



# Slow Amyloid Nucleation via $\alpha$ -Helix-Rich Oligomeric Intermediates in Short Polyglutamine-Containing Huntingtin Fragments

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The 17-amino-acid N-terminal segment (htt<sup>NT</sup>) that leads into the polyglutamine (polyQ) segment in the Huntington's disease protein huntingtin (htt) dramatically increases aggregation rates and changes the aggregation mechanism, compared to a simple polyQ peptide of similar length. With polyQ segments near or above the pathological repeat length threshold of about 37, aggregation of htt N-terminal fragments is so rapid that it is difficult to tease out mechanistic details. We describe here the use of very short polyQ repeat lengths in htt N-terminal fragments to slow this disease-associated aggregation. Although all of these peptides, in addition to htt<sup>NT</sup> itself, form  $\alpha$ -helix-rich oligomeric intermediates, only peptides with Q<sub>N</sub> of eight or longer mature into amyloid-like aggregates, doing so by a slow increase in  $\beta$ -structure. Concentration-dependent circular dichroism and analytical ultracentrifugation suggest that the htt<sup>NT</sup> sequence, with or without added glutamine residues, exists in solution as an equilibrium between disordered monomer and  $\alpha$ -helical tetramer. Higher order,  $\alpha$ -helix rich oligomers appear to be built up *via* these tetramers. However, only htt<sup>NT</sup>Q<sub>N</sub> peptides with N=8 or more undergo conversion into polyQ  $\beta$ -sheet aggregates. These final amyloid-like aggregates not only feature the expected high  $\beta$ -sheet content but also retain an element of solvent-exposed  $\alpha$ -helix. The  $\alpha$ -helix-rich oligomeric intermediates appear to be both on- and off-pathway, with some oligomers serving as the pool from within which nuclei emerge, while those that fail to undergo amyloid nucleation serve as a reservoir for release of monomers to support fibril elongation. Based on a regular pattern of multimers observed in analytical ultracentrifugation, and a concentration dependence of  $\alpha$ -helix formation in CD spectroscopy, it is likely that these oligomers assemble via a four-helix assembly unit. PolyQ expansion in these peptides appears to enhance the rates of both oligomer

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Abbreviations used: polyQ, polyglutamine; htt, huntingtin; AUC, analytical ultracentrifugation; SEC, size-exclusion chromatography; PBS, phosphate-buffered saline; MW, molecular weight; FTIR, Fourier transform infrared; EM, electron micrograph; ThT, thioflavin T; DLS, dynamic light scattering; TFA, trifluoroacetic acid; PBSA, PBS plus sodium azide.

formation and nucleation from within the oligomer population, by structural mechanisms that remain unclear.

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## Introduction

There are  $9^1$  or  $10^2$  different expanded CAG repeat diseases in which a polyglutamine (polyQ) repeat expansion in a particular disease protein is associated with a neurodegenerative disorder.<sup>1</sup> Intra-neuronal polyQ-rich aggregates are found in patient brains on autopsy, and the polyQ repeat length dependence of aggregation rates *in vivo*<sup>3</sup> and *in vitro*<sup>4,5</sup> intriguingly mirrors the repeat length dependence of disease risk and age of onset in the diseases.<sup>1</sup> Therefore, it has been of great interest to elucidate the mechanisms by which polyQ aggregation is initiated.

For simple polyQ sequences, aggregation rates increase with increasing repeat length,<sup>5</sup> and the aggregation reaction follows classical nucleated growth polymerization kinetics.<sup>6–9</sup> With such peptides, while short polyQs in the  $Q_{20}$  range require multimeric critical nuclei for aggregation initiation and consequently aggregate relatively slowly, polyQ sequences of  $Q_{26}$  or longer exhibit a critical nucleus of  $\sim 1$ , aggregating relatively quickly.<sup>6,9</sup> PolyQ aggregation behavior can be further complicated by the presence of flanking amino acid sequences such as those found in disease proteins.<sup>10–17</sup> Some flanking sequences modestly affect rates but do not fundamentally change the nucleated growth aggregation mechanism.<sup>9,13,15</sup> Other flanking sequences, however, produce a profound change in mechanism.<sup>15,18–20</sup>

In a striking example of a flanking sequence effect, the presence of a short, 17-amino-acid N-terminal sequence ("htt<sup>NT</sup>") adjacent to the polyQ at the N-terminus of the protein huntingtin (htt) leads to an enormous rate acceleration while fundamentally changing the spontaneous aggregation mechanism.<sup>15</sup> In this mechanism, a small portion of monomers self-associates to form roughly spherical oligomers in which all or part of the htt<sup>NT</sup> segment is packed into the oligomer core, while the polyQ portion remains disordered and accessible to antibody binding.<sup>15</sup> In a subsequent phase, the rate of aggregation of the remaining monomers dramatically increases, consistent with the operation of a nucleation event. Aggregates recovered from the reaction mixture just at the time of this rate increase exhibit evidence of a remarkable, apparently concerted transformation to more amyloid-like structure.<sup>15</sup> The results are consistent with models in which oligomer formation contributes to polyQ amyloid formation by providing a scaffold that locally concentrates disordered polyQ segments.<sup>15,21</sup>

Many details of this mechanism, however, are yet to be elucidated. Thus, in analogy to simple polyQ

peptides, aggregation rates of htt<sup>NT</sup>Q<sub>N</sub> peptides increase with increasing polyQ repeat length,<sup>4,15</sup> but the mechanism of this repeat length effect on htt<sup>NT</sup>Q<sub>N</sub> aggregation is not well understood. Furthermore, while recent data suggest that htt<sup>NT</sup>  $\alpha$ -helix formation is part of the oligomer formation process,<sup>21</sup> many questions remain, including the role and timing of  $\alpha$ -helix formation, and the role(s) of the  $\alpha$ -helical oligomers, in spontaneous amyloid assembly from htt<sup>NT</sup>Q<sub>N</sub> peptides. Elucidating these details is difficult with disease-associated polyQ lengths, however, since the nucleation event that triggers the rapid elongation phase likely occurs stochastically within only a small percentage of oligomers, leading to runaway elongation that obscures details of the early assembly mechanism.

We report here the results of a systematic study of htt<sup>NT</sup>Q<sub>N</sub>K<sub>2</sub> peptides with short polyQ repeat lengths designed to slow the early phases of the aggregation reaction. While aggregation rates are quite slow for all of these peptides, we find some significant differences in behavior within this repeat length range. For peptides of  $Q_7$  or lower, the aggregation reaction does not progress beyond the  $\alpha$ -helical oligomer stage. In contrast, peptides with  $Q_8$  or above are capable of moving on to the amyloid fibril stage, while nonetheless retaining an element of  $\alpha$ -helix in the amyloid-like aggregates. Analytical ultracentrifugation (AUC) and circular dichroism (CD) measurements suggest that  $\alpha$ -helix-rich oligomers build up in an organized, hierarchical fashion through tetrameric assemblies that appear to exist in equilibrium with the monomer under normal solution conditions. Other data show that oligomer dissociation rates are similar to association rates, suggesting that monomers can be generated from oligomers to support continued elongation late in the fibril growth reaction. The data shed light onto how polyQ repeat length contributes to aggregation rates in these htt<sup>NT</sup>Q<sub>N</sub> peptides and reveal an unprecedented case of a nucleation event associated with a net decrease in aggregation rate.

## Results

### Solution properties of htt<sup>NT</sup>Q<sub>N</sub>K<sub>2</sub> peptides with a wide range of polyQ repeat length

In order to address several unresolved aspects of the aggregation mechanism of htt fragments (Introduction), we purified and characterized the solution structures of a series of htt<sup>NT</sup>Q<sub>N</sub>K<sub>2</sub> peptides with *N*

**Table 1.** Structures of peptides used in this study

Name	Sequence
htt <sup>NT</sup>	MATLEKLMKA FESLKSF-amide
htt <sup>NT</sup> Q	MATLEKLMKA FESLKSQ
htt <sup>NT</sup> Q <sub>3</sub>	MATLEKLMKA FESLKSQQQ
htt <sup>NT</sup> Q <sub>4</sub> K <sub>2</sub>	MATLEKLMKA FESLKSQQQ QKK
htt <sup>NT</sup> Q <sub>5</sub> K <sub>2</sub>	MATLEKLMKA FESLKSQQQ QKK
htt <sup>NT</sup> Q <sub>6</sub> K <sub>2</sub>	MATLEKLMKA FESLKSQQQ QQQK
htt <sup>NT</sup> Q <sub>7</sub> K <sub>2</sub>	MATLEKLMKA FESLKSQQQ QQQK
htt <sup>NT</sup> Q <sub>8</sub> K <sub>2</sub>	MATLEKLMKA FESLKSQQQ QQQQK
htt <sup>NT</sup> Q <sub>9</sub> K <sub>2</sub>	MATLEKLMKA FESLKSQQQ QQQQK
htt <sup>NT</sup> Q <sub>10</sub> K <sub>2</sub>	MATLEKLMKA FESLKSQQQ QQQQK
htt <sup>NT</sup> Q <sub>15</sub> K <sub>2</sub>	MATLEKLMKA FESLKSQQQ QQQQQQQQ QKK
htt <sup>NT</sup> Q <sub>25</sub> K <sub>2</sub>	MATLEKLMKA FESLKSQQQ QQQQQQQQQ QQQQQQQQ QKK
htt <sup>NT</sup> Q <sub>35</sub> K <sub>2</sub>	MATLEKLMKA FESLKSQQQ QQQQQQQQQ QQQQQQQQQ QQQQQQQQ QKK
htt <sup>NT</sup> Q <sub>37</sub> P <sub>10</sub> K <sub>2</sub>	MATLEKLMKA FESLKSQQQ QQQQQQQQQ QQQQQQQQQ QQQQQPPPPP PPPPK
K <sub>2</sub> Q <sub>37</sub> K <sub>2</sub>	KKQQQQQQQQ QQQQQQQQQ QQQQQQQQQ QQQQQQQQ K

ranging from 3 to 35 (Table 1). Peptides were subjected to a stringent disaggregation protocol (Materials and Methods) and were immediately subjected to analytical size-exclusion chromatography (SEC) in phosphate-buffered saline (PBS) at 23 °C. This analysis indicated only one detectible form, the monomer, in each case (Fig. 1a).

One peptide, htt<sup>NT</sup>Q<sub>10</sub>K<sub>2</sub>, was further analyzed by AUC. A sedimentation velocity experiment on this peptide in PBS yielded data that fit well to a predominantly monomeric solution, with molecular weight (MW)=3500 (calculated MW=3512) (Fig. 1b). However, low levels of species with MWs of 14,300, 29,100, 45,200, and so forth, were also revealed in the best fit of the data. A shape change appears to occur upon assembly of the oligomers. The frictional coefficient decreases from 1.58 for the monomer to 1.24 for the oligomers, consistent with oligomers being more compact and/or more spherical than isolated monomers. The AUC results confirm the monomeric nature of the bulk of the sample, while at the same time suggesting that, under mild conditions, monomers can form low-MW oligomers whose sizes roughly correspond to 4-mers, 8-mers, 12-mers, and so forth, of the monomeric htt<sup>NT</sup>Q<sub>10</sub>K<sub>2</sub> peptide. We observe a similar series of small oligomers in the AUC analysis of htt<sup>NT</sup> itself (data not shown), indicating that polyQ interactions are not required for formation of tetramers or higher-order oligomeric intermediates. Although we observed no corresponding oligomeric forms on injection of disaggregated peptides into SEC (Fig. 1a), these apparently fail to appear because they do not chromatograph well.

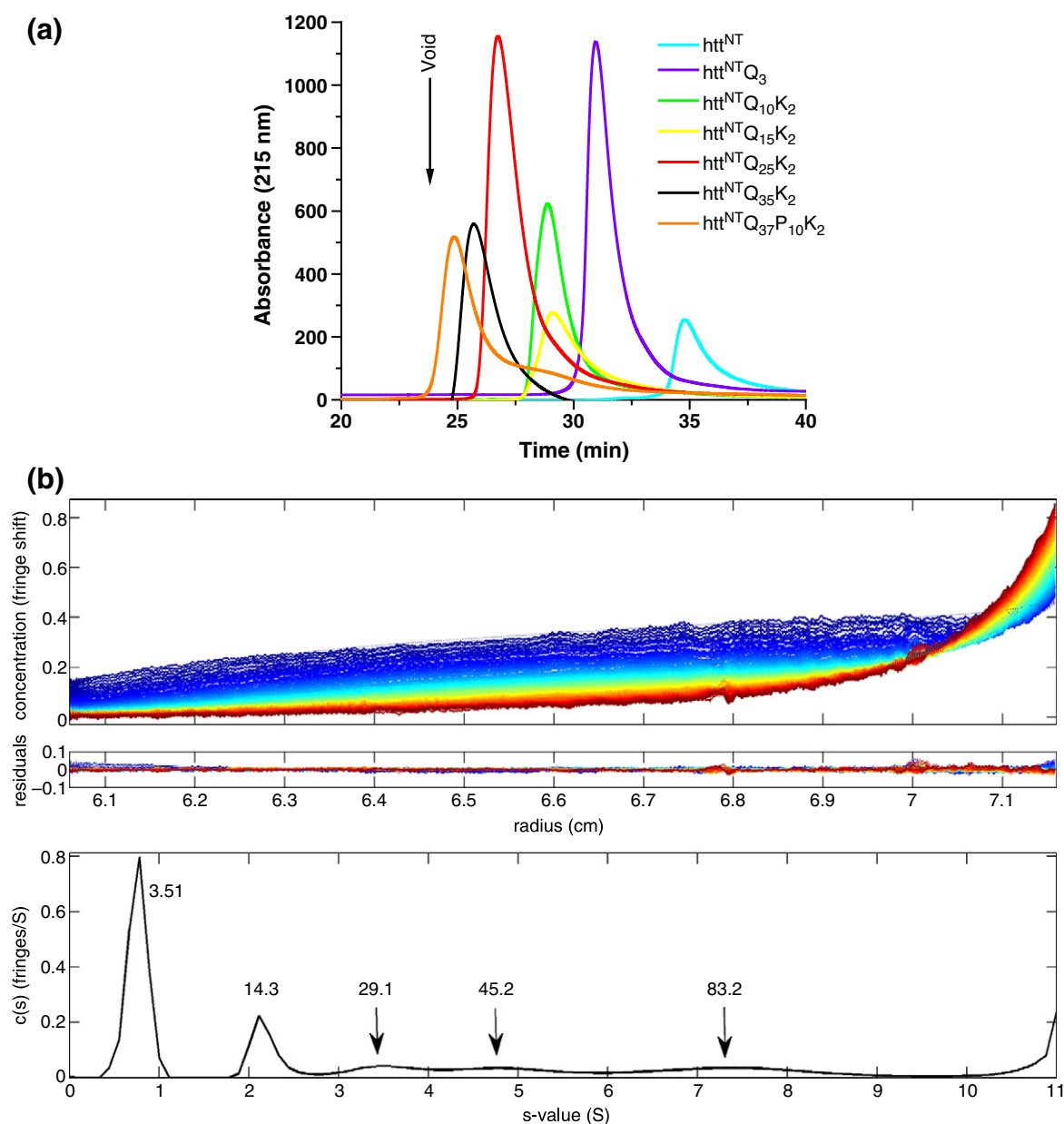
We obtained independent data consistent with an equilibrium distribution of oligomers in non-incubated samples of htt<sup>NT</sup> by CD analysis. The  $\alpha$ -helical bands in CD spectra of an htt<sup>NT</sup>Q peptide exhibit a marked concentration dependence consistent with a self-associating system (Fig. 2a). The high  $\alpha$ -helix content of samples analyzed in the 1-mM range is completely and essentially instantly

