± 0.08 | $7.34 \cdot 10^{-6} - 2.88 \cdot 10^{-3}$ | 2.05 | 0.059 |
| | | | | 3.45 | 0.200 |
| | | | | 6.10 | 28.601 |
| <i>Pseudoscleropodium purum</i> | | 6.77 ± 0.05 | $7.19 \cdot 10^{-6} - 2.85 \cdot 10^{-3}$ | 2.25 | 0.095 |
| | | | | 5.45 | 7.118 |
| <i>Brachytecium rutabulum</i> | | 6.78 ± 0.06 | $7.04 \cdot 10^{-6} - 2.78 \cdot 10^{-3}$ | 2.40 | 0.116 |
| | | | | 4.65 | 2.070 |

Table 4. Langmuir parameters computed from the experiments at different aqueous metal concentration (Langmuirian isotherm).

| Specie | Metal studied | q_{\max} mmol gr ⁻¹ | K_L gr mmol ⁻¹ |
|---------------------------------|---------------|-------------------------------------|--------------------------------|
| <i>Hypnum sp.</i> | Copper | 0.994 | 0.437 |
| <i>Sphagnum sp.</i> | | 1.288 | 0.443 |
| <i>Pseudoscleropodium purum</i> | | 1.356 | 0.417 |
| <i>Brachytecium rutabulum</i> | | 0.782 | 0.970 |
| <i>Hypnum sp.</i> | Cadmium | 0.318 | 1.771 |
| <i>Sphagnum sp.</i> | | 0.729 | 0.785 |
| <i>Pseudoscleropodium purum</i> | | 0.263 | 2.047 |
| <i>Brachytecium rutabulum</i> | | 0.456 | 1.385 |
| <i>Hypnum sp.</i> | Nickel | 0.089 | 9.155 |
| <i>Sphagnum sp.</i> | | 0.107 | 7.081 |
| <i>Pseudoscleropodium purum</i> | | 0.062 | 11.360 |
| <i>Brachytecium rutabulum</i> | | 0.071 | 10.454 |
| <i>Hypnum sp.</i> | Lead | 1.509 | 0.531 |
| <i>Sphagnum sp.</i> | | 1.109 | 0.746 |
| <i>Pseudoscleropodium purum</i> | | 0.880 | 0.775 |
| <i>Brachytecium rutabulum</i> | | 2.560 | 0.305 |
| <i>Hypnum sp.</i> | Zinc | 0.702 | 0.765 |
| <i>Sphagnum sp.</i> | | 0.929 | 0.456 |
| <i>Pseudoscleropodium purum</i> | | 0.901 | 0.344 |
| <i>Brachytecium rutabulum</i> | | 1.049 | 0.388 |

