

Accepted Manuscript

Title: Capability of microalgae-based wastewater treatment systems to remove emerging organic contaminants: a pilot-scale study

Author: Víctor Matamoros Raquel Gutiérrez Ivet Ferrer Joan García Josep M Bayona



PII: S0304-3894(15)00083-7
DOI: <http://dx.doi.org/doi:10.1016/j.jhazmat.2015.02.002>
Reference: HAZMAT 16573

To appear in: *Journal of Hazardous Materials*

Received date: 4-7-2014
Revised date: 12-1-2015
Accepted date: 1-2-2015

Please cite this article as: Víctor Matamoros, Raquel Gutiérrez, Ivet Ferrer, Joan García, Josep M Bayona, Capability of microalgae-based wastewater treatment systems to remove emerging organic contaminants: a pilot-scale study, Journal of Hazardous Materials <http://dx.doi.org/10.1016/j.jhazmat.2015.02.002>

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

Capability of microalgae-based wastewater treatment systems to remove emerging organic contaminants: a pilot-scale study

Víctor Matamoros^{1*}, Raquel Gutiérrez², Ivet Ferrer², Joan García², Josep M Bayona¹

¹Department of Environmental Chemistry, IDAEA-CSIC, c/Jordi Girona, 18-26, E-08034, Barcelona, Spain.

²GEMMA-Group of Environmental Engineering and Microbiology, Department of Hydraulic, Maritime and Environmental Engineering, Universitat Politècnica de Catalunya BarcelonaTech, c/ Jordi Girona, 1-3, Building D1, E-08034, Barcelona, Spain.

**Corresponding author: victor.matamoros@idaea.csic.es*

22 **Highlights** (max. 85 characters per highlight including spaces):

- 23 • The effect of hydraulic retention time and seasonality has been evaluated.
- 24 • Removal efficiency ranged from undetectable removal to more than 90%.
- 25 • Biodegradation and photodegradation were the most important removal pathways.
- 26 • We suggested that microalgae enhance the biodegradation of emerging contaminants.
- 27 • Up to 90% of the contaminant toxicity risk was removed by microalgae treatment.

29 0. Abstract

30 The effect of hydraulic retention time (HRT) and seasonality on the removal efficiency of 26
 31 organic microcontaminants from urban wastewater was studied in two pilot high-rate algal ponds
 32 (HRAPs). The targeted compounds included pharmaceuticals and personal care products, fire
 33 retardants, surfactants, anticorrosive agents, pesticides and plasticizers, among others. The pilot
 34 plant, which was fed at a surface loading rate of 7-29 g of COD m⁻² d⁻¹, consisted of a
 35 homogenisation tank and two parallel lines, each one with a primary settler and an HRAP with a
 36 surface area of 1.5 m² and a volume of 0.5 m³. The two HRAPs were operated with different HRTs
 37 (4 and 8 d). The removal efficiency ranged from negligible removal to more than 90% depending on
 38 the compound. Microcontaminant removal efficiencies were enhanced during the warm season,
 39 while the HRT effect on microcontaminant removal was only noticeable in the cold season. Our
 40 results suggest that biodegradation and photodegradation are the most important removal pathways,
 41 whereas volatilization and sorption were solely achieved for hydrophobic compounds (log K_{ow}>4)
 42 with a moderately high Henry's law constant values (11-12 Pa m⁻³ mol⁻¹) such as musk fragrances.
 43 Whereas acetaminophen, ibuprofen and oxybenzone presented ecotoxicological hazard quotients
 44 (HQs) higher than 1 in the influent wastewater samples, the HQs for the effluent water samples
 45 were always below 1.

49

50

51 **Keywords:** *emerging organic contaminants; microalgae; high-rate algal pond; photodegradation;*
52 *biodegradation; volatilization.*

53 **1. Introduction**

54 Emerging organic contaminants (EOCs) include a wide range of compounds belonging to different
55 chemical classes, such as pharmaceuticals, personal care products, plasticizers, flame retardants,
56 surfactants, and certain pesticides, among others, the ecotoxicological effects of which are relatively
57 unknown [1]. Since conventional wastewater treatment plants (WWTPs) are not designed to remove
58 emerging and related contaminants, many of these compounds occur at different concentrations in
59 natural water bodies [2], where they may exert ecotoxicological effects at relatively low
60 concentrations [3, 4]. Although some of the compounds have been proposed for inclusion on
61 regulatory lists of contaminants (European Commission, 2006), there is relatively little information
62 on the ecotoxicological effects of complex mixtures at environmental levels, and, to date, they have
63 not been regulated [1]. Known environmental effects of some EOCs include the reduction of
64 macroinvertebrate diversity in rivers [3], behavioural changes in mosquito fish [4] and reproductive
65 disruption in fish [5], among others. Due to the difficulty of assessing the effects of EOCs on
66 ecosystems, the use of hazard quotients (HQs) based on the chemical composition of water samples
67 and tabulated predicted non-effect concentrations (PNECs) for different aquatic organisms has been
68 postulated as a good screening strategy [6].

69 Microalgae-based wastewater treatment technologies such as high-rate algal ponds (HRAPs) have
70 received considerable attention in recent years due to the resource recovery of algal biomass, for use
71 as fertilizer, protein-rich feed or biofuel, and a high-quality effluent (treated wastewater)[7]. HRAPs
72 are shallow raceway reactors in which microalgae and bacteria grow in symbiosis. In such systems,

73 organic matter is degraded by heterotrophic bacteria, which consume oxygen provided by
74 microalgal photosynthesis; therefore, no aeration is needed [8]. Although the capability of
75 microalgae wastewater treatment systems to remove nutrients, heavy metals, bacteria, and
76 helminthic eggs has been studied since the 1950s, few studies have focused on the removal of
77 organic contaminants, namely, phenolic compounds, surfactants, biocides and polycyclic aromatic
78 hydrocarbons [9-12]. Indeed, no attention has been paid to the effectiveness of HRAPs for
79 removing EOCs of environmental concern.

80 The removal of EOCs by conventional activated sludge WWTPs has been widely studied, but the
81 effectiveness of HRAPs for removing EOCs from wastewater has not yet been addressed. There is
82 only one study dealing with HRAPs' capacity to remove tetracyclines, and it was performed at
83 laboratory-scale with synthetic wastewater [13]. Other studies dealing with microalgae's capacity to
84 remove organic contaminants, such as polycyclic aromatic hydrocarbons (PAHs), biocides (e.g.
85 organotin compounds), surfactants and phenolic compounds, suggest that microalgae-based
86 wastewater technologies may remove microcontaminants by both abiotic (sorption, volatilization or
87 photodegradation) and biotic (biodegradation, microalgae uptake or metabolization) processes [14-
88 16].

89 The aim of this study was to evaluate for the first time, the effect of hydraulic retention time (HRT)
90 and ambient temperature / sunlight irradiation (seasonality) on the removal efficiency of 26 EOCs
91 in two HRAP pilot plants fed with real urban wastewater. The selected compounds were high
92 production volume chemicals (e.g. fire retardants, surfactants, anticorrosive agents, pesticides,
93 plasticizers, pharmaceuticals and personal care products, among others). Finally, aquatic risk
94 assessment was performed based on the concentrations of the detected EOCs in the influent and
95 effluent water samples, and the listed EC50 values for *Daphnia magna*.

96

97 2. Material and Methods

98 2.1. Chemicals and reagents

99 Gas chromatography (GC) grade (Suprasolv) hexane, methanol, and ethyl acetate were obtained
 100 from Merck (Darmstadt, Germany). Analytical-grade hydrogen chloride was obtained from Panreac
 101 (Barcelona, Spain). Caffeine, acetaminophen, ibuprofen, methyl dihydrojasmonate, oxybenzone,
 102 ketoprofen, hydrocinnamic acid, 5-methylbenzotriazole, naproxen, carbamazepine, galaxolide,
 103 benzothiazole, diclofenac, methylparaben, benzotriazole, tonalide, OH-benzothiazole, tributyl
 104 phosphate, tris(2-chloroethyl) phosphate, triphenyl phosphate, triclosan, cashmeran, octylphenol,
 105 diazinon, celestolide, atrazine, bisphenol A, 2,4-D, atrazine D5, mecoprop D3, tonalide D3 and
 106 dihydrocarbamazepine were purchased from Sigma-Aldrich (Steinheim, Germany).
 107 Trimethylsulfonium hydroxide (TMSH) was obtained from Fluka (Buchs, Switzerland). Strata-X
 108 polymeric SPE cartridges (200 mg) were purchased from Phenomenex (Torrance, CA, USA) and
 109 the 0.7 μm glass fibre filters (ϕ 47 mm) were obtained from Whatman (Maidstone, UK).

110

111 2.2. Description of the HRAP pilot plant

112 The experimental set-up was located outdoors at the laboratory of the GEMMA research group
 113 (Universitat Politècnica de Catalunya-BarcelonaTech, Spain). The system has been operated since
 114 March 2010. The microalgae production system was composed of a screening pre-treatment and
 115 two identical parallel lines, each one equipped with a primary settler, a pilot high-rate algal pond
 116 and a final settler for biomass separation (Fig. 1). Paddle wheel was set at 5 rpm giving mixed
 117 liquor with a linear velocity of recirculation of 11 cm s^{-1} , enough to ensure complete mixing. Urban
 118 wastewater was pumped from a municipal sewer to a homogenisation tank (1.2 m^3), which was
 119 continuously stirred to avoid solids sedimentation. From there, the wastewater was pre-treated and
 120 conveyed to each line. The primary treatment included a settler with an internal diameter of 0.3 m, a
 121 total height of 0.4 m and an effective volume of 7 L that was operated at an HRT of 0.9 h. Primary
 122 effluent from the settlers was pumped to the HRAPs by means of peristaltic pumps. The

123 experimental HRAPs were PVC raceway ponds equipped with a paddle wheel for stirring the mixed
124 liquor (Fig.1). The two HRAPs had a nominal volume of 0.47 m^3 , a surface area of 1.54 m^2 and a
125 water depth of 0.3 m, and they were operated simultaneously with different HRTs (4 and 8 days
126 corresponding to 117.5 and 58.8 L d^{-1} respectively). The final settlers for biomass separation had an
127 internal diameter of 0.15 m, a total height of 0.3 m and an effective volume of 3.5 L that were
128 operated at an HRT of 0.7 and 1.4 h for the HRAP set at 4 days HRT and 8 days, respectively. Note
129 that these settlers were only used for biomass separation, which was not recycled back to the
130 HRAPs.

131

132 2.3. Sampling strategy

133 Two sampling campaigns were carried out, one in July 2013 (warm season) and the other in
134 December 2013 (cold season). In each campaign, influent and effluent grab samples were collected
135 from both HRAPs at the same time each day (9:00 am) for a period of 10 days ($n=8$), from Monday
136 to the Wednesday of the next week, Saturday and Sunday were not sampled. The samples were
137 collected in the primary effluent from the settler and at the effluent from both HRAPs (Fig. 1). No
138 rainfall events were recorded at any time during the sampling period. All water samples were
139 collected in 1000 mL amber glass bottles, which were transported under refrigeration to the
140 laboratory, where they were stored at 4°C until analysis. The sample holding time was less than 12
141 hours.

142

143 2.4. Analytical procedures

144 Conventional wastewater quality parameters, including ammonium nitrogen ($\text{NH}_4\text{-N}$), total
145 suspended solids (TSS) and chemical oxygen demand (COD), were determined using the Standard
146 Methods (APHA, 2001). Onsite measurements of water temperature, dissolved oxygen (DO) and

147 pH were taken using a Checktemp-1 Hanna thermometer, an Eutech Ecoscan DO6 oxygen meter
148 and a Crison pH-meter, respectively.

149 For each campaign, 2 well-mixed 25 mL samples from each HRAP were examined by light
150 microscopy and the predominant microalgae were identified and quantified. Microalgae genus were
151 identified from classical specific literature [17, 18].

152 All water samples were filtered and processed as previously reported [19]. A 100 mL sample was
153 spiked with 50 ng of a surrogate standard (atrazine D5, mecoprop D3, tonalide D3, and
154 dihydrocarbamazepine). The spiked sample was percolated through a previously activated
155 polymeric solid-phase extraction cartridge (200 mg Strata X). Elution was performed with 10 mL of
156 hexane/ethyl acetate (1:1). The eluted extract was evaporated under a gentle nitrogen stream until
157 ca. 100 μ L remained, at which point 20 ng of triphenylamine was added as an internal standard.
158 Finally, the vial was reconstituted to 300 μ L with ethyl acetate.

159 The TSS collected in the glass fibre filters (0.7 μ m) were processed according to a previously
160 reported analytical method [19]. Briefly, the filters were freeze-dried and extracted in an ultrasonic
161 bath with hexane/acetone (3:1) for 15 minutes. The extracts were then further processed as water
162 samples.

163 Methylation of the acidic carboxyl group was performed in a hot GC PTV injector (270 °C) by
164 adding 10 μ L of TMSH solution (0.25 mol L⁻¹ in methanol) to a 50 μ L sample before injection.
165 Derivatized samples were analysed into a Bruker 450-GC gas chromatograph coupled to a Bruker
166 320-MS triple quadrupole mass spectrometer (Bruker Daltonics Inc., Billerica, MA, USA) fitted
167 with a 20 m \times 0.18 mm, 0.18 μ m film thickness Sapiens X5-MS capillary column coated with 5%
168 diphenyl 95% dimethyl polysiloxane from Teknokroma (Sant Cugat del Vallès, Spain) operated in
169 the multiple reaction mode (MRM). Validation of the analytical methodology has been described
170 elsewhere [20]. The limit of detection (LOD) and limit of quantification (LOQ) of the analytical
171 methodology were determined (using ultra-pure water) based on the mean background noise plus 3

or 10 times the standard deviation of the background noise, respectively. The LOD and LOQ ranged from 1 to 40 ng L⁻¹ and from 3 to 80 ng L⁻¹, respectively. Recoveries and repeatability were always higher than 80% and lower than 20%, respectively.