reversible, since a sample diluted from a high-concentration solution exhibits a stable CD spectrum identical with that of a freshly made up solution of the same final concentration (Fig. 2a). This result has several important implications. First, it suggests that the oligomers observed in the AUC most likely exist in a rapid dynamic equilibrium with the monomer. Second, it suggests that the oligomers observed in the AUC almost certainly are  $\alpha$ -helix rich. Third, it supports previous data<sup>15</sup> in indicating that the htt<sup>NT</sup> monomer in solution possesses no stable  $\alpha$ -helix; rather, the bulk of the  $\alpha$ -helix that is observed in CD analysis, at least at some concentrations, is probably due to reversible oligomer formation. Our previous CD examination of htt<sup>NT</sup> peptides did not reveal such a concentration dependence,<sup>15</sup> most likely because high concentrations in the 1-mM range were not analyzed.

We also determined the CD spectra for each of the htt<sup>NT</sup>Q<sub>N</sub> peptides analyzed in Fig. 1a. Analysis of these spectra (Materials and Methods) showed only minor differences in secondary structure between the different htt<sup>NT</sup> peptides, with the exception of a trend of increasing  $\alpha$ -helix with increasing polyQ repeat length (Fig. 2b). Part of this trend may be due to contributions of  $\alpha$ -helix from the polyQ sequence.<sup>22</sup> It is also possible that some additional  $\alpha$ -helix forms within the htt<sup>NT</sup> segment in response to polyQ expansion. Unfortunately, CD spectroscopy is incapable of localizing secondary structure to particular sequence elements. It is clear that there are no dramatic increases in  $\beta$ -content as polyQ repeat length increases in this family of monomers, similar to previous results on simple polyQ peptides,<sup>5,23,24</sup> and inconsistent with models positing large amounts of  $\beta$ -structure in polyQ molecules in solution.<sup>25</sup>

### Aggregation properties of htt<sup>NT</sup>Q<sub>M</sub>K<sub>2</sub> peptides with a wide range of polyQ repeat length

We also determined the aggregation kinetics of this set of htt<sup>NT</sup>Q<sub>N</sub> peptides. As reported previously,<sup>15</sup>



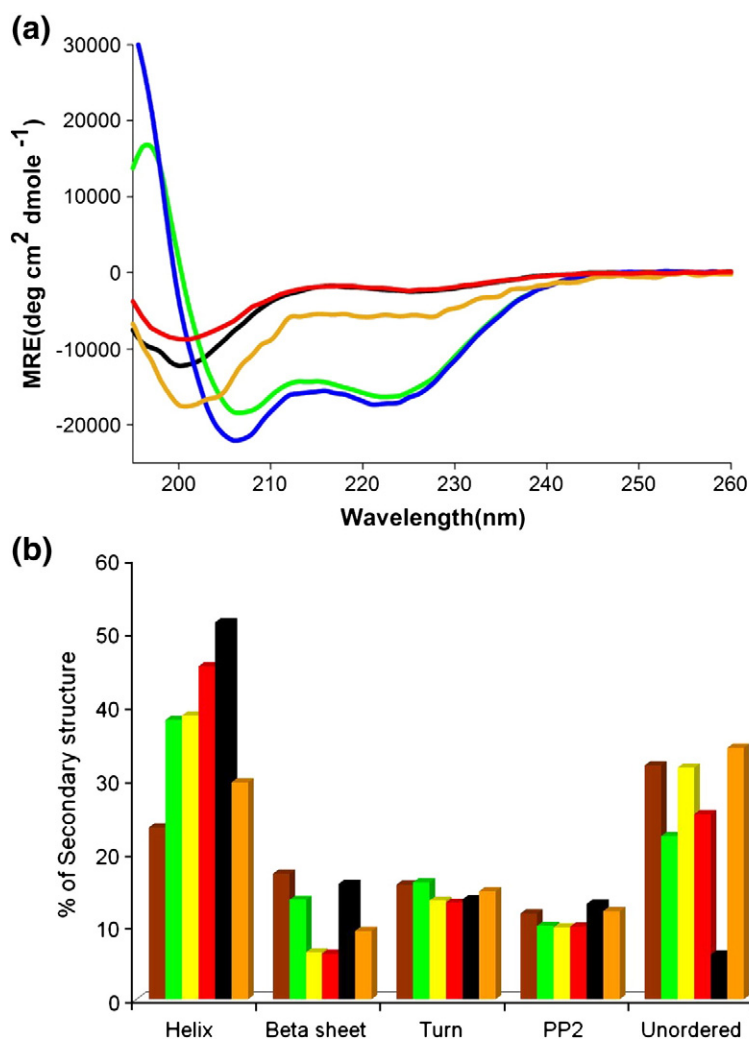
**Fig. 1.** Molecular size distribution of htt<sup>NT</sup>Q<sub>N</sub> peptides in PBS, pH 7.5. (a) Analytical SEC (at 23 °C) of peptides. (b) Sedimentation velocity analysis of 50 μM htt<sup>NT</sup>Q<sub>10</sub>K<sub>2</sub> at 20 °C. Data were analyzed using a continuous *c(s)* distribution model implemented in the program SEDFIT. The frictional ratio for each sample was a fitted parameter in the analysis, yielding 1.58 for the monomer and 1.24 for the oligomeric species. Sedimentation data, with global fits, are shown for htt<sup>NT</sup>Q<sub>10</sub>K<sub>2</sub> in the top frame, and resulting residuals for the analysis are given below the experimental data in the middle frame. The *c(s)* profile for the peptide species is shown in the bottom frame.

aggregation rates of a series of htt<sup>NT</sup>Q<sub>N</sub>K<sub>2</sub> peptides are strongly dependent on polyQ repeat length (Fig. 3a). While htt<sup>NT</sup>Q<sub>3</sub> (—Δ—) aggregates very slowly, and rates increase only modestly for htt<sup>NT</sup>Q<sub>10</sub>K<sub>2</sub> (—○—) and htt<sup>NT</sup>Q<sub>15</sub>K<sub>2</sub> (—◇—), there are substantial increases in rate as repeat lengths increase further to htt<sup>NT</sup>Q<sub>25</sub>K<sub>2</sub> (—□—) and htt<sup>NT</sup>Q<sub>35</sub>K<sub>2</sub> (—▼—). As reported previously,<sup>15</sup> polyQ peptides that mimic the htt exon1 fragment by containing both the htt<sup>NT</sup>

segment and a Pro<sub>10</sub> segment also aggregate very much faster than simple polyQ of comparable repeat length (htt<sup>NT</sup>Q<sub>37</sub>P<sub>10</sub>K<sub>2</sub>, ☆).

Elongation rates can have an enormous influence on overall aggregation rates.<sup>8</sup> We assessed the ability of each of these monomers to undergo seeded elongation using preformed fibrils of htt<sup>NT</sup>Q<sub>37</sub>P<sub>10</sub>K<sub>2</sub> as seeds (Fig. 3a, continuous lines). Although htt<sup>NT</sup>Q<sub>3</sub> monomers (▲) experience very little rate





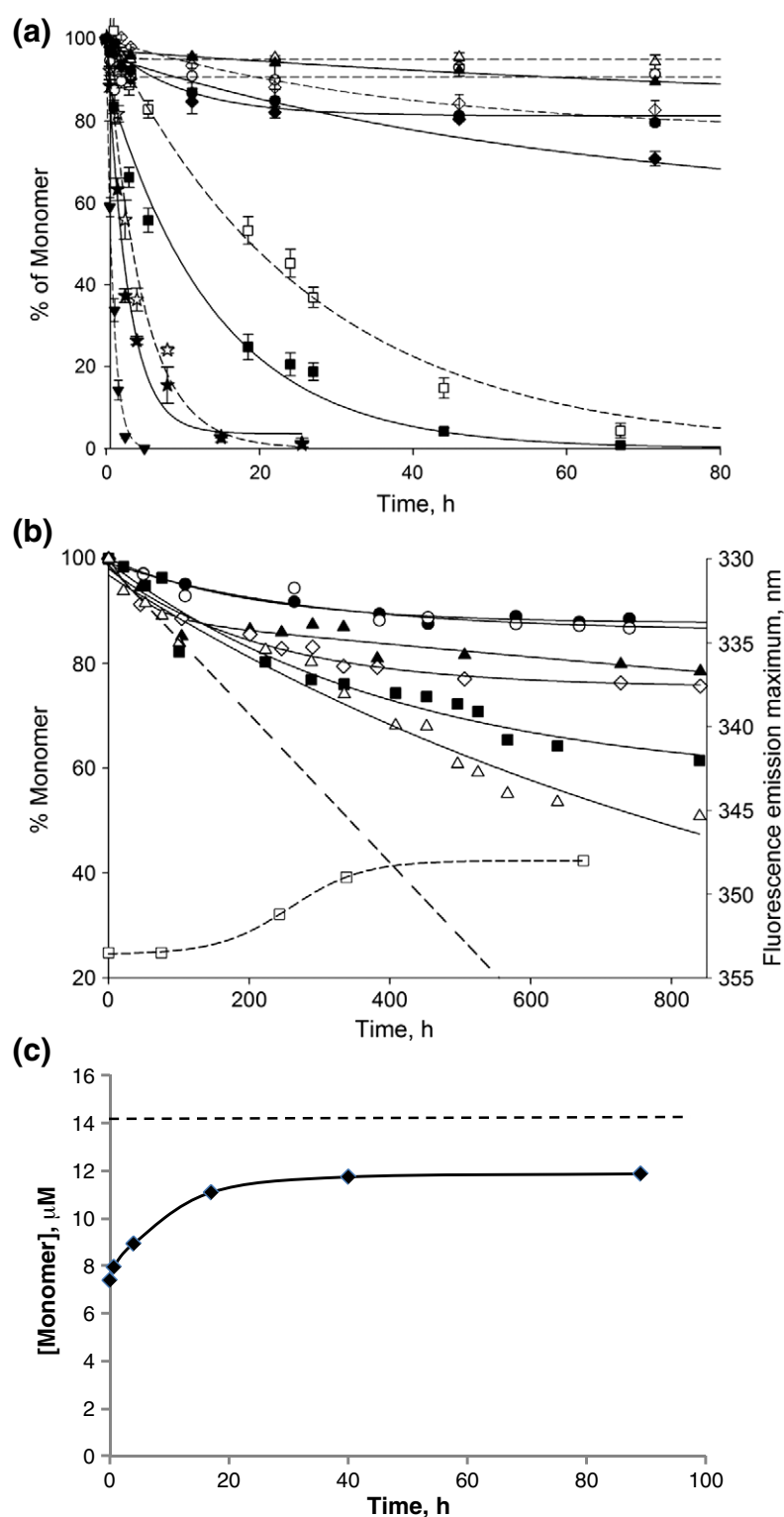
**Fig. 2.** CD analysis of monomeric  $\text{htt}^{\text{NT}}\text{Q}_N$  peptides in Tris-TFA, pH 7.4. (a) Concentration dependence of the CD spectra of  $\text{htt}^{\text{NT}}\text{Q}$ : green, 1.3 mM; blue, 0.67 mM; orange, 0.33 mM; black, 0.15 mM; red, 0.10 mM obtained by dilution from a 0.76-mM solution. All samples were analyzed in a 0.1-mm-path-length cell, except for the 0.10-mM sample, which was analyzed in a 0.1-cm cell. (b) Secondary-structure contents calculated (Materials and Methods) from CD curves, obtained in a 0.1-cm-path-length cell, of the following samples:  $\text{htt}^{\text{NT}}\text{Q}_3$ , 36  $\mu\text{M}$  (brown);  $\text{htt}^{\text{NT}}\text{Q}_{10}\text{K}_2$ , 32  $\mu\text{M}$  (green);  $\text{htt}^{\text{NT}}\text{Q}_{15}\text{K}_2$ , 28  $\mu\text{M}$  (yellow);  $\text{htt}^{\text{NT}}\text{Q}_{25}\text{K}_2$ , 25  $\mu\text{M}$  (red);  $\text{htt}^{\text{NT}}\text{Q}_{35}\text{K}_2$ , 27  $\mu\text{M}$  (black);  $\text{htt}^{\text{NT}}\text{Q}_{37}\text{P}_{10}\text{K}_2$  20  $\mu\text{M}$  (orange).

enhancement from seeding, monomers of  $\text{htt}^{\text{NT}}\text{Q}_{10}\text{K}_2$  (●),  $\text{htt}^{\text{NT}}\text{Q}_{15}\text{K}_2$  (◆), and  $\text{htt}^{\text{NT}}\text{Q}_{25}\text{K}_2$  (■) all respond to the presence of seed fibrils. While these experiments are by no means definitive, the data suggest that, with the exception of  $\text{htt}^{\text{NT}}\text{Q}_3$ ,  $\text{htt}^{\text{NT}}\text{Q}_N$  monomers are all capable of undergoing seeded elongation.

