



Concept and development of an autonomous wearable micro-fluidic platform for real time pH sweat analysis

Vincenzo F. Curto, S. Coyle, R. Byrne, N. Angelov, D. Diamond, F. Benito-Lopez*

CLARITY: Centre for Sensor Web Technologies, National Centre for Sensor Research, School of Chemical Sciences, Dublin City University, Dublin 9, Ireland

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ABSTRACT

In this work the development of an autonomous, robust and wearable micro-fluidic platform capable of performing on-line analysis of pH in sweat is discussed. Through the means of an optical detection system based on a surface mount light emitting diode (SMD LED) and a light photo sensor as a detector, a wearable system was achieved in which real-time monitoring of sweat pH was performed during 55 min of cycling activity. We have shown how through systems engineering, integrating miniaturised electrical components, and by improving the micro-fluidic chip characteristics, the wearability, reliability and performance of the micro-fluidic platform was significantly improved.

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1. Introduction

Sweat is a body fluid naturally produced during physical exercise and emotional stress, and it is essentially a filtrate of blood plasma containing many substances such as sodium, chloride, potassium, bicarbonate, calcium, ammonia and organic compounds, such as glucose and lactate [1]. Sweating is primarily a mechanism of body thermoregulation to avoid dangerous rise in body temperature that correlates with the increases in metabolic rate of the individual. During this physiological process, electrolytes and liquid are excreted by sweat glands *via* a duct to the outer skin surface. [2] Through the analysis of its composition it is possible to obtain useful information regarding the physiological condition of the body and provides as well information about the health and well-being of the individual, especially during sport activities.

One of the key points that have to be taken into account when sweat analysis want to be carried out it is the way sweat is collected, especially during sport activities. Different sweat collection techniques have been employed over the pass years, including whole body wash down technique, where the whole body sweat loss is determined weighing the subject before and after exercise and all fluid lost is collected and stored [3]. Others sweat collection techniques involve the use of patches [4] and even a capsule configuration, made by a flexible adhesive membrane that is covered with an impermeable laboratory film paper, such as Parafilm

[5]. However, these techniques are not able to give real-time information about the physical condition of the body, mainly due to a substantial sampling to analysis delay. Moreover, there is high risk of cross-contamination of the samples during sampling, handling and posterior analysis. Therefore, real-time sweat analysis when performed during exercise is a great challenge for sensor fabrication due to the need for on-body fluid handling, sensor deployment and data management. If all these issues can be accomplished, the obtained devices will be capable of provide immediate feedback of fluid loss and variations of sweat analytes, giving prompt and reliable information of athlete performance and/or general health.

Sweat analysis has been employed in the diagnosis of diseases [6], drugs abuse [7] and in the optimisation of the performance of athletes [8]. Using a gold disc microelectrode De Souza et al. [9] develop a rapid and straightforward method for copper ions detection for clinical diagnosis of Wilson disease. Also, inorganic cations, amines and amino acids in human sweat were detected by capillary electrophoresis [10], while odour active compounds of human male armpit sweat, after fenugreek ingestion, were investigated by gas chromatography coupled to mass spectrometry and olfactometry [11]. Real-time measurements of sodium concentration in sweat was developed by Schazmann et al. [12] using a sodium sensor belt (SSB) based on Ion Selective Electrodes (ISE). An application of this SSB was found for the diagnosis of Cystic Fibrosis (CF) disease.

In our laboratories we are interested in pH since it is a very important, but at the same time, easy to measure physiological parameter in sweat. Many research groups including us are studying ways of analysing pH in sweat. It was investigated before that there are several factors that correlate sweat pH with health. For instance, Patterson et al. [13] found a correlation between sweat pH and sodium and chloride concentration in sweat, suggesting

* Corresponding author. Tel.: +353 1 700 7603; fax: +353 1 7007995.

E-mail addresses: vincenzo.curto2@mail.dcu.ie (V.F. Curto), shirley.coile@dcu.ie (S. Coyle), robert.byrne@intel.com (R. Byrne), dermot.diamond@dcu.ie (D. Diamond), fernando.lopez@dcu.ie (F. Benito-Lopez).

that decreasing of sodium and chloride concentrations in sweat is followed by a decrease of its pH. Furthermore, it was demonstrated that the ingestion of sodium bicarbonate led to the increase in blood and sweat pH's [14]. In addition, it was found that changes in the pH of the skin play a role in the pathogenesis of skin diseases, such as dermatitis and acne [15].

Autonomous wearable sensors to monitor sport activities should consist of reliable systems capable of monitoring physical and/or bio-chemical conditions in real time so different factors need to be considered during the design of these systems. Firstly, sampling is crucial since sample needs to be collected and delivered to the sensing active area where a signal will be generated. A failure on this stage will compromise answer and reliability of the whole sensor. Moreover, other important requirements to take into account are: (1) low cost of manufacture, (2) affordable market price, (3) flexibility of the device in order to be wearable and adaptable to the body contours (minimising the discomfort to the wearer), (4) easy signal to wearer interface and (5) long term stability.

In our laboratories, as part of BIOTEX (Biosensing textile for health management) project (<http://www.biotex-eu.com/>), the first generation of a wearable, wireless sweat analysis system was successfully fabricated and tested [16]. This sensor was integrated into a wearable platform consisting of a black masked case containing a fluid-handling system and an optical detector. A textile-based fluidic system was used to draw and deliver sweat to the sensing area where a pH sensitive dye was incorporated in one end of the polyester/lycra® blends wicking textile. The colorimetric response of the pH sensitive dye was detected using Light Emitted Diodes (LEDs) integrated and positioned over the fabric channel into the device holder.

In this paper, we present the technological achievements in performing real-time pH sweat analysis in real time coming from the basic concept of the textile-based fluid handling device towards a miniaturised wearable micro-fluidic device. We focus our investigations in two main streams: the size reduction of the electronic components that compose the detection system of the device and a smart micro-fluidic chip design and fabrication, capable of improving sample collection, minimise dead volumes and so sample cross-contamination and decrease detection times. Following both approaches we were successful in the generation of a fully autonomous and functional wearable micro-fluidic platform for real-time sweat analysis.

2. Experimental

2.1. Materials

The fabrication of the micro-fluidic devices was carried out using a laser ablation system-excimer/CO₂ laser (Optec Laser Micromachining Systems, Belgium) and a thermal roller laminator (Titan-110, GBC Films, USA). Poly(methyl-methacrylate) (PMMA) 50 µm slides were purchased from Goodfellow, UK. 80 µm double-sided pressure sensitive adhesive film (PSA-AR8890) was obtained from Adhesives Research, Ireland.

