



Contents lists available at ScienceDirect

Life Sciences

journal homepage: [www.elsevier.com/locate/lifescie](http://www.elsevier.com/locate/lifescie)

## Minireview

## Control of respiratory and cardiovascular functions by leptin

M. Bassi<sup>a,\*</sup>, I.F. Werner<sup>a</sup>, D.B. Zoccal<sup>a</sup>, J.V. Menani<sup>a</sup>, E. Colombari<sup>a</sup>, J.E. Hall<sup>b</sup>, A.A. da Silva<sup>b</sup>,  
J.M. do Carmo<sup>b</sup>, D.S.A. Colombari<sup>a</sup>

<sup>a</sup> Department of Physiology and Pathology, School of Dentistry, São Paulo State University (UNESP), Araraquara, SP, Brazil

<sup>b</sup> Department of Physiology and Biophysics, University of Mississippi Medical Center, Jackson, MS, USA

## ARTICLE INFO

## Article history:

Received 18 September 2014

Accepted 23 January 2015

Available online xxxx

## Keywords:

Obesity

Leptin

Chemoreflex

Sympathetic nerve activity

Breathing

Blood pressure

## ABSTRACT

Leptin, a peptide hormone produced by adipose tissue, acts in brain centers that control critical physiological functions including metabolism, breathing and cardiovascular function. The importance of leptin for respiratory control is evident by the fact that leptin deficient mice exhibit impaired ventilatory responses to carbon oxide (CO<sub>2</sub>), which can be corrected by intracerebroventricular leptin replacement therapy. Leptin is also recognized as an important link between obesity and hypertension. Humans and animal models lacking either leptin or functional leptin receptors exhibit many characteristics of the metabolic syndrome, including hyperinsulinemia, insulin resistance, hyperglycemia, dyslipidemia and visceral adiposity, but do not exhibit increased sympathetic nerve activity (SNA) and have normal to lower blood pressure (BP) compared to lean controls. Even though previous studies have extensively focused on the brain sites and intracellular signaling pathways involved in leptin effects on food intake and energy balance, the mechanisms that mediate the actions of leptin on breathing and cardiovascular function are only beginning to be elucidated. This mini-review summarizes recent advances on the effects of leptin on cardiovascular and respiratory control with emphasis on the neural control of respiratory function and autonomic activity.

© 2015 Published by Elsevier Inc.

## Contents

Introduction	0
Leptin and breathing control	0
Leptin and central chemoreception	0
Involvement of melanocortin system in mediating leptin's effects on ventilation	0
Leptin and peripheral control of breathing function	0
Breathing disorders and impairment of leptin function in humans	0
Leptin and cardiovascular function	0
Leptin regulates sympathetic outflow and blood pressure	0
Leptin acts in different brain regions to regulate SNS activity and blood pressure	0
Intracellular signaling and specific CNS areas that may mediate differential control of cardiovascular and metabolic functions by leptin	0
Role of the CNS melanocortin system in mediating the effects of leptin on SNS activity and BP regulation	0
Perspectives and conclusion	0
Conflict of interest statement	0
Uncited references	0
Acknowledgements	0
References	0

## Introduction

Obesity is a major public health problem worldwide. The genesis of obesity is multifactorial involving genetic, metabolic and environmental aspects. Progress in endocrinology research shows that the adipocyte

\* Corresponding author at: Department of Physiology and Pathology, School of Dentistry of Araraquara, São Paulo State University, 1680 Humaitá St., Araraquara, São Paulo CEP: 14801-903, Brazil.

E-mail address: [mbassi@foar.unesp.br](mailto:mbassi@foar.unesp.br) (M. Bassi).

is an endocrine tissue producing several active substances, such as interleukin-6, tumor necrosis factors- $\alpha$ , adiponectin and leptin, which modulate many physiological functions. In this review, we focus on the cardiorespiratory actions of leptin.

Leptin circulates freely in the plasma and crosses the blood–brain barrier via a saturable receptor-mediated transport system [64] to enter central nervous system centers (CNSs) where it regulates neural pathways that control appetite [37], sympathetic nerve activity (SNA) and thermogenesis [58,75]. In addition, previous studies have suggested that leptin stimulates chemorespiratory responses [4,6,45].

Leptin receptors (LRs) belong to the class I cytokine receptor superfamily [50,83]. Alternative splicing of the LR gene generates 6 leptin receptor isoforms, termed from Ob-Ra to Ob-Rf, which have an identical extracellular N-terminal. Ob-Re is the only soluble receptor form, probably binding circulating leptin and affecting its stability and availability [32,88]. Four of the remaining 5 isoforms have short C-terminal domains and are considered to be mainly involved in endocytosis and transport of leptin across the blood–brain barrier [3]. The isoform Ob-Rb, however, has a long intracellular domain and is essential for mediating leptin's intracellular signal transduction [84].

The hypothalamic arcuate nucleus (ARC) was initially considered the main site of leptin actions, however, increasing evidences suggest that leptin acts on a more extensive brain network (Grill 2006). For example, functional LRs are present in the nucleus of the solitary tract (NTS) [43,60], an important center involved in cardiorespiratory function.

Stimulation of LR by leptin activates janus tyrosine kinases (JAK), especially JAK2 [33]. In the central nervous system (CNS), leptin increases the activity of JAK2 to trigger three major intracellular pathways: 1) phosphorylation of tyrosine (Tyr) residue 1138 to recruit latent signal transducers and activators of transcription 3 (STAT3) to the LR-JAK2 complex, resulting in the phosphorylation and nuclear translocation of STAT3 to regulate transcription; 2) insulin receptor substrate (IRS2) phosphorylation which activates phosphatidylinositol 3-kinase (PI3K) which appears to be involved in regulating rapid non-genomic events affecting neuronal activity and neuropeptide release; and 3) Tyr985 phosphorylation which recruits the tyrosine phosphatase (SHP2) to activate ERK (MAPK). Although the roles of these intracellular signaling pathways in mediating the various actions of leptin are the subject of intense investigation, especially on appetite behavior [27], their importance in SNA and breathing control is only beginning to be elucidated.

Strong evidence shows that leptin requires activation of the brain melanocortin system, including activation of proopiomelanocortin (POMC) neurons and melanocortin 4 receptors (MC4R) to exert most of its effects on blood pressure (BP) and ventilatory function [5,21,72]. Thus, the focus of this mini-review is on the brain circuits and potential mechanisms that mediate the effects of leptin on respiratory function and cardiovascular regulation.