For the reactions shown in Fig. 3a, we also analyzed aggregates by Fourier transform infrared (FTIR) spectroscopy after the reaction appeared to have reached completion. As previously described for other polyQ peptides containing an  $\text{htt}^{\text{NT}}$  flanking sequence,<sup>15</sup> we found that aggregates of  $\text{htt}^{\text{NT}}\text{Q}_N\text{K}_2$  peptides with  $N=10$  or more Gln residues exhibit three major bands in the amide I region in second-derivative (Fig. 4a) and primary (Fig. 4b) FTIR spectra: a band  $\sim 1605\text{ cm}^{-1}$ , assigned to the  $\text{NH}_2$  deformation vibrations of the Gln side chain;<sup>26</sup> a band at  $1625\text{--}1630\text{ cm}^{-1}$ , assigned to a  $\beta$ -sheet;<sup>27</sup> and a band at  $1655\text{--}1660\text{ cm}^{-1}$ , assigned to C=O stretching of the Gln side chains<sup>26</sup> (Table 2). In contrast, aggregates of the  $\text{htt}^{\text{NT}}$  and  $\text{htt}^{\text{NT}}\text{Q}_3$  peptides exhibit

a single, broad band at  $1655\text{ cm}^{-1}$  (Fig. 4a; Table 2). The  $1648\text{--}1660\text{ cm}^{-1}$  amide I region is normally associated with  $\alpha$ -helix.<sup>27</sup> The  $\beta$ -sheet band is absent in the  $\text{htt}^{\text{NT}}$  and  $\text{htt}^{\text{NT}}\text{Q}_3$  aggregates.

Other analyses are consistent with the above results. A CD spectrum (Fig. 5a) of partially aggregated  $\text{htt}^{\text{NT}}\text{Q}_3$  obtained after 600 h of incubation (red) exhibits an increase in  $\alpha$ -helix content compared with the starting, freshly disaggregated peptide (blue), consistent with the FTIR data showing dominant  $\alpha$ -helix in the isolated oligomeric aggregates. In addition, electron micrographs (EMs) of  $\text{htt}^{\text{NT}}\text{Q}_3$  aggregates exhibit clusters of oligomers (Fig. 6a), while EMs of aggregates of  $\text{htt}^{\text{NT}}\text{Q}_N\text{K}_2$  peptides with  $N=10\text{--}35$  all exhibit fibrillar morphologies (Fig. 6h–k). Finally, isolated aggregates of the  $\text{Q}_{10}$  to  $\text{Q}_{25}$  peptides were all found to be good seeds for the elongation of monomeric  $\text{htt}^{\text{NT}}\text{Q}_{37}\text{P}_{10}\text{K}_2$  (Table 3), consistent with an amyloid-like structure in these aggregates.<sup>15,20,28</sup> In contrast,  $\text{htt}^{\text{NT}}\text{Q}_3$  aggregates not only do not stimulate  $\text{htt}^{\text{NT}}\text{Q}_{37}\text{P}_{10}\text{K}_2$  aggregation compared to the unseeded reaction but

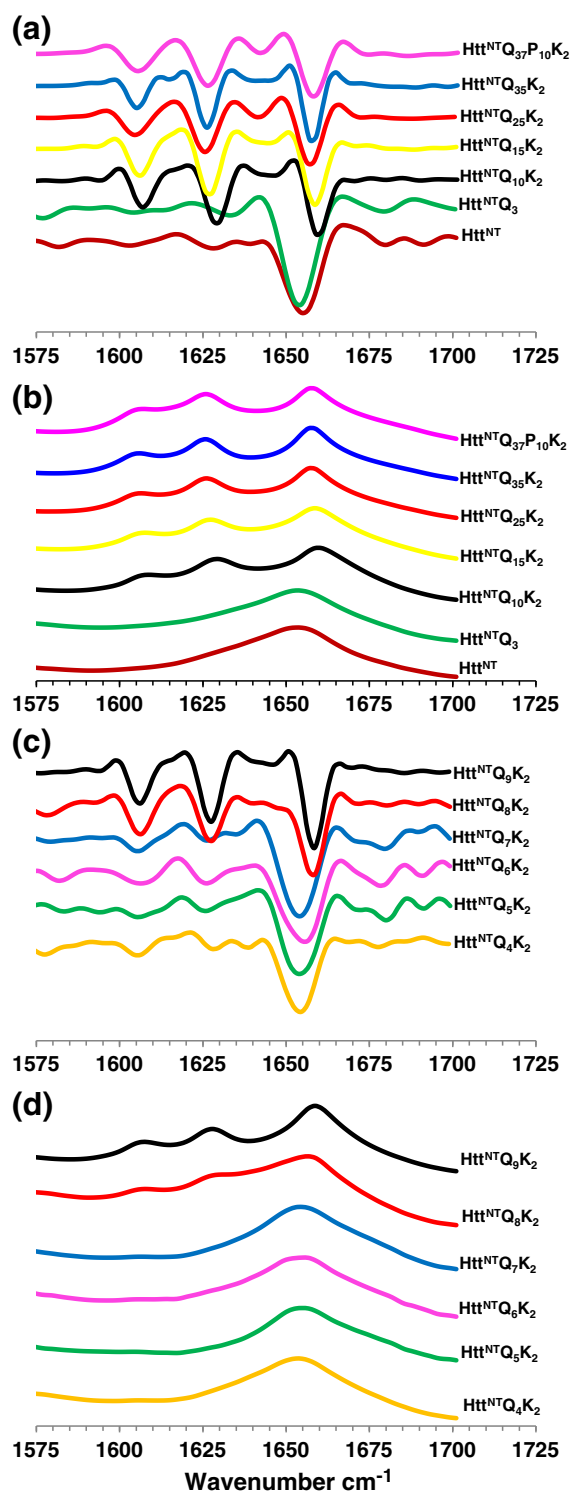


**Fig. 3.** Aggregation kinetics of htt<sup>NT</sup>Q<sub>N</sub> peptides in PBS at 37 °C. (a) ~5 μM monomer with (continuous lines) and without (broken lines) 7.5 wt% htt<sup>NT</sup>Q<sub>37</sub>P<sub>10</sub>K<sub>2</sub> aggregate seeds: htt<sup>NT</sup>Q<sub>3</sub> (Δ,▲), htt<sup>NT</sup>Q<sub>10</sub>K<sub>2</sub> (○,●), htt<sup>NT</sup>Q<sub>15</sub>K<sub>2</sub> (◇,◆), htt<sup>NT</sup>Q<sub>25</sub>K<sub>2</sub> (□,■), htt<sup>NT</sup>Q<sub>35</sub>K<sub>2</sub> (▼), and htt<sup>NT</sup>Q<sub>37</sub>P<sub>10</sub>K<sub>2</sub> (☆,★). The figure was modified from Ref.15. (b) Monomer loss (continuous lines) and shift in the Trp fluorescence emission maximum of isolated aggregates (curved broken line) for incubation of ~5 μM htt<sup>NT</sup>Q<sub>4</sub>K<sub>2</sub> (●), htt<sup>NT</sup>Q<sub>5</sub>K<sub>2</sub> (○), htt<sup>NT</sup>Q<sub>6</sub>K<sub>2</sub> (▲), htt<sup>NT</sup>Q<sub>7</sub>K<sub>2</sub> (◇), htt<sup>NT</sup>Q<sub>8</sub>K<sub>2</sub> (■,□), and htt<sup>NT</sup>Q<sub>9</sub>K<sub>2</sub> (Δ); the straight broken line is an extrapolation of the initial oligomer formation kinetics for htt<sup>NT</sup>Q<sub>9</sub>K<sub>2</sub>. (c) Dissociation kinetics of htt<sup>NT</sup>Q<sub>8</sub>K<sub>2</sub> oligomers collected after 120 h of incubation. The broken line represents total peptide (aggregate plus monomer) in the incubated sample.

also actually appear to inhibit the reaction somewhat (Table 3), similar to previously reported<sup>29</sup> analogous results with the Aβ<sub>40</sub> peptide. Together, these results suggest that aggregates from Q<sub>10</sub> and higher

peptides are amyloid-like, while htt<sup>NT</sup>Q<sub>3</sub> aggregates have a fundamentally different structure.

Overall, this analysis of a broad range of polyQ repeat lengths suggested that there is a critical

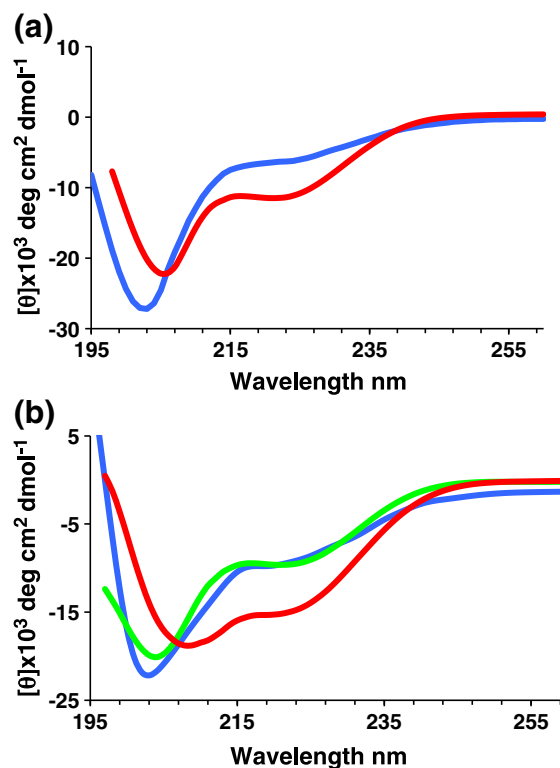


**Fig. 4.** FTIR spectra of isolated final aggregates. Second-derivative (a) and primary (b) spectra of final aggregates of  $\text{htt}^{\text{NT}}$  (2160 h),  $\text{htt}^{\text{NT}}\text{Q}_3$  (2160 h),  $\text{htt}^{\text{NT}}\text{Q}_{10}\text{K}_2$  (1540 h),  $\text{htt}^{\text{NT}}\text{Q}_{15}\text{K}_2$  (1080 h),  $\text{htt}^{\text{NT}}\text{Q}_{25}\text{K}_2$  (80 h),  $\text{htt}^{\text{NT}}\text{Q}_{35}\text{K}_2$  (24 h), and  $\text{htt}^{\text{NT}}\text{Q}_{37}\text{P}_{10}\text{K}_2$  (24 h). Second-derivative (c) and primary (d) spectra of final aggregates of  $\text{htt}^{\text{NT}}\text{Q}_4\text{K}_2$  (1020 h),  $\text{htt}^{\text{NT}}\text{Q}_5\text{K}_2$  (1140 h),  $\text{htt}^{\text{NT}}\text{Q}_6\text{K}_2$  (960 h),  $\text{htt}^{\text{NT}}\text{Q}_7\text{K}_2$  (1040 h),  $\text{htt}^{\text{NT}}\text{Q}_8\text{K}_2$  (950 h), and  $\text{htt}^{\text{NT}}\text{Q}_9\text{K}_2$  (950 h).

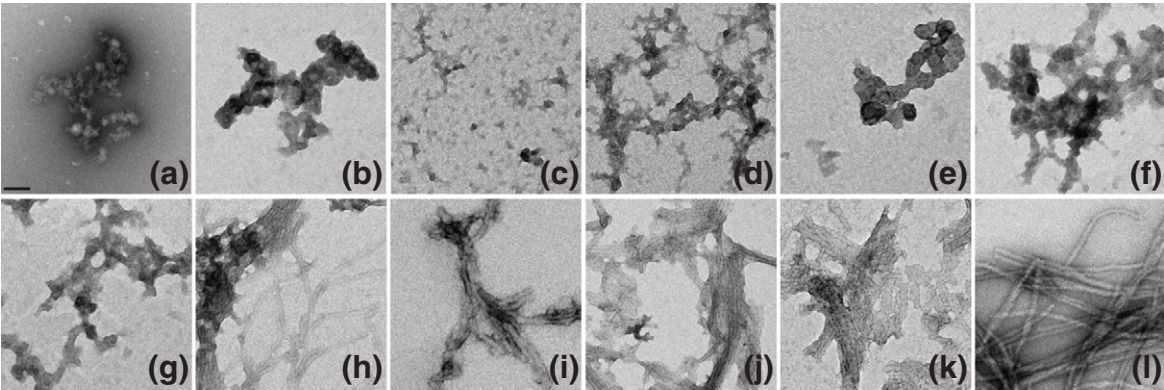
**Table 2.** Major FTIR amide I bands of isolated  $\text{htt}^{\text{NT}}\text{Q}_N$  final aggregates

Peptide	Gln N-H bending	$\beta$ -Sheet	$\alpha$ -Helix	Gln C=O stretch
$\text{htt}^{\text{NT}}$			1654.8	
$\text{htt}^{\text{NT}}\text{Q}_3$			1655.4	
$\text{htt}^{\text{NT}}\text{Q}_4\text{K}_2$	1604.2		1654.5	
$\text{htt}^{\text{NT}}\text{Q}_5\text{K}_2$	1604.4		1654.8	
$\text{htt}^{\text{NT}}\text{Q}_6\text{K}_2$	1606.6		1654.8	
$\text{htt}^{\text{NT}}\text{Q}_7\text{K}_2$	1606.6		1653.5	
$\text{htt}^{\text{NT}}\text{Q}_8\text{K}_2$	1606.6	1627.8		1656.7
$\text{htt}^{\text{NT}}\text{Q}_9\text{K}_2$	1606.3	1627.5		1658.4
$\text{htt}^{\text{NT}}\text{Q}_{10}\text{K}_2$	1606.6	1630.0		1658.9
$\text{htt}^{\text{NT}}\text{Q}_{15}\text{K}_2$	1606.8	1627.8		1658.7
$\text{htt}^{\text{NT}}\text{Q}_{25}\text{K}_2$	1604.3	1625.5		1656.3
$\text{htt}^{\text{NT}}\text{Q}_{35}\text{K}_2$	1605.0	1625.9		1659.0
$\text{htt}^{\text{NT}}\text{Q}_{37}\text{P}_{10}\text{K}_2$	1605.2	1626.4		1658.9

polyQ repeat length, somewhere between  $\text{Q}_3$  and  $\text{Q}_{10}$ , at which  $\text{htt}^{\text{NT}}\text{Q}_N\text{K}_2$  peptides are able to grow into amyloid-like fibrils. Based on this hypothesis, we carried out a systematic examination of an additional set of peptides exploring in detail this repeat length range.