Artificial sweat was prepared according to the standard ISO 3160-2 (20 g/L NaCl, 17.5 g/L NH₄OH, 5 g/L acetic acid and 15 g/L lactic acid) (Sigma-Aldrich, St. Louis, USA). Bromocresol purple dye (BCP), hydrochloric acid, sodium hydroxide and tetraoctyl ammonium bromide were obtained from Sigma-Aldrich, St. Louis, USA. Cotton gauze was purchased from Boots Pharmaceuticals, Ireland.

Yellow surface mount LEDs (SMD LEDs) of peak wavelength 590 nm (KP 2012SYC, Kingbright) and surface mount light photo sensor modules (APDS-9004, Avago Technologies) were purchased from Farnell Ireland. Temperature sensors (Analog Devices, ADT

7301) and humidity sensors (Sensirion, SHT 11) were purchased from Radionics. Arduino microcontrollers and Xbee wireless modules were purchased from Sparkfun Electronics, Boulder, Colorado, USA.

The in-house-designed case and micro-fluidic holder of wired and wireless detectors were fabricated using a 3D printer (Stratasys, USA) in acrylonitrile butadiene styrene co-polymer (ABS) plastic in order to protect the electronics from moisture and liquid and to minimise interferences from ambient light during the operation of the device. The printed parts were designed using ProEngineer CAD/CAM software package®.

2.2. Textile-based fluid handling device

The textile-based fluid handling device used a fabric based fluidic system, where sweat entered and flowed through a channel by capillary action and collected by an absorbent patch at the end of the channel. The channel was created by coating regions of the fabric with hydrophobic materials. The channel width was 8 mm and overall length was over 30 mm. This was designed to accommodate multiple sensors. A cover was held 5 mm above the fabric layer by means of a rubber gasket. The fabric patch covered an area of 40 mm × 50 mm. pH sensitive BCP dye was applied to the fabric channel and optical sensing components were positioned in the plastic cover. A paired emitter-detector LED configuration was used to measure the colour change of the fabric. Two red LED's (λ = 660 nm) were positioned at an angle above the fabric in a reflectance mode configuration. Black silicone was placed around the sides of the detector LED to block ambient light (Fig. 3a). The LEDs were chosen because they have a narrow viewing angle (34°) to help focus the light on a smaller sensing region. A minimum sensing region of 5 mm × 5 mm was needed, based on the height and angles that the LEDs were positioned. As the fabric channel was 8 mm wide, based on the requirements of other sensors placed on the channel, the pH dye was printed onto a 8 mm × 7 mm area [17].

2.3. Micro-fluidic platform

All the micro-fluidic chips, Fig. 1a and c, were fabricated using multilayer lamination protocol. In brief, a CO₂ laser system was used to cut the various polymer layers. Micro-fluidic channels were cut from 80 µm thick layer of PSA and laminated onto PMMA 50 µm thick, using the thermal roller laminator. Fig. 1b shows the fabrication of the two generations of the micro-fluidic chips. The sensing area is a piece of nylon lycra® textile (3 mm × 3 mm) embedded in the middle of the device with a pH sensitive dye, bromocresol purple (BCP), which varies colour according to the pH of the sweat. The immobilisation of BCP is performed in an ethanol solution containing tetraoctyl ammonium bromide to avoid leaching of the dye during the contact with aqueous solutions and sweat. BCP dye colour range varies in the physiological pH range of sweat (pH 5–7).

The first generation of the micro-fluidic chip consists on a small structure of 20 by 10 mm, as shown in Fig. 1a. A round inlet of 2 mm in diameter is placed at the top of the channel that has an overall length of 8 mm. The first part of the channel is characterised by a drop shape (length of 6 mm) structure, which acts as a collector of fresh sweat coming from the skin. The collector is followed by a 2 mm length channel, while the channel following the sensing area (exhaustion channel) presents a length of 2 mm. Both channels have a width and depth equal to 400 µm and 80 µm, respectively. At the end of the channel a 5 mm diameter chamber contains the absorbent material (4 mm × 3 mm).

The design of the second generation of the micro-fluidic chip takes into consideration other parameters like long-term operability and flow control by the integration of a cotton thread inside the

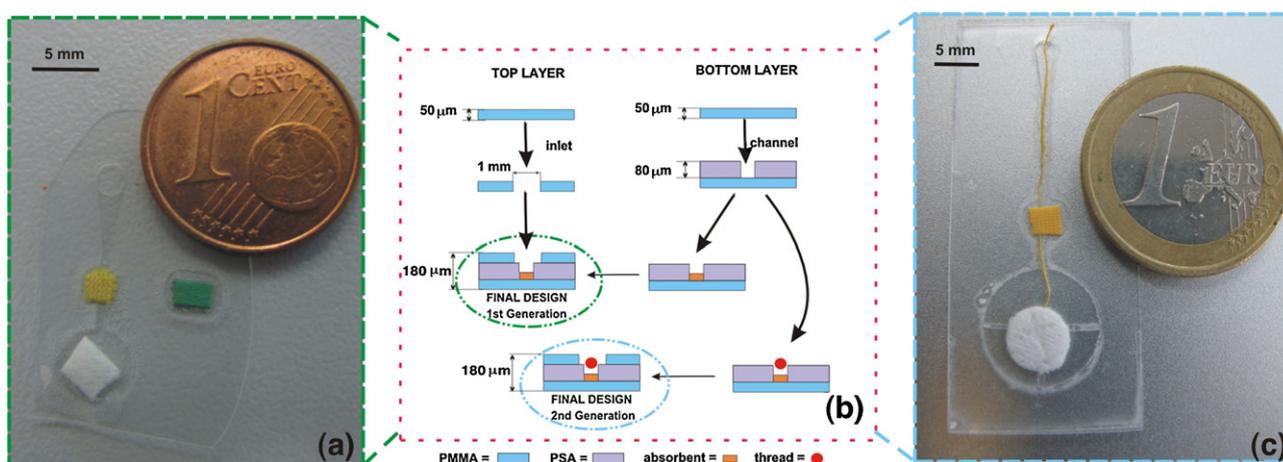


Fig. 1. First generation of the micro-fluidic chip (a). Schematic representation of the fabrication steps for first and second generation of the micro-fluidic chip (b). Second generation of the micro-fluidic chip (c).

micro-channel to ensure a homogeneous sweat flow rate through the channels (Fig. 1b and c). The overall dimensions of the micro-fluidic chip are 45 mm × 20 mm. Inlet diameter, channel width, depth and collector are the same than in the first generation, while the exhaustion channel has a length of 4 mm. The absorbent material has a diameter of 6 mm and it is placed into a round chamber of 12 mm in diameter.