175

2.5. Data analysis

The removal efficiencies of conventional water quality parameters and EOCs were calculated as follows (equation 1):

179

180

$$Removal = \frac{1}{n} \sum_{i=1}^n \frac{C'a - (Ci - Ci \times \frac{EVR}{HLR})}{C'a} \times 100 \quad (Equation 1)$$

182

where $C'a$ is the average concentration of a selected compound in the HRAP influents in each sampling campaign, Ci is the concentration in the HRAP effluents on each sampling day, and n is the number of samples collected per sampling campaign ($n=8$). HLR are 83 or 43 L m⁻² d⁻¹ at a HRT of 8 and 4 days, respectively. Evaporation rates (EVRs) are 21 and 9 L m⁻² d⁻¹ in warm and cold season, respectively calculated from Turc's equation.

188

The experimental results were statistically evaluated using the SPSS v.13 package (Chicago, IL, USA). According with the data set size, non-parametric statistics were applied. The comparison of means was conducted by means of the Kruskal–Wallis test. Spearman's coefficients were used for correlations between variables (physicochemical parameters, removal efficiencies and influent concentration). Significance was defined as $p < 0.05$.

194

195

196

197

198 **3. Results and Discussion**199 *3.1 Assessment of conventional water quality parameters*

200 The HRAPs' performance was monitored in warm and cold seasons (Table 1). COD, TSS and NH₄-
 201 N concentrations in the primary settlers' effluents were within the typical range of a conventional
 202 primary effluent [21]. Note that DO concentration was quite high for a primary effluent due to
 203 wastewater stirring in the homogenisation tank. The actual surface organic loading rates (OLRs)
 204 applied to the HRAPs were, on average, 13 and 26 g total COD m⁻² d⁻¹ (HRT=8 and 4 d
 205 respectively) in the warm season and 29 and 58 g total COD m⁻² d⁻¹ (HRT=8 and 4 d respectively)
 206 in the cold season.

207 The microalgae present in the HRAPs were mostly species of the Phylum *Chlorophyta*, and varied
 208 depending on seasonal conditions. In the warm season, predominant algae species were
 209 *Stigeoclonium* sp. (filamentous algae); diatoms, *Chlorella* sp. and *Monoraphidium* sp. In the cold
 210 season, predominant algae species were *Chlorella* sp.; diatoms, *Stigeoclonium* sp. In both
 211 campaigns, the number of species was higher in the HRAP set at HRT of 8 days.

212 In the present study, mixed liquor TSS was analysed as an indicator of biomass concentration in the
 213 HRAP (Table 1). Note that biomass in such type of systems corresponds to microalgae as well as
 214 bacteria, and the relative proportion of them can be approached by means of mass balances on an
 215 estimated net yield of heterotrophic bacteria [22]. Thus, if the yield for domestic wastewater aerobic
 216 treatment is around 0.25 g VSS/g bCOD (biodegradable COD), assuming bCOD = 0.8 COD and
 217 TSS = 1.35 VSS; the heterotrophic bacteria concentration expected is approximately 30 mg TSS L⁻¹
 218 in summer and 75 mg TSS L⁻¹ in winter (for both HRT). This then implies an algae biomass
 219 concentration near 290 (HRT 4 days) -320 (HRT 8 days) mg TSS L⁻¹ in summer against 35 (HRT 4
 220 days) -75 (HRT 8 days) mg TSS L⁻¹ in winter. These numbers agree with experimental observations

221 and meteorological data: average microalgal biomass concentration and production were clearly
 222 higher in the warm season than in the cold season in relation with the higher solar radiation (Table
 223 1). In addition, put into evidence that algal activity was clearly lower in winter than in summer,
 224 although enough to produce changes in the DO concentration and the pH of the mixed liquor (Table
 225 1). Biomass concentration was slightly higher in the HRAP with a HRT 8 days where lower flow-
 226 rate gave place to less biomass wash-up, as already observed in other studies [23-25]. Despite this,
 227 biomass production was higher in the HRAP operated at a HRT of 4 days, in these systems biomass
 228 production usually increases inversely with the HRT [26].

229 Biomass production values are in accordance with those previously reported [27]. De Godos et al.
 230 [28] observed a biomass production of 21.3-27.7 g TSS m⁻² d⁻¹ in summer (average daily solar
 231 radiation of 282 W m⁻²) for HRAPs operating at 10 HRT fed with diluted swine manure. In winter
 232 period (average daily solar radiation of 74 W m⁻²) biomass production decreased to 5.7-6.1 g TSS
 233 m⁻² d⁻¹. García et al. [29] using the same HRAPs as in the present study reported a production
 234 between 12.7 and 14.8 m⁻² d⁻¹. The HRAPs' performance (Table 1) was consistent throughout the
 235 experimental period, with removal efficiencies similar to those reported in previous studies for this
 236 pilot plant [8] and others previously reported [27]. COD removal was moderate (66-85%) and its
 237 removal in these systems depends on influent concentration because background concentration
 238 remains around 50-70 mg L⁻¹ [29]. Up to 99% of NH₄-N was removed in the HRAPs in the warm
 239 season at both HRTs, whereas the removal rate was lower and different at both HRTs (90 vs 98%) in
 240 the cold season. Hence, environmental conditions (i.e. temperature and solar radiation) played an
 241 important role in NH₄-N removal, whereas HRT was only relevant in the cold season. Mechanisms
 242 for nitrogen removal have been studied in detail in the past and the most predominant include
 243 volatilisation, biological uptake and nitrification [27, 30]. NH₄-N removal values were similar to
 244 those found in the literature. De Godos et al. [28] found in summer a COD and NH₄-N removal of
 245 76 and 96% respectively, whereas in winter those removal decreased to 57 and 92% for HRAPs

operating at a HRT of 10 days. Sutherland et al. [31] found a decrease on $\text{NH}_4\text{-N}$ removal between summer (77%) and winter (53%) in a HRAP operating at a HRT of 4 and 9 days respectively.

3.2. Occurrence and removal efficiency of EOCs

For this study, we selected the emerging contaminants with the highest concentrations that are most often detected in raw wastewaters [32]. The concentration of EOCs in HRAP influent (primary effluent) ranged from undetected to $24 \mu\text{g L}^{-1}$ (Fig. 2). Caffeine, acetaminophen and ibuprofen were usually detected at concentrations higher than $9 \mu\text{g L}^{-1}$, in keeping with the reported literature [33]. Although the influent COD showed a high seasonal variability (Table 1), the concentration of the studied EOCs showed no statistically significant seasonal difference ($p=0.84$). This may be explained by the presence of a primary treatment that favours a more constant influent wastewater in terms of quality, as can be seen in the low variability of the EOC concentrations (Fig. 2).

Table 2 shows the removal efficiencies of the target EOCs. They can be classified into four groups in accordance with the corresponding overall average removal efficiency in the HRAPs: high removal ($>90\%$: caffeine, acetaminophen, ibuprofen, methyl dihydrojasmonate and hydrocinnamic acid), moderate-to-high removal (from 60% to 90%: oxybenzone, ketoprofen, 5-methyl/benzotriazole, naproxen, galaxolide, tonalide, tributyl phosphate, triclosan, bisphenol A and octylphenol), moderate-to-low removal (from 40 to 60%: diclofenac, benzotriazole, OH-benzothiazole, triphenyl phosphate, cashmeran, diazinon, benzothiazole, celestolide, 2,4-D and atrazine) and poor or no removal ($<40\%$, carbamazepine, methyl paraben, tris(2-chloroethyl) phosphate).

Taking into account the configuration of the HRAPs, the most relevant removal processes that may occur in these systems can be biodegradation, photodegradation, volatilization and sorption to microalgae biomass. Uptake by microalgae is an important removal process, and it was assessed by analysing the occurrence of EOCs in the TSS (solids retained in the filters). Table 1 in the Supplementary Material (SM) shows that the most abundant compounds in the biomass (mostly

271 microalgae) were the most hydrophobic ones, such as galaxolide and tonalide ($\log K_{ow} > 5$). Hence,
 272 HRAPs may remove hydrophobic compounds by sorption, similarly to other wastewater treatment
 273 technologies such as constructed wetlands (CWs) and activated sludge systems [34, 35]. Despite the
 274 higher concentration of TSS in the HRAPs during the warm season, due to the greater biomass
 275 production and evaporation losses (Table 1), the concentration of musk fragrances in the TSS was
 276 higher in winter. Therefore, it may be postulated that the increase in biomass (microalgae,
 277 heterotrophs and non-photosynthetic autotrophs organisms) improved the biodegradability of these
 278 compounds or that the higher sunlight irradiation and temperatures in warm season improved the
 279 volatilization rates. This is in keeping with the moderate biodegradability found for musk fragrances
 280 ($> 75\%$) in lab-scale activated sludge reactors [36] and the tabulated high Henry's law constants (a
 281 measure of air-water partitioning) for musk fragrances ($11\text{--}12 \text{ Pa m}^{-3} \text{ mol}^{-1}$). The occurrence of most
 282 of the studied EOCs in the filters was below their LOD. This may be due to the fact that these EOCs
 283 were not uptaken by microalgae or because they were removed by microalgal metabolism. A
 284 microalgae removal effect due to the release of exudates likewise cannot be disregarded [37]. In
 285 fact, it has been proved that the consortia of cyanobacteria/microalgae and bacteria can be efficient
 286 in detoxification of organic and inorganic pollutants, and removal of nutrients from wastewaters,
 287 compared to the individual microorganisms. Cyanobacterial/algal photosynthesis provides oxygen
 288 and organic exudates that serves to the pollutant-degrading heterotrophic bacteria [38].

289 The overall average removal efficiencies of the studied EOCs were plotted against their
 290 physicochemical properties ($\log K_{ow}$, molecular weight (MW) and Henry's law constant) as is
 291 shown in Fig. 1 SM. Although the plots seem to show a relationship between the EOC removal
 292 efficiencies and MW (Spearman's correlation coefficient = -0.197), $\log K_{ow}$ (Spearman's
 293 correlation coefficient = -0.080) and Henry's law constant (Spearman's correlation coefficient = $-$
 294 0.075), no significant correlations were found (significance level > 0.05). This may be explained by
 295 the complexity of the chemical compounds studied as well as the fact that different removal

processes occurred simultaneously. Conversely, a statistically significant relationship between influent concentration and removal efficiency was obtained (Spearman's correlation coefficient = 0.627, significance level = 0.002). This may be explained by the fact that biodegradation needs a certain compound concentration before microbial degradation is stimulated. Nevertheless, this general rule must be applied with care and further work is necessary in this field.

The removal efficiency of HRAPs is comparable to that of conventional activated sludge WWTPs, as can be seen in Table 2 (overall removal efficiencies of 84% and 59% in warm and cold season respectively). Hijosa-Valsero et al. [39] found that waste stabilization ponds (WSPs) were capable of moderately removing pharmaceuticals such as naproxen (33%) and ibuprofen (56%). Matamoros et al. [40] found that unsaturated CWs were capable of removing up to 90% of pharmaceuticals and personal care products such as ibuprofen, naproxen and galaxolide, but did not remove carbamazepine. Hence, HRAPs seem to be as or more efficient than other biological wastewater treatment technologies, such as CWs or WSPs, with regard to EOC removal. Therefore, HRAPs can be considered a suitable technology for the treatment of wastewaters containing EOCs, with the added advantage that they produce microalgal biomass, do not require aeration and have smaller land area requirements than other engineered natural wastewater treatment technologies (i.e. CWs and WSPs). Nevertheless, since the effectiveness of engineered natural wastewater treatment technologies for removing EOCs has been shown to rely on different key design and environmental factors, such as HRT and seasonality [41], the next two sections will explore the influence of these factors on HRAPs.

316

3.2.1. *Effect of hydraulic retention time (HRT)*

HRT is a key design parameter for achieving proper removal efficiency of biodegradable organic contaminants from wastewaters engineered natural treatment systems such as constructed wetlands and waste stabilization ponds [33]. In fact, it has already proved that EOC removal in engineered

321 natural treatment systems and activated sludge WWTPs increases as HRT increases due to the
 322 increase of biodegradation and sorption processes [34, 42]. No significant differences in their
 323 performance were observed between HRTs in the warm season ($p>0.05$), but significant differences
 324 were found in the cold season for those compounds that has already been described in the literature
 325 [41] as been removed by biodegradation (i.e. caffeine, 4%; ibuprofen, 7%; methyl
 326 dihydrojasmonate, 5%; oxybenzone, 13%; naproxen, 8% and triphenyl phosphate, 44%),
 327 photodegradation (i.e. ketoprofen; 25% and triclosan, 20%) and sorption or volatilization (i.e.
 328 galaxolide, 24% and tonalide, 16%). As already noted, conventional water quality parameters such
 329 as COD and $\text{NH}_4\text{-N}$ behaved similarly. From these results, it can be postulated that biodegradation,
 330 photodegradation, sorption and volatilization removal mechanism were likely affected by the
 331 increase of HRT in the cold season. García-Rodríguez et al. [41] reported that biological wastewater
 332 treatment technologies for removing EOCs are highly dependent on HRT because it enhances
 333 biodegradation, photodegradation and sorption removal processes. In general, the higher the HRT,
 334 the greater the EOC removal efficiency. However, our results suggest that an HRT of 4 days is
 335 enough to remove most of the compounds in both seasons. Therefore, while this technology is
 336 competitive in terms of HRT compared to CWs and WSPs, activated sludge WWTPs are generally
 337 set at an HRT of 12-24 hours or lower. Notwithstanding the foregoing, activated sludge WWTPs
 338 also have higher energy requirements (0.6 kWh m^{-3} for activated sludge WWTPs vs. 0.02 kWh m^{-3}
 339 for HRAPs). Finally, the lower microcontaminant sorption onto the biomass (table 1-SM) than in
 340 conventional activated sludge WWTPs biosolids [43] is relevant for risk management and sludge
 341 valorisation. This low bioaccumulation of microcontaminants into the biomass have already been
 342 reported for vegetables [44], but this is the first time that it has been assessed for microalgae.