**Fig. 5.** CD analysis of  $\text{htt}^{\text{NT}}\text{Q}_N$  aggregation. CD curves of peptides incubated in PBS at 37 °C. (a) 74  $\mu\text{M}$   $\text{htt}^{\text{NT}}\text{Q}_3$  for 0 h (blue) and 600 h (red). (b) 32.6  $\mu\text{M}$   $\text{htt}^{\text{NT}}\text{Q}_7\text{K}_2$  for 0 h (blue), 48 h (green), and 120 h (red). Note that spectra reflect the mixture of monomers and aggregates at each time point.



**Fig. 6.** Negative stained EMs of  $\text{htt}^{\text{NT}}\text{Q}_N$  peptide aggregates:  $\text{htt}^{\text{NT}}\text{Q}_3$  (a, 2160 h),  $\text{htt}^{\text{NT}}\text{Q}_4\text{K}_2$  (b, 912 h),  $\text{htt}^{\text{NT}}\text{Q}_5\text{K}_2$  (c, 1210 h),  $\text{htt}^{\text{NT}}\text{Q}_6\text{K}_2$  (d, 800 h),  $\text{htt}^{\text{NT}}\text{Q}_7\text{K}_2$  (e, 820 h),  $\text{htt}^{\text{NT}}\text{Q}_8\text{K}_2$  (f, 840 h),  $\text{htt}^{\text{NT}}\text{Q}_9\text{K}_2$  (g, 860 h),  $\text{htt}^{\text{NT}}\text{Q}_{10}\text{K}_2$  (h, 800 h),  $\text{htt}^{\text{NT}}\text{Q}_{15}\text{K}_2$  (i, 1080 h),  $\text{htt}^{\text{NT}}\text{Q}_{25}\text{K}_2$  (j, 82 h),  $\text{htt}^{\text{NT}}\text{Q}_{35}\text{K}_2$  (k, 24 h), and  $\text{htt}^{\text{NT}}\text{Q}_{37}\text{P}_{10}\text{K}_2$  (l, 20 h). The scale bar represents 50 nm.

**Aggregation of a narrow range of short polyQ repeat lengths in the  $\text{htt}^{\text{NT}}\text{Q}_N\text{K}_2$  series**

We obtained and purified the polyQ repeat length series  $\text{htt}^{\text{NT}}\text{Q}_4\text{K}_2$  to  $\text{htt}^{\text{NT}}\text{Q}_9\text{K}_2$  and analyzed their aggregation kinetics (Fig. 3b). After about 800 h of incubation, the  $\text{htt}^{\text{NT}}\text{Q}_4\text{K}_2$  (●) and  $\text{htt}^{\text{NT}}\text{Q}_5\text{K}_2$  (○) peptides reach an apparent plateau of ~12% aggregation, while the  $\text{htt}^{\text{NT}}\text{Q}_6\text{K}_2$  (▲) and  $\text{htt}^{\text{NT}}\text{Q}_7\text{K}_2$  (◇) peptides reach about 20% aggregation. The  $\text{htt}^{\text{NT}}\text{Q}_8\text{K}_2$  (■) and  $\text{htt}^{\text{NT}}\text{Q}_9\text{K}_2$  (Δ) peptides reach substantially higher aggregation levels of 40% ( $\text{htt}^{\text{NT}}\text{Q}_8\text{K}_2$ ) and 50% ( $\text{htt}^{\text{NT}}\text{Q}_9\text{K}_2$ ) by 800 h.

The structures of the aggregated products of these peptides exhibit a polyQ repeat length dependence that roughly correlates with the different aggregation kinetics. Second-derivative (Fig. 4c) and primary (Fig. 4d) FTIR spectra in the amide I region show a dominant  $\alpha$ -helix band (Table 2) for the late-stage aggregates of the  $\text{Q}_4$ ,  $\text{Q}_5$ ,  $\text{Q}_6$ , and  $\text{Q}_7$  peptides, in analogy to the aggregates of  $\text{htt}^{\text{NT}}$  and  $\text{htt}^{\text{NT}}\text{Q}_3$ . In contrast, aggregates of the  $\text{Q}_8$  and  $\text{Q}_9$  peptides isolated at 800 h exhibit spectra essentially identical with those of all polyQ amyloid fibrils, featuring strong Gln side-chain bands and a strong  $\beta$ -structure band (Fig. 4c and d; Table 2). In the EM, the  $\text{Q}_4$ ,  $\text{Q}_5$ ,  $\text{Q}_6$ , and  $\text{Q}_7$  (Fig. 6b–e) peptides exhibit only non-fibrillar aggregates. Although EMs of the

$\text{Q}_8$  and  $\text{Q}_9$  peptide aggregates (Fig. 6f and g) do not exhibit clear fibrillar morphologies, there is a suggestion of a filamentous substructure that may be obscured due to the greater relative mass from the non-amyloid (see below)  $\text{htt}^{\text{NT}}$  component in these peptides. A non-amyloid structure for the  $\text{htt}^{\text{NT}}\text{Q}_7\text{K}_2$  aggregates is also indicated by the negligible thioflavin T (ThT) fluorescent signal from these aggregates (Fig. 7c, red curve), compared to a typical amyloid-like response from aggregates of  $\text{htt}^{\text{NT}}\text{Q}_9\text{K}_2$  (Fig. 7c, blue curve). In addition,  $\text{htt}^{\text{NT}}\text{Q}_7\text{K}_2$  solutions grow in  $\alpha$ -helix content by CD over the aggregation reaction time course (Fig. 5b), and dynamic light scattering (DLS) data (Fig. 7a and b) yield a narrow range of relatively small, oligomer-sized particles throughout this time course.

Overall, these data suggest that  $\text{htt}^{\text{NT}}\text{Q}_N\text{K}_2$  peptides with polyQ segments of seven or less grow into aggregates that are incapable of nucleating a  $\beta$ -rich amyloid structure and therefore accumulate as stable,  $\alpha$ -helix-rich, ThT-negative aggregates. If the polyQ segment is of  $\text{Q}_8$  or longer, however,  $\beta$ -rich aggregates slowly develop, with overall aggregation levels increasing as polyQ repeat lengths increase.

**Time course of  $\text{htt}^{\text{NT}}\text{Q}_8\text{K}_2$  aggregation**

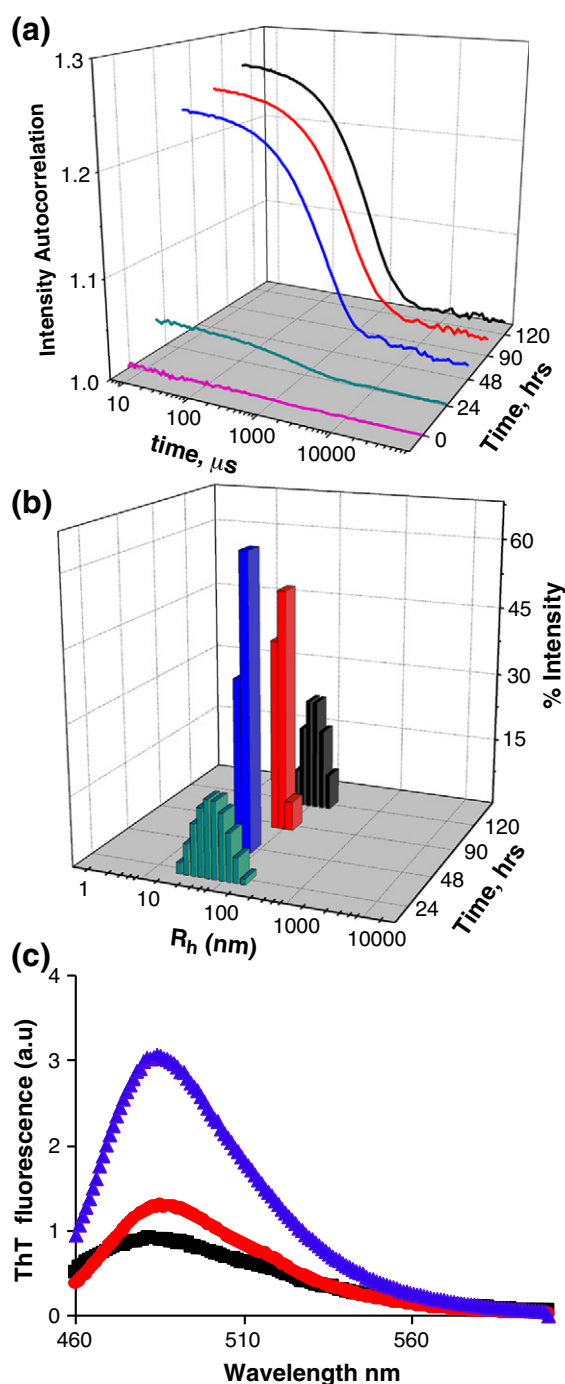
The above data define a sharp repeat length threshold in  $\text{htt}^{\text{NT}}\text{Q}_N$  peptides for their ability to convert from  $\alpha$ -rich oligomers to  $\beta$ -rich fibrils. While suggestive, these data do not formally address whether or not those peptides that develop into  $\beta$ -rich aggregates do so via an  $\alpha$ -rich oligomeric intermediate. To address this question, we isolated  $\text{htt}^{\text{NT}}\text{Q}_8\text{K}_2$  aggregates at different reaction times to spectroscopically examine their secondary structures. We found that aggregates isolated at 75 h exhibit a very strong

**Table 3.** Seeded elongation rate constants for  $\text{htt}^{\text{NT}}\text{Q}_{37}\text{P}_{10}$  monomers

	Seed aggregate				No seed
	$\text{htt}^{\text{NT}}\text{Q}_3$	$\text{htt}^{\text{NT}}\text{Q}_{10}$	$\text{htt}^{\text{NT}}\text{Q}_{15}$	$\text{htt}^{\text{NT}}\text{Q}_{25}$	
Rate constant ( $\text{h}^{-1}$ ) <sup>a</sup>	0.085	0.249	0.236	0.316	0.171

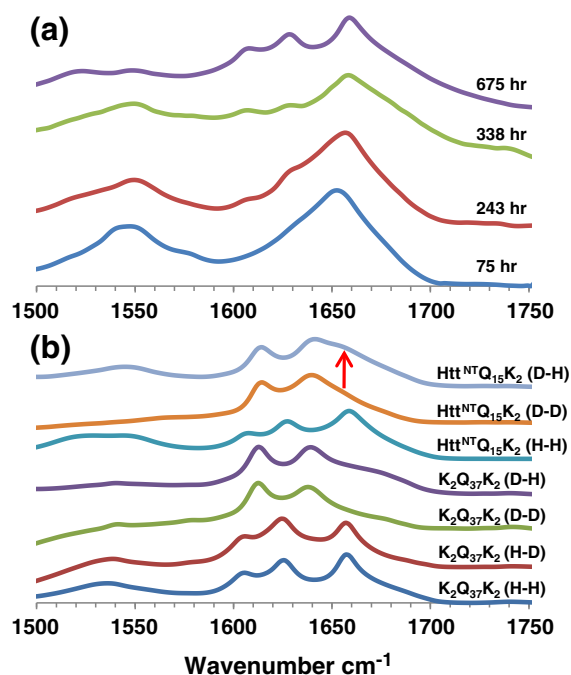
<sup>a</sup> Pseudo-first-order rate constants from reactions seeded with 7.5 wt% of aggregates.





**Fig. 7.** Analysis of  $\text{htt}^{\text{NT}}\text{Q}_N\text{K}_2$  aggregation. Autocorrelation functions (a) and particle size distributions (b) obtained from DLS measurements of  $32.6 \mu\text{M}$   $\text{htt}^{\text{NT}}\text{Q}_7\text{K}_2$  incubated in PBS at  $37^\circ\text{C}$  for the times indicated. (c) Fluorescence emission spectra of ThT alone (black) or bound to final aggregates of  $\text{htt}^{\text{NT}}\text{Q}_7\text{K}_2$  (red) or  $\text{htt}^{\text{NT}}\text{Q}_9\text{K}_2$  (blue) at monomer-equivalent concentrations of  $30 \mu\text{M}$ .

$\alpha$ -helix band in the FTIR at  $1653 \text{ cm}^{-1}$  (Fig. 8a). In addition, the fluorescence emission maximum for Trp17 in the isolated aggregates occurs at  $353.5 \text{ nm}$



**Fig. 8.** FTIR structural and mechanistic analysis. (a) Primary FTIR spectra of  $\text{htt}^{\text{NT}}\text{Q}_8\text{K}_2$  aggregates collected at the times indicated. (b) Primary FTIR spectra of  $\text{K}_2\text{Q}_{37}\text{K}$  and  $\text{htt}^{\text{NT}}\text{Q}_{15}\text{K}_2$  aggregates with different exposures to  $\text{H}_2\text{O}$  ("H") and  $\text{D}_2\text{O}$  ("D"). The first letter indicates the solvent used for aggregate growth, and the second letter denotes the solvent used for measurement.