2.4. Detection system

2.4.1. Textile-based fluid handling device

The LEDs were controlled by a central control unit that was supplied by CSEM. The detector LED was reverse-biased at +5 V, which charges the capacitance across it. This is discharged by the photocurrent generated upon incident light. The discharge rate is proportional to the intensity of the light reaching the detector. A digital output can be obtained by using a basic detection/timer circuit, which measures the time it takes the photocurrent to discharge the voltage from +5 V (logic 1) to +1.7 V (logic 0). Data was wirelessly transmitted to a laptop for analysis via Bluetooth.

2.4.2. Wired device

By reducing the fluidic channel from a width of 8 mm to 400 μm and reducing the sensing region from 56 mm² to 4 mm² the optical detection components also needed to be miniaturised and be aligned to focus on a smaller region. For this reason super-bright yellow SMD LEDs ($\lambda = 590$ nm) were chosen.

2.4.2.1. Wired system 1 (not shown here). The first miniaturised system used the same principle as the textile-based fluid handling device and a pair of SMD LEDs were placed above and below the sensing area of the micro-fluidic chip in a transmission configuration. In first trials black masking tape was used to position the SMD LEDs and also to block ambient light. One of the SMD LEDs acted as light source while the other was reverse biased and acted as a detector. A resistor was connected in series with the source SMD LED to attenuate the light to prevent saturation of the detector. Both SMD LEDs were controlled by a microcontroller platform, Lilypad Arduino, which is designed for wearable applications. The detector SMD LED was connected to the digital I/O pin of the Arduino and a timing routine was used to measure the discharge time from logic '1' to '0'. Data was sampled at 2 Hz and transferred to a laptop by an RS232 serial link.

2.4.2.2. Wired system 2. In this system SMD LED was used as light source, while surface mount light photo sensor (APDS-9004, Avago

Technologies) was chosen as a detector. The circuit diagram of the detector is shown in Fig. 2. The SMD LED intensity was controlled by a current limiting resistor (R_1). 30 awg wire leads were soldered to the pins of the components and the devices subsequently coated in silicone (Dow Corning, Sylgard 184) to protect the components and also to prevent short circuits between the pins. The output pin of the light photo sensor module was connected to the analogue input channel of the microcontroller. A capacitor (C) was used as a low pass filter to remove high frequency noise, and a load resistor (R_2) was used to control the current to voltage output signal of the light photo sensor.

A transmission configuration was used, where the light passes directly through the sensing material. The SMD LED and light photo sensor were placed on either side of the chip. For the on-body trials a robust system was required to position the optical components onto the micro-fluidic chip. Plastic 3D printed case was designed to hold the components in place, as shown in Fig. 3b. The data was sampled at 2 Hz using either an Arduino Lilypad or Arduino Pro and transferred to a laptop by an RS232 serial link.

2.4.3. Wireless device

In this system wired connection to the laptop was replaced with a wireless link. One option was to attach a Bluetooth[®] modem (BlueSMiRF silver) to the Arduino device. At the time of development a new Arduino Funnel IO was released, which presents an Xbee socket. This device provided a more compact solution rather than adding a separate modem to the existing configuration. There

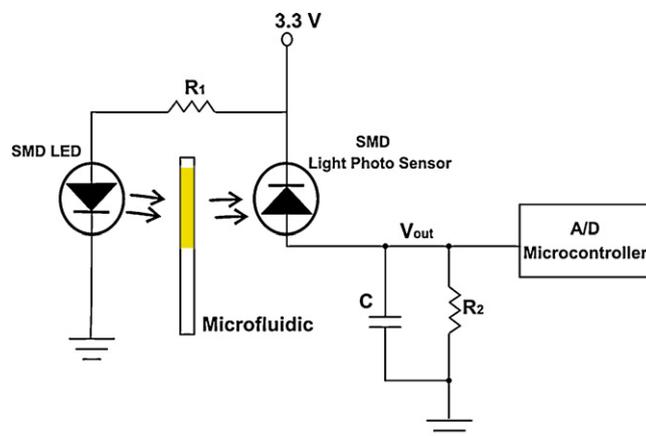


Fig. 2. Circuit diagram of the detection system in transmittance mode.

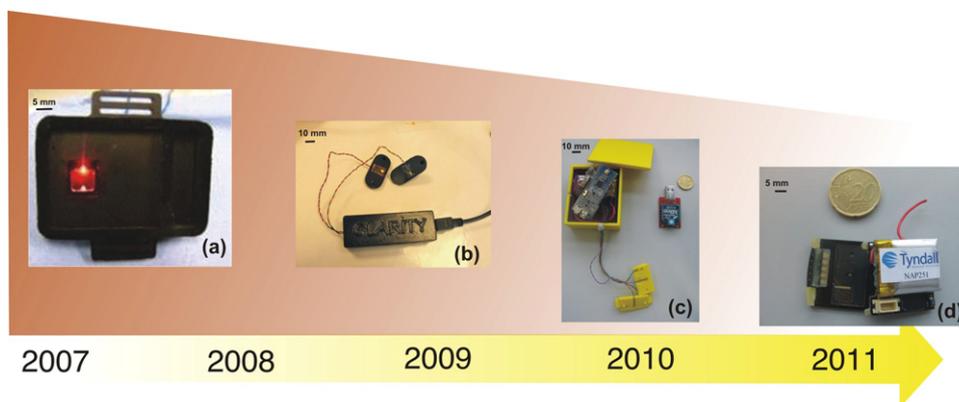


Fig. 3. Development of the electronics in the autonomous wearable micro-fluidic platform for sweat pH analysis. Textile-based fluid handling device (a), Wired detection system 2 (b), Wireless detection system (c) and miniaturised wearable micro-fluidic detector (d).

is also an option of a longer range using Xbee Pro modules and lower power consumption for shorter ranges compared to Bluetooth. However, this system requires an Xbee base-station to be connected to a laptop, while many devices would have Bluetooth capability in-built. At the end, an Xbee module was chosen.