343

344 3.2.2 Seasonality (environmental conditions)

345 Seasonality is relevant to achieving adequate EOC removal efficiency in mild climates such as that

of the NW Mediterranean because it affects temperature, daylight duration and intensity, and biomass production, four important factors influencing biodegradation, photodegradation, volatilization and sorption EOC removal processes [41]. Significant differences ($p < 0.05$) between the warm and cold seasons were observed for the removal of above described biodegradable compounds (i.e. caffeine, ibuprofen, methyl dihydrojasmonate, oxybenzone, naproxen, benzothiazole, methylparaben, benzotriazole, 5-methyl-benzotriazole, OH-benzothiazole, and triphenyl phosphate), photodegradable compounds (i.e. ketoprofen, diclofenac, and triclosan) and highly hydrophobic / moderately volatile compounds (i.e. galaxolide and tonalide). The higher temperature (11 vs. 26 °C, on a daily average) and greater average daily solar radiation (74 vs. 282 W m^{-2}) in the warm season may explain these differences. It should be noted that the effect of seasonality on the pollutant removal performance of this technology for the most abundant compounds was low (around 10-20%) or null (i.e. for caffeine, acetaminophen, ibuprofen and methyl dihydrojasmonate). In contrast, various authors [45-47] have reported higher seasonal variability for EOC removal by other engineered natural wastewater treatment technologies (CWs or WSPs). Hence, although the HRAP technology seems to be a robust and reliable wastewater treatment technology in terms of EOC removal efficiency, further studies are required to provide more insight.

363

3.3. Aquatic risk assessment

Aquatic risk assessment throughout the HRAP treatment was performed based on the concentrations of the detected EOCs in the influent and effluent water samples, and the listed EC50 values for *Daphnia magna*. Hazard quotient indexes (HQs) were calculated according to the following equation (2):

$$HQ = \frac{MEC}{PNEC} \quad (\text{Equation 2})$$

371

372 where PNEC is the predicted non-effect concentration and MEC is the measured environmental
 373 concentration at the influent or effluent of each HRAP reactor. PNEC values were estimated for
 374 *Daphnia magna*, dividing the EC50 values (48 hours) by a recommended arbitrary safety factor of
 375 1000 [48]. The EC50 values used in this study were collected from the literature and are
 376 summarized in Table 2SM. When more than one EC50 value was reported for a single compound,
 377 the lowest value was used. When no experimental values were available, the EC50 values were
 378 estimated with ECOSAR v1.10 (EPI Suite software, US EPA).

379 Fig. 3 shows the individual HQ for each of the studied EOCs in the HRAP influent and effluent
 380 water samples. As the difference between the EOC removal efficiencies at both of the studied HRTs
 381 was minimal, the risk assessment was only performed at an HRT of 4 days (under the most critical
 382 operating conditions). Acetaminophen, ibuprofen and oxybenzone exhibited higher HQs in influent
 383 wastewater samples ($HQ > 1$), mainly due to their high concentration. Following the treatment in the
 384 HRAP system, all of the studied EOCs had an $HQ < 1$. The most relevant compounds in the treated
 385 wastewater effluents, apart from those observed at the influent, were triclosan and galaxolide. These
 386 HQ values for the EOCs are in keeping with those reported in the literature for treated wastewater
 387 effluents [42]. In fact, triclosan has already been postulated as a critical compound in terms of
 388 contribution and environmental risk in wastewater effluents [49]. Hence, von der Ohe et al. [50]
 389 argued that triclosan should be seriously considered as a candidate for regulatory monitoring and
 390 prioritization on a European scale on the basis of realistic PNECs.

391 As previously described by various authors, HQs should follow an additive model [6]. Hence, the
 392 final HQ for each water sample can be calculated as the sum of each individual HQ. The cumulative
 393 HQs for the influent wastewater samples were 8.45 and 6.20 for the summer and winter campaigns,
 394 respectively. However, they fell 93% (warm season) and 72% (cold season) following the treatment
 395 with the HRAPs. Consequently, in summer the cumulative HQ was less than 1 (0.62), whereas in

396 winter it was significantly higher ($HQ=1.73$). These results are in keeping with the high reduction in
397 acute toxicity achieved by other biological wastewater treatment technologies, such as CWs [51].
398 Nevertheless, this ecological risk assessment was only performed for the target EOCs; therefore,
399 further studies may be needed to include other EOCs and related transformation products.

400

401 **4. Conclusions**

402 This study has shown that microalgae-based wastewater treatment systems (such as HRAPs) enable
403 the removal of a wide range of EOCs from urban wastewater. Removal efficiency ranged from none
404 to up to 99%. The EOCs were classified into four groups in accordance with their average removal
405 efficiency in HRAPs: high removal ($>90\%$: caffeine, acetaminophen, ibuprofen, methyl
406 dihydrojasmonate and hydrocinnamic acid), moderate-high removal (from 60% to 90%:
407 oxybenzone, ketoprofen, 5-methyl/benzotriazole, naproxen, galaxolide, tonalide, tributyl phosphate,
408 triclosan, bisphenol A and octylphenol), moderate-low removal (from 40 to 60%: diclofenac,
409 benzotriazole, OH-benzothiazole, triphenyl phosphate, cashmeran, diazinon, celestolide and
410 atrazine) and poor or no removal ($<30\%$, carbamazepine, benzothiazole, methyl paraben, tris(2-
411 chloroethyl) phosphate, and 2,4-D). The removal of emerging contaminants in HRAPs was only
412 affected by the HRT during the cold season, whereas no differences were observed in the warm
413 season. The most frequently occurring compounds (caffeine, acetaminophen and ibuprofen) had
414 removal efficiencies of up to 90% that were minimally affected by seasonality and HRT. The
415 ecotoxicological risk assessment study revealed that the HQ for the influent wastewater was
416 removed by up to 90%, indicating no acute toxicity risk associated with the studied EOCs at the
417 water effluents.

418

419 **5. Acknowledgments**

420 The authors gratefully acknowledge the financial support of the Spanish Ministry of Economy and
 421 Competitiveness through projects CTM2012-33547 and CTM2012-37860 (DIPROBIO). Dr V. M.
 422 would like to acknowledge a JAE-Doc contract from the Spanish National Research Council (CSIC)
 423 and the European Social Fund.

424

425 6. References

- 426 [1] K.E. Murray, S.M. Thomas, A.A. Bodour, Prioritizing research for trace pollutants and emerging
 427 contaminants in the freshwater environment, *Environ. Pollut.*, 158 (2010) 3462-3471.
- 428 [2] T.A. Ternes, A. Joss, H. Siegrist, Scrutinizing pharmaceuticals and personal care products in
 429 wastewater treatment, *Environ. Sci. Technol.*, 38 (2004) 392A-399A.
- 430 [3] I. Muñoz, J.C. López-Doval, M. Ricart, M. Villagrasa, R. Brix, A. Geiszinger, A. Ginebreda, H.
 431 Guasch, M.J.L. de Alda, A.M. Romaní, S. Sabater, D. Barceló, Bridging levels of
 432 pharmaceuticals in river water with biological community structure in the Llobregat river basin
 433 (northeast Spain), *Environ. Toxicol. Chem.*, 28 (2009) 2706-2714.
- 434 [4] T. Henry, M. Black, Acute and Chronic Toxicity of Fluoxetine (Selective Serotonin Reuptake
 435 Inhibitor) in Western Mosquitofish, *Arch. Environ. Contam. Toxicol.*, 54 (2008) 325-330.
- 436 [5] A.M. Vajda, L.B. Barber, J.L. Gray, E.M. Lopez, J.D. Woodling, D.O. Norris, Reproductive
 437 Disruption in Fish Downstream from an Estrogenic Wastewater Effluent, *Environ. Sci.*
 438 *Technol.*, 42 (2008) 3407-3414.
- 439 [6] A. Ginebreda, I. Muñoz, M.L. de Alda, R. Brix, J. López-Doval, D. Barceló, Environmental risk
 440 assessment of pharmaceuticals in rivers: Relationships between hazard indexes and aquatic
 441 macroinvertebrate diversity indexes in the Llobregat River (NE Spain), *Environ. Int.*, 36 (2010)
 442 153-162.
- 443 [7] R. Craggs, D. Sutherland, H. Campbell, Hectare-scale demonstration of high rate algal ponds for
 444 enhanced wastewater treatment and biofuel production, *J. Appl. Phycol.*, 24 (2012) 329-337.
- 445 [8] F. Passos, H.-M. Mariona, J. García, I. Ferrer, Long-term anaerobic digestion of microalgae
 446 grown in HRAP for wastewater treatment. Effect of microwave pretreatment, *Water Res.*, 49
 447 (2014) 351-359.
- 448 [9] B. El Hamouri, K. Khallayoune, K. Bouzoubaa, N. Rhallabi, M. Chalabi, High-rate algal pond
 449 performances in faecal coliforms and helminth egg removals, *Water Res.*, 28 (1994) 171-174.
- 450 [10] R. Muñoz, B. Guieysse, Algal-bacterial processes for the treatment of hazardous contaminants:
 451 A review, *Water Res.*, 40 (2006) 2799-2815.
- 452 [11] N. Abdel-Raouf, A.A. Al-Homaidan, I.B.M. Ibraheem, Microalgae and wastewater treatment,
 453 *Saudi J. Biol. Sci.*, 19 (2012) 257-275.
- 454 [12] K. Lika, I.A. Papadakis, Modeling the biodegradation of phenolic compounds by microalgae, *J.*
 455 *Sea Res.*, 62 (2009) 135-146.
- 456 [13] I. de Godos, R. Muñoz, B. Guieysse, Tetracycline removal during wastewater treatment in
 457 high-rate algal ponds, *J. Hazard. Mater.*, 229-230 (2012) 446-449.
- 458 [14] M.R. Abargues, J. Ferrer, A. Bouzas, A. Seco, Removal and fate of endocrine disruptors
 459 chemicals under lab-scale posttreatment stage. Removal assessment using light, oxygen and
 460 microalgae, *Bioresour. Technol.*, 149 (2013) 142-148.
- 461 [15] A.K. Haritash, C.P. Kaushik, Biodegradation aspects of Polycyclic Aromatic Hydrocarbons
 462 (PAHs): A review, *J. Hazard. Mater.*, 169 (2009) 1-15.

- 463 [16] L.E. de-Bashan, Y. Bashan, Immobilized microalgae for removing pollutants: Review of
464 practical aspects, *Bioresour. Technol.*, 101 (2010) 1611-1627.
- 465 [17] J.J. Lee, G.F. Leedale, P. Bradbury, *The illustrated guide to the protozoa*, Second edition,
466 Wiley-Blackwell, 2000.
- 467 [18] D.M. John, B.A. Whitton, A.J. Brook, *The Freshwater Algal Flora of the British Isles: An*
468 *Identification Guide to Freshwater and Terrestrial Algae*, 2nd edition ed., Cambridge University
469 Press, 2011.
- 470 [19] V. Matamoros, J.M. Bayona, Elimination of pharmaceuticals and personal care products in
471 subsurface flow constructed wetlands, *Environ. Sci. Technol.*, 40 (2006) 5811-5816.
- 472 [20] V. Matamoros, E. Jover, J.M. Bayona, Part-per-Trillion determination of pharmaceuticals,
473 pesticides, and related organic contaminants in river water by solid-phase extraction followed
474 by comprehensive two-dimensional gas chromatography time-of-flight mass spectrometry,
475 *Anal. Chem.*, 82 (2010) 699-706.
- 476 [21] A. Pedescoll, A. Corzo, E. Álvarez, J. Puigagut, J. García, Contaminant removal efficiency
477 depending on primary treatment and operational strategy in horizontal subsurface flow
478 treatment wetlands, *Ecol. Eng.*, 37 (2011) 372-380.
- 479 [22] M.a.E. Inc., *Wastewater Engineering. Treatment and Reuse*, 4th ed., McGraw-Hill, New York,
480 2003.
- 481 [23] Z. Arbib, J. Ruiz, P. Álvarez-Díaz, C. Garrido-Pérez, J. Barragan, J. Perales, Long term
482 outdoor operation of a tubular airlift pilot photobioreactor and a high rate algal pond as tertiary
483 treatment of urban wastewater, *Ecol. Eng.*, 52 (2013) 143-153.
- 484 [24] J.B.K. Park, R.J. Craggs, Wastewater treatment and algal production in high rate algal ponds
485 with carbon dioxide addition, *Water Sci. Technol.*, (2010) 633-639.
- 486 [25] D.L. Sutherland, M.H. Turnbull, R.J. Craggs, Increased pond depth improves algal productivity
487 and nutrient removal in wastewater treatment high rate algal ponds, *Water Res.*, 53 (2014) 271-
488 281.
- 489 [26] Y. Azov, G. Shelef, Operation of high-rate oxidation ponds: theory and experiments, *Water*
490 *Res.*, 16 (1982) 1153-1160.
- 491 [27] J.B.K. Park, R.J. Craggs, Nutrient removal in wastewater treatment high rate algal ponds with
492 carbon dioxide addition, *Water Sci. Technol.*, 63 (2011) 1758-1764.
- 493 [28] I.d. Godos, S. Blanco, P.A. García-Encina, E. Becares, R. Muñoz, Long-term operation of high
494 rate algal ponds for the bioremediation of piggery wastewaters at high loading rates, *Bioresour.*
495 *Technol.*, 100 (2009) 4332-4339.
- 496 [29] J. García, B.F. Green, T. Lundquist, R. Mujeriego, M. Hernández-Mariné, W.J. Oswald, Long
497 term diurnal variations in contaminant removal in high rate ponds treating urban wastewater,
498 *Bioresour. Technol.*, 97 (2006) 1709-1715.
- 499 [30] J. García, R. Mujeriego, M. Hernández-Mariné, High rate algal pond operating strategies for
500 urban wastewater nitrogen removal, *J. Appl. Phycol.*, 12 (2000) 331-339.
- 501 [31] D.L. Sutherland, C. Howard-Williams, M.H. Turnbull, P.A. Broady, R.J. Craggs, Seasonal
502 variation in light utilisation, biomass production and nutrient removal by wastewater
503 microalgae in a full-scale high-rate algal pond, *J. Appl. Phycol.*, 26 (2014) 1317-1329.
- 504 [32] J.Q. Jiang, Z. Zhou, V.K. Sharma, Occurrence, transportation, monitoring and treatment of
505 emerging micro-pollutants in waste water - A review from global views, *Microchem. J.*, 110
506 (2013) 292-300.
- 507 [33] Y. Luo, W. Guo, H.H. Ngo, L.D. Nghiem, F.I. Hai, J. Zhang, S. Liang, X.C. Wang, A review on
508 the occurrence of micropollutants in the aquatic environment and their fate and removal during
509 wastewater treatment, *Sci. Total Environ.*, 473-474 (2014) 619-641.
- 510 [34] V. Matamoros, J.M. Bayona, D.B.a.S.P.r. Mira Petrovic, Chapter 12 - Removal of
511 Pharmaceutical Compounds from Wastewater and Surface Water by Natural Treatments, in:
512 *Comprehensive Analytical Chemistry*, Elsevier, 2013, pp. 409-433.