(Fig. 3b,  $\square$ ), identical with that for the monomeric peptide and consistent with a Trp residue that is highly solvent exposed.<sup>15</sup> Aggregates isolated at 243 h exhibit not only a complex FTIR spectrum with shoulders at positions of the dominant bands of mature  $\text{htt}^{\text{NT}}\text{Q}_N$  amyloid fibrils (Fig. 8a) but also a shift in Trp fluorescence consistent with partial burial in solvent-excluded structure (Fig. 3b). By 338 h, the growth of  $\beta$ -structure in the FTIR (Fig. 8a) and the shift in Trp fluorescence (Fig. 3b) are nearly complete. By 675 h, the FTIR spectrum of the  $\text{htt}^{\text{NT}}\text{Q}_8\text{K}_2$  aggregates (Fig. 8a) is identical with spectra of  $\text{htt}^{\text{NT}}\text{Q}_N$  amyloid fibrils (Fig. 4b), and the Trp fluorescence has reached a plateau at  $348 \text{ nm}$  (Fig. 3b). These data support a model in which  $\alpha$ -helix-rich oligomers serve as intermediates in the formation of  $\text{htt}^{\text{NT}}\text{Q}_N$  amyloid. Interestingly, the FTIR spectra (Fig. 8a) also show that the N-H bending resonance at  $\sim 1605 \text{ cm}^{-1}$  that is a common feature of all polyQ amyloid aggregates is negligible or not present in the  $\alpha$ -helix-rich  $\text{htt}^{\text{NT}}\text{Q}_8\text{K}_2$  oligomeric intermediate but develops over time, in parallel with  $\beta$ -sheet (in contrast to a recent description of ataxin-3 aggregation<sup>30</sup>). This suggests that this relatively sharp band is specifically associated with

conformationally restricted, presumably H-bonded Gln side chains.<sup>31</sup> This interpretation in turn supports previous data suggesting that the polyQ segments are disordered in the htt<sup>NT</sup>Q<sub>N</sub> oligomeric intermediates.<sup>15</sup>

The inability of polyQ htt<sup>NT</sup>Q<sub>N</sub> molecules with  $N < 8$  to aggregate to completion suggests that oligomer formation is reversible and is controlled by a characteristic critical concentration or  $K_{eq}$ . To test this hypothesis, we monitored the ability of isolated htt<sup>NT</sup>Q<sub>8</sub>K<sub>2</sub> oligomers to dissociate (Materials and Methods). The results (Fig. 3c) show that oligomers in PBS at 37 °C dissociate within 20 h to an equilibrium concentration, confirming their relatively low stability and relatively rapid dissociation kinetics, on a par with their association kinetics (Fig. 3b).

### Hydrogen–deuterium exchange FTIR on final aggregates

Recent solid-state NMR experiments show that the htt<sup>NT</sup> segment of htt<sup>NT</sup>Q<sub>30</sub>P<sub>6</sub>K<sub>2</sub> amyloid fibrils contains a well-formed  $\alpha$ -helical segment in residues 4–11.<sup>21</sup> Given the strong secondary structural features observed in the FTIR spectra of the oligomeric intermediates and amyloid final products described above, one might have expected to observe an FTIR confirmation of this residual  $\alpha$ -helical component. Unfortunately, the C=O stretch band from the Gln side chains in the final amyloid product directly overlaps the  $\alpha$ -helix position (Table 2).

We used isotope-edited FTIR spectroscopy<sup>32</sup> to investigate the possible existence of  $\alpha$ -helix in the final fibril product. Figure 8b shows a typical FTIR spectrum in the amide I and II regions for an amyloid-like aggregate of a simple polyQ peptide (K<sub>2</sub>Q<sub>37</sub>K<sub>2</sub>) grown and examined in H<sub>2</sub>O (designated the “H–H” form in Fig. 8b). This spectrum exhibits the three bands for the Gln side chain and the  $\beta$ -sheet amide groups characteristic of all  $\beta$ -rich polyQ aggregates (Table 2). The assignment of these bands to the Gln side chains (1605 and 1655–1660 cm<sup>−1</sup>) and to the  $\beta$ -sheet (1625–1630 cm<sup>−1</sup>) was based on literature reports.<sup>26,27,30</sup> None of these bands changes position or significant intensity when this aggregate is transferred to D<sub>2</sub>O (Fig. 8b, H–D), suggesting that the protons associated with all three of these amide resonances are strongly protected from hydrogen–deuterium exchange.

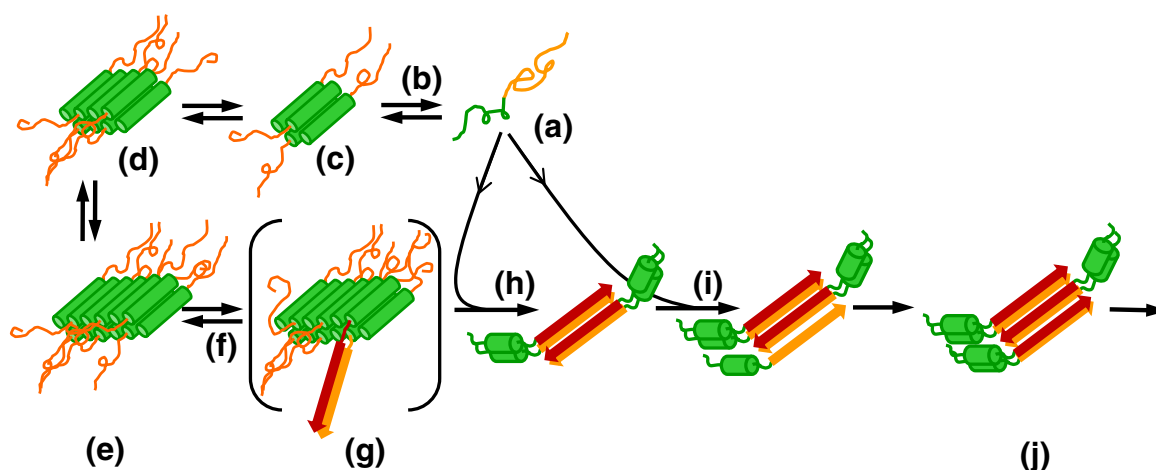
When K<sub>2</sub>Q<sub>37</sub>K<sub>2</sub> is aggregated and analyzed in D<sub>2</sub>O, the above three bands disappear and are replaced by two new ones (Fig. 8b, D–D). [These new band locations were assigned to the  $\beta$ -sheet (~1612 cm<sup>−1</sup>) and to the C=O stretch from the Gln side chain (~1638 cm<sup>−1</sup>) as follows. It was previously reported that deuteration of Gln analogs has the effect of moving the C=O stretch band from

around 1672 cm<sup>−1</sup><sup>26</sup> to about 1640 cm<sup>−1</sup>.<sup>33</sup> Furthermore, deuteration, as expected,<sup>32</sup> has been observed to shift the Gln side-chain N–H band completely out of the amide I region in a Gln-containing lipopeptide.<sup>34</sup> The amide II band at 1535 cm<sup>−1</sup> is also expected to disappear; deuteration of proteins typically shifts the amide II bands from ~1550 cm<sup>−1</sup> to ~1450 cm<sup>−1</sup>.<sup>35</sup>] Reanalyzing this deuterated K<sub>2</sub>Q<sub>37</sub>K<sub>2</sub> aggregate in H<sub>2</sub>O (Fig. 8b, D–H) produces no change, once again indicating that these FTIR bands are associated with amides that are strongly resistant to hydrogen–deuterium exchange.

A similar series of spectra of htt<sup>NT</sup>Q<sub>15</sub>K<sub>2</sub> final aggregates exhibit some subtle but important differences. The spectrum of htt<sup>NT</sup>Q<sub>15</sub>K<sub>2</sub> aggregates grown and analyzed in H<sub>2</sub>O (H–H) is remarkably similar to the corresponding H–H spectrum for K<sub>2</sub>Q<sub>37</sub>K<sub>2</sub> (Fig. 8b). Furthermore, when htt<sup>NT</sup>Q<sub>15</sub>K<sub>2</sub> is grown in D<sub>2</sub>O and analyzed in D<sub>2</sub>O, its D–D FTIR spectrum is also essentially identical with the corresponding D–D K<sub>2</sub>Q<sub>37</sub>K<sub>2</sub> spectrum (Fig. 8b). However, when htt<sup>NT</sup>Q<sub>15</sub>K<sub>2</sub> aggregates grown in D<sub>2</sub>O are transferred into H<sub>2</sub>O, this D–H spectrum is altered to produce two additional resonances, a band in the amide II region centered at 1545 cm<sup>−1</sup> and a shoulder in the amide I region at ~1655 cm<sup>−1</sup> (Fig. 8b, arrow). The emergence of these bands suggests that htt<sup>NT</sup>Q<sub>15</sub>K<sub>2</sub> aggregates possess an exchangeable, non- $\beta$ -element of regular secondary structure that is not present in K<sub>2</sub>Q<sub>37</sub>K<sub>2</sub> aggregates. Interestingly, the secondary structural feature associated with both of these band positions is the  $\alpha$ -helix.<sup>27</sup> The simplest explanation for this newly observed structural element is that it belongs to the htt<sup>NT</sup> portion of the htt<sup>NT</sup>Q<sub>15</sub>K<sub>2</sub> peptide. This suggestion that elements of the htt<sup>NT</sup> sequence reside in flexible, solvent-accessible  $\alpha$ -helix in the amyloid fibril is consistent with recent solid-state NMR results showing the same thing.<sup>21</sup>

## Discussion

Our model for the nucleation mechanism for amyloid formation by htt<sup>NT</sup>Q<sub>N</sub>-type peptides suggested by these and previous experiments is shown in Fig. 9. In htt<sup>NT</sup>Q<sub>N</sub> monomers in solution (Fig. 9a), the polyQ portion (orange) exists in a compact coil state<sup>36–39</sup> with no stable, regular secondary structure.<sup>5,6,23,24</sup> The htt<sup>NT</sup> segment (green) in isolation is also disordered but compact, with a modest tendency toward  $\alpha$ -helix formation<sup>15</sup> that is greatly enhanced in certain environments and solvents.<sup>15,40,41</sup> Based on data presented here, it appears that the htt<sup>NT</sup> segment becomes engaged in stable  $\alpha$ -helix immediately upon assembly into oligomers and retains at least a portion of this  $\alpha$ -helical character throughout the nucleation and elongation processes (Fig. 8a) as well as in the final



**Fig. 9.** Model for  $\text{htt}^{\text{NT}}\text{Q}_N$  aggregation. Disordered monomeric  $\text{htt}$  N-terminal fragments (a) ( $\text{htt}^{\text{NT}}$ , green; polyQ, orange or red) spontaneously and reversibly assemble (b) into oligomeric intermediates in which portions of  $\text{htt}^{\text{NT}}$  assemble into a bundled  $\alpha$ -helical core (c–e), and these assemblies grow larger upon incubation. Oligomer formation results in a high local concentration of polyQ segments, increasing the frequency of formation (f) of nuclei with segments of polyQ in  $\beta$ -sheet structure (g). Elongation of nuclei (h) and fibrils (i) by monomer addition generates growing amyloid-like aggregates (j). As fibrils grow and monomer is depleted, oligomers that have not undergone a productive nucleation reaction dissociate (b) to provide additional monomers to support fibril elongation. According to the mechanism in this schematic, nucleation driven by enhanced local polyQ concentrations might occur within oligomers of any size. The four-helix assemblies serving as the basic units of oligomer formation in this model are depicted as being antiparallel structures, but there is no direct evidence for this.

$\text{htt}^{\text{NT}}\text{Q}_N$  amyloid fibrils (Fig. 8b).<sup>21</sup> Based on the high  $\alpha$ -helix content of the oligomeric intermediates (Figs. 5 and 8), the tetrameric building block suggested by the AUC experiment (Fig. 1b), the concentration dependence of  $\alpha$ -helix formation in a freshly solubilized  $\text{htt}^{\text{NT}}$  peptide, and the helical packing of  $\text{htt}^{\text{NT}}$  domains in crystals of some  $\text{htt}$  N-terminal fragment derivatives,<sup>41</sup> we believe that some sort of packed four-helix assembly unit (Fig. 9c) mediates oligomer assembly.

The ability of the  $\text{htt}^{\text{NT}}$  segment to mediate a regular tetramerization of  $\text{htt}$  N-terminal fragments has not been previously recognized and may have significant implications for the cellular life of this poorly understood protein. Low MW oligomers of  $\text{htt}$  have been observed in several cell biology studies.<sup>42</sup> Whether these multimers reflect the operation of a generalized, reversible  $\alpha$ -helix rich oligomer formation, or are associated with specific, perhaps pathological, processes, remains to be determined.

The timing of the structural consolidation that occurs within the aggregate pool in these and previous<sup>15</sup> studies is consistent with nucleation occurring as a structural rearrangement within the oligomer fraction (Fig. 9g). At the same time, the poor stability of oligomers relative to fibrils (Fig. 3b), as well as the ability of oligomers to relatively rapidly dissociate (Fig. 3c), suggests that most oligomers that have not undergone this rearrangement/nucleation event ultimately serve as a reser-

voir for releasing monomers (Fig. 9b) to support continued fibril elongation (Fig. 9i). Given that nucleation of amyloid formation for at least some polyQ repeat lengths tends to occur at a point where the reaction mixture is still  $\sim 80\%$  monomeric<sup>15</sup> (Fig. 3b), it is likely that the bulk of the final amyloid mass (Fig. 9j) is formed via elongation by monomer addition of a relatively small number of oligomer-derived nuclei and fibrils (Fig. 9h and i). The pivotally important role in the aggregation mechanism of the  $\text{htt}^{\text{NT}}$  segment and its ability to engage in  $\alpha$ -helix-rich oligomer structure are further supported by experiments characterizing the abilities of  $\text{htt}^{\text{NT}}$  analogs to inhibit the oligomer-dependent aggregation pathway.<sup>43</sup>

Figure 9 shows nucleation occurring within a 12-mer, but this schematic is not meant to suggest that sporadic nucleation events are confined to a particular oligomeric species. In fact, in the short polyQ  $\text{htt}$  N-terminal fragments studied here, the aggregation kinetics suggest that significant nucleated growth of amyloid does not begin until after 200 h of incubation, at a time when about 20% of the sample consists of pelletable (i.e., much larger than 12-mer)  $\alpha$ -helix-rich oligomers (Fig. 3b). On the other hand, it is likely that nucleation occurs within much smaller oligomers in peptides such as  $\text{htt}^{\text{NT}}\text{Q}_{35}\text{K}_2$ , where amyloid formation occurs rapidly with little, if any, apparent lag time<sup>15</sup> (Fig. 3a).