The optical components and associated electronics were assembled in the same way as the previous wired system (Fig. 2). In addition, a temperature sensor and humidity sensor were integrated into the device to measure skin temperature and the humidity of the surrounding environment. Digital sensors were used as these could be easily configured using the Arduino FIO. A plastic holder was made using 3D printer to hold the temperature sensor (Analog Devices, ADT 7301), humidity sensor (Sensirion, SHT 11) sensors, SMD light photo sensor and LED and the pH micro-fluidic chip. A separate box was designed to hold the Arduino funnel, battery and control circuitry (Fig. 3c).

2.4.4. Portable device

The portable device was designed to improve the wearability of the system by reducing the size of the overall design and customise a wireless system based on the sweat sensor requirements. The sampling point, sensing elements and associated electronics were combined into a single device, Fig. 3d. The device once again uses a transmission mode configuration with a SMD LED and light photo sensor to detect colour changes related to pH. A temperature sensor was included in the device to measure skin temperature. The wireless capability is based on the Tyndall 25 mm mote. This device operates in the 2.4 GHz ISM Band and uses ZigBee protocols for communication. A base-station mote is connected to a laptop to receive the transmitted data and log this for analysis.

3. Results and discussion

3.1. Detection system

Colorimetric detection is performed as the sensing material changes colour according to the pH of sweat. A light source is needed to illuminate the fabric and an optical detector is needed to measure the transmitted or reflected light. In the textile-based fluid handling device, the optical sensing configuration used a reflectance measurement where the light source and the detector were positioned at an angle above the sensing layer, as shown in Fig. 4a. This meant that an adequate sensing region (56 mm^2) was needed to ensure accurate measurements, so that the detector LED would only receive light reflected from the sensing region of the fabric and not from the surrounding textile. This was important considering possible movement effects during wear. 3 mm LEDs were used and were encased in clear silicone to avoid the effects

of sweat condensation. A reverse-biased LED was used as a detector. This arrangement used the inherent capacitance of the LED and measured, the discharge time using a microcontroller. The discharge rate is proportional to the intensity of the light reaching the detector.

Despite the good performance obtained by the textile-based fluid handling device, the bulkiness of the LED-LED system makes difficult to place and adapt the sensor in different parts of the body. Therefore, we decided to miniaturise the detection system by replacing the 3 mm LEDs by a SMD LEDs. This reduced the bulkiness of the detection system considerably since the SMD LED is $1 \text{ mm} \times 1 \text{ mm}$ and flat ($500 \mu\text{m}$) ensuring a more compact and compatible detector for wearable applications.

The first detector realised using SMD LED was the wired system 1, where a reverse biased SMD LED in transmittance was used as detector. During preliminary experiments this system demonstrated good response for changes in pH from pH 4 to pH 7. However, the detection system did not have sufficient sensitivity when calibrations were performed using artificial sweat solutions between these pH ranges [18]. Therefore a SMD light photo sensor (APDS-9004, Avago Technologies) was chosen as detector, with the aim to increase its sensitivity respect the reverse biased SMD LED.

The SMD LED and light photo sensor were placed above and below the sensing area of the micro-fluidic chip in a tidy and compact structure as shown in Fig. 4b. The light emitted by the SMD LED is attenuated as it passes through the textile that presents different colour depending on the pH of the sample. This change in light intensity is sensed by the light photo sensor operating in photoconductive mode [19] where the V_{out} (Fig. 2) is proportional to the photocurrent generated by the transmitted light. A black plastic holder for the wired system 2 was made using the 3D printer to align the SMD LED and the light photo sensor. Initially, the SMD LED and the light photo sensor were integrated in a wired configuration (Fig. 3b), wherein both were controlled by an Arduino Pro platform, which uses the ATmega168 microcontroller.

Therefore, in the second generation of the device the replacement of an Arduino Pro by an Arduino Funnel IO, which uses the ATmega328P and contains an Xbee socket, enabled a wireless system to be realised through which data was wirelessly transmitted to an Xbee base-station connected to a remote laptop via a USB serial link. With this configuration it was possible to provide more freedom of movements to the wearer and increasing the spatial operation of the system.

In spite of the good performance of the detector, its packaging was performed in such a way that it was too big and uncomfortable to carry during the exercise period. A more compact structure will provide a better adaptability of the whole system to the body

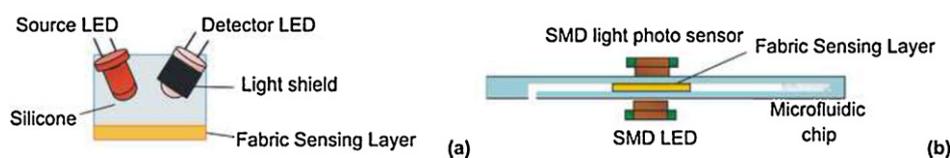


Fig. 4. Schematic representation of the alignment of LED-LED detector system in textile-based fluid handling device (a), and the alignment of SMD LED and light photo sensor in the transmittance detectors (b).

contours. That is why we decided to work in the fabrication of a miniaturised system.

Fig. 3d shows the third generation of the wearable detector device, in which all the electronic components are integrated in a much smaller cordless platform (2.5 mm × 3 cm) capable of wireless transmission of pH sweat data directly to a laptop. This device is based on the Tyndall 25 mm mote, which is a modular stackable layer solution (<http://www.tyndall.ie/mai/>) [20,21]. This design opens the possibility of performing real-time sweat analysis, minimising the discomfort of the wearer while simultaneously providing information about the wearer's physiological conditions during exercise session. Moreover, due to its small dimensions, several devices could be used in different parts of the body simultaneously, to map pH sweat composition over the whole body.

3.2. Micro-fluidic chip fabrication

The employment of a micro-fluidic chip to perform sweat analysis gives several advantages over previous systems presented in literature [3,17]. For instance, in the textile-based fluid handling device it is employed a "fluid handling system" based on fabrics with inherent moisture wicking properties. Despite of the good performance and the inherent capability of the system to collect fresh sample over a period of time, the system needs high volume of liquid (millilitre) to carry out the analysis and activate the passive pump system prior use. These factors limit the continuous operability of the sensor, in fact a decrease in the sweat rate compromises its real-time operation.

To overcome these limitations, the development of micro-fluidic systems allows a substantially reduction of the volume that is needed to monitor certain analytes in sweat, *i.e.* pH, because of the drastic reduction of channel width and length and therefore the sensing area. Moreover, since the sensing area is enclosed inside the micro-fluidic device cross-contamination from other skin areas and the surrounding environment is very unlikely.