- 513 [35] K.Y. Bell, J. Bandy, B.J. Finnegan, O. Keen, M.S. Mauter, A.M. Parker, L.C. Sima, H.A.
514 Stretz, Emerging pollutants - Part II: Treatment, *Water Environ. Res.*, 85 (2013) 2022-2071.
- 515 [36] S. Suarez, J.M. Lema, F. Omil, Removal of Pharmaceutical and Personal Care Products
516 (PPCPs) under nitrifying and denitrifying conditions, *Water Res.*, 44 (2010) 3214-3224.
- 517 [37] V.V. Unnithan, A. Unc, G.B. Smith, Mini-review: A priori considerations for bacteria-algae
518 interactions in algal biofuel systems receiving municipal wastewaters, *Algal Res.*, 4 (2014) 35-
519 40.
- 520 [38] S.R. Subashchandrabose, B. Ramakrishnan, M. Megharaj, K. Venkateswarlu, R. Naidu,
521 Consortia of cyanobacteria/microalgae and bacteria: Biotechnological potential, *Biotechnol.*
522 *Adv.*, 29 (2011) 896-907.
- 523 [39] M. Hijosa-Valsero, V. Matamoros, J. MartÍN-Villacorta, E. BÉcares, J.M. Bayona, Assessment
524 of full-scale natural systems for the removal of PPCPs from wastewater in small communities,
525 *Water Res.*, 44 (2010) 1429-1439.
- 526 [40] V. Matamoros, C. Arias, H. Brix, J.M. Bayona, Removal of pharmaceuticals and personal care
527 products (PPCPs) from urban wastewater in a pilot vertical flow constructed wetland and a
528 sand filter, *Environ. Sci. Technol.*, 41 (2007) 8171-8177.
- 529 [41] A. Garcia-Rodríguez, V. Matamoros, C. Fontàs, V. Salvadó, The ability of biologically based
530 wastewater treatment systems to remove emerging organic contaminants-a review, *Environ.*
531 *Sci. Pollut. Res.*, (2014) 1-21.
- 532 [42] M. Gros, M. Petrovic, A. Ginebreda, D. Barceló, Removal of pharmaceuticals during
533 wastewater treatment and environmental risk assessment using hazard indexes, *Environ. Int.*,
534 36 (2010) 15-26.
- 535 [43] A. Garcia-Rodríguez, E. Sagristà, V. Matamoros, C. Fontàs, M. Hidalgo, V. Salvadó,
536 Determination of pharmaceutical compounds in sewage sludge using a standard addition
537 method approach, *Int. J. Environ. Anal. Chem.*, (2014) 1-11.
- 538 [44] X. Wu, J.L. Conkle, F. Ernst, J.J. Gan, Treated Wastewater Irrigation: Uptake of
539 Pharmaceutical and Personal Care Products by Common Vegetables under Field Conditions,
540 *Environ. Sci. Technol.*, (2014).
- 541 [45] C. Reyes-Contreras, M. Hijosa-Valsero, R. Sidrach-Cardona, J.M. Bayona, E. BÉcares,
542 Temporal evolution in PPCP removal from urban wastewater by constructed wetlands of
543 different configuration: A medium-term study, *Chemosphere*, 88 (2012) 161-167.
- 544 [46] M.E. Hoque, F. Cloutier, C. Arcieri, M. McInnes, T. Sultana, C. Murray, P.A. Vanrolleghem,
545 C.D. Metcalfe, Removal of selected pharmaceuticals, personal care products and artificial
546 sweetener in an aerated sewage lagoon, *Sci. Total Environ.*, (2014).
- 547 [47] A. Dordio, A.J.P. Carvalho, D.M. Teixeira, C.B. Dias, A.P. Pinto, Removal of pharmaceuticals
548 in microcosm constructed wetlands using *Typha* spp. and LECA, *Bioresour. Technol.*, 101
549 (2010) 886-892.
- 550 [48] H. Sanderson, D.J. Johnson, T. Reitsma, R.A. Brain, C.J. Wilson, K.R. Solomon, Ranking and
551 prioritization of environmental risks of pharmaceuticals in surface waters, *Regul. Toxicol.*
552 *Pharmacol.*, 39 (2004) 158-183.
- 553 [49] C.I. Kosma, D.A. Lambropoulou, T.A. Albanis, Investigation of PPCPs in wastewater
554 treatment plants in Greece: Occurrence, removal and environmental risk assessment, *Sci. Total*
555 *Environ.*, 466-467 (2014) 421-438.
- 556 [50] P.C. von der Ohe, M. Schmitt-Jansen, J. Slobodnik, W. Brack, Triclosan-the forgotten priority
557 substance?, *Environ. Sci. Pollut. Res.*, 19 (2012) 585-591.
- 558 [51] C. Ávila, V. Matamoros, C. Reyes-Contreras, B. Piña, M. Casado, L. Mita, C. Rivetti, C.
559 Barata, J. García, J.M. Bayona, Attenuation of emerging organic contaminants in a hybrid
560 constructed wetland system under different hydraulic loading rates and their associated
561 toxicological effects in wastewater, *Sci. Total Environ.*, 470-471 (2014) 1272-1280.

Capability of microalgae-based wastewater treatment systems to remove emerging organic contaminants: a pilot-scale study

Víctor Matamoros^{1*}, Raquel Gutiérrez², Ivet Ferrer², Joan García², Josep M Bayona¹

¹Department of Environmental Chemistry, IDAEA-CSIC, c/Jordi Girona, 18-26, E-08034, Barcelona, Spain.

²GEMMA-Group of Environmental Engineering and Microbiology, Department of Hydraulic, Maritime and Environmental Engineering, Universitat Politècnica de Catalunya BarcelonaTech, c/ Jordi Girona, 1-3, Building D1, E-08034, Barcelona, Spain.

**Corresponding author: victor.matamoros@idaea.csic.es*

584 **0. Abstract**

585 The effect of hydraulic retention time (HRT) and seasonality on the removal efficiency of 26
586 organic microcontaminants from urban wastewater was studied in two pilot high-rate algal ponds
587 (HRAPs). The targeted compounds included pharmaceuticals and personal care products, fire
588 retardants, surfactants, anticorrosive agents, pesticides and plasticizers, among others. The pilot
589 plant, which was fed at a surface loading rate of 7-29 g of COD m⁻² d⁻¹, consisted of a
590 homogenisation tank and two parallel lines, each one with a primary settler and an HRAP with a
591 surface area of 1.5 m² and a volume of 0.5 m³. The two HRAPs were operated with different HRTs
592 (4 and 8 d). The removal efficiency ranged from negligible removal to more than 90% depending on
593 the compound. Microcontaminant removal efficiencies were enhanced during the warm season,
594 while the HRT effect on microcontaminant removal was only noticeable in the cold season. Our
595 results suggest that biodegradation and photodegradation are the most important removal pathways,
596 whereas volatilization and sorption were solely achieved for hydrophobic compounds (log K_{ow}>4)
597 with a moderately high Henry's law constant values (11-12 Pa m⁻³ mol⁻¹) such as musk fragrances.
598 Whereas acetaminophen, ibuprofen and oxybenzone presented ecotoxicological hazard quotients
599 (HQs) higher than 1 in the influent wastewater samples, the HQs for the effluent water samples
600 were always below 1.

601

602

603

604

605

606 **Keywords:** *emerging organic contaminants; microalgae; high-rate algal pond; photodegradation;*
607 *biodegradation; volatilization.*

608 1. Introduction

609 Emerging organic contaminants (EOCs) include a wide range of compounds belonging to different
610 chemical classes, such as pharmaceuticals, personal care products, plasticizers, flame retardants,
611 surfactants, and certain pesticides, among others, the ecotoxicological effects of which are relatively
612 unknown [1]. Since conventional wastewater treatment plants (WWTPs) are not designed to remove
613 emerging and related contaminants, many of these compounds occur at different concentrations in
614 natural water bodies [2], where they may exert ecotoxicological effects at relatively low
615 concentrations [3, 4]. Although some of the compounds have been proposed for inclusion on
616 regulatory lists of contaminants (European Commission, 2006), there is relatively little information
617 on the ecotoxicological effects of complex mixtures at environmental levels, and, to date, they have
618 not been regulated [1]. Known environmental effects of some EOCs include the reduction of
619 macroinvertebrate diversity in rivers [3], behavioural changes in mosquito fish [4] and reproductive
620 disruption in fish [5], among others. Due to the difficulty of assessing the effects of EOCs on
621 ecosystems, the use of hazard quotients (HQs) based on the chemical composition of water samples
622 and tabulated predicted non-effect concentrations (PNECs) for different aquatic organisms has been
623 postulated as a good screening strategy [6].

624 Microalgae-based wastewater treatment technologies such as high-rate algal ponds (HRAPs) have
625 received considerable attention in recent years due to the resource recovery of algal biomass, for use
626 as fertilizer, protein-rich feed or biofuel, and a high-quality effluent (treated wastewater)[7]. HRAPs
627 are shallow raceway reactors in which microalgae and bacteria grow in symbiosis. In such systems,
628 organic matter is degraded by heterotrophic bacteria, which consume oxygen provided by
629 microalgal photosynthesis; therefore, no aeration is needed [8]. Although the capability of
630 microalgae wastewater treatment systems to remove nutrients, heavy metals, bacteria, and
631 helminthic eggs has been studied since the 1950s, few studies have focused on the removal of
632 organic contaminants, namely, phenolic compounds, surfactants, biocides and polycyclic aromatic

633 hydrocarbons [9-12]. Indeed, no attention has been paid to the effectiveness of HRAPs for
634 removing EOCs of environmental concern.

635 The removal of EOCs by conventional activated sludge WWTPs has been widely studied, but the
636 effectiveness of HRAPs for removing EOCs from wastewater has not yet been addressed. There is
637 only one study dealing with HRAPs' capacity to remove tetracyclines, and it was performed at
638 laboratory-scale with synthetic wastewater [13]. Other studies dealing with microalgae's capacity to
639 remove organic contaminants, such as polycyclic aromatic hydrocarbons (PAHs), biocides (e.g.
640 organotin compounds), surfactants and phenolic compounds, suggest that microalgae-based
641 wastewater technologies may remove microcontaminants by both abiotic (sorption, volatilization or
642 photodegradation) and biotic (biodegradation, microalgae uptake or metabolization) processes [14-
643 16].

644 The aim of this study was to evaluate for the first time, the effect of hydraulic retention time (HRT)
645 and ambient temperature / sunlight irradiation (seasonality) on the removal efficiency of 26 EOCs
646 in two HRAP pilot plants fed with real urban wastewater. The selected compounds were high
647 production volume chemicals (e.g. fire retardants, surfactants, anticorrosive agents, pesticides,
648 plasticizers, pharmaceuticals and personal care products, among others). Finally, aquatic risk
649 assessment was performed based on the concentrations of the detected EOCs in the influent and
650 effluent water samples, and the listed EC50 values for *Daphnia magna*.