## Leptin and breathing control

### *Leptin and central chemoreception*

Accumulated evidence suggests a role for leptin in control of breathing. Initial studies evaluating the ventilatory responses to CO<sub>2</sub> in leptin-deficient (ob/ob) mice demonstrated impairment of breathing function in these mice [68,82]. This attenuated hypercapnic ventilatory response observed in ob/ob mice was improved after 3 days of systemic leptin administration suggesting an important stimulatory effect of leptin on breathing [68]. In addition, a study performed in anesthetized rats showed that acute systemic infusion of leptin (for 90 min) elicited a long-lasting increase in the amplitude of phrenic nerve discharge that remained elevated for over 1 h after terminating the leptin infusion [11]. Moreover, we demonstrated that 4th ventricle leptin administration

for 3 days also enhanced the ventilatory responses to CO<sub>2</sub> indicating that the central action of leptin facilitates the central chemoreflex [4].

In order to better understand the CNS mechanisms activated by leptin that modulate chemosensory control of ventilation, previous studies investigated the effects of leptin administration into specific medullary brain areas involved with breathing control. Leptin administration into the NTS, a primary site of peripheral chemorespiratory afferents of the brainstem of anesthetized rats increased respiratory motor output and ventilatory response to CO<sub>2</sub> potentially via inhibition of the Hering–Breuer reflex [44,45]. It was hypothesized that elevated PaCO<sub>2</sub> reduces the effectiveness of the Breuer–Hering modulation of respiratory pattern that facilitates elimination of CO<sub>2</sub> (as described by [63]) and that the stimulatory effect of leptin on chemoreflex responses may depend on a reduction of the effectiveness of Breuer–Hering reflex.

Leptin injections into the NTS also attenuate the cardiovagal component of the baroreceptor reflex [1] and potentiate the sympathoexcitatory responses evoked by the activation of the chemoreflex [14]. In addition, systemic administration of leptin increases c-fos expression in the neurons of the caudal NTS that express LR [29]; Elmquist et al., 1998; [37], indicating that leptin may activate NTS neurons involved with the cardiorespiratory reflex.

In addition to its effects in the NTS, leptin may also contribute to the chemoreflex by acting in the ventral surface of the medulla where several nuclei involved in breathing control are located. For instance, administration of leptin for 3 consecutive days into the rostral ventrolateral region of the medulla increased baseline ventilation and hypercapnic ventilatory response in ob/ob mice [5]. Although multiple mechanisms involved in chemoreception at level of the ventral surface of the medulla have been described including modulation of glutamatergic neurons of the retrotrapezoid nuclei (RTN) [40] and purinergic glial cells that release adenosine 5'-triphosphate (ATP) in response to CO<sub>2</sub> stimulation [66,87], the mechanisms by which leptin contributes to the chemoreflex is still unclear and remains an important area for investigation.

### *Involvement of melanocortin system in mediating leptin's effects on ventilation*

Leptin depolarizes POMC neurons leading to the release of alpha-melanocyte stimulating hormone ( $\alpha$ -MSH) which, in turn, activates the MC3/4R located in several hypothalamic nuclei as well as in the brainstem [17,65].

Only a few studies have examined the participation of the melanocortin system in mediating the effects of leptin on ventilation. Polotsky et al. [72] investigated the ventilatory responses of obese agouti yellow mice, a model that overexpresses the agouti protein which inhibits MC3/4R. They reported that agouti yellow mice exhibited attenuated ventilatory responses to CO<sub>2</sub> but a normal ventilatory response to hypoxia, suggesting that the melanocortin system may play an important role in mediating the ventilatory responses to hypercapnia.

We found that chronic central MC3/4R antagonism for 6 days reduced the ventilatory response to hypercapnia in rats and abolished leptin's ability to increase baseline ventilation. Our data suggest that the effects of leptin on ventilation depend on the activation of the brain-melanocortin system. We also demonstrated attenuated ventilatory responses to CO<sub>2</sub> in mice with LR deficiency specifically in POMC neurons, reinforcing the concept that leptin-induced improvement of ventilatory function is mediated by the brain melanocortin system [5].

Besides the CNS action of leptin in modulating ventilation, leptin has an important role in controlling bronchial diameter [2,10,47,78]. Previous studies showed that the absence of leptin action is the main cause of increased airway resistance present in obese leptin-deficient (ob/ob) mice and leptin receptor-deficient (db/db) mice [2]. It is important to note that leptin administration in trachea rings evoked no changes in the bronchial diameter [67] whereas intracerebroventricular (i.c.v.) administration of leptin for 5 days decreased airway resistance

[2]. These findings suggest that the effects of leptin on airway resistance may also be mediated by leptin's actions on the CNS. Moreover, leptin-induced modulation of respiratory resistance appears to be independent of the brain melanocortin system since mice lacking MC3/4R exhibit normal airway resistance [2].

#### *Leptin and peripheral control of breathing function*

In addition to leptin's CNS action to modulate respiratory function, previous studies suggest direct effects of leptin on peripheral tissues involved with ventilatory control, including arterial chemoreceptors and lung tissue [16,38,62]. Leptin appears to be secreted by various epithelial tissues including bronchial epithelial cells (BECs) and type II pneumocytes [85] and high levels of LRs have been observed in proximal airway biopsies [85] where leptin is thought to modulate inflammatory response [56].

Peripheral chemoreceptors localized predominantly within the carotid bodies also present LR isoform b in type-1 cells [73]. These cells play an integral role in detecting changes in PO<sub>2</sub> by transducing this chemical signal to sensory afferent neurons within the petrosal (PG) and nodose (NG) ganglia to trigger brainstem autonomic reflex pathways [31]. Previous studies demonstrated that not only carotid body glomus cells express LR, but that LRs are also present in neurons within both the PG and NG [62]. In the same study intravenous injections of leptin were shown to induce phosphorylation of signal transducer and activator of transcription 3 (pSTAT3), fos and Fra-1 within carotid body cells, similar to the response produced by hypoxia. Taken together, these observations also point toward a potential contribution of leptin in the peripheral chemoreflex response.