In Section 2.3 two generations of micro-fluidic chips were described, in where a the main advantage was the reduction of the sample volume to less than 10 μl . [18] In both micro-fluidic chip configurations the sweat is drawn into the sensing area by an absorbent fibre placed at the end of the channel that acts as a passive pump system. Fig. 5 shows four pump-less micro-fluidic

devices and the relation of channel length with the time necessary for the fluid to reach the sensing area. The channel of 2 mm ensures fast sensor response reducing the delay time (t_d) between the generation of the sweat in the skin and the sensor response to less than 20 s.

Nevertheless, a better control of the flow rate in the micro-fluidic was obtained by the integration of a cotton thread inside the micro-channel. The use of cotton threads textiles into micro-fluidic systems has been explored by Li et al. [22], who demonstrated that using the intrinsic capillarity action of a thread textile it is possible to fabricate low-cost and easy-to-use devices for sensor purposes. Here the wicking property of a thread was used to transport sweat along the micro-fluidic channel, ensuring a homogeneous sweat flow rate, improving response time and platform robustness of the micro-fluidic device.

Although it was previously shown that the first generation of the micro-fluidic chip presents a good performance using a channel length of 2 mm, in the second generation the length of the channel was increased seven times. The reason for this new design was mainly to ensure better flexibility and body adaptability of the inlet of the micro-fluidic chip when in contact with the skin with respect to the sensor electronics. A defined distance, see Section 2.3, was necessary to avoid inlet detachment from the skin due to the tight position between micro-fluidic chip and electronics.

3.3. Micro-fluidic chip performance

3.3.1. Passive pump

Fig. 6 shows the autonomous pumping rate of the second generation of the micro-fluidic chip over time. The flow rate was calculated by adding a 10 μl drop of DI water on the inlet of the micro-fluidic chip and recording the time the water is completely absorbed by the passive pump (absorbent). According to Morris et al. [17] it is possible to define three operative regimes where the fluid rate is consistently different. When the system is in its dry state, the rate of fluid transports through the channel is due to the natural adsorption of the cotton thread placed into the channel. In our experiments is in the range of 1 $\mu\text{l min}^{-1}$. Considering the volume of the micro-channel from the inlet to the sensing material (approximately 0.928 μl) and assuming a linear increment of the pumping rate during the first 10 min, it has been calculated that the

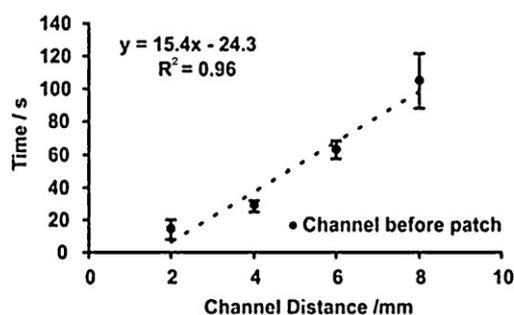


Fig. 5. Relation between channel length and the time that sweat needs to reach the sensing area using the passive pump in the chip (left) ($n=5$), picture of four micro-fluidic devices with different channel length (right).

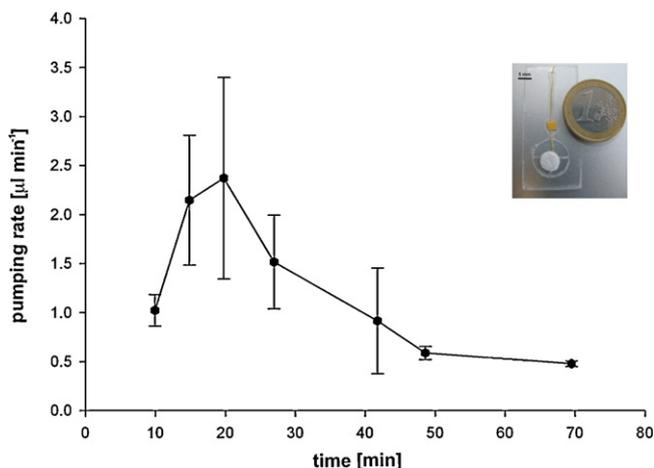


Fig. 6. Average of the pumping rate of the passive pumping system, $n = 4$.

average time for the fluid to reach the sensing area is around 3 min. The second regime is characterised by an increasing of the pumping rate when the fluid reaches the super absorbent. The flow is controlled by the absorbing capacity of the absorbent material placed at the end of the channel. The pumping rate presents a maximum value of $2.37 \mu\text{l min}^{-1}$ after 20 min, although the range of operation at this regime can be defined between minutes 15 and 40, as shown in Fig. 6. There is a quite large variation of the pumping rate over time according to the error shown in Fig. 6, e.g. $\pm 1 \mu\text{l min}^{-1}$ for the worst scenario. The error is the average of four independent micro-fluidic chips and it can be attributed to the differences in the fabrication of the device since is manual. We have observed that the position of the thread in the channel is of extreme importance for the homogeneous flow behaviour during testing.

Finally the last regime found during passive pumping starts when the absorbent gets saturated and so the flow rate slowed down considerably, stabilising a flow rate of around $0.5 \mu\text{l min}^{-1}$ till full absorbent saturation.

3.3.2. Loading capacity of the micro-fluidic chip

The loading capacity of the micro-fluidic chip, which is the maximum amount of sweat that the absorbent can uptake before the passive flow reaches zero, was calculated to be $68.7 \pm 6 \mu\text{l}$. Therefore, the calculated life-time of the device, taking into account an average of sweat flow ($1.29 \pm 0.4 \mu\text{l min}^{-1}$) was found to be around 53 min. Since in a healthy individual the sweating process starts between the 10 and 15 min from the beginning of the sport activity [17], the whole operation time of the device can be approximately considered 1 h in continuous mode. Moreover, due to the easy fabrication protocol of the device, multiple copies can be prepared in a single batch and replacement of an exhausted chip by a new fully operative device will increase the operation time.

However, when longer life-times are required, a careful redesign of the micro-fluidic can be performed to extend the device operation time. For example, changing the amount of the absorbent material placed at the end of the channel, the second operative regime can be easily extended. Other option is, as well, to vary the channel dimensions to reduce the sweat flow rate.

These results suggested that the passive pump system is able to provide fresh sweat into the channel towards the sensing area without using any mechanical components, what it makes this device an effective low-cost pumping system for wearable applications.