There is an important distinction to be made between how amyloid structure is proposed to be

nucleated in the model summarized in Fig. 9 and classical nucleation theory. In the classical theory, the nucleus, its oligomeric precursors, and its polymeric products, are all formed via a continuous series of discrete homologous equilibria.<sup>44</sup> In contrast, the Fig. 9 model consists of a series of abrupt, differentiated steps: (a) tetramer formation from monomers, (b) packing of tetramers into higher oligomers, (c) nucleation within individual oligomers, and (d) elongation of nuclei into amyloid fibrils by monomer addition. Both the classical mechanism and the oligomer-mediated nucleation mechanism, however, share the central feature of a critical, energetically unfavorable step that must be traversed before strongly thermodynamically favored amyloid formation reaction can proceed. It is important to stress that the  $\alpha$ -helix-rich oligomers are themselves not nuclei for amyloid growth. Thus, contrary to expected behavior of nuclei,  $\alpha$ -helical oligomers (a) exhibit a finite stability allowing characterization by AUC, CD, FTIR, DLS, EM, and sedimentation; (b) do not structurally resemble the final amyloid product; and (c) are incapable of “seeding” amyloid growth.<sup>15</sup> Instead, the nucleus is an ephemeral species (formed within the oligomer fraction) that is uniquely capable of initiating the propagation of amyloid structure. As previously pointed out,<sup>15</sup> the model shown in Fig. 9 is similar in some respects to the “nucleated conformational conversion” model<sup>45</sup> for initiation of yeast prion aggregation under certain conditions. Only further studies will determine whether such models will ultimately be adequate to describe other amyloid formation reactions that feature oligomeric intermediates.

### Are the $\alpha$ -helical oligomers on- or off-pathway?

The role of non-amyloid oligomers in the amyloid assembly process has been much debated,<sup>46</sup> with claims that oligomers are either obligate intermediates for fibril formation (on-pathway)<sup>45,47</sup> or not required for fibril formation (off-pathway).<sup>48</sup> The concept of intermediates being on- or off-pathway derives from the protein folding field, where the issue is whether observed intermediates are obligatory stages through which all protein molecules must pass during productive assembly, or whether their formation merely represents a shunt that must ultimately be reversed before their molecules can productively fold. Indeed, it can be very difficult to kinetically distinguish on-pathway folding intermediates from off-pathway, since both can be seen to build up and decay during the folding reaction.<sup>49,50</sup>

In analogy, in many amyloid formation reactions, including those of the htt N-terminal fragments described here, non-amyloid aggregates are formed early but are no longer evident by the end of the

reaction. In the model shown in Fig. 9, the fate of some of the initial  $\alpha$ -helical oligomers is to undergo a nucleation process to initiate amyloid growth. At the same time, a significant portion of oligomers probably do not undergo nucleation and persist only until depletion of the monomer pool, by fibril elongation, creates a thermodynamic driving force for their dissociation to monomers. This dynamic is suggested by the very slow buildup of  $\beta$ -structure in a persistent field of  $\alpha$ -helical oligomers (Figs. 3b and 8) coupled with the relatively rapid dissociation of  $\alpha$ -oligomers back to monomers (Figs. 2a and 3c). While those oligomers that play a role in nucleation are clearly on-pathway for amyloid fibril formation, the oligomers that serve only as a reservoir for monomers can be considered off-pathway, in strict analogy to the protein folding experience.<sup>51</sup> Thus, at least for htt N-terminal fragment amyloid formation, the  $\alpha$ -helix-rich oligomeric intermediates serve both on-pathway and off-pathway roles. It would be premature to extend these observations to all oligomeric intermediates in amyloid formation reactions, however. Many such oligomers appear to possess significant levels of  $\beta$ -structure, and both their roles in nucleation and their abilities to dissociate have been little investigated.

This dual role for oligomers should have different consequences *in vitro* and *in vivo*. *In vitro*, as monomers are depleted during elongation, those oligomers that have not undergone amyloid nucleation should tend to dissociate and disappear according to their characteristic critical concentrations. In cells continuously producing new monomers, however, this mechanism will essentially lead to a steady-state concentration of oligomers, as has been observed.<sup>42,52</sup>

### Structures and role in amyloid nucleation of $\alpha$ -helical oligomeric intermediates

Several models exist in the literature for how htt<sup>NT</sup> and polyQ might interact to provide the rapid polyQ amyloid formation kinetics observed in such molecules. Based on domain accessibility measurements of isolated oligomeric intermediates, we proposed that htt<sup>NT</sup> is buried, and polyQ is disordered and solvent exposed, in oligomers, leading to a model conceptually like that shown in Fig. 9.<sup>15</sup> A similar oligomer-mediated model was also proposed by Tam *et al.*,<sup>16</sup> based primarily on photocross-linking data; this group also proposed the existence of non-covalent, long-range htt<sup>NT</sup>–polyQ interactions that might enhance nucleation of amyloid growth by favoring particular polyQ conformations. Quite different models for oligomer formation and structure, in which the oligomers are held together primarily by intermolecular polyQ interactions, have been proposed based on simulations<sup>53</sup> or



experimental data.<sup>54</sup> We think that our observation that the htt<sup>NT</sup> segment alone forms oligomeric species whether or not it is connected to a polyQ chain, and that htt<sup>NT</sup> by itself exhibits a concentration dependence in forming  $\alpha$ -helix in CD measurements, argues for htt<sup>NT</sup> playing a leading (rather than a following) role in oligomerization. At the same time, it remains possible that intermolecular polyQ association might play a supporting role in helping to stabilize oligomers initially brought together by htt<sup>NT</sup> interactions. This hypothesis remains to be tested experimentally.

Because of this structural role,  $\alpha$ -helix-rich oligomers also play an important functional role in greatly stimulating overall amyloid formation rates. This is illustrated by a comparison of the aggregation kinetics of molecules containing a Q<sub>25</sub> repeat. While a 5- $\mu$ M solution of htt<sup>NT</sup>Q<sub>25</sub>K<sub>2</sub> requires only ~20 h to reach 50% amyloid formation (Fig. 3a), under the same conditions, a 10-times higher concentration of K<sub>2</sub>Q<sub>25</sub>K<sub>2</sub> reaches 50% amyloid formation only after a 10-times longer incubation time (~250 h).<sup>9</sup> The prime importance of htt<sup>NT</sup> self-association in driving amyloid formation kinetics in this system is also supported by our observations, reported elsewhere,<sup>43</sup> of strong aggregation inhibition by htt<sup>NT</sup> and various derivatives. One interesting outcome of the htt<sup>NT</sup>-mediated nucleation mechanism is that the observed rate of spontaneous amyloid formation is driven by a portion of the peptide sequence that never actually engages amyloid structure.

This pivotal role of an  $\alpha$ -helix-rich oligomeric intermediate in amyloid nucleation may not be unique to the htt system. Similar mechanisms have been suggested for other amyloid systems,<sup>55</sup> including  $\alpha$ -synuclein,<sup>56</sup> amyloid  $\beta$ ,<sup>57</sup> islet amyloid polypeptide,<sup>58</sup> and several model systems.<sup>59,60</sup> The htt system, however, appears to be unique in that a substantial portion of the  $\alpha$ -helix that is the basis of oligomer formation is actually retained in the final fibrils<sup>21</sup> (Fig. 8b). The unusual retention of  $\alpha$ -helix in htt<sup>NT</sup> may be partly due to its intrinsic  $\alpha$ -helical propensity, to a constitutive inability of elements of the htt<sup>NT</sup> sequence to form amyloid,<sup>61</sup> or to a fastidious discrimination by the polyQ segment against incorporating substantial portions of non-Gln sequence into the growing amyloid core. The clean compartmentalization of the htt mechanism, which can be teased apart by damping down aggregation kinetics through adjustments in polyQ repeat length, has allowed us to elucidate critical aspects of this general mechanism.

### Mechanistic source of expanded polyQ acceleration of htt<sup>NT</sup>Q<sub>N</sub> aggregation

Previously, we observed that expanded polyQ sequences in htt<sup>NT</sup>Q<sub>N</sub> peptides tended to increase

the efficiency with which these peptides form oligomers.<sup>15</sup> We suggested that this trend might play a role in polyQ expansion enhancement of amyloid formation rates. We also provided FRET data consistent with the induction of a conformational expansion within the htt<sup>NT</sup> segment by polyQ repeat expansion that could be the source of increased oligomer formation.<sup>15</sup> Predictions from simulations, while differing considerably in detail, tend to support this conformational effect.<sup>53,62,63</sup>

At the same time, we believe that further studies are required to elucidate the basis of the polyQ expansion effect on oligomer formation. Data presented here indicate that polyQ expansion also contributes to the rate of amyloid formation coming out of the  $\alpha$ -helix oligomer pool. Thus, amyloid nucleation in htt<sup>NT</sup>Q<sub>N</sub> peptides does not occur, even at long incubation times, unless the polyQ repeat length is at least Q<sub>8</sub> (Figs. 3b and 4c). Furthermore, while oligomers of htt<sup>NT</sup>Q<sub>8</sub>K<sub>2</sub> populated by 100 h require another 300 h before the isolated aggregates are completely amyloid-like (Fig. 3b), fibril formation (via oligomers) from htt<sup>NT</sup>Q<sub>35</sub>K<sub>2</sub> monomers is complete after only a few hours under the same conditions (Fig. 3a). This great rate enhancement of the monomer-to-fibril transformation cannot be explained by rate effects on oligomer formation alone and must consider the nucleation and elongation events. PolyQ repeat length dependent secondary nucleation effects<sup>44,64–66</sup> might contribute to the repeat length dependence of overall aggregation rates, but there is no indication that polyQ amyloid formation is susceptible to this process under typical growth conditions.<sup>6–9</sup> In combination with other methodologies, reducing polyQ repeat length in order to slow aggregation rates may lead to future improvements in our understanding of the polyQ repeat length effect.

### Amyloid formation by very short polyQ sequences

Our finding that htt<sup>NT</sup>Q<sub>N</sub> peptides as short as Q<sub>8</sub> are capable of making amyloid is consistent with X-ray diffraction<sup>67</sup> and mutational<sup>68</sup> analyses, suggesting  $\beta$ -sheet widths of ~7 amino acids in simple polyQ amyloids.  $\beta$ -Sheet widths in the htt<sup>NT</sup>Q<sub>8</sub>K<sub>2</sub> peptide may be as much as 9 or 10 residues, since ssNMR of fibrils of a longer polyQ htt N-terminal fragment showed that Ser16 and Phe17 also exist in  $\beta$ -sheet.<sup>21</sup> A recent analysis of the repeat length dependence of short polyQ solution conformations suggests that simple polyQ peptides in the Q<sub>8</sub>–Q<sub>12</sub> repeat length range should not be capable of making amyloid.<sup>39</sup> As shown herein, however, the biophysics appears to change significantly with the addition of flanking sequences that can, for example, contribute at least a few residues to the  $\beta$ -sheet<sup>21</sup> and hence to its energetics.

## The driving force for nucleated growth

Although nucleation events are normally associated with enhanced aggregation kinetics, the defining feature of nucleation is more accurately characterized as the overcoming of a significant kinetic barrier to a thermodynamically favorable transformation. This subtle distinction is illustrated in Fig. 3b, where it can be seen that the nucleation and elongation phases of amyloid formation by the htt<sup>NT</sup>Q<sub>8</sub>K<sub>2</sub> and htt<sup>NT</sup>Q<sub>9</sub>K<sub>2</sub> peptides exhibit kinetics that are actually slower than the preceding rates of oligomer formation (indicated by the broken line extrapolation). Nucleated growth of amyloid by these peptides occurs in spite of the slow kinetics because of the thermodynamic driving force for conversion of monomers and oligomers into amyloid.

## Materials and Methods

### Materials

Water (HPLC grade), acetonitrile (99.8% HPLC grade), hexafluoro-2-propanol (99.5%, spectrophotometric grade), and formic acid were from Acros Organics; trifluoroacetic acid (TFA) (99.5%, Sequanal Grade) was from Pierce; and ThT was from Sigma. Chemically synthesized peptides (Table 1) were obtained from either the Keck Biotechnology Center at Yale University† or GenScript, Inc. In general, peptides were purified and subjected to rigorous disaggregation in volatile solvents (hexafluoro-2-propanol/TFA) for 16 h, followed by evaporation of solvents, resuspending the peptide film in TFA–water (pH 3) solution and ultracentrifugation at 435,680g, as previously described.<sup>9,15,69</sup> Peptides were used immediately, without storage, after disaggregation. Most peptides contained a conservative Phe→Trp replacement at position 17 (Table 1) previously shown to have little effect on aggregation rates.<sup>15</sup> For FTIR and ThT binding characterization and for use in seeding kinetics, aggregates were isolated from ongoing reactions and resuspended, and aggregate concentrations were determined as previously described.<sup>69</sup>

### Determination of concentrations

Determination of monomer and aggregate concentrations is critical to obtaining interpretable data. We used analytical HPLC with integration of the peak from an A<sub>215</sub> trace to quantify peptides. Although the A<sub>215</sub> absorption is largely determined by the peptide bond, different sequences can exhibit significantly different extinction coefficients because of side-chain contributions. We calculated the expected extinction coefficient in the HPLC solvent for each sequence using a modification of the method of Kuipers and Gruppen.<sup>70</sup> After HPLC purification of each peptide, the A<sub>215</sub> absorbance of a

portion of the pool was determined, and the concentration of the peptide was calculated from the extinction coefficient calculated from the Kuipers and Gruppen values.<sup>70</sup> Calibrated peptide solutions were then adjusted in concentration for use as an HPLC standard, and a standard curve of peak area with respect to injected mass was generated for each peptide, allowing future determinations of mass for each peptide. These modifications of the published<sup>70</sup> procedure compensate for observed sequence-dependent recovery efficiencies in analytical reverse-phase HPLC.

### Secondary structure of the monomeric htt<sup>NT</sup> peptides by CD spectroscopy

Far-UV CD measurements were performed on a JASCO J-810 spectropolarimeter using 0.1-cm- and 0.1-mm-path length-cuvettes. For the concentration dependence of htt<sup>NT</sup>Q (Fig. 2a), lyophilized purified peptides were dissolved in pH 3 (TFA) water and centrifuged at 435,680g for 2 h; the top 50% of the supernatant was removed, adjusted to 10 mM Tris–HCl, pH 7.4, and diluted with the same buffer to the desired concentrations. Spectra were collected at 1-nm resolution at a scan rate of 100 nm min<sup>−1</sup>. Four spectra were collected and averaged for each sample and corrected for the buffer blank. For measuring the conformations of monomeric htt<sup>NT</sup>Q<sub>N</sub> peptides (Fig. 2b), peptides were disaggregated as described in Materials, and centrifugation supernatants were adjusted to 10 mM Tris buffer, pH 7.4, and then filtered through 20-nm filters (Anotop inorganic membrane disc filter, Whatman) and immediately subjected to CD measurements. For aggregation reactions (Fig. 5), samples were prepared in PBS, pH 7.4, and filtered through the 20-nm filter, and then 400- $\mu$ l aliquots of the ongoing reaction were subjected to CD measurements at 37 °C. Spectra were recorded from 195 to 260 nm at 1-nm resolution with a scan rate of 50 nm min<sup>−1</sup>. Six scans were averaged and the buffer spectrum was subtracted. For Fig. 2b, CD spectra were analyzed using the CONTINLL program from the CDPro package (lamar.colostate.edu/~sreeram/CDPro) in which the SP37A reference set (ibasis 5) was used to estimate the amount of secondary structure.<sup>71</sup>

### Size-exclusion chromatography

To assess the aggregation state of peptides prior to spectroscopic analysis, we used a Superdex peptide column with a MW range of 7000 (GE Health Sciences) equilibrated in PBS on an Agilent 1200 isocratic HPLC system with a flow rate of 0.4 ml min<sup>−1</sup>. Freshly disaggregated peptide samples in TFA–water, pH 3.0, were adjusted to PBS, pH 7.4, using 10× PBS. Samples (100  $\mu$ l of ~10–15  $\mu$ M) were immediately injected with elution monitored at 214 nm.