651

652 **2. Material and Methods**

653 *2.1. Chemicals and reagents*

654 Gas chromatography (GC) grade (Suprasolv) hexane, methanol, and ethyl acetate were obtained
655 from Merck (Darmstadt, Germany). Analytical-grade hydrogen chloride was obtained from Panreac
656 (Barcelona, Spain). Caffeine, acetaminophen, ibuprofen, methyl dihydrojasmonate, oxybenzone,

ketoprofen, hydrocinnamic acid, 5-methylbenzotriazole, naproxen, carbamazepine, galaxolide, benzothiazole, diclofenac, methylparaben, benzotriazole, tonalide, OH-benzothiazole, tributyl phosphate, tris(2-chloroethyl) phosphate, triphenyl phosphate, triclosan, cashmeran, octylphenol, diazinon, celestolide, atrazine, bisphenol A, 2,4-D, atrazine D5, mecoprop D3, tonalide D3 and dihydrocarbamazepine were purchased from Sigma-Aldrich (Steinheim, Germany). Trimethylsulfonium hydroxide (TMSH) was obtained from Fluka (Buchs, Switzerland). Strata-X polymeric SPE cartridges (200 mg) were purchased from Phenomenex (Torrance, CA, USA) and the 0.7 μm glass fibre filters (\varnothing 47 mm) were obtained from Whatman (Maidstone, UK).

665

666 2.2. Description of the HRAP pilot plant

The experimental set-up was located outdoors at the laboratory of the GEMMA research group (Universitat Politècnica de Catalunya-BarcelonaTech, Spain). The system has been operated since March 2010. The microalgae production system was composed of a screening pre-treatment and two identical parallel lines, each one equipped with a primary settler, a pilot high-rate algal pond and a final settler for biomass separation (Fig. 1). Paddle wheel was set at 5 rpm giving mixed liquor with a linear velocity of recirculation of 11 cm s^{-1} , enough to ensure complete mixing. Urban wastewater was pumped from a municipal sewer to a homogenisation tank (1.2 m^3), which was continuously stirred to avoid solids sedimentation. From there, the wastewater was pre-treated and conveyed to each line. The primary treatment included a settler with an internal diameter of 0.3 m, a total height of 0.4 m and an effective volume of 7 L that was operated at an HRT of 0.9 h. Primary effluent from the settlers was pumped to the HRAPs by means of peristaltic pumps. The experimental HRAPs were PVC raceway ponds equipped with a paddle wheel for stirring the mixed liquor (Fig.1). The two HRAPs had a nominal volume of 0.47 m^3 , a surface area of 1.54 m^2 and a water depth of 0.3 m, and they were operated simultaneously with different HRTs (4 and 8 days corresponding to 117.5 and 58.8 L d^{-1} respectively). The final settlers for biomass separation had an

682 internal diameter of 0.15 m, a total height of 0.3 m and an effective volume of 3.5 L that were
683 operated at an HRT of 0.7 and 1.4 h for the HRAP set at 4 days HRT and 8 days, respectively. Note
684 that these settlers were only used for biomass separation, which was not recycled back to the
685 HRAPs.

686

687 2.3. Sampling strategy

688 Two sampling campaigns were carried out, one in July 2013 (warm season) and the other in
689 December 2013 (cold season). In each campaign, influent and effluent grab samples were collected
690 from both HRAPs at the same time each day (9:00 am) for a period of 10 days (n=8), from Monday
691 to the Wednesday of the next week, Saturday and Sunday were not sampled. The samples were
692 collected in the primary effluent from the settler and at the effluent from both HRAPs (Fig. 1). No
693 rainfall events were recorded at any time during the sampling period. All water samples were
694 collected in 1000 mL amber glass bottles, which were transported under refrigeration to the
695 laboratory, where they were stored at 4 °C until analysis. The sample holding time was less than 12
696 hours.

697

698 2.4. Analytical procedures

699 Conventional wastewater quality parameters, including ammonium nitrogen ($\text{NH}_4\text{-N}$), total
700 suspended solids (TSS) and chemical oxygen demand (COD), were determined using the Standard
701 Methods (APHA, 2001). Onsite measurements of water temperature, dissolved oxygen (DO) and
702 pH were taken using a Checktemp-1 Hanna thermometer, an Eutech Ecoscan DO6 oxygen meter
703 and a Crison pH-meter, respectively.

704 For each campaign, 2 well-mixed 25 mL samples from each HRAP were examined by light
705 microscopy and the predominant microalgae were identified and quantified. Microalgae genus were
706 identified from classical specific literature [17, 18].

707 All water samples were filtered and processed as previously reported [19]. A 100 mL sample was
708 spiked with 50 ng of a surrogate standard (atrazine D5, mecoprop D3, tonalide D3, and
709 dihydrocarbamazepine). The spiked sample was percolated through a previously activated
710 polymeric solid-phase extraction cartridge (200 mg Strata X). Elution was performed with 10 mL of
711 hexane/ethyl acetate (1:1). The eluted extract was evaporated under a gentle nitrogen stream until
712 ca. 100 μ L remained, at which point 20 ng of triphenylamine was added as an internal standard.
713 Finally, the vial was reconstituted to 300 μ L with ethyl acetate.

714 The TSS collected in the glass fibre filters (0.7 μ m) were processed according to a previously
715 reported analytical method [19]. Briefly, the filters were freeze-dried and extracted in an ultrasonic
716 bath with hexane/acetone (3:1) for 15 minutes. The extracts were then further processed as water
717 samples.

718 Methylation of the acidic carboxyl group was performed in a hot GC PTV injector (270 $^{\circ}$ C) by
719 adding 10 μ L of TMSH solution (0.25 mol L⁻¹ in methanol) to a 50 μ L sample before injection.
720 Derivatized samples were analysed into a Bruker 450-GC gas chromatograph coupled to a Bruker
721 320-MS triple quadrupole mass spectrometer (Bruker Daltonics Inc., Billerica, MA, USA) fitted
722 with a 20 m \times 0.18 mm, 0.18 μ m film thickness Sapiens X5-MS capillary column coated with 5%
723 diphenyl 95% dimethyl polysiloxane from Teknokroma (Sant Cugat del Vallès, Spain) operated in
724 the multiple reaction mode (MRM). Validation of the analytical methodology has been described
725 elsewhere [20]. The limit of detection (LOD) and limit of quantification (LOQ) of the analytical
726 methodology were determined (using ultra-pure water) based on the mean background noise plus 3
727 or 10 times the standard deviation of the background noise, respectively. The LOD and LOQ ranged
728 from 1 to 40 ng L⁻¹ and from 3 to 80 ng L⁻¹, respectively. Recoveries and repeatability were always

729 higher than 80% and lower than 20%, respectively.

730

731 2.5. Data analysis

732 The removal efficiencies of conventional water quality parameters and EOCs were calculated as
733 follows (equation 1):

734

735

$$736 \text{ Removal} = \frac{1}{n} \sum_{i=1}^n \frac{C'a - (C_i - C_i \times \frac{EVR}{HLR})}{C'a} \times 100 \quad (\text{Equation 1})$$

737

738 where $C'a$ is the average concentration of a selected compound in the HRAP influents in each
739 sampling campaign, C_i is the concentration in the HRAP effluents on each sampling day, and n is
740 the number of samples collected per sampling campaign ($n=8$). HLR are 83 or 43 L m⁻² d⁻¹ at a HRT
741 of 8 and 4 days, respectively. Evaporation rates (EVRs) are 21 and 9 L m⁻² d⁻¹ in warm and cold
742 season, respectively calculated from Turc's equation.

743

744 The experimental results were statistically evaluated using the SPSS v.13 package (Chicago, IL,
745 USA). According with the data set size, non-parametric statistics were applied. The comparison of
746 means was conducted by means of the Kruskal–Wallis test. Spearman's coefficients were used for
747 correlations between variables (physicochemical parameters, removal efficiencies and influent
748 concentration). Significance was defined as $p < 0.05$.

749

750

751

752

753 3. Results and Discussion

754 3.1 Assessment of conventional water quality parameters

755 The HRAPs' performance was monitored in warm and cold seasons (Table 1). COD, TSS and $\text{NH}_4\text{-}$
 756 N concentrations in the primary settlers' effluents were within the typical range of a conventional
 757 primary effluent [21]. Note that DO concentration was quite high for a primary effluent due to
 758 wastewater stirring in the homogenisation tank. The actual surface organic loading rates (OLRs)
 759 applied to the HRAPs were, on average, 13 and 26 g total COD $\text{m}^{-2} \text{d}^{-1}$ (HRT=8 and 4 d
 760 respectively) in the warm season and 29 and 58 g total COD $\text{m}^{-2} \text{d}^{-1}$ (HRT=8 and 4 d respectively)
 761 in the cold season.

762 The microalgae present in the HRAPs were mostly species of the Phylum *Chlorophyta*, and varied
 763 depending on seasonal conditions. In the warm season, predominant algae species were
 764 *Stigeoclonium* sp. (filamentous algae); diatoms, *Chlorella* sp. and *Monoraphidium* sp. In the cold
 765 season, predominant algae species were *Chlorella* sp.; diatoms, *Stigeoclonium* sp. In both
 766 campaigns, the number of species was higher in the HRAP set at HRT of 8 days.

767 In the present study, mixed liquor TSS was analysed as an indicator of biomass concentration in the
 768 HRAP (Table 1). Note that biomass in such type of systems corresponds to microalgae as well as
 769 bacteria, and the relative proportion of them can be approached by means of mass balances on an
 770 estimated net yield of heterotrophic bacteria [22]. Thus, if the yield for domestic wastewater aerobic
 771 treatment is around 0.25 g VSS/g bCOD (biodegradable COD), assuming bCOD = 0.8 COD and
 772 TSS = 1.35 VSS; the heterotrophic bacteria concentration expected is approximately 30 mg TSS L^{-1}
 773 in summer and 75 mg TSS L^{-1} in winter (for both HRT). This then implies an algae biomass
 774 concentration near 290 (HRT 4 days) -320 (HRT 8 days) mg TSS L^{-1} in summer against 35 (HRT 4
 775 days) -75 (HRT 8 days) mg TSS L^{-1} in winter. These numbers agree with experimental observations
 776 and meteorological data: average microalgal biomass concentration and production were clearly
 777 higher in the warm season than in the cold season in relation with the higher solar radiation (Table

1). In addition, put into evidence that algal activity was clearly lower in winter than in summer, although enough to produce changes in the DO concentration and the pH of the mixed liquor (Table 1). Biomass concentration was slightly higher in the HRAP with a HRT 8 days where lower flow-rate gave place to less biomass wash-up, as already observed in other studies [23-25]. Despite this, biomass production was higher in the HRAP operated at a HRT of 4 days, in these systems biomass production usually increases inversely with the HRT [26].

Biomass production values are in accordance with those previously reported [27]. De Godos et al. [28] observed a biomass production of 21.3-27.7 g TSS m⁻² d⁻¹ in summer (average daily solar radiation of 282 W m⁻²) for HRAPs operating at 10 HRT fed with diluted swine manure. In winter period (average daily solar radiation of 74 W m⁻²) biomass production decreased to 5.7-6.1 g TSS m⁻² d⁻¹. García et al. [29] using the same HRAPs as in the present study reported a production between 12.7 and 14.8 m⁻² d⁻¹. The HRAPs' performance (Table 1) was consistent throughout the experimental period, with removal efficiencies similar to those reported in previous studies for this pilot plant [8] and others previously reported [27]. COD removal was moderate (66-85%) and its removal in these systems depends on influent concentration because background concentration remains around 50-70 mg L⁻¹ [29]. Up to 99% of NH₄-N was removed in the HRAPs in the warm season at both HRTs, whereas the removal rate was lower and different at both HRTs (90 vs 98%) in the cold season. Hence, environmental conditions (i.e. temperature and solar radiation) played an important role in NH₄-N removal, whereas HRT was only relevant in the cold season. Mechanisms for nitrogen removal have been studied in detail in the past and the most predominant include volatilisation, biological uptake and nitrification [27, 30]. NH₄-N removal values were similar to those found in the literature. De Godos et al. [28] found in summer a COD and NH₄-N removal of 76 and 96% respectively, whereas in winter those removal decreased to 57 and 92% for HRAPs operating at a HRT of 10 days. Sutherland et al. [31] found a decrease on NH₄-N removal between summer (77%) and winter (53%) in a HRAP operating at a HRT of 4 and 9 days respectively.

803 3.2. Occurrence and removal efficiency of EOCs

804 For this study, we selected the emerging contaminants with the highest concentrations that are most
 805 often detected in raw wastewaters [32]. The concentration of EOCs in HRAP influent (primary
 806 effluent) ranged from undetected to $24 \mu\text{g L}^{-1}$ (Fig. 2). Caffeine, acetaminophen and ibuprofen were
 807 usually detected at concentrations higher than $9 \mu\text{g L}^{-1}$, in keeping with the reported literature [33].
 808 Although the influent COD showed a high seasonal variability (Table 1), the concentration of the
 809 studied EOCs showed no statistically significant seasonal difference ($p=0.84$). This may be
 810 explained by the presence of a primary treatment that favours a more constant influent wastewater
 811 in terms of quality, as can be seen in the low variability of the EOC concentrations (Fig. 2).

812 Table 2 shows the removal efficiencies of the target EOCs. They can be classified into four groups
 813 in accordance with the corresponding overall average removal efficiency in the HRAPs: high
 814 removal ($>90\%$: caffeine, acetaminophen, ibuprofen, methyl dihydrojasmonate and hydrocinnamic
 815 acid), moderate-to-high removal (from 60% to 90%: oxybenzone, ketoprofen, 5-
 816 methyl/benzotriazole, naproxen, galaxolide, tonalide, tributyl phosphate, triclosan, bisphenol A and
 817 octylphenol), moderate-to-low removal (from 40 to 60%: diclofenac, benzotriazole, OH-
 818 benzothiazole, triphenyl phosphate, cashmeran, diazinon, benzothiazole, celestolide, 2,4-D and
 819 atrazine) and poor or no removal ($<40\%$, carbamazepine, methyl paraben, tris(2-chloroethyl)
 820 phosphate).