#### *Breathing disorders and impairment of leptin function in humans*

Increased leptin levels have been reported in obese subject's leading to a state of leptin resistance. In obese patients, high concentrations of serum leptin are associated with reduced respiratory drive and impaired hypercapnic responses in men and women, suggesting resistance to the effects of leptin on respiratory function [8,55,70]. However, hypoxemia stimulates leptin secretion [39], suggesting that leptin resistance and hyperleptinemia might be caused by hypoventilation. In support of this concept, patients with obstructive sleep apnea syndrome (OSAS) who have high levels of leptin presented normal plasma leptin levels after nasal continuous positive airway pressure (NCPAP), suggesting that once the hypoxemia is corrected, leptin levels return to normal [12,71]. Similar results were found in patients with obese hypoventilation syndrome (OHS) who used non-invasive ventilation. The reduction of the leptin levels after the treatment in this case appears to be independent of any change in body weight [89].

Obesity-induced breathing disorders also lead to cardiovascular complications, including arrhythmias and hypertension. Hypoxia, resulting from obstructive apneic episodes, is a potent stimulator of SNA via a complex reflex mechanism that alters heart rate and BP. During the apneic episode, the combination of hypoxia and an absence of airflow result in carotid body chemoreceptor stimulation, leading to reflex bradycardia via vagal afferents [18,19]. However, in the presence of airflow, in the postapneic ventilation phase, a tachycardia occurs due to the inhibition of parasympathetic outflow and unopposed sympathetic outflow to the heart [51]. The long-term effects of OSA are not well understood, although autonomic nervous system dysregulation with chronic sympathetic activation and development of systemic hypertension are usually present.

Leptin may contribute to the development of hypertension caused by hypoxia. As mentioned, hypoxia increases leptin release from adipocytes. Chronic leptin infusion raises BP due to the activation of renal sympathetic nerve activity [41]. This effect of hyperleptinemia on BP seems opposite to the resistance to leptin's anorexic and respiratory

effects. While the excess of leptin fails to modulate appetite and ventilation, its action on sympathetic activity appears to remain effective.

#### **Leptin and cardiovascular function**

##### *Leptin regulates sympathetic outflow and blood pressure*

Leptin not only plays a role in the modulation of breathing and regulation of SNA to tissues involved in the breathing process but also modulates SNA to other organs, some of which contribute to the regulation of BP. For instance, acute intravenous or i.c.v. administration of leptin increased SNA to the brown adipose tissue, kidneys and adrenal gland in lean rats [28,42]. Acute hyperleptinemia also increases muscle SNA, as assessed by microneurography [54]. Chronic infusions of leptin to produce increases in circulating leptin levels comparable to those found in severe obesity evoked sustained increases in BP that can be completely prevented by  $\alpha$  and  $\beta$  adrenergic receptor blockade [9]. Leptin-mediated increases in BP are gradual and occur over several days, indicating a slow-acting mechanism consistent with the modest increases in renal SNA and increased renal tubular sodium reabsorption [79]. Although the chronic hypertensive effects of leptin in lean animals are modest, they are more significant when taking into account the accompanying marked decreases in food intake and weight loss which would normally tend to lower SNA and BP.

A major role for leptin in contributing to increased BP also comes from the studies of Lim and colleagues who showed that increases in BP and renal SNA in obese rabbits fed with a high fat diet were attenuated by acute (90 min) i.c.v. administration of a selective leptin receptor antagonist [53]. Thus, blockade of the actions of endogenous leptin lowers BP in obese animals, further supporting the concept that leptin, at physiological concentrations, can cause chronic increases in BP, at least in experimental animals, and may contribute to obesity induced hypertension. Moreover, mice with leptin deficiency (ob/ob mice) are extremely obese and have many metabolic abnormalities, including insulin resistance, hyperinsulinemia, and dyslipidemia which have been suggested to raise BP. However, mice with leptin deficiency are not hypertensive and tend to have lower BP and reduced SNA compared to lean control mice [23,57]. Similar findings are observed in humans with leptin deficiency who also exhibit early-onset morbid obesity and many characteristics of the metabolic syndrome but these individuals usually are not hypertensive and do not have evidence of increased SNA [69]. In fact, humans with leptin gene mutation show postural hypotension and attenuated renin-angiotensin-aldosterone system responses to upright posture [69]. Collectively, these observations support a role for leptin as a link between obesity, increased SNA and elevated BP.

The effects of leptin to increase SNA and BP however are partially counterbalanced by metabolic actions of leptin. For example, leptin decreases appetite and increases energy expenditure which tend to reduce adiposity and cause rapid weight loss, at least in lean subjects who are sensitive to the metabolic effects of leptin. These effects would tend to reduce BP. In addition, leptin also stimulates endothelial-derived nitric oxide (NO) formation, at least in subjects with normal endothelial function. Frühbeck [30] showed, for example, that acute infusion of leptin increased serum NO concentrations and after the inhibition of NO synthesis leptin significantly raised BP. After SNA blockade, however, acute leptin infusion reduced BP [30]. Blockade of NO synthesis also greatly exacerbated the chronic effects of leptin to raise BP and heart rate (HR) [49]. Thus, to the extent that obesity causes endothelial dysfunction and impaired NO formation, one might expect greater leptin-mediated increases in BP than in lean subjects, especially if obesity does not induce resistance to the SNA responses to leptin. Moreover, if obesity is associated with resistance to the anorexic effects of leptin with preserved effects on SNA, as previously suggested [59], this would amplify the hypertensive effects of leptin since the effects of leptin to cause weight loss and associated decreases in BP might be attenuated.



# Leptin acts in different brain regions to regulate SNS activity and blood pressure

High levels of leptin receptor mRNA and protein are expressed in the forebrain, especially in the ventromedial hypothalamus, arcuate nucleus and dorsomedial areas of the hypothalamus, as well as in vasomotor centers of the brainstem [29,59]. Although the brain centers that mediate leptin's action on SNA and BP have not been precisely mapped, hypothalamic centers as well as certain extra-hypothalamic regions (e.g. brainstem, subfornical organ – SFO) appear to be important in mediating the effects of leptin on SNA and BP [59]. Acute microinjections of leptin into the ARC increase SNA to the kidneys and to brown adipose tissue (BAT) [74], while site specific ARC deletion of LR markedly attenuates the rise in renal and BAT SNA evoked by leptin, suggesting that the ARC is an important site for leptin-mediated modulation of SNSA to several tissues [74]. In fact, deletion of leptin receptors only in POMC neurons, which comprise an important portion of the neuronal types within the ARC, prevents the rise in BP evoked by chronic hyperleptinemia [24, 25]. Other nuclei in the hypothalamus have also been implicated in the effects of leptin on SNA. The ventromedial and dorsomedial hypothalamus, for example, appear to contribute to leptin-mediated increases in SNA to the kidneys, skeletal muscle and BAT [59].