### Analytical ultracentrifugation

Sample preparation was tailored to insure a rigorous match to the buffer reference. Thus, freshly disaggregated htt<sup>NT</sup>Q<sub>10</sub>K<sub>2</sub> in pH 3 TFA–water was transported

† <http://keck.med.yale.edu/ssps/>

on ice to the AUC laboratory (30 min). The sample was then adjusted to PBS buffer using a 10 $\times$  PBS stock and dialyzed for 2 h against PBS buffer at 23 °C. The dialysate was centrifuged 20–30 min in a refrigerated Eppendorf centrifuge (4 °C) to clear any pelletable aggregates. The recovered supernatant was used in the analytical centrifuge run, which was started approximately 30 min after the end of the centrifugation time. A reference buffer sample was prepared by following the above procedure, but with no peptide present, using a common dialysis buffer.

Sedimentation velocity experiments in a Beckman Optima XL-I analytical ultracentrifuge were used to evaluate the distribution of species in the peptide preparations. Epon double-sector centerpieces were filled with 400  $\mu$ l of samples. For all experiments, the reference buffer sample was loaded into the reference sector. Samples were centrifuged at a rotor speed of 50,000 rpm, and data were collected in fringes using interference optics. Sedimentation velocity analyses were performed with the program SEDFIT<sup>‡</sup> (as described in detail in Ref. 72) that carry out size distribution analyses based on Lamm equation modeling.<sup>73–75</sup> Briefly, the analysis of velocity profiles was performed by direct boundary modeling by distributions of Lamm equation solutions  $c(s)$  with

$$a(r, t) \equiv \int_{s_{\min}}^{s_{\max}} c(s) \chi_1(s, F, r, t) ds + b(r) + \beta(t) \quad (1)$$

where  $a(r, t)$  denotes the measured absorbance or interference profiles at position  $r$  at time  $t$ ,  $b(r)$  and  $\beta(t)$  are the characteristic systematic offsets,<sup>73</sup> and  $\chi_1(s, F, r, t)$  are solutions of the Lamm equation

$$\frac{\partial \chi}{\partial t} = \frac{1}{r} \frac{\partial}{\partial r} \left[ r D \frac{\partial \chi}{\partial r} - s \omega^2 r^2 \chi \right] \quad (2)$$

(with  $\omega$  denoting the rotor angular velocity,  $D$  the diffusion coefficient, and  $s$  the sedimentation coefficient) at unit loading concentration.<sup>76</sup>  $D$  and  $s$  are related via the frictional ratio  $F = (f/f_0)$  and the hydrodynamic scale relationship

$$D(s) = \frac{\sqrt{2}}{18\pi} kT s^{-1/2} (\eta F)^{-3/2} ((1 - \bar{v}\rho/\bar{v})^{-1/2}) \quad (3)$$

(with  $h$  and  $r$  denoting the solvent viscosity and density, respectively, and  $\bar{v}$  the partial specific volume of the macromolecules, calculated using the software SEDNTERP<sup>73</sup>). The partial specific volume for the htt peptides differs from that of globular proteins due to its unusual amino acid sequence composition and was determined to be 0.655 ml g<sup>-1</sup>. Initial fits to the data on htt<sup>NT</sup>Q<sub>10</sub>K<sub>2</sub> with a variety of values for  $\bar{v}$  in the calculated range and a single  $F$  value gave incompatible results, either with monomer MWs at expected values and oligomer MWs that were too large or monomer MWs that were unexpectedly low in fits with oligomer

MWs that were expected. From these data fits, it was clear that  $F$  was different comparing the monomer and multimer. For this reason, a fit to a model that allows for a variable  $F$  value for monomer *versus* oligomers was employed (i.e., a model that assumes a conformational change in assembly of oligomers from monomers), taking advantage of the known MW of the monomer as the basis for assembly of the oligomers in the fitting. Satisfactory fits to the data were generated with this approach, yielding an  $F$  value of 1.58 for the monomer and an  $F$  value of 1.24 for the oligomers.

## Aggregation and seeded elongation kinetics

Htt peptides were tested for their aggregation propensity and cross-seeding ability using an analytical HPLC sedimentation assay.<sup>69</sup> Disaggregated htt<sup>NT</sup>Q<sub>N</sub> peptides were incubated in PBSA (PBS plus 0.05% sodium azide, w/v) at 37 °C, aliquots were periodically removed and centrifuged, and the supernatants were analyzed by analytical HPLC.

For seeded elongation kinetics, disaggregated peptides were incubated at 37 °C in PBS either alone or in the presence of the 7.5% freshly grown aggregates delivered from the stock suspension described in Materials. The rate of monomer loss over incubation period was calculated and compared with the spontaneous aggregation rate in order to estimate the seeding efficiency.

For the oligomer stability experiment, 445  $\mu$ l of 620  $\mu$ M freshly disaggregated htt<sup>NT</sup>Q<sub>8</sub>K<sub>2</sub> in PBSA was incubated at 37 °C for 120 h. A 300- $\mu$ l aliquot was removed and centrifuged at 14,000 rpm in a tabletop centrifuge. The supernatant was carefully removed, and the pellet was suspended in 200  $\mu$ l of PBSA with gentle vortexing. A 5- $\mu$ l aliquot of the resulting suspension was removed and added to 30  $\mu$ l of formic acid and incubated for 30 min and then analyzed by analytical HPLC to determine the total peptide concentration (monomer plus aggregates) in the sample. For the dissociation kinetics, 25- $\mu$ l aliquots were removed and centrifuged 30 min at 14,000 rpm, and 5  $\mu$ l of each supernatant was added to 30  $\mu$ l of formic acid, and the amount of peptide in the sample was determined by analytical HPLC to yield the monomer concentration at each time point.

## ThT fluorescence

Aggregates were isolated as described above and resuspended in 350  $\mu$ l of PBS, to which a ThT stock solution was added to give a final ThT concentration of 40  $\mu$ M. Fluorescence emission spectra were collected using a Fluorolog-3 Research Spectrofluorometer (Horiba Jobin-Yvon). Samples were excited at 445 nm and emission spectra were recorded from 460 to 600 nm. Excitation and emission slits were at 2 nm and 5 nm, respectively.

## Tryptophan fluorescence measurements

Tryptophan fluorescence measurements on aggregates were conducted as previously described, using isolated aggregates resuspended in PBS.<sup>15</sup>

<sup>‡</sup> Available online at <http://www.AnalyticalUltracentrifugation.com>



## DLS measurements

DLS measurements were carried out on a Wyatt DynaPro light-scattering instrument. Approximately 80- $\mu$ l aliquots of ongoing aggregation reactions were transferred to wells of a 386-well plate and the autocorrelation functions were measured. The correlation curves were deconvoluted using Dynamics V6 software (Wyatt Technologies Corp.) to obtain size distribution and hydrodynamic radii ( $R_h$ ).

## Transmission electron microscopy

Aliquots of aggregation reaction of various htt peptide samples were placed on freshly glow-discharged carbon-coated grids (Electron Microscopy Sciences, Hatfield, PA) and incubated for 1 min, and the excess sample was wicked away with filter paper. The sample grid was then washed with deionized water, stained with 1% uranyl acetate (w/v) solution for 3 s, and blotted. Grids were imaged using Tecnai T12 microscope (FEI Co., Hillsboro, OR) operating at 120 kV and 30,000 $\times$  magnification and equipped with an ultrascan 1000 CCD camera (Gatan, Pleasanton, CA) with post-column magnification of 1.4 $\times$ . Aggregates visualized by this protocol almost certainly are present in the sampled solutions and are not induced artifacts; previously, we showed that a solution of an htt N-terminal fragment monomer solution, added to a grid and stained by the above procedure, is free of aggregates and identical in appearance to an empty mock-stained grid.<sup>15</sup>

## FTIR spectroscopy

To provide sufficient aggregates for analysis, we conducted reactions with monomers in the 0.5–1 mM range. This did not appreciably alter aggregation rates, however, presumably because of the low concentration dependence of the formation of these non-amyloid intermediates.<sup>15</sup> Aggregates were harvested by centrifugation at 14,000 rpm for 45 min on a bench top centrifuge, and the pellets were washed three times with PBS to remove traces of TFA and the other solutes. Pellets containing aggregates were then resuspended in 4  $\mu$ l of PBS at around 10 mg ml<sup>-1</sup> concentration, and spectra were acquired by placing the aggregate suspension between two polished CaF<sub>2</sub> windows using a BioCell module (BioTools, Inc.) on an ABB Bomem FTIR instrument. Data from a total of 400 scans were collected with 4 cm<sup>-1</sup> resolution at room temperature and averaged for each sample. Residual buffer absorption was subtracted and spectral components were identified from second-derivative minima using PROTA software.

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## References

1. Bates, G. P. & Benn, C. (2002). The polyglutamine diseases. In *Huntington's Disease* (Bates, G. P., Harper, P. S. & Jones, L., eds), pp. 429–472, Oxford University Press, Oxford, UK.
2. Wilburn, B., Rudnicki, D. D., Zhao, J., Weitz, T. M., Cheng, Y., Gu, X. F. *et al.* (2011). An antisense CAG repeat transcript at JPH3 locus mediates expanded polyglutamine protein toxicity in Huntington's disease-like 2 mice. *Neuron*, **70**, 427–440.
3. Morley, J. F., Brignull, H. R., Weyers, J. J. & Morimoto, R. I. (2002). The threshold for polyglutamine-expansion protein aggregation and cellular toxicity is dynamic and influenced by aging in *Caenorhabditis elegans*. *Proc. Natl Acad. Sci. USA*, **99**, 10417–10422.
4. Scherzinger, E., Sittler, A., Schweiger, K., Heiser, V., Lurz, R., Hasenbank, R. *et al.* (1999). Self-assembly of polyglutamine-containing huntingtin fragments into amyloid-like fibrils: implications for Huntington's disease pathology. *Proc. Natl Acad. Sci. USA*, **96**, 4604–4609.
5. Chen, S., Berthelie, V., Yang, W. & Wetzel, R. (2001). Polyglutamine aggregation behavior in vitro supports a recruitment mechanism of cytotoxicity. *J. Mol. Biol.* **311**, 173–182.
6. Chen, S., Ferrone, F. & Wetzel, R. (2002). Huntington's disease age-of-onset linked to polyglutamine aggregation nucleation. *Proc. Natl Acad. Sci. USA*, **99**, 11884–11889.
7. Bhattacharyya, A. M., Thakur, A. K. & Wetzel, R. (2005). Polyglutamine aggregation nucleation: thermodynamics of a highly unfavorable protein folding reaction. *Proc. Natl Acad. Sci. USA*, **102**, 15400–15405.
8. Slepko, N., Bhattacharyya, A. M., Jackson, G. R., Steffan, J. S., Marsh, J. L., Thompson, L. M. & Wetzel, R. (2006). Normal-repeat-length polyglutamine peptides accelerate aggregation nucleation and cytotoxicity of expanded polyglutamine proteins. *Proc. Natl. Acad. Sci. USA*, **103**, 14367–14372.
9. Kar, K., Jayaraman, M., Sahoo, B., Kodali, R. & Wetzel, R. (2011). Critical nucleus size for disease-related polyglutamine aggregation is repeat-length dependent. *Nat. Struct. Mol. Biol.* **18**, 328–336.
10. Nozaki, K., Onodera, O., Takano, H. & Tsuji, S. (2001). Amino acid sequences flanking polyglutamine stretches influence their potential for aggregate formation. *NeuroReport*, **12**, 3357–3364.
11. Dehay, B. & Bertolotti, A. (2006). Critical role of the proline-rich region in Huntingtin for aggregation and cytotoxicity in yeast. *J. Biol. Chem.* **281**, 35608–35615.
12. Duennwald, M. L., Jagadish, S., Muchowski, P. J. & Lindquist, S. (2006). Flanking sequences profoundly alter polyglutamine toxicity in yeast. *Proc. Natl Acad. Sci. USA*, **103**, 11045–11050.