821 Taking into account the configuration of the HRAPs, the most relevant removal processes that may
 822 occur in these systems can be biodegradation, photodegradation, volatilization and sorption to
 823 microalgae biomass. Uptake by microalgae is an important removal process, and it was assessed by
 824 analysing the occurrence of EOCs in the TSS (solids retained in the filters). Table 1 in the
 825 Supplementary Material (SM) shows that the most abundant compounds in the biomass (mostly
 826 microalgae) were the most hydrophobic ones, such as galaxolide and tonalide ($\log K_{ow}>5$). Hence,
 827 HRAPs may remove hydrophobic compounds by sorption, similarly to other wastewater treatment

828 technologies such as constructed wetlands (CWs) and activated sludge systems [34, 35]. Despite the
829 higher concentration of TSS in the HRAPs during the warm season, due to the greater biomass
830 production and evaporation losses (Table 1), the concentration of musk fragrances in the TSS was
831 higher in winter. Therefore, it may be postulated that the increase in biomass (microalgae,
832 heterotrophs and non-photosynthetic autotrophs organisms) improved the biodegradability of these
833 compounds or that the higher sunlight irradiation and temperatures in warm season improved the
834 volatilization rates. This is in keeping with the moderate biodegradability found for musk fragrances
835 (>75%) in lab-scale activated sludge reactors [36] and the tabulated high Henry's law constants (a
836 measure of air-water partitioning) for musk fragrances ($11\text{-}12\text{ Pa m}^{-3}\text{ mol}^{-1}$). The occurrence of most
837 of the studied EOCs in the filters was below their LOD. This may be due to the fact that these EOCs
838 were not uptaken by microalgae or because they were removed by microalgal metabolism. A
839 microalgae removal effect due to the release of exudates likewise cannot be disregarded [37]. In
840 fact, it has been proved that the consortia of cyanobacteria/microalgae and bacteria can be efficient
841 in detoxification of organic and inorganic pollutants, and removal of nutrients from wastewaters,
842 compared to the individual microorganisms. Cyanobacterial/algal photosynthesis provides oxygen
843 and organic exudates that serves to the pollutant-degrading heterotrophic bacteria [38].

844 The overall average removal efficiencies of the studied EOCs were plotted against their
845 physicochemical properties (log K_{ow} , molecular weight (MW) and Henry's law constant) as is
846 shown in Fig. 1 SM. Although the plots seem to show a relationship between the EOC removal
847 efficiencies and MW (Spearman's correlation coefficient = -0.197), log K_{ow} (Spearman's
848 correlation coefficient = -0.080) and Henry's law constant (Spearman's correlation coefficient = -
849 0.075), no significant correlations were found (significance level >0.05). This may be explained by
850 the complexity of the chemical compounds studied as well as the fact that different removal
851 processes occurred simultaneously. Conversely, a statistically significant relationship between
852 influent concentration and removal efficiency was obtained (Spearman's correlation coefficient =

0.627, significance level = 0.002). This may be explained by the fact that biodegradation needs a certain compound concentration before microbial degradation is stimulated. Nevertheless, this general rule must be applied with care and further work is necessary in this field.

The removal efficiency of HRAPs is comparable to that of conventional activated sludge WWTPs, as can be seen in Table 2 (overall removal efficiencies of 84% and 59% in warm and cold season respectively). Hijosa-Valsero et al. [39] found that waste stabilization ponds (WSPs) were capable of moderately removing pharmaceuticals such as naproxen (33%) and ibuprofen (56%). Matamoros et al. [40] found that unsaturated CWs were capable of removing up to 90% of pharmaceuticals and personal care products such as ibuprofen, naproxen and galaxolide, but did not remove carbamazepine. Hence, HRAPs seem to be as or more efficient than other biological wastewater treatment technologies, such as CWs or WSPs, with regard to EOC removal. Therefore, HRAPs can be considered a suitable technology for the treatment of wastewaters containing EOCs, with the added advantage that they produce microalgal biomass, do not require aeration and have smaller land area requirements than other engineered natural wastewater treatment technologies (i.e. CWs and WSPs). Nevertheless, since the effectiveness of engineered natural wastewater treatment technologies for removing EOCs has been shown to rely on different key design and environmental factors, such as HRT and seasonality [41], the next two sections will explore the influence of these factors on HRAPs.

3.2.1. *Effect of hydraulic retention time (HRT)*

HRT is a key design parameter for achieving proper removal efficiency of biodegradable organic contaminants from wastewaters engineered natural treatment systems such as constructed wetlands and waste stabilization ponds [33]. In fact, it has already proved that EOC removal in engineered natural treatment systems and activated sludge WWTPs increases as HRT increases due to the increase of biodegradation and sorption processes [34, 42]. No significant differences in their

performance were observed between HRTs in the warm season ($p>0.05$), but significant differences were found in the cold season for those compounds that has already been described in the literature [41] as been removed by biodegradation (i.e. caffeine, 4%; ibuprofen, 7%; methyl dihydrojasmonate, 5%; oxybenzone, 13%; naproxen, 8% and triphenyl phosphate, 44%), photodegradation (i.e. ketoprofen; 25% and triclosan, 20%) and sorption or volatilization (i.e. galaxolide, 24% and tonalide, 16%). As already noted, conventional water quality parameters such as COD and $\text{NH}_4\text{-N}$ behaved similarly. From these results, it can be postulated that biodegradation, photodegradation, sorption and volatilization removal mechanism were likely affected by the increase of HRT in the cold season. García-Rodríguez et al. [41] reported that biological wastewater treatment technologies for removing EOCs are highly dependent on HRT because it enhances biodegradation, photodegradation and sorption removal processes. In general, the higher the HRT, the greater the EOC removal efficiency. However, our results suggest that an HRT of 4 days is enough to remove most of the compounds in both seasons. Therefore, while this technology is competitive in terms of HRT compared to CWs and WSPs, activated sludge WWTPs are generally set at an HRT of 12-24 hours or lower. Notwithstanding the foregoing, activated sludge WWTPs also have higher energy requirements (0.6 kWh m^{-3} for activated sludge WWTPs vs. 0.02 kWh m^{-3} for HRAPs). Finally, the lower microcontaminant sorption onto the biomass (table 1-SM) than in conventional activated sludge WWTPs biosolids [43] is relevant for risk management and sludge valorisation. This low bioaccumulation of microcontaminants into the biomass have already been reported for vegetables [44], but this is the first time that it has been assessed for microalgae.

898

899 3.2.2 Seasonality (environmental conditions)

Seasonality is relevant to achieving adequate EOC removal efficiency in mild climates such as that of the NW Mediterranean because it affects temperature, daylight duration and intensity, and biomass production, four important factors influencing biodegradation, photodegradation,

903 volatilization and sorption EOC removal processes [41]. Significant differences ($p < 0.05$) between
 904 the warm and cold seasons were observed for the removal of above described biodegradable
 905 compounds (i.e. caffeine, ibuprofen, methyl dihydrojasmonate, oxybenzone, naproxen,
 906 benzothiazole, methylparaben, benzotriazole, 5-methyl-benzotriazole, OH-benzothiazole, and
 907 triphenyl phosphate), photodegradable compounds (i.e. ketoprofen, diclofenac, and triclosan) and
 908 highly hydrophobic / moderately volatile compounds (i.e. galaxolide and tonalide). The higher
 909 temperature (11 vs. 26 °C, on a daily average) and greater average daily solar radiation (74 vs. 282
 910 W m^{-2}) in the warm season may explain these differences. It should be noted that the effect of
 911 seasonality on the pollutant removal performance of this technology for the most abundant
 912 compounds was low (around 10-20%) or null (i.e. for caffeine, acetaminophen, ibuprofen and
 913 methyl dihydrojasmonate). In contrast, various authors [45-47] have reported higher seasonal
 914 variability for EOC removal by other engineered natural wastewater treatment technologies (CWs
 915 or WSPs). Hence, although the HRAP technology seems to be a robust and reliable wastewater
 916 treatment technology in terms of EOC removal efficiency, further studies are required to provide
 917 more insight.

918

919 3.3. Aquatic risk assessment

920 Aquatic risk assessment throughout the HRAP treatment was performed based on the
 921 concentrations of the detected EOCs in the influent and effluent water samples, and the listed EC50
 922 values for *Daphnia magna*. Hazard quotient indexes (HQs) were calculated according to the
 923 following equation (2):

$$924 \quad HQ = \frac{MEC}{PNEC} \quad (Equation$$

925 2)

926

927 where PNEC is the predicted non-effect concentration and MEC is the measured environmental

concentration at the influent or effluent of each HRAP reactor. PNEC values were estimated for *Daphnia magna*, dividing the EC50 values (48 hours) by a recommended arbitrary safety factor of 1000 [48]. The EC50 values used in this study were collected from the literature and are summarized in Table 2SM. When more than one EC50 value was reported for a single compound, the lowest value was used. When no experimental values were available, the EC50 values were estimated with ECOSAR v1.10 (EPI Suite software, US EPA).

Fig. 3 shows the individual HQ for each of the studied EOCs in the HRAP influent and effluent water samples. As the difference between the EOC removal efficiencies at both of the studied HRTs was minimal, the risk assessment was only performed at an HRT of 4 days (under the most critical operating conditions). Acetaminophen, ibuprofen and oxybenzone exhibited higher HQs in influent wastewater samples ($HQ > 1$), mainly due to their high concentration. Following the treatment in the HRAP system, all of the studied EOCs had an $HQ < 1$. The most relevant compounds in the treated wastewater effluents, apart from those observed at the influent, were triclosan and galaxolide. These HQ values for the EOCs are in keeping with those reported in the literature for treated wastewater effluents [42]. In fact, triclosan has already been postulated as a critical compound in terms of contribution and environmental risk in wastewater effluents [49]. Hence, von der Ohe et al. [50] argued that triclosan should be seriously considered as a candidate for regulatory monitoring and prioritization on a European scale on the basis of realistic PNECs.

As previously described by various authors, HQs should follow an additive model [6]. Hence, the final HQ for each water sample can be calculated as the sum of each individual HQ. The cumulative HQs for the influent wastewater samples were 8.45 and 6.20 for the summer and winter campaigns, respectively. However, they fell 93% (warm season) and 72% (cold season) following the treatment with the HRAPs. Consequently, in summer the cumulative HQ was less than 1 (0.62), whereas in winter it was significantly higher ($HQ = 1.73$). These results are in keeping with the high reduction in acute toxicity achieved by other biological wastewater treatment technologies, such as CWs [51].

953 Nevertheless, this ecological risk assessment was only performed for the target EOCs; therefore,
 954 further studies may be needed to include other EOCs and related transformation products.

955

956 **4. Conclusions**

957 This study has shown that microalgae-based wastewater treatment systems (such as HRAPs) enable
 958 the removal of a wide range of EOCs from urban wastewater. Removal efficiency ranged from none
 959 to up to 99%. The EOCs were classified into four groups in accordance with their average removal
 960 efficiency in HRAPs: high removal (>90%: caffeine, acetaminophen, ibuprofen, methyl
 961 dihydrojasmonate and hydrocinnamic acid), moderate-high removal (from 60% to 90%:
 962 oxybenzone, ketoprofen, 5-methyl/benzotriazole, naproxen, galaxolide, tonalide, tributyl phosphate,
 963 triclosan, bisphenol A and octylphenol), moderate-low removal (from 40 to 60%: diclofenac,
 964 benzotriazole, OH-benzothiazole, triphenyl phosphate, cashmeran, diazinon, celestolide and
 965 atrazine) and poor or no removal (<30%, carbamazepine, benzothiazole, methyl paraben, tris(2-
 966 chloroethyl) phosphate, and 2,4-D). The removal of emerging contaminants in HRAPs was only
 967 affected by the HRT during the cold season, whereas no differences were observed in the warm
 968 season. The most frequently occurring compounds (caffeine, acetaminophen and ibuprofen) had
 969 removal efficiencies of up to 90% that were minimally affected by seasonality and HRT. The
 970 ecotoxicological risk assessment study revealed that the HQ for the influent wastewater was
 971 removed by up to 90%, indicating no acute toxicity risk associated with the studied EOCs at the
 972 water effluents.

973

974 **5. Acknowledgments**

975 The authors gratefully acknowledge the financial support of the Spanish Ministry of Economy and
 976 Competitiveness through projects CTM2012-33547 and CTM2012-37860 (DIPROBIO). Dr V. M.

would like to acknowledge a JAE-Doc contract from the Spanish National Research Council (CSIC) and the European Social Fund.