Extra-hypothalamic regions may also play a role in mediating leptin's effect on SNA. Microinjection of leptin into the NTS in the brainstem increases renal SNA and acutely raised BP [58]. Additionally, intracarotid injection of leptin excited presympathetic neurons of the rostral ventrolateral medulla (RVLM) increasing the renal SNA, suggesting that leptin has a direct action on ventral centers of the medulla [91]. Young and colleagues [90] showed that mice with specific deletion of LR in SFO neurons had normal BAT SNA responses to systemic or i.c.v. administration of leptin but did not exhibit the expected increase in renal SNA. Collectively, these studies suggest that leptin may act on several brain regions in concert to regulate SNA.

## Intracellular signaling and specific CNS areas that may mediate differential control of cardiovascular and metabolic functions by leptin

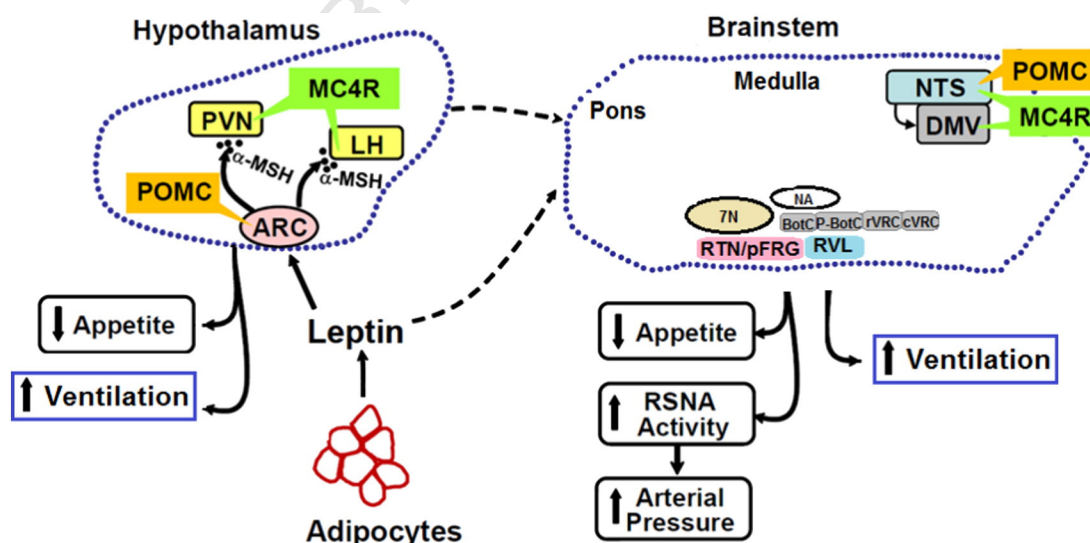
Deletion of STAT3 specifically in POMC neurons attenuated leptin's ability to raise BP but had only minor effects on leptin's actions on food intake and energy expenditure [27,34].

Previous acute studies also indicate that the IRS2–PI3K pathway may contribute to leptin's effect on SNA and BP. For instance, pharmacological blockade of PI3K abolished the acute effects of leptin to increase renal SNA [76]. To our knowledge, however, no long-term studies have tested whether chronic blockade of the IRS2–PI3K pathway abolishes or attenuates the long-term effects of sustained hyperleptinemia to increase SNA and BP. Deletion of IRS2 in the entire CNS causes only moderate obesity and slight hyperphagia associated with normal anorexic and weight loss responses to leptin [7,13]. These observations suggest that IRS2–PI3K signaling contributes modestly to body weight regulation but may mediate, at least in part, the action of leptin on SNA.

The SHP2–MAPK pathway has been shown to participate in energy balance and metabolism as neuronal deletion of SHP2 causes obesity associated with hyperphagia and diabetes [48]. Chronic effects of hyperleptinemia to increase BP were attenuated in mice with forebrain deletion of SHP2 [26] suggesting that SHP2 signaling may also be important in mediating the effects of leptin on SNA and BP. Further studies are needed, however, to assess the role of these pathways in mediating the chronic effects of leptin on renal SNA and BP in obesity.

## Role of the CNS melanocortin system in mediating the effects of leptin on SNS activity and BP regulation

Although the precise intracellular events and brain regions by which leptin regulates body weight homeostasis and cardiovascular function are not completely understood, strong evidence shows that leptin requires activation of the brain melanocortin system, including activation of POMC neurons and MC4R, to exert most of its effects on renal SNA and BP regulation [21,81]. Activation of LR in POMC neurons is critical for leptin's ability to increase SNA and BP [24,25], while activation of MC4R using synthetic agonists increases renal SNA, BP and HR in experimental animal models as well as in humans ([21,35,46]; Sayk et al., 2010). Furthermore, mice with whole body MC4R deficiency are hyperphagic and obese, and have many characteristics of metabolic syndrome including hyperglycemia, hyperinsulinemia, visceral adiposity and dyslipidemia despite markedly elevated blood leptin levels, but are also completely unresponsive to the effects of leptin to increase renal SNA and raise BP [77,81]. In addition, mutations in POMC or MC4R genes lead to severe early-onset obesity and dysregulation of appetite in humans who, despite pronounced obesity, exhibit reduced BP, HR and



24-h urinary catecholamine excretion, lower prevalence of hypertension, and reduced SNA in response to acute stress [35,36]. Taken together, these studies strongly suggest that a functional MC4R is necessary for obesity and hyperleptinemia to increase SNA and cause hypertension. Although previous studies suggest that MC4R in the PVN and pre-ganglionic sympathetic neurons in the brainstem modulate SNA and BP [52,80], additional long-term studies are needed to examine the brain regions where MC4R regulates SNA and cardiovascular function.

## Perspectives and conclusion

In this review we highlight recent advances on leptin's role in regulating cardiorespiratory physiology. Leptin has emerged as a multifunctional peptide able to not only mediate energy balance, but also regulate cardiovascular and respiratory functions. Although the precise mechanism by which leptin exerts its effects on respiratory and cardiovascular functions are still under investigation, strong evidence suggests an important involvement of the CNS and the brain melanocortin system.