3.3.3. Sensor response

The dynamic response of the sensing area of the micro-fluidic chip was investigated using solutions of different pH's. A $40 \mu\text{l}$

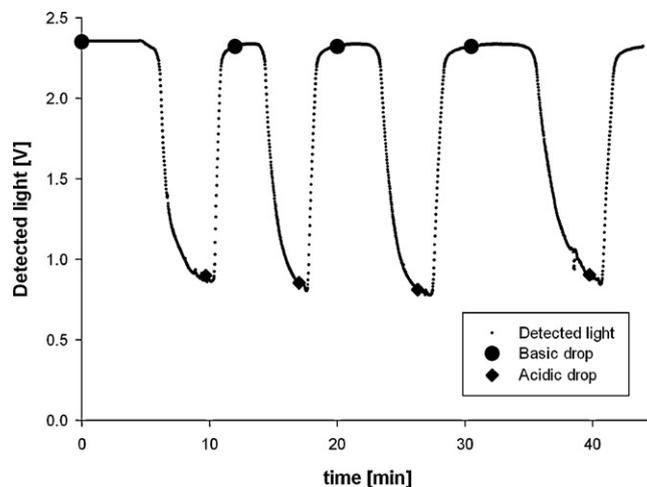


Fig. 7. Dynamic response of the micro-fluidic chip when alternatively is exposed to pH 1 and pH 12 solutions.

drop of HCl ($[\text{H}^+] = 0.1 \text{ M}$) and NaOH ($[\text{OH}^-] = 0.01 \text{ M}$) were alternatively placed in the inlet and they were let flow till the sensing area was reached, then the colour variation generated by the pH was recorded using the SMD LED-light photo sensor system.

Fig. 7 shows the response when the system is alternatively exposed to pH 1 and 12 for four times. Briefly, low voltage detection of the light photo sensor corresponds to a less detecting light due to a darker colour of the sensing material. Because BCP dye changes colour from yellow (acidic form) to blue (basic form), a value of $\sim 0.8 \text{ V}$ has to be interpreted as the high pH, e.g. 12. Meanwhile, value of $\sim 2.35 \text{ V}$ corresponds to acidic pH.

The sensing area generates reproducible signals for both acidic and basic pH conditions. Two main parameters can be obtained from Fig. 7, the “delay time” and the “response time” summarised in Table 1.

The “delay time” (t_d) corresponds to the time that sweat takes to reach the sensor area (textile), flat regions in Fig. 7.

The “response time” (t_r) corresponds to the time that sweat of certain pH fills the full sensing area and generates a stable signal in the detector, half parabolic regions in Fig. 7.

As observed in Table 1, both basic t_d and t_r in cycle 1, showed substantial higher values than in the rest of the cycles. These results are in accordance with the pumping rate data presented in Fig. 6. In fact, when the system is on its first regime, the micro-fluidic chip is dry, the flow rate is slow because the passive pump system is not still active and the thread needs to hydrate. In the subsequent cycles acidic and basic t_d are found to be lower than the first basic t_d (cycle 1) but the values are progressively increasing with the number of cycles. This expected behaviour is following the trend of Fig. 6 where the pumping flow, once it has reached its maximum rate, starts to decrease gradually till regime three where the absorbent is completely full with sweat and the passive flow stops. It is interesting to point out that since concentrations of acid and base are not equal ($\text{HCl}_{\text{sol}} 0.1 \text{ M}$ and $\text{NaOH}_{\text{sol}} 0.01 \text{ M}$) the sensor was able to get the two different t_d coming from the acid and the basic solution. The acidic solution has a proton concentration ten times

Table 1
Basic/acidic delay and response times of the micro-fluidic chip.

	Basic t_d (min)	Acidic t_d (min)	Basic t_r (min)	Acidic t_r (min)
Cycle 1	4.74	0.60	4.96	1.7
Cycle 2	1.77	0.69	3.21	2.28
Cycle 3	2.37	1.13	3.97	3.06
Cycle 4	3.70	1.29	5.30	3.17

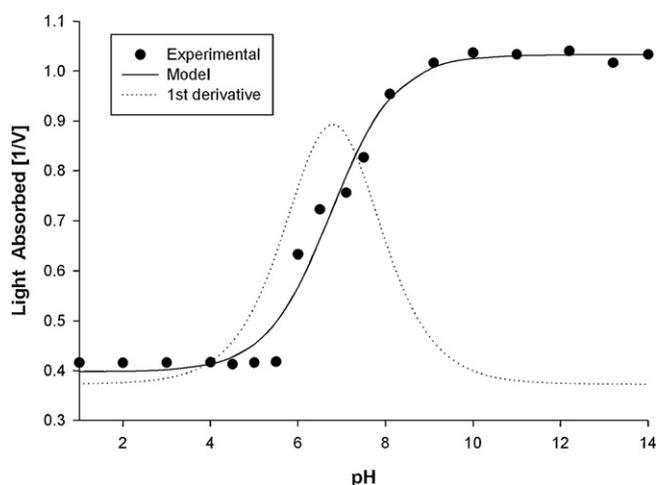


Fig. 8. Calibration curve of the fabric textile immobilised BCP dye.

bigger than the hydroxide concentration of the basic solution, so neutralisation of the excess of acid in the textile needs to be performed before the sensor area starts to change colour and so to be observed by the SMD LED-light photo sensor. In addition, experiments carried out with the same acidic and basic concentrations solutions showed that the t_d is equal for a defined passive flow rate. The t_r also presented the same trend than t_d for each consecutive cycle. It is possible to observe that during the last basic t_r the micro-fluidic chip is reaching the last regime where the passive pump flow is reduced significantly.

In short, these results confirm the reproducibility of the colorimetric detection during several cycles and the capability of the thread to draw the fluid towards the sensing area, generating a stable signal.

3.3.4. Sensor calibration

Fig. 8 shows the calibration curve of the fabric textile containing the pH sensitive dye embedded as explained in Section 2.1. The sensor material was tested using artificial sweat as standard solutions, where its pH was systematically varied from 1 to 14 adding an exact amount of 1 M NaOH solution. The textile exhibits a colour change from yellow to blue in this pH region and the change in colour was detected by the SMD LED-light photo sensor detector. The absorbed light is plotted as the inverse of the detected voltage. The experimental data are fit by a sigmoidal curve [17] following the eq.1.

$$I = \left[\frac{a}{1 + e^{b(\text{pH}-z)}} \right] \times c + d \quad (1)$$

where I is the detected light intensity, a is the peak height, b is the slope coefficient, z is the point of inflection, c the symmetry parameter for the sigmoid and d accounts for a baseline offset.

