

Review

Epigenetics, hippocampal neurogenesis, and neuropsychiatric disorders: Unraveling the genome to understand the mind

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ABSTRACT

In mature, differentiated neurons in the central nervous system (CNS), epigenetic mechanisms—including DNA methylation, histone modification, and regulatory noncoding RNAs—play critical roles in encoding experience and environmental stimuli into stable, behaviorally meaningful changes in gene expression. For example, epigenetic changes in mature hippocampal neurons have been implicated in learning and memory and in a variety of neuropsychiatric disorders, including depression. With all the recent (and warranted) attention given to epigenetic modifications in mature neurons, it is easy to forget that epigenetic mechanisms were initially described for their ability to promote differentiation and drive cell fate in embryonic and early postnatal development, including neurogenesis. Given the discovery of ongoing neurogenesis in the adult brain and the intriguing links among adult hippocampal neurogenesis, hippocampal function, and neuropsychiatric disorders, it is timely to complement the ongoing discussions on the role of epigenetics in mature neurons with a review on what is currently known about the role of epigenetics in adult hippocampal neurogenesis. The process of adult hippocampal neurogenesis is complex, with neural stem cells (NSCs) giving rise to fate-restricted progenitors and eventually mature dentate gyrus granule cells. Notably, neurogenesis occurs within an increasingly well-defined “neurogenic niche”, where mature cellular elements like vasculature, astrocytes, and neurons release signals that can dynamically regulate neurogenesis. Here we review the evidence that key stages and aspects of adult neurogenesis are driven by epigenetic mechanisms. We discuss the intrinsic changes occurring within NSCs and their progeny that are critical for neurogenesis. We also discuss how extrinsic changes occurring in cellular components in the niche can result in altered neurogenesis. Finally we describe the potential relevance of epigenetics for understanding the relationship between hippocampal neurogenesis in neuropsychiatric disorders. We propose that a more thorough understanding of the molecular and genetic mechanisms that control the complex process of neurogenesis, including the proliferation and differentiation of NSCs, will lead to novel therapeutics for the treatment of neuropsychiatric disorders.

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Adult neurogenesis and the neurogenic niche

Two regions in the adult mammalian brain retain the ability to generate neurons: the subgranular zone (SGZ) of the hippocampal dentate gyrus (DG), which is the focus of this review, and the more anterior subventricular zone (SVZ). Nestled within their discrete microenvironments or “niches”, SGZ and SVZ resident neural stem cells (NSCs) undergo self-renewal to maintain a lifelong supply of mature hippocampal DG granule neurons and olfactory bulb interneurons, respectively (Fig. 1). Much is now known about the “process” of adult neurogenesis. For example, in the SGZ, Type 1 NSCs present a characteristic radial morphology and stem-like protein expression (GFAP, nestin, BLP, Sox2), and appear to give rise to non-radial Type 2 progenitors that maintain the expression of nestin and Sox2 but downregulate GFAP (Suh et al., 2007). We also know that different stages of neurogenesis are regulated by discrete environmental and physiological stimuli (Eisch et al., 2008; Ming and Song, 2005). For example, the number of Type 2 progenitors is increased by voluntary exercise, extended exposure to antidepressant drugs, and seizures, but is decreased with age and extended exposure to drugs of abuse like nicotine, opiates, and psychostimulants. As discussed below, this dynamic regulation of neurogenesis was among the first clue that adult-generated neurons might be relevant for neuropsychiatric disorders, like depression and addiction.

Despite the identification of morphology and marker expression in NSCs and their progeny and increased understanding of how they are regulated by discrete stimuli, our specific understanding of the molecular and genetic basis for how NSCs self-renew and generate neurons *in vivo* is still very limited. This is largely due to two related factors. First, there is an inherent difficulty in unambiguously tracking

adult-generated neurons *in vivo* and identifying and isolating NSCs *in vitro* (Morrison and Spradling, 2008). Recent technical advances in viral-mediated gene transfer and transgenic mouse development now allow the study and manipulation of NSCs and their progeny both *in vivo* and *in vitro*, and much of the work reviewed here relies on these new approaches (Imayoshi et al., 2009; Ming and Song, 2005).

A second factor limiting our knowledge about the genetic mechanisms that drive adult neurogenesis is the demonstrated reliance of the process of adult neurogenesis on the