Leptin has a stimulatory effect on ventilatory response to CO<sub>2</sub> and these responses are likely mediated by leptin's action in hypothalamic and brainstem nuclei (Fig. 1). In the hypothalamus, leptin's effect on ventilation appears to be mediated by the melanocortin system. However, the role of the melanocortin system in contributing to the brainstem (e.g. NTS and rostral ventrolateral medulla) actions of leptin on ventilatory function has not been investigated.

In addition to its effects on respiratory function, leptin also plays an important role on cardiovascular regulation and is an important link between excess weight gain and increased SNA and hypertension. Although the precise mechanisms by which leptin regulates SNA and BP are still unclear and represent an area of intense investigation, strong evidence suggests a critical role of the brain melanocortin system and the activation of LR in various areas of the CNS including forebrain (e.g. hypothalamus) as well as brainstem centers (Fig. 1). Therefore, future investigations are needed to unravel the areas of the brain and signaling pathways by which the leptin–melanocortin system affects respiratory and cardiovascular functions.

## Conflict of interest statement

We have no conflict of interest to disclose.

## Q16 Uncited references

[15,20,22,61,86]

## Acknowledgements

The authors' work was supported by grants from the FAPESP (09/54888-7), CNPq, Capes, NIH (NHLBI P01 HL51971 and NIGMS-P20GM104357).

## References

- [1] A.C. Arnold, H.A. Shaltout, P.E. Gallagher, D.I. Diz, Leptin impairs cardiopulmonary baroreflex function at the level of the solitary tract nucleus, *Hypertension* 54 (2009) 1001–1008.
- [2] E. Artega-Sollis, T. Zee, C.W. Emala, C. Vinson, J. Wess, G. Karsenty, Inhibition of leptin regulation of parasympathetic signaling as a cause of extreme body weight-associated asthma, *Cell Metab.* 17 (2013) 35–48.
- [3] W.A. Banks, A.J. Kastin, W. Huang, J.B. Jassan, L.M. Maness, Leptin enters the brain by a saturable system independent of insulin, *Peptides* 17 (1996) 305–311.
- [4] M. Bassi, H. Giusti, C.M. Leite, J.A. Anselmo-Franci, J.M. do Carmo, A.A. da Silva, J.E. Hall, E. Colombari, M.L. Glass, Central leptin replacement enhances chemorespiratory responses in leptin-deficient mice independent of changes in body weight, *Pflügers Arch. Eur. J. Physiol.* 464 (2012) 145–153.
- [5] M. Bassi, W.I. Furuya, J.V. Menani, D.S. Colombari, J.M. do Carmo, A.A. da Silva, J.E. Hall, T.S. Moreira, I.C. Wenker, D.K. Mulkey, E. Colombari, Leptin in the ventrolateral medulla facilitates chemorespiratory response in leptin deficient (ob/ob) mice, *Acta Physiol. (Oxf.)* (Feb 13 2014). <http://dx.doi.org/10.1111/apha.12257>.
- [6] M. Bassi, N.B. Nakamura, W.I. Furuya, D.S.A. Colombari, J.V. Menani, J.M. do Carmo, A.A. da Silva, J.E. Hall, E. Colombari, Activation of the brain melanocortin system is required for leptin-induced modulation of chemorespiratory function, *Acta Physiol. (Oxf.)* (2015) (under review).
- [7] D. Burks, J.F. de Mora, M. Schubert, D.J. Withers, M.G. Myers, H.H. Towery, S.L. Altamuro, C.L. Flint, M.F. White, IRS-2 pathways integrate female reproduction and energy homeostasis, *Nature* 407 (2000) 377–382.
- [8] A. Campo, G. Frühbeck, J.J. Zulueta, J. Iriarte, L.M. Seijo, A.B. Alcáide, J.B. Galdiz, J. Salvador, Hyperleptinaemia, respiratory drive and hypercapnic response in obese patients, *Eur. Respir. J.* 30 (2007) 223–231.
- [9] M. Carlyle, O.B. Jones, J.J. Kuo, J.E. Hall, Chronic cardiovascular and renal actions of leptin of adrenergic activity, *Hypertension* 39 (2002) 496–501.
- [10] J.C. Celedón, L.J. Palmer, A.A. Litonjua, S.T. Weiss, B. Wang, Z. Fang, X. Xu, Body mass index and asthma in adults in families of subjects with asthma in Anqing, China, *Am. J. Respir. Crit. Care Med.* 164 (2001) 1835–1840.
- [11] Z. Chang, E. Ballou, W. Jiao, K.E. McKenna, S.F. Morrison, D.R. McCrimmon, Systemic leptin produces a long-lasting increase in respiratory motor output in rats, *Front. Physiol.* (2013). <http://dx.doi.org/10.3389/fphys.2013.00016>.
- [12] K. Chin, K. Shimizu, T. Nakamura, N. Narai, et al., Changes in intra-abdominal visceral fat and serum leptin levels in patients with obstructive sleep apnea syndrome following nasal continuous positive airway pressure therapy, *Circulation* 100 (1999) 706–712.
- [13] A.I. Choudhury, H. Heffron, M.A. Smith, H. Al-Qassab, A.W. Xu, C. Selman, M. Simmen, M. Clements, M. Claret, G. Maccoll, D.C. Bedford, K. Hisadome, I. Diakonov, V. Moosajee, J.D. Bell, J.R. Speakman, R.L. Batterham, G.S. Barsh, M.L. Ashford, D.J. Withers, The role of insulin receptor substrate 2 in hypothalamic and beta cell function, *J. Clin. Invest.* 115 (2005) 940–950.