The sensor presents a pK_a of 6.8 which is slightly higher than the one from literature for the same dye in solution, pK_a 6.2 [23]. However, this effect has been previously observed by others concluding that immobilised dye in a solid support varies the pK_a value due to a change of the microenvironment where the dye is immobilised [24].

3.3.5. On-body trial

During on-body-trials the athlete was equipped with a Velcro belt located around the pelvis and the micro-fluidic device was fixed to the lumbar region of his back, as shown in the picture of Fig. 9. The trial was carried out using the wireless detector (located in a pocket) and the micro-fluidic chip containing the thread. The pH of sweat was monitored in real-time for 55 min during a cycling

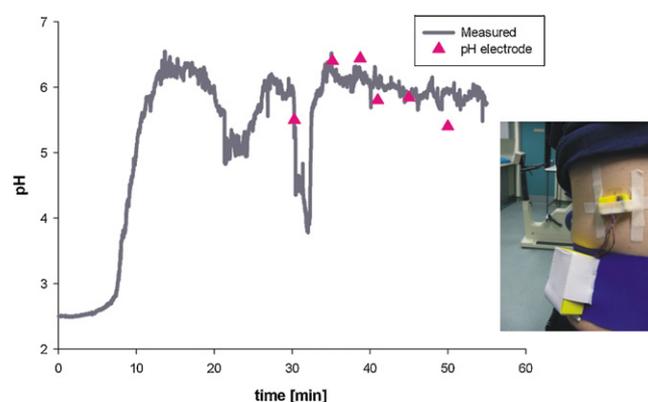


Fig. 9. pH of sweat data recorded during 55 min of cycling exercise using the wireless detector and the micro-fluidic chip containing the thread.

trial. To verify the reliability of the measurements of the micro-fluidic device, a conventional glass pH electrode was used to carry out reference measurements.

In the trial the athlete worked at different loading intensity in particular light, moderate and high. During the warm-up period (between 0 and 10 min) the athlete cycled in a moderate intensity mode for the first 5 min, then changes at high intensity regime with the aim to generate sweat as fast as possible. In this time interval it is possible to appreciate a drastic increase of the pH, from 2.5 to 6.2 as shown in Fig. 9. Considering that the chip was placed on the body without any previous precondition, the first 5 min reading was generated by a dry chip, in its acidic state. After the initial 5 warm-up minutes the athlete started to sweat, therefore sweat is driven from the skin to the reservoir by the passive pump system that worked on its first regime (Fig. 6). From minutes 5 to 8 there is an increase in pH towards a stable value of 6.2. This reading is in accordance with the estimation made in Section 3.3.1, where it was calculated that the average time for the fluid to reach the sensing area is around 3 min. Finally 6.2 pH value is reached when fresh the sweat is covering the entire sensing area and a homogeneous colour was generated giving a stable signal.

In Fig. 9 it is possible to appreciate two big drops of the pH value. During this time intervals the athlete changed alternatively the work intensity from high to moderate and it was observed a substantial increase on the sweat rate. The reason why these two big drops in pH value were generated can be attributed to physiology reasons, high sweat flow rate, variation of the pH value during the moderate exercise regime and an error occurred during the reading by the detector. At the best of our knowledge, we are not able to justify this pH drops so far; further investigations are being carried out in our laboratories to clarify this behaviour. In addition, the measurements made thorough a standard pH electrode showed a good correlation with the real-time pH data obtained using the micro-fluidic device.

The last pH meter reading value differs 0.5 pH units with the pH electrode value. This can be attributed to a decreasing of the fluid flow rate in the micro-fluidic chip when the absorbent material started to be saturated and the system was working on the last regime.

4. Conclusions

In this work we have presented significant improvements made in the realisation of a fully autonomous wearable sensor capable of performing real-time chemical analysis of sweat composition, in particular pH, during exercise events. The micro-fluidic system was developed using low-cost and flexible materials, and the passive pump system was developed through an absorbent material

and a cotton thread inside the channel, improving robustness and dynamic response. Improvements in the detection system were gained through miniaturisation of its components as such detection system, wireless platform, battery and package. All these improvements have been made in order to have a platform that was physically compatible with the needs of wearable applications, while still providing a chemical analysis capability through the monitoring of reactive indicator colour (pH in this case) using a SMD LED-light photo sensor detector.

Future work will be focused on the extension of the life-time of the micro-fluidic chip and on the use of the portable device and its possible integration in T-shirt, avoiding the use of external belt used for the placement of the whole system.