13. Bhattacharyya, A., Thakur, A. K., Chellgren, V. M., Thiagarajan, G., Williams, A. D., Chellgren, B. W. *et al.* (2006). Oligoproline effects on polyglutamine conformation and aggregation. *J. Mol. Biol.* **355**, 524–535.
14. Darnell, G., Orgel, J. P., Pahl, R. & Meredith, S. C. (2007). Flanking polyproline sequences inhibit beta-sheet structure in polyglutamine segments by inducing PPII-like helix structure. *J. Mol. Biol.* **374**, 688–704.
15. Thakur, A. K., Jayaraman, M., Mishra, R., Thakur, M., Chellgren, V. M., Byeon, I. J. *et al.* (2009). Polyglutamine disruption of the huntingtin exon 1 N terminus triggers a complex aggregation mechanism. *Nat. Struct. Mol. Biol.* **16**, 380–389.
16. Tam, S., Spiess, C., Auyeung, W., Joachimiak, L., Chen, B., Poirier, M. A. & Frydman, J. (2009). The chaperonin TRiC blocks a huntingtin sequence element that promotes the conformational switch to aggregation. *Nat. Struct. Mol. Biol.* **16**, 1279–1285.
17. Saunders, H. M. & Bottomley, S. P. (2009). Multi-domain misfolding: understanding the aggregation pathway of polyglutamine proteins. *Protein Eng. Des. Sel.* **22**, 447–451.
18. de Chiara, C., Menon, R. P., Dal Piaz, F., Calder, L. & Pastore, A. (2005). Polyglutamine is not all: the functional role of the AXH domain in the ataxin-1 protein. *J. Mol. Biol.* **354**, 883–893.
19. Ellisdon, A. M., Thomas, B. & Bottomley, S. P. (2006). The two-stage pathway of ataxin-3 fibrillogenesis involves a polyglutamine-independent step. *J. Biol. Chem.* **281**, 16888–16896.
20. Ignatova, Z., Thakur, A. K., Wetzel, R. & Gierasch, L. M. (2007). In-cell aggregation of a polyglutamine-containing chimera is a multistep process initiated by the flanking sequence. *J. Biol. Chem.* **282**, 36736–36743.
21. Sivanandam, V. N., Jayaraman, M., Hoop, C. L., Kodali, R., Wetzel, R. & van der Wel, P. C. (2011). The aggregation-enhancing huntingtin N-terminus is helical in amyloid fibrils. *J. Am. Chem. Soc.* **133**, 4558–4566.
22. Chellgren, B. W., Miller, A. F. & Creamer, T. P. (2006). Evidence for polyproline II helical structure in short polyglutamine tracts. *J. Mol. Biol.* **361**, 362–371.
23. Masino, L., Kelly, G., Leonard, K., Trotter, Y. & Pastore, A. (2002). Solution structure of polyglutamine tracts in GST–polyglutamine fusion proteins. *FEBS Lett.* **513**, 267–272.
24. Klein, F. A., Pastore, A., Masino, L., Zeder-Lutz, G., Nierengarten, H., Oulad-Abdelghani, M. *et al.* (2007). Pathogenic and non-pathogenic polyglutamine tracts have similar structural properties: towards a length-dependent toxicity gradient. *J. Mol. Biol.* **371**, 235–244.
25. Zhang, Q. C., Yeh, T. L., Leyva, A., Frank, L. G., Miller, J., Kim, Y. E. *et al.* (2011). A compact beta model of huntingtin toxicity. *J. Biol. Chem.* **286**, 8188–8196.
26. Venyaminov, S. & Kalnin, N. N. (1990). Quantitative IR spectrophotometry of peptide compounds in water (H<sub>2</sub>O) solutions. I. Spectral parameters of amino acid residue absorption bands. *Biopolymers*, **30**, 1243–1257.
27. Jackson, M. & Mantsch, H. H. (1995). The use and misuse of FTIR spectroscopy in the determination of protein structure. *Crit. Rev. Biochem. Mol. Biol.* **30**, 95–120.
28. Jayaraman, M., Thakur, A. K., Kar, K., Kodali, R. & Wetzel, R. (2011). Assays for studying nucleated aggregation of polyglutamine proteins. *Methods*, **53**, 246–254.
29. Wood, S. J., Maleeff, B., Hart, T. & Wetzel, R. (1996). Physical, morphological and functional differences between pH 5.8 and 7.4 aggregates of the Alzheimer's peptide Ab. *J. Mol. Biol.* **256**, 870–877.
30. Natalello, A., Frana, A. M., Relini, A., Apicella, A., Invernizzi, G., Casari, C. *et al.* (2011). A major role for side-chain polyglutamine hydrogen bonding in irreversible ataxin-3 aggregation. *PLoS ONE*, **6**.
31. Bader, R., Seeliger, M. A., Kelly, S. E., Ilag, L. L., Meersman, F., Limones, A. *et al.* (2006). Folding and fibril formation of the cell cycle protein Cks1. *J. Biol. Chem.* **281**, 18816–18824.
32. Arkin, I. T. (2006). Isotope-edited IR spectroscopy for the study of membrane proteins. *Curr. Opin. Chem. Biol.* **10**, 394–401.
33. Chirgadze, Y. N., Fedorov, O. V. & Trushina, N. P. (1975). Estimation of amino acid residue side-chain absorption in the infrared spectra of protein solutions in heavy water. *Biopolymers*, **14**, 679–694.
34. Besson, F., Raimbault, C., Hourdou, M. & Buchet, R. (1996). Solvent-induced conformational modifications of iturin A: an infrared and circular dichroism study of a L,D-lipopeptide of *Bacillus subtilis*. *Spectrochim. Acta, Part A*, **52**, 793–803.
35. Stuart, B. H. (2004). *Infrared Spectroscopy: Fundamentals and Applications*. Wiley, Chichester, UK.
36. Crick, S. L., Jayaraman, M., Frieden, C., Wetzel, R. & Pappu, R. V. (2006). Fluorescence correlation spectroscopy shows that monomeric polyglutamine molecules form collapsed structures in aqueous solutions. *Proc. Natl Acad. Sci. USA*, **103**, 16764–16769.
37. Wang, X., Vitalis, A., Wyczalkowski, M. A. & Pappu, R. V. (2006). Characterizing the conformational ensemble of monomeric polyglutamine. *Proteins*, **63**, 297–311.
38. Vitalis, A., Wang, X. & Pappu, R. V. (2007). Quantitative characterization of intrinsic disorder in polyglutamine: insights from analysis based on polymer theories. *Biophys. J.* **93**, 1923–1937.
39. Walters, R. H. & Murphy, R. M. (2009). Examining polyglutamine peptide length: a connection between collapsed conformations and increased aggregation. *J. Mol. Biol.* **393**, 978–992.
40. Atwal, R. S., Xia, J., Pinchev, D., Taylor, J., Epand, R. M. & Truant, R. (2007). Huntingtin has a membrane association signal that can modulate huntingtin aggregation, nuclear entry and toxicity. *Hum. Mol. Genet.* **16**, 2600–2615.
41. Kim, M. W., Chelliah, Y., Kim, S. W., Otwinowski, Z. & Bezprozvanny, I. (2009). Secondary structure of Huntingtin amino-terminal region. *Structure*, **17**, 1205–1212.
42. Ossato, G., Digman, M. A., Aiken, C., Lukacovich, T., Marsh, J. L. & Gratton, E. (2010). A two-step path to inclusion formation of huntingtin peptides revealed by number and brightness analysis. *Biophys. J.* **98**, 3078–3085.
43. Mishra, R., Jayaraman, M., Roland, B. P., Landrum, E., Fullam, T., Kodali, R. *et al.* (2012). Inhibiting the nucleation of amyloid structure in a huntingtin fragment by targeting  $\alpha$ -helix-rich oligomeric intermediates. *J. Mol. Biol.* **415**, 120–137.

44. Ferrone, F. (1999). Analysis of protein aggregation kinetics. *Methods Enzymol.* **309**, 256–274.
45. Serio, T. R., Cashikar, A. G., Kowal, A. S., Sawicki, G. J., Moslehi, J. J., Serpell, L. *et al.* (2000). Nucleated conformational conversion and the replication of conformational information by a prion determinant. *Science*, **289**, 1317–1321.
46. Kodali, R. & Wetzel, R. (2007). Polymorphism in the intermediates and products of amyloid assembly. *Curr. Opin. Struct. Biol.* **17**, 48–57.
47. Mukhopadhyay, S., Krishnan, R., Lemke, E. A., Lindquist, S. & Deniz, A. A. (2007). A natively unfolded yeast prion monomer adopts an ensemble of collapsed and rapidly fluctuating structures. *Proc. Natl Acad. Sci. USA*, **104**, 2649–2654.
48. Chen, Y. R. & Glabe, C. G. (2006). Distinct early folding and aggregation properties of Alzheimer amyloid-beta peptides Abeta40 and Abeta42: stable trimer or tetramer formation by Abeta42. *J. Biol. Chem.* **281**, 24414–24422.
49. Wildegger, G. & Kiefhaber, T. (1997). Three-state model for lysozyme folding: triangular folding mechanism with an energetically trapped intermediate. *J. Mol. Biol.* **270**, 294–304.
50. Capaldi, A. P., Shastry, M. C., Kleanthous, C., Roder, H. & Radford, S. E. (2001). Ultrarapid mixing experiments reveal that Im7 folds via an on-pathway intermediate. *Nat. Struct. Biol.* **8**, 68–72.
51. Powers, E. T. & Ferrone, F. (2009). Kinetic models for protein misfolding and association. In *Protein Misfolding Diseases: Current and Emerging Principles and Therapies* (Dobson, C. M., Kelly, J. W. & Ramirez-Alvarado, M., eds), pp. 73–92, Wiley, New York, NY.
52. Olshina, M. A., Angley, L. M., Ramdzan, Y. M., Tang, J., Bailey, M. F., Hill, A. F. & Hatters, D. M. (2010). Tracking mutant huntingtin aggregation kinetics in cells reveals three major populations that include an invariant oligomer pool. *J. Biol. Chem.* **285**, 21807–21816.
53. Williamson, T. E., Vitalis, A., Crick, S. L. & Pappu, R. V. (2010). Modulation of polyglutamine conformations and dimer formation by the N-terminus of huntingtin. *J. Mol. Biol.* **396**, 1295–1309.
54. Legleiter, J., Mitchell, E., Lotz, G. P., Sapp, E., Ng, C., DiFiglia, M. *et al.* (2010). Mutant huntingtin fragments form oligomers in a polyglutamine length-dependent manner in vitro and in vivo. *J. Biol. Chem.* **285**, 14777–14790.
55. Abedini, A. & Raleigh, D. P. (2009). A role for helical intermediates in amyloid formation by natively unfolded polypeptides? *Phys. Biol.* **6**, 015005.
56. Anderson, V. L., Ramlall, T. F., Rospigliosi, C. C., Webb, W. W. & Eliezer, D. (2010). Identification of a helical intermediate in trifluoroethanol-induced alpha-synuclein aggregation. *Proc. Natl Acad. Sci. USA*, **107**, 18850–18855.
57. Kirkitadze, M. D., Condrón, M. M. & Teplow, D. B. (2001). Identification and characterization of key kinetic intermediates in amyloid beta-protein fibrillogenesis. *J. Mol. Biol.* **312**, 1103–1119.
58. Liu, G., Prabhakar, A., Aucoin, D., Simon, M., Sparks, S., Robbins, K. J. *et al.* (2010). Mechanistic studies of peptide self-assembly: transient alpha-helices to stable beta-sheets. *J. Am. Chem. Soc.* **132**, 18223–18232.
59. Sekhar, A. & Udgaonkar, J. B. (2011). Fluoroalcohol-induced modulation of the pathway of amyloid protofibril formation by Barstar. *Biochemistry*, **50**, 805–819.
60. Singh, Y., Sharpe, P. C., Hoang, H. N., Lucke, A. J., McDowall, A. W., Bottomley, S. P. & Fairlie, D. P. (2011). Amyloid formation from an alpha-helix peptide bundle is seeded by 3(10)-helix aggregates. *Chemistry*, **17**, 151–160.
61. West, M. W., Wang, W., Patterson, J., Mancias, J. D., Beasley, J. R. & Hecht, M. H. (1999). De novo amyloid proteins from designed combinatorial libraries. *Proc. Natl Acad. Sci. USA*, **96**, 11211–11216.
62. Lakhani, V. V., Ding, F. & Dokholyan, N. V. (2010). Polyglutamine induced misfolding of huntingtin exon1 is modulated by the flanking sequences. *PLoS Comput. Biol.* **6**, e1000772.
63. Dlugosz, M. & Trylska, J. (2011). Secondary structures of native and pathogenic huntingtin N-terminal fragments. *J. Phys. Chem. B*, **115**, 11597–11608.
64. Bishop, M. F. & Ferrone, F. A. (1984). Kinetics of nucleation-controlled polymerization. A perturbation treatment for use with a secondary pathway. *Biophys. J.* **46**, 631–644.
65. Padrick, S. B. & Miranker, A. D. (2002). Islet amyloid: phase partitioning and secondary nucleation are central to the mechanism of fibrillogenesis. *Biochemistry*, **41**, 4694–4703.
66. Tanaka, M., Collins, S. R., Toyama, B. H. & Weissman, J. S. (2006). The physical basis of how prion conformations determine strain phenotypes. *Nature*, **442**, 585–589.
67. Sharma, D., Shinchuk, L. M., Inouye, H., Wetzel, R. & Kirschner, D. A. (2005). Polyglutamine homopolymers having 8–45 residues form slablike beta-crystallite assemblies. *Proteins*, **61**, 398–411.
68. Thakur, A. & Wetzel, R. (2002). Mutational analysis of the structural organization of polyglutamine aggregates. *Proc. Natl Acad. Sci. USA*, **99**, 17014–17019.
69. O'Nuallain, B., Thakur, A. K., Williams, A. D., Bhattacharyya, A. M., Chen, S., Thiagarajan, G. & Wetzel, R. (2006). Kinetics and thermodynamics of amyloid assembly using a high-performance liquid chromatography-based sedimentation assay. *Methods Enzymol.* **413**, 34–74.
70. Kuipers, B. J. & Gruppen, H. (2007). Prediction of molar extinction coefficients of proteins and peptides using UV absorption of the constituent amino acids at 214 nm to enable quantitative reverse phase high-performance liquid chromatography-mass spectrometry analysis. *J. Agric. Food Chem.* **55**, 5445–5451.
71. Sreerama, N. & Woody, R. W. (2000). Estimation of protein secondary structure from circular dichroism spectra: comparison of CONTIN, SELCON, and CDSSTR methods with an expanded reference set. *Anal. Biochem.* **287**, 252–260.
72. Dam, J. & Schuck, P. (2004). Calculating sedimentation coefficient distributions by direct modeling of sedimentation velocity concentration profiles. *Methods Enzymol.* **384**, 185–212.
73. Schuck, P. (2000). Size-distribution analysis of macromolecules by sedimentation velocity ultracentrifugation

- and lamm equation modeling. *Biophys. J.* **78**, 1606–1619.
74. Lebowitz, J., Lewis, M. S. & Schuck, P. (2002). Modern analytical ultracentrifugation in protein science: a tutorial review. *Protein Sci.* **11**, 2067–2079.
75. Schuck, P. (2003). On the analysis of protein self-association by sedimentation velocity analytical ultracentrifugation. *Anal. Biochem.* **320**, 104–124.
76. Lamm, O. (1929). Die Differentialgleichung der Ultrazentrifugierung. *Ark. Mat., Astron. Fys.* **21B**, 1–4.