6. References

- [1] K.E. Murray, S.M. Thomas, A.A. Bodour, Prioritizing research for trace pollutants and emerging contaminants in the freshwater environment, *Environ. Pollut.*, 158 (2010) 3462-3471.
- [2] T.A. Ternes, A. Joss, H. Siegrist, Scrutinizing pharmaceuticals and personal care products in wastewater treatment, *Environ. Sci. Technol.*, 38 (2004) 392A-399A.
- [3] I. Muñoz, J.C. López-Doval, M. Ricart, M. Villagrasa, R. Brix, A. Geiszinger, A. Ginebreda, H. Guasch, M.J.L. de Alda, A.M. Romaní, S. Sabater, D. Barceló, Bridging levels of pharmaceuticals in river water with biological community structure in the Llobregat river basin (northeast Spain), *Environ. Toxicol. Chem.*, 28 (2009) 2706-2714.
- [4] T. Henry, M. Black, Acute and Chronic Toxicity of Fluoxetine (Selective Serotonin Reuptake Inhibitor) in Western Mosquitofish, *Arch. Environ. Contam. Toxicol.*, 54 (2008) 325-330.
- [5] A.M. Vajda, L.B. Barber, J.L. Gray, E.M. Lopez, J.D. Woodling, D.O. Norris, Reproductive Disruption in Fish Downstream from an Estrogenic Wastewater Effluent, *Environ. Sci. Technol.*, 42 (2008) 3407-3414.
- [6] A. Ginebreda, I. Muñoz, M.L. de Alda, R. Brix, J. López-Doval, D. Barceló, Environmental risk assessment of pharmaceuticals in rivers: Relationships between hazard indexes and aquatic macroinvertebrate diversity indexes in the Llobregat River (NE Spain), *Environ. Int.*, 36 (2010) 153-162.
- [7] R. Craggs, D. Sutherland, H. Campbell, Hectare-scale demonstration of high rate algal ponds for enhanced wastewater treatment and biofuel production, *J. Appl. Phycol.*, 24 (2012) 329-337.
- [8] F. Passos, H.-M. Mariona, J. García, I. Ferrer, Long-term anaerobic digestion of microalgae grown in HRAP for wastewater treatment. Effect of microwave pretreatment, *Water Res.*, 49 (2014) 351-359.
- [9] B. El Hamouri, K. Khallayoune, K. Bouzoubaa, N. Rhallabi, M. Chalabi, High-rate algal pond performances in faecal coliforms and helminth egg removals, *Water Res.*, 28 (1994) 171-174.
- [10] R. Muñoz, B. Guieysse, Algal-bacterial processes for the treatment of hazardous contaminants: A review, *Water Res.*, 40 (2006) 2799-2815.
- [11] N. Abdel-Raouf, A.A. Al-Homaidan, I.B.M. Ibraheem, Microalgae and wastewater treatment, *Saudi J. Biol. Sci.*, 19 (2012) 257-275.
- [12] K. Lika, I.A. Papadakis, Modeling the biodegradation of phenolic compounds by microalgae, *J. Sea Res.*, 62 (2009) 135-146.
- [13] I. de Godos, R. Muñoz, B. Guieysse, Tetracycline removal during wastewater treatment in high-rate algal ponds, *J. Hazard. Mater.*, 229-230 (2012) 446-449.
- [14] M.R. Abargues, J. Ferrer, A. Bouzas, A. Seco, Removal and fate of endocrine disruptors chemicals under lab-scale posttreatment stage. Removal assessment using light, oxygen and microalgae, *Bioresour. Technol.*, 149 (2013) 142-148.
- [15] A.K. Haritash, C.P. Kaushik, Biodegradation aspects of Polycyclic Aromatic Hydrocarbons (PAHs): A review, *J. Hazard. Mater.*, 169 (2009) 1-15.
- [16] L.E. de-Bashan, Y. Bashan, Immobilized microalgae for removing pollutants: Review of practical aspects, *Bioresour. Technol.*, 101 (2010) 1611-1627.
- [17] J.J. Lee, G.F. Leedale, P. Bradbury, *The illustrated guide to the protozoa*, Second edition, Wiley-Blackwell, 2000.

- [18] D.M. John, B.A. Whitton, A.J. Brook, *The Freshwater Algal Flora of the British Isles: An Identification Guide to Freshwater and Terrestrial Algae*, 2nd edition ed., Cambridge University Press, 2011.
- [19] V. Matamoros, J.M. Bayona, Elimination of pharmaceuticals and personal care products in subsurface flow constructed wetlands, *Environ. Sci. Technol.*, 40 (2006) 5811-5816.
- [20] V. Matamoros, E. Jover, J.M. Bayona, Part-per-Trillion determination of pharmaceuticals, pesticides, and related organic contaminants in river water by solid-phase extraction followed by comprehensive two-dimensional gas chromatography time-of-flight mass spectrometry, *Anal. Chem.*, 82 (2010) 699-706.
- [21] A. Pedescoll, A. Corzo, E. Álvarez, J. Puigagut, J. García, Contaminant removal efficiency depending on primary treatment and operational strategy in horizontal subsurface flow treatment wetlands, *Ecol. Eng.*, 37 (2011) 372-380.
- [22] M.a.E. Inc., *Wastewater Engineering. Treatment and Reuse*, 4th ed., McGraw-Hill, New York, 2003.
- [23] Z. Arbib, J. Ruiz, P. Álvarez-Díaz, C. Garrido-Pérez, J. Barragan, J. Perales, Long term outdoor operation of a tubular airlift pilot photobioreactor and a high rate algal pond as tertiary treatment of urban wastewater, *Ecol. Eng.*, 52 (2013) 143-153.
- [24] J.B.K. Park, R.J. Craggs, Wastewater treatment and algal production in high rate algal ponds with carbon dioxide addition, *Water Sci. Technol.*, (2010) 633-639.
- [25] D.L. Sutherland, M.H. Turnbull, R.J. Craggs, Increased pond depth improves algal productivity and nutrient removal in wastewater treatment high rate algal ponds, *Water Res.*, 53 (2014) 271-281.
- [26] Y. Azov, G. Shelef, Operation of high-rate oxidation ponds: theory and experiments, *Water Res.*, 16 (1982) 1153-1160.
- [27] J.B.K. Park, R.J. Craggs, Nutrient removal in wastewater treatment high rate algal ponds with carbon dioxide addition, *Water Sci. Technol.*, 63 (2011) 1758-1764.
- [28] I.d. Godos, S. Blanco, P.A. García-Encina, E. Becares, R. Muñoz, Long-term operation of high rate algal ponds for the bioremediation of piggery wastewaters at high loading rates, *Bioresour. Technol.*, 100 (2009) 4332-4339.
- [29] J. García, B.F. Green, T. Lundquist, R. Mujeriego, M. Hernández-Mariné, W.J. Oswald, Long term diurnal variations in contaminant removal in high rate ponds treating urban wastewater, *Bioresour. Technol.*, 97 (2006) 1709-1715.
- [30] J. García, R. Mujeriego, M. Hernández-Mariné, High rate algal pond operating strategies for urban wastewater nitrogen removal, *J. Appl. Phycol.*, 12 (2000) 331-339.
- [31] D.L. Sutherland, C. Howard-Williams, M.H. Turnbull, P.A. Broady, R.J. Craggs, Seasonal variation in light utilisation, biomass production and nutrient removal by wastewater microalgae in a full-scale high-rate algal pond, *J. Appl. Phycol.*, 26 (2014) 1317-1329.
- [32] J.Q. Jiang, Z. Zhou, V.K. Sharma, Occurrence, transportation, monitoring and treatment of emerging micro-pollutants in waste water - A review from global views, *Microchem. J.*, 110 (2013) 292-300.
- [33] Y. Luo, W. Guo, H.H. Ngo, L.D. Nghiem, F.I. Hai, J. Zhang, S. Liang, X.C. Wang, A review on the occurrence of micropollutants in the aquatic environment and their fate and removal during wastewater treatment, *Sci. Total Environ.*, 473-474 (2014) 619-641.
- [34] V. Matamoros, J.M. Bayona, D.B.a.S.Pr. Mira Petrovic, Chapter 12 - Removal of Pharmaceutical Compounds from Wastewater and Surface Water by Natural Treatments, in: *Comprehensive Analytical Chemistry*, Elsevier, 2013, pp. 409-433.
- [35] K.Y. Bell, J. Bandy, B.J. Finnegan, O. Keen, M.S. Mauter, A.M. Parker, L.C. Sima, H.A. Stretz, Emerging pollutants - Part II: Treatment, *Water Environ. Res.*, 85 (2013) 2022-2071.
- [36] S. Suarez, J.M. Lema, F. Omil, Removal of Pharmaceutical and Personal Care Products (PPCPs) under nitrifying and denitrifying conditions, *Water Res.*, 44 (2010) 3214-3224.

- 1072 [37] V.V. Unnithan, A. Unc, G.B. Smith, Mini-review: A priori considerations for bacteria-algae
1073 interactions in algal biofuel systems receiving municipal wastewaters, *Algal Res.*, 4 (2014) 35-
1074 40.
- 1075 [38] S.R. Subashchandrabose, B. Ramakrishnan, M. Megharaj, K. Venkateswarlu, R. Naidu,
1076 Consortia of cyanobacteria/microalgae and bacteria: Biotechnological potential, *Biotechnol.*
1077 *Adv.*, 29 (2011) 896-907.
- 1078 [39] M. Hijosa-Valsero, V. Matamoros, J. MartÍN-Villacorta, E. BÉcares, J.M. Bayona, Assessment
1079 of full-scale natural systems for the removal of PPCPs from wastewater in small communities,
1080 *Water Res.*, 44 (2010) 1429-1439.
- 1081 [40] V. Matamoros, C. Arias, H. Brix, J.M. Bayona, Removal of pharmaceuticals and personal care
1082 products (PPCPs) from urban wastewater in a pilot vertical flow constructed wetland and a
1083 sand filter, *Environ. Sci. Technol.*, 41 (2007) 8171-8177.
- 1084 [41] A. Garcia-Rodríguez, V. Matamoros, C. Fontàs, V. Salvadó, The ability of biologically based
1085 wastewater treatment systems to remove emerging organic contaminants-a review, *Environ.*
1086 *Sci. Pollut. Res.*, (2014) 1-21.
- 1087 [42] M. Gros, M. Petrovic, A. Ginebreda, D. Barceló, Removal of pharmaceuticals during
1088 wastewater treatment and environmental risk assessment using hazard indexes, *Environ. Int.*,
1089 36 (2010) 15-26.
- 1090 [43] A. Garcia-Rodríguez, E. Sagristà, V. Matamoros, C. Fontàs, M. Hidalgo, V. Salvadó,
1091 Determination of pharmaceutical compounds in sewage sludge using a standard addition
1092 method approach, *Int. J. Environ. Anal. Chem.*, (2014) 1-11.
- 1093 [44] X. Wu, J.L. Conkle, F. Ernst, J.J. Gan, Treated Wastewater Irrigation: Uptake of
1094 Pharmaceutical and Personal Care Products by Common Vegetables under Field Conditions,
1095 *Environ. Sci. Technol.*, (2014).
- 1096 [45] C. Reyes-Contreras, M. Hijosa-Valsero, R. Sidrach-Cardona, J.M. Bayona, E. Bécares,
1097 Temporal evolution in PPCP removal from urban wastewater by constructed wetlands of
1098 different configuration: A medium-term study, *Chemosphere*, 88 (2012) 161-167.
- 1099 [46] M.E. Hoque, F. Cloutier, C. Arcieri, M. McInnes, T. Sultana, C. Murray, P.A. Vanrolleghem,
1100 C.D. Metcalfe, Removal of selected pharmaceuticals, personal care products and artificial
1101 sweetener in an aerated sewage lagoon, *Sci. Total Environ.*, (2014).
- 1102 [47] A. Dordio, A.J.P. Carvalho, D.M. Teixeira, C.B. Dias, A.P. Pinto, Removal of pharmaceuticals
1103 in microcosm constructed wetlands using *Typha* spp. and LECA, *Bioresour. Technol.*, 101
1104 (2010) 886-892.
- 1105 [48] H. Sanderson, D.J. Johnson, T. Reitsma, R.A. Brain, C.J. Wilson, K.R. Solomon, Ranking and
1106 prioritization of environmental risks of pharmaceuticals in surface waters, *Regul. Toxicol.*
1107 *Pharmacol.*, 39 (2004) 158-183.
- 1108 [49] C.I. Kosma, D.A. Lambropoulou, T.A. Albanis, Investigation of PPCPs in wastewater
1109 treatment plants in Greece: Occurrence, removal and environmental risk assessment, *Sci. Total*
1110 *Environ.*, 466-467 (2014) 421-438.
- 1111 [50] P.C. von der Ohe, M. Schmitt-Jansen, J. Slobodnik, W. Brack, Triclosan-the forgotten priority
1112 substance?, *Environ. Sci. Pollut. Res.*, 19 (2012) 585-591.
- 1113 [51] C. Ávila, V. Matamoros, C. Reyes-Contreras, B. Piña, M. Casado, L. Mita, C. Rivetti, C.
1114 Barata, J. García, J.M. Bayona, Attenuation of emerging organic contaminants in a hybrid
1115 constructed wetland system under different hydraulic loading rates and their associated
1116 toxicological effects in wastewater, *Sci. Total Environ.*, 470-471 (2014) 1272-1280.

Table 1. Average concentration and standard deviation of conventional water quality parameters (n=8 per campaign). Removal efficiencies for COD and NH₄-N are shown in brackets for the two HRAP (4 and 8 d HRT). **COD (chemical oxygen demand); DO (dissolved oxygen); TSS (total suspended solids).**

	Warm season			Cold season		
	Primary effluent	HRT = 4 d	HRT = 8d	Primary effluent	HRT = 4 d	HRT = 8d
Average daily solar radiation (W m ⁻²)		282			74	
T (°C)	28±1	25±1	25±1	16±1	13±1	13±1
DO (mg L ⁻¹)	2±1	6±1	8±1	7±2	10±1	12±2
pH	8±1	8±1	9±1	8±1	8±1	9±1
TSS (mg L ⁻¹)	118±112	316±50	346±38	-	110±23	149±15
Biomass production (gTSS m ⁻² d ⁻¹)	-	24±8	13±2	-	8±2	6±0.3
COD (mg L ⁻¹)	156±79	52±9* (75**)	52±12* (84**)	342±107	67±7* (83**)	52±7* (88**)
NH ₄ -N (mg L ⁻¹)	81±9	0.6±0.3(99**)	0.7±0.5(99**)	19±4	2±1 (90**)	0.4±0.1 (98**)

* soluble COD; ** calculation corrected for evaporation water losses.