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References

- [1] A.G.R. Whitehouse, The dissolved constituents of human sweat, *Proc. R. Soc. Lond. B: Biol. Sci.* 117 (1935) 139–154.
- [2] K. Wilke, A. Martin, L. Terstegen, S.S. Biel, A short history of sweat gland biology, *Int. J. Cosmetic Sci* 29 (2007) 169–179.
- [3] S.M. Shirreffs, R.J. Maughan, Whole body sweat collection in humans: an improved method with preliminary data on electrolyte content, *J. Appl. Physiol.* 82 (1997) 336–341.
- [4] G. Hayden, H.C. Milne, M.J. Patterson, M.A. Nimmo, The reproducibility of closed-pouch sweat collection and thermoregulatory responses to exercise—heat stress, *Eur. J. Appl. Physiol.* 91 (2004) 748–751.
- [5] G.R. Brisson, P. Boisvert, F. Peronnet, H. Perrault, D. Boisvert, J.S. Lafond, A simple and disposable sweat collector, *Eur. J. Appl. Physiol. Occup. Physiol.* 63 (1991) 269–272.
- [6] L. Webster, H. Lochlin, Cystic fibrosis screening by sweat analysis: a critical review of techniques, *Med. J. Aust.* 1 (1977) 923–927.
- [7] D.A. Kidwell, J.C. Holland, S. Athanaselis, Testing for drugs of abuse in saliva and sweat, *J. Chromatogr. B* 713 (1998) 111–135.
- [8] R.J. Maughan, S.M. Shirreffs, Development of Individual Hydration Strategies for Athletes, *Int. J. Sport Nutr. Exerc. Metab.* 18 (2008) 457–472.
- [9] A.P.R. De Souza, A.S. Lima, M.O. Salles, A.N. Nascimento, M. Bertotti, The use of a gold disc microelectrode for the determination of copper in human sweat, *Talanta* 83 (2010) 167–170.
- [10] T. Hirokawa, H. Okamoto, Y. Gosyo, T. Tsuda, A.R. Timerbaev, Simultaneous monitoring of inorganic cations, amines and amino acids in human sweat by capillary electrophoresis, *Anal. Chim. Acta* 581 (2007) 83–88.
- [11] R. Mebazaa, B. Rega, V. Camel, Analysis of human male armpit sweat after fenugreek ingestion: characterisation of odour active compounds by gas chromatography coupled to mass spectrometry and olfactometry, *Food Chem.* 128 (2011) 227–235.
- [12] B. Schazmann, D. Morris, C. Slater, S. Beirne, C. Fay, R. Reuveny, N. Moyna, D. Diamond, A wearable electrochemical sensor for the real-time measurement of sweat sodium concentration, *Anal. Methods* 2 (2010) 342–348.
- [13] M.J. Patterson, S.D.R. Galloway, M.A. Nimmo, Variations in regional sweat composition in normal human males, *Exp. Physiol.* 85 (2000) 869–875.
- [14] M.J. Patterson, S.D.R. Galloway, M.A. Nimmo, Effect of induced metabolic alkalosis on sweat composition in men, *Acta Physiol. Scand.* 174 (2002) 41–46.
- [15] M.H. Schmid-Wendner, H.C. Korting, The pH of the skin surface and its impact on the barrier function, *Skin Pharmacol. Physiol.* 19 (2006) 296–302.
- [16] S. Coyle, et al., BIOTEX – biosensing textiles for personalised healthcare management, *IEEE Trans. Inf. Technol. Biomed.* 14 (2010) 364–370.
- [17] D. Morris, S. Coyle, Y. Wu, K.T. Lau, G. Wallace, D. Diamond, Bio-sensing textile based patch with integrated optical detection system for sweat monitoring, *Sens. Actuators B* 139 (2009) 231–236.
- [18] F. Benito-Lopez, S. Coyle, B. Byrne, A.F. Smeaton, N.E. O'Connor, D. Diamond, Pump less wearable microfluidic device for real time pH sweat monitoring, *Proc. Chem.* 1 (2009) 1103–1106.
- [19] J. Fraden, *Handbook of Modern Sensors – Physics, Design, and Applications*, 4th ed., Springer, San Diego, 2010.
- [20] B. O'Flynn, S. Bellis, K. Mahmood, M. Morris, G. Duffy, K. Delaney, C. O'Mathuna, A 3-D miniaturised programmable transceiver, *Microelectron. Int.* 22 (2005) 8–12.
- [21] J. Barton, G. Hynes, B. O'Flynn, K. Aherne, A. Norman, A. Morrissey, 25 mm sensor-actuator layer: a miniature, highly adaptable interface layer, *Sens. Actuators A* 132 (2006) 362–369.
- [22] X. Li, J. Tian, W. Shen, Thread as a versatile material for low-cost microfluidic diagnostics, *ACS Appl. Mater. Interface* 2 (2010) 1–6.
- [23] R.W. Sabnis, *Handbook of Acid-Base Indicators*, 1st ed., CRC Press, San Francisco, 2008.
- [24] B.R. Soller, Design of intravascular fibre optic blood gas sensors, *IEEE Eng. Med. Biol.* 13 (1994) 327–335.

Biographies

Vincenzo Fabio Curto studied chemical engineering at University of Palermo, Italy (MSc Hons 2010). In 2010 he joined the Adaptive Sensors Group at Dublin City University where she is currently pursuing a PhD degree under the supervision of Prof. Dermot Diamond and Dr. Fernando Benito-Lopez. His research interests include the development of wearable micro-fluidic system to perform real-time analysis.

Shirley Coyle is a researcher/designer in the field of wearable technologies and smart textiles. She has combined expertise in Biomedical Engineering and Fashion Design. She received her BEng in Electronic Engineering in 2000 from Dublin City University, Ireland. She then worked in the Information and Communications division in Siemens Ltd. for 2 years before commencing a Ph.D. study to develop the first optical brain computer interface. She received her PhD from the National University of Ireland Maynooth in 2005. Studying by night she graduated from the Grafton Academy of Fashion Design in 2008. She has worked on the EU FP6 'Biotex' project, a European-wide multi-partner research effort to merge sensing capabilities with fabrics and textiles. She currently works within CLARITY: Centre for Sensor Web Technologies investigating ways to improve personal health and fitness using textile technologies.

Robert Byrne studied pure and applied chemistry at Dublin City University (BSc Hons 2004) and received his PhD from Dublin City University (Materials Chemistry, 2008). His research interests include controlling molecular actuation by external stimuli for biomimetic applications.

Nikolay Angelov is pursuing his postgraduate degree in Mobile and Satellite Communications at the University of Surrey (United Kingdom). He was born in Sofia, Bulgaria where he completed his secondary education in a bilingual Spanish high-school. Nikolay graduated from Jacobs University (Bremen, Germany) with a B.Sc. in Electrical and Computer Engineering in 2010 and prior to commencing his current studies he worked as an intern at the CLARITY Centre for Sensor Web Technologies in Dublin, Ireland.

Dermot Diamond received his PhD and DSc from Queen's University Belfast (Chemical Sensors, 1987, Internet Scale Sensing, 2002), and was VP for Research at Dublin City University (2002–2004). He has published over 200 peer-reviewed papers in international journals, is a named inventor in 13 patents, and is co-author and editor of three books. He is currently director of the National Centre for Sensor Research (www.ncsr.ie) and a Principle Investigator in CLARITY (www.clarity-centre.com/), a major research initiative focused on wireless sensor networks. In 2002 he was awarded the inaugural silver medal for Sensor Research by the RSC, London.

Fernando Benito López studied chemistry at the Universidad Autonoma de Madrid and completed his master studies in the Department of Inorganic Chemistry in 2002. He obtained his PhD at the University of Twente, The Netherlands, under the supervision of Prof. David N. Reinhoudt and Dr. Willem Verboom in 2007. He carried out his postdoctoral research in the group of Prof. Dermot Diamond at Dublin City University, Dublin, Ireland. From 2010, he is Team Leader in polymer micro-fluidics at CLARITY: Centre for Sensor Web Technology, National Centre for Sensor Research, Dublin City University.