- [14] J. Ciriello, J.M. Moreau, Leptin signaling in the nucleus of the solitary tract alters the cardiovascular responses to activation of the chemoreceptor reflex, *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 303 (2012) R727–R736.
- [15] J. Ciriello, J.M. Moreau, Systemic administration of leptin potentiates the response of neurons in the nucleus of the solitary tract to chemoreceptor activation in the rat, *Neuroscience* 229 (2013) 88–99.
- [16] J. Ciriello, Leptin in nucleus of the solitary tract alters the cardiovascular responses to aortic baroreceptor activation, *Peptides* 44 (2013) 1–7.
- [17] M.A. Cowley, J.L. Smart, M. Rubinstein, M.G. Cerdán, S. Diano, T.L. Horvath, R.D. Cone, M.J. Low, *Nature* 411 (6836) (2001) 480–484.
- [18] M.B. Daly, M.J. Scott, The effects of stimulation of the carotid body chemoreceptors on heart rate in the dog, *J. Physiol.* 144 (1985) 148–166.
- [19] M.D.B. Daly, M.J. Scott, The cardiovascular responses to stimulation of the carotid body chemoreceptors in the dog, *J. Physiol.* 165 (1963) 179–197.
- [20] A.A. da Silva, J.J. Kuo, J.E. Hall, Role of hypothalamic melanocortin3/4 receptors in mediating chronic cardiovascular, renal and metabolic actions of leptin, *Hypertension* 43 (2004) 1312–1317.
- [21] A.A. da Silva, J.M. do Carmo, Z. Wang, J.E. Hall, The brain melanocortin system, sympathetic control, and obesity hypertension, *Physiology (Bethesda)* 29 (2014) 196–202.
- [22] A.A. da Silva, J.M. do Carmo, J.N. Freeman, L.S. Tallam, J.E. Hall, A functional melanocortin system may be required for chronic CNS-mediated antidiabetic and cardiovascular actions of leptin, *Diabetes* 58 (8) (2009) 1749–1756.
- [23] J.M. do Carmo, J.E. Hall, A.A. da Silva, Chronic central leptin infusion restores cardiac sympathetic-vagal balance and baroreflex sensitivity in diabetic rats, *Am. J. Physiol. Heart Circ. Physiol.* 295 (2008) H1974–H1981.
- [24] J.M. do Carmo, A.A. da Silva, Z. Cai, S. Lin, J.H. Dubinon, J.E. Hall, Receptors in proopiomelanocortin neurons control of blood pressure, appetite, and glucose by leptin in mice lacking leptin, *Hypertension* 57 (2011) 918–926.
- [25] J.M. do Carmo, A.A. da Silva, Z. Cai, S. Lin, J.H. Dubinon, J.E. Hall, Control of blood pressure, appetite, and glucose by leptin in mice lacking leptin receptors in proopiomelanocortin neurons, *Hypertension* 57 (2011) 918–926.
- [26] J.M. do Carmo, A.A. da Silva, P.O. Sessums, S.H. Ebaady, B.R. Pace, J.S. Rushing, M.T. Davis, J.E. Hall, Role of Shp2 in forebrain neurons in regulating metabolic and cardiovascular functions and responses to leptin, *Int. J. Obes.* (2013) (epub ahead of print).
- [27] J.H. Dubinon, A.A. da Silva, J.E. Hall, Enhanced blood pressure and appetite responses to chronic central melanocortin-3/4 receptor blockade in dietary-induced obesity, *Hypertension* 28 (7) (2010) 1466–1470.
- [28] J.C. Dunbar, Y. Hu, H. Lu, Intracerebroventricular leptin increases lumbar and renal sympathetic nerve activity and blood pressure in normal rats, *Diabetes* 46 (1997) 2040–2043.
- [29] C.F. Elias, J.F. Kelly, C.E. Lee, R.S. Ahima, D.J. Drucker, C.B. Saper, J.K. Elmquist, Chemical characterization of leptin-activated neurons in the rat brain, *J. Comp. Neurol.* 423 (2000) 261–281.
- [30] G. Frühbeck, Pivotal role of nitric oxide in the control of blood pressure after leptin administration, *Diabetes* 48 (1999) 903–908.
- [31] M.L. Fung, S.Y. Lam, Y. Chen, X. Dong, P.S. Leung, Functional expression of angiotensin II receptors in type-I cells of the rat carotid body, *Pflügers Arch.* 441 (4) (2001) 474–480.
- [32] O. Gavrilova, V. Barr, B. Marcus-Samuels, M. Reitman, Hyperleptinemia of pregnancy associated with the appearance of a circulating form of the leptin receptor, *J. Biol. Chem.* 272 (1997) 30546–30551.
- [33] N. Ghilardi, R.C. Skoda, The leptin receptor activates janus kinase 2 and signals for proliferation in a factor-dependent cell line, *Mol. Endocrinol.* 11 (1997) 393–399.
- [34] N. Ghilardi, R.C. Skoda, J.H. J.M. do Carmo, A. Adi, S. Hamza, A.A. da Silva, J.E. Hall, Role of signal transducer and activator of transcription 3 in proopiomelanocortin neurons in cardiovascular and metabolic actions of leptin, *Hypertension* 61 (2013) 1066–1074.
- [35] J.R. Greenfield, Melanocortin signaling and the regulation of blood pressure in human obesity, *J. Neuroendocrinol.* 23 (2011) 186–193.