1149
1150
1151
1152
1153
1154
1155
1156
1157
1158
1159
1160
1161
1162

Table 2. Seasonal removal efficiency (%) of target EOCs in the HRAPs operated at HRT of 4 and 8 days. EOCs have been sorted by their abundance in Fig. 2.

Name	Warm season		Cold season		Activate d sludge WWTPs
	HRT 4 d	HRT 8d	HRT 4d	HRT 8 d	
Caffeine	97±1	98±1	85±2 ^{ab}	91±2 ^{ab}	50-99 ^c
Acetaminophen	99±1	99±1	99±1	99±1	99-100 ^c
Ibuprofen	99±1	99±1	86±4 ^{ab}	93±3 ^{ab}	72-100 ^c
Methyl dihydrojasmonate	99±1	99±1	92±2 ^{ab}	97±1 ^{ab}	98 ^d
Oxybenzone	97±1	99±1	75±10 ^{ab}	88±4 ^{ab}	63-98 ^c
Ketoprofen	87±6 ^b	95±4	50±17 ^{ab}	75±9 ^{ab}	11-100 ^c
Hydrocinnamic acid	99±1	99±1	99±1	99±1	-
5-methyl benzotriazole	83±16	95±8	74±5	77±2 ^a	60 ^e
Naproxen	83±4	89±4	48±5 ^a	60±3 ^a	43-99 ^c
Carbamazepine	46±9	62±15	15±19 ^a	34±15 ^a	<nr-62 ^c
Galaxolide	94±1b	97±1	47±1 ^{ab}	71±2 ^{ab}	88 ^c
Benzothiazole	70±6	78±7	13±8 ^a	30±14 ^a	40-60 ^d
Diclofenac	82±6	92±3	21±29 ^a	29±14 ^a	<0-81 ^c
Methylparaben	59±12	75±8	12±9 ^a	25±11 ^a	82-91 ^h
Benzotriazole	74±7	84±4	33±10 ^a	41±5 ^a	60 ^e
Tonalide	84±1b	90±1	51±5 ^a	67±7 ^a	85 ^c
OH-Benzothiazole	80±3	82±5	20±17 ^a	37±11 ^a	50-70 ^d
Tributyl phosphate	82±5	86±8	69±8	78±2	55-86 ^g
Tris(2- chloroethyl)phosphate	39±28	63±12	15±23	21±19	nr ^c
Triphenyl phosphate	82±2b	89±1	24±6 ^{ab}	68±10 ^{ab}	40 ^g
Triclosan	93±1	95±1	49±5 ^{ab}	69±2 ^{ab}	71-99 ^c
Cashmeran	70±5	79±5	61±3	64±8	50 ^f
Octylphenol	90±6	93±4	58±12 ^a	74±5 ^a	<nr-97 ^c
Diazinon	61±4	63±1	-	-	nr ^c
Celestolide	52±1	53±1	-	-	59 ^f
Atrazine	76±6	85±3	41±7 ^{ab}	69±6 ^{ab}	nr-25 ^c
Bisphenol A	72±14	85±8	66±16	78±6	63-99 ^c
2,4-D	22±10	32±26	-	-	-

^a seasonal statistical difference at p=0.05; ^b HRT statistical difference at p=0.05, ^c [32]; d [51]; d [52]; e [53] ; f[54]; g[55]; h [56]

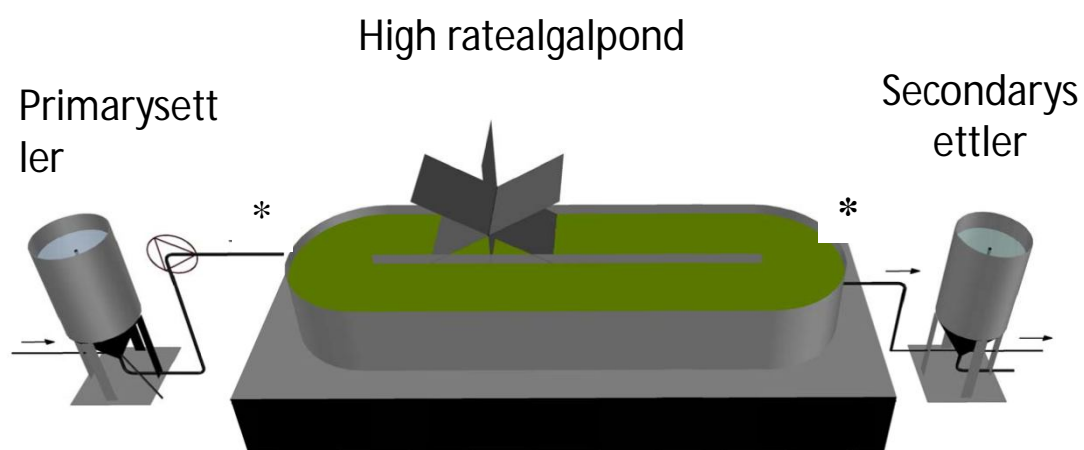
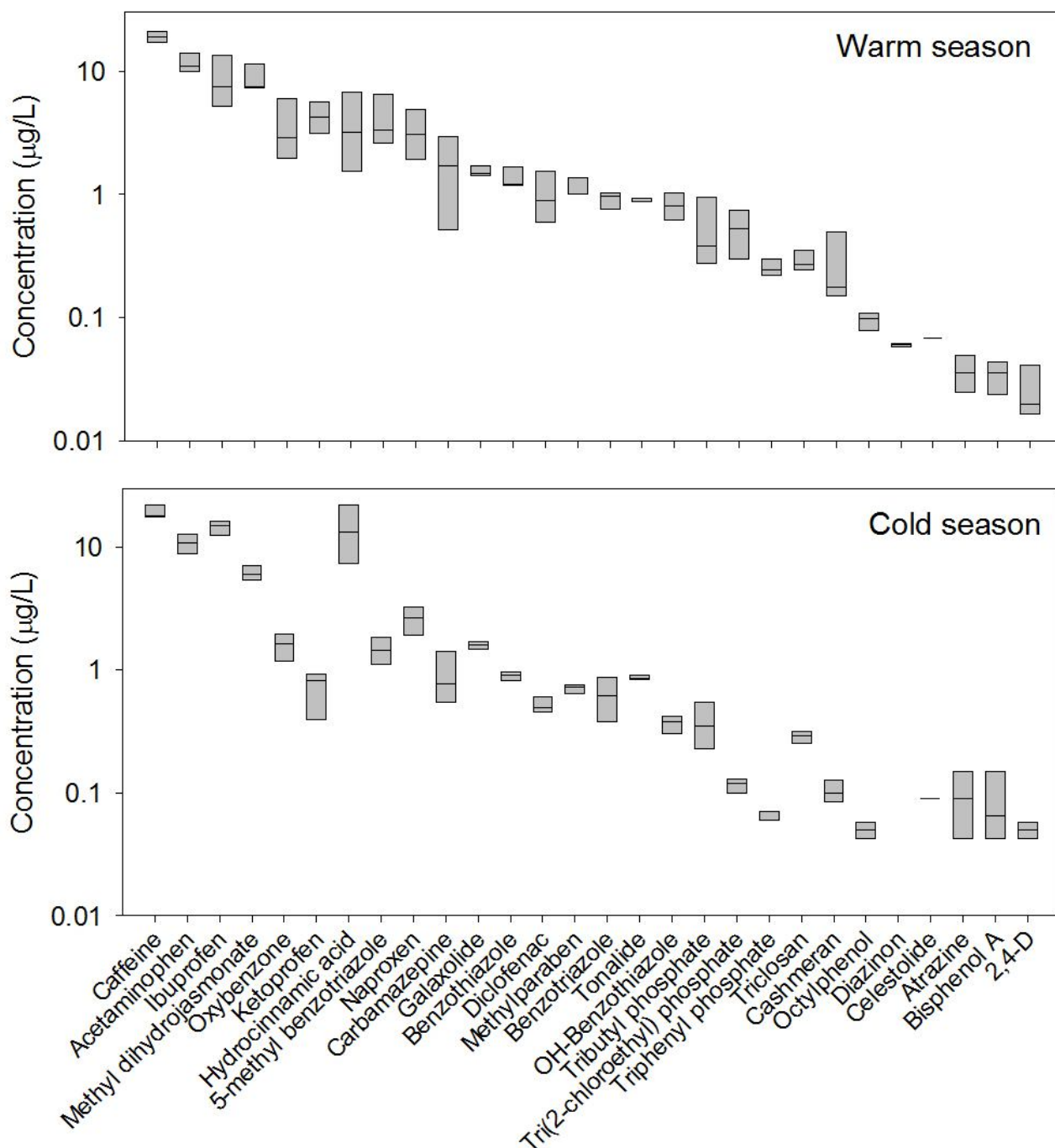


Figure 1. 3D view of treatment units of one line. Primary settler is fed with screened wastewater. Secondary settler allows separation of the biomass produced in the HRAP. Sampling points are indicated (*).

1209



1210

1211

1212

1213 **Figure 2.** Logarithmic box-plot of the seasonal occurrence of emerging contaminants in the primary
1214 effluent wastewater (n=8). The box plots indicate the median, and the 25th and 75th percentiles for
1215 each compound. Note that particulate and dissolved phase were both included. Similar profiles were
1216 obtained when molar concentrations were compared (not shown).

1217

1218

1219

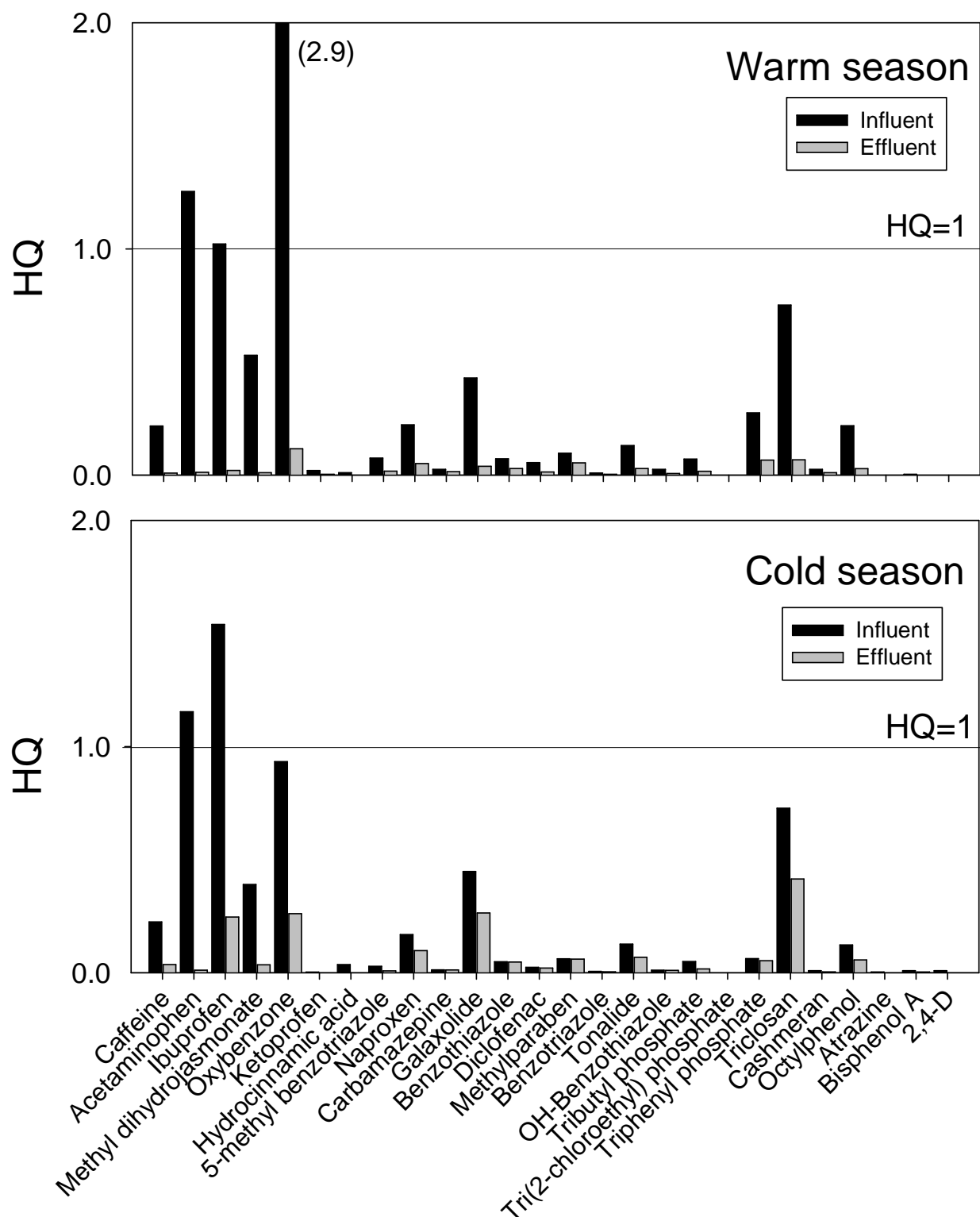


Figure 3. Seasonal hazard quotients (HQs) for the influent and effluent water samples collected from the HRAP set at a HRT of 4days.