- [36] J.R. Greenfield, J.W. Miller, J.M. Keogh, E. Henning, J.H. Satterwhite, G.S. Cameron, B. Astruc, J.P. Mayer, S. Brage, T.C. See, D.J. Lomas, S. O'Rahilly, I.S. Farooqi, Modulation of blood pressure by central melanocortinergic pathways, *N. Engl. J. Med.* 360 (2009) 44–52.
- [37] H.J. Grill, M.W. Schwartz, J.M. Kaplan, J.S. Foxhall, J. Breininger, D.G. Baskin, Evidence that the caudal brainstem is a target for the inhibitory effect of leptin on food intake, *Endocrinology* 143 (2002) 239–246.
- [38] H. Groeben, S. Meier, R.H. Brown, C.P. O'Donnell, W. Mitzner, C.G. Tankersley, The effect of leptin on the ventilator response to hyperoxia, *Exp. Lung Res.* 30 (2004) 559–570.
- [39] A. Grosfeld, V. Zilberfarb, S. Turban, J. Andre, M. Guerre-Millo, T. Issad, Hypoxia increase leptin expression in human PAZ6 adipose cells, *Diabetologia* (March 27 2002) (Published online).
- [40] P.G. Guyenet, R.L. Stornetta, D.A. Bayliss, Retrotrapezoid nucleus and central chemoreception, *J. Physiol.* 586 (2008) 2043–2048.
- [41] J.E. Hall, A.A. da Silva, J.M. do Carmo, J. Dubinien, S. Hamza, S. Munusamy, G. Smith, D.E. Stec, Obesity-induced hypertension: role of sympathetic nervous system, leptin and melanocortins, *J. Biol. Chem.* 285 (23) (2010) 17271–17276.
- [42] W.G. Haynes, D.A. Morgan, S.A. Walsh, A.L. Mark, W.I. Sivitz, Receptor-mediated regional sympathetic nerve activation by leptin, *J. Clin. Invest.* 100 (1997) 270–278.
- [43] T. Hosoi, T. Kawagishi, Y. Okuma, J. Tanaka, Y. Nomura, Brain stem is a direct target for leptin's action in the central nervous system, *Endocrinology* 143 (2002) 3498–3504.
- [44] A.N. Inyushkin, E.M. Inyushkina, N.A. Merkulova, Respiratory responses to microinjections of leptin into the solitary tract nucleus, *Neurosci. Behav. Physiol.* 39 (3) (2009) 231–240.
- [45] E.M. Inyushkina, N.A. Merkulova, A.N. Inyushkin, Mechanisms of the respiratory activity of leptin at the level of the solitary tract nucleus, *Neurosci. Behav. Physiol.* 40 (7) (2010) 707–713.
- [46] P. Kievit, H. Halem, D.L. Markers, J.Z. Dong, M.M. Glavas, P. Sinnayah, L. Pranger, M.A. Cowley, K.L. Grove, M.D. Culler, Chronic treatment with a melanocortin 4 receptor agonist causes weight loss, reduces insulin resistance, and improves cardiovascular function in diet-induced obese rhesus macaques, *Diabetes* 62 (2013) 490–497.
- [47] G.G. King, N.J. Brown, C. Diba, C.W. Thorpe, P. Munoz, G.B. Marks, B. Toelle, K. Ng, N. Berend, C.M. Salome, The effects of body weight on airway calibre, *Eur. Respir. J.* 25 (2005) 896–901.
- [48] M. Krajewska, S. Banares, E.E. Zhang, X. Huang, M. Scadeng, U.S. Jhala, G.S. Feng, S. Krajewski, Development of diabetes in mice with neuronal deletion of Shp2 tyrosine phosphatase, *Am. J. Pathol.* 172 (2008) 1312–1324.
- [49] J.J. Kuo, O.B. Jones, J.E. Hall, Inhibition of NO synthesis enhances chronic cardiovascular and renal actions of leptin, *Hypertension* 37 (2001) 670–676.
- [50] G.H. Lee, R. Proenca, J.M. Montez, K.M. Carroll, J.G. Darvishzadeh, J.I. Lee, J.M. Friedman, Abnormal splicing of the leptin receptor in diabetic mice, *Nature* 379 (6566) (1996) 632–635.
- [51] R.S. Leung, T.D. Bradley, Sleep apnea and cardiovascular disease, *Am. J. Respir. Crit. Care Med.* 164 (12) (2001) 2147–2165.
- [52] P. Li, B.P. Cui, L.L. Zhang, H.J. Sun, T.Y. Liu, G.Q. Zhu, Melanocortin 3/4 receptors in paraventricular nucleus modulate sympathetic outflow and blood pressure, *Exp. Physiol.* 98 (2013) 435–443.
- [53] K. Lim, S.L. Burke, G.A. Head, Obesity-related hypertension and the role of insulin and leptin in high-fat-fed rabbits, *Hypertension* 61 (2013) 628–634.
- [54] F. Machleidt, P. Simon, A.F. Krapalis, M. Hallschmid, H. Lehnert, F. Sayk, Experimental hyperleptinemia acutely increases vasoconstrictory sympathetic nerve activity in healthy humans, *J. Clin. Endocrinol. Metab.* 98 (2013) E491–E496.
- [55] K. Makinodan, M. Yoshikawa, A. Fukuoaka, S. Tamaki, N. Koyama, M. Yamauchi, K. Tomoda, K. Hamada, H. Kimura, Effect of serum leptin levels on hypercapnic ventilatory response in obstructive sleep apnea, *Respiration* 75 (3) (2008) 257–264.
- [56] F. Malli, A.I. Papaioannou, K.I. Gourgoulanis, Z. Daniil, The role of leptin in the respiratory system: an overview, *Respir. Res.* 11 (2010) 152.
- [57] A.L. Mark, R.A. Shafer, M.L. Correia, D.A. Morgan, C.D. Sigmund, W.G. Haynes, Contrasting blood pressure effects of obesity in leptin deficient ob/ob mice and agouti yellow obese mice, *J. Hypertens.* 17 (1999) 1949–1953.
- [58] A.L. Mark, K. Agassandian, D.A. Morgan, X. Liu, M.D. Cassel, K. Rahmouni, Leptin signaling in the nucleus tractus solitarius increases sympathetic nerve activity to the kidney, *Hypertension* 53 (2009) 375–380.
- [59] A.L. Mark, Selective leptin resistance revisited, *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 305 (2013) R566–R581.
- [60] J.G. Mercer, K.M. Moar, N. Hoggard, Localization of leptin receptor (Ob-R) messenger ribonucleic acid in the rodent hindbrain, *Endocrinology* 139 (1998) 29–34.
- [61] E.G. Merrill, L. Fedorko, Monosynaptic inhibition of phrenic motoneurons: a long descending projection from Böttinger neurons, *J. Neurosci.* 4 (9) (1984) 2350–2353.
- [62] S.A. Messenger, J. Moreau, J. Ciriello, Intermittent hypoxia and systemic leptin administration induces pSTAT3 and Fos/Fra-1 in the carotid body, *Brain Res.* 1446 (2012) 56–70.
- [63] G.S. Mitchell, B.D. Selby, Effects of carotid denervation on interactions between lung inflation and PaCO<sub>2</sub> in modulating phrenic activity, *Respir. Physiol.* 67 (3) (1987) 367–378.
- [64] D.L. Morris, L. Rui, Recent advances in understanding leptin signaling and leptin resistance, *Am. J. Physiol. Endocrinol. Metab.* 297 (6) (2009) E1247–E1259.
- [65] G.J. Morton, M.W. Schwartz, Leptin and CNS control of glucose metabolism, *Physiol. Rev.* 91 (2) (2011) 389–411.
- [66] D.K. Mulkey, A.M. Mistry, P.G. Guyenet, D.A. Bayliss, Purinergic P2 receptors modulate excitability but do not mediate pH sensitivity of RTN respiratory chemoreceptors, *J. Neurosci.* 26 (2006) 7230–7233.
- [67] P. Nair, K. Radford, A. Fanat, L.J. Janssen, M. Peters-Golden, P.G. Cox, The effects of leptin on airway smooth muscle responses, *Am. J. Respir. Cell Mol. Biol.* 39 (2008) 475–481.
- [68] C.P. O'Donnell, C.G. Tankersley, V.P. Polotsky, A.R. Schwartz, P.L. Smith, Leptin, obesity, and respiratory function, *Respir. Physiol.* 119 (2000) 173–180.
- [69] M. Ozata, I.C. Ozdemir, J. Licinio, Human leptin deficiency caused by a missense mutation: multiple endocrine defects, decreased sympathetic tone, and immune system dysfunction indicate new targets for leptin action, greater central than peripheral resistance to the effects of leptin, and spontaneous correction of leptin-mediated defects, *J. Clin. Endocrinol. Metab.* 10 (1999) 3686–3695.
- [70] L. Öztürk, M. Ünal, L. Tamer, F. Çelikoglu, The association of the severity of obstructive sleep apnea with plasma leptin levels, *Arch. Otolaryngol. Head Neck Surg.* 129 (2003) 538–540.
- [71] A.J. Piper, C.E. Sullivan, Effects of short-term NIPPV in the treatment of patients with severe obstructive sleep apnea and hypercapnia, *Chest* 105 (2) (1994) 434–440.
- [72] V.Y. Polotsky, M.C. Saldone, M.T. Scharf, J. Li, C.G. Tankersley, P.L. Smith, A.R. Schwartz, C.P. O'Donnell, Impact of interrupted leptin pathways on ventilatory control, *J. Appl. Physiol.* 96 (2004) 991–998.
- [73] A. Porzionato, M. Rucinski, V. Macchi, C. Stecco, I. Castagliuolo, L.K. Malendowicz, R. De Caro, Expression of leptin and leptin receptor isoforms in the rat and human carotid body, *Brain Res.* 18 (1385) (2011) 56–67.
- [74] K. Rahmouni, D.A. Morgan, Hypothalamic arcuate nucleus mediates the sympathetic and arterial pressure responses to leptin, *Hypertension* 49 (2007) 647–652.
- [75] K. Rahmouni, D.A. Morgan, G.M. Morgan, A.L. Mark, W.G. Haynes, Role of selective leptin resistance in diet-induced obesity hypertension, *Diabetes* 54 (2005) 2012–2018.
- [76] K. Rahmouni, W.G. Haynes, D.A. Morgan, A.L. Mark, Intracellular mechanisms involved in leptin regulation of sympathetic outflow, *Hypertension* 41 (2002) 763–767.
- [77] K. Rahmouni, W.G. Haynes, D.A. Morgan, A.L. Mark, Role of melanocortin-4 receptors in mediating renal sympathoactivation to leptin and insulin, *J. Neurosci.* 23 (2003) 5998–6004.
- [78] Y.M. Rivera-Sanchez, R.A. Johnston, I.N. Schwartzman, J. Valone, E.S. Silverman, J.J. Fredberg, S.A. Shore, Differential effects of ozone on airway and tissue mechanics in obese mice, *J. Appl. Physiol.* 96 (2004) 2200–2206.
- [79] E.W. Shek, M.W. Brands, J.E. Hall, Chronic leptin infusion increases arterial pressure, *Hypertension* 31 (1998) 409–414.
- [80] J.W. Sohn, L.E. Harris, E.D. Berglund, T. Liu, L. Vong, B.B. Lowell, N. Balthasar, K.W. Williams, J.K. Elmquist, Melanocortin 4 receptors reciprocally regulate sympathetic and parasympathetic preganglionic neuron, *Cell* 152 (2013) 612–619.
- [81] L.S. Tallam, A.A. da Silva, J.E. Hall, Melanocortin-4 receptor mediates chronic cardiovascular and metabolic actions of leptin, *Hypertension* 48 (2006) 58–64.
- [82] C. Tankersley, S. Kleiberger, B. Russ, A. Schwartz, P. Smith, Modified control of breathing in genetically obese (ob/ob) mice, *J. Appl. Physiol.* 81 (1996) 716–723.
- [83] L.A. Tartaglia, The leptin receptor, *J. Biol. Chem.* 272 (1997) 6093–6096.
- [84] H. Tu, A.J. Kastin, H. Hsueh, W. Pan, Soluble receptor inhibits leptin transport, *J. Cell. Physiol.* 214 (2008) 301–305.
- [85] J.H.J. Vernooij, N.E.A. Drummen, R.J. van Suylen, R.H. Cloots, G.M. Möller, K.R. Bracke, S. Zuyderduyn, M.A. Dentener, G.G. Brusselle, P.S. Hiemstra, E.F. Wouters, Enhanced pulmonary leptin expression in patients with severe COPD and asymptomatic smokers, *Thorax* 64 (2009) 26–32.
- [86] J.H.J. Vernooij, K.R. Bracke, N.E.A. Drummen, N.S.A. Pauwels, L. Zabeau, R.J. van Suylen, J. Tavernier, G.F. Joos, E.F.M. Wouters, G.G. Brusselle, Leptin modulates innate and adaptive immune cell recruitment after cigarette smoke exposure in mice, *J. Immunol.* 184 (2010) 7169–7177.
- [87] I.C. Wenker, O. Kréneisz, A. Nishiyama, D.K. Mulkey, Astrocytes in the retrotrapezoid nucleus sense H<sub>2</sub> by inhibition of a Kir4.1–Kir5.1-like current and may contribute to chemoreception by a purinergic mechanism, *J. Neurophysiol.* 104 (2010) 3042–3052.
- [88] G. Yang, H. Ge, A. Boucher, X. Yu, C. Li, Modulation of direct leptin signaling by soluble leptin receptor, *Mol. Endocrinol.* 18 (2004) 1354–1362.
- [89] B.J. Yee, J. Cheung, P. Phipps, D. Banerjee, A.J. Piper, R.R. Grunstein, Treatment of obesity hypoventilation syndrome and serum leptin, *Respiration* 73 (2) (2006) 209–212.
- [90] C.N. Young, D.A. Morgan, S.D. Butler, A.L. Mark, R.L. Davisson, The brain subfornical organ mediates leptin-induced increases in renal sympathetic activity but not its metabolic effects, *Hypertension* 61 (2013) 737–744.
- [91] Z.-H. Zhang, R.B. Felder, Melanocortin receptors mediate the excitatory effects of blood-borne murine leptin on hypothalamic paraventricular neurons in rat, *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 286 (2004) R303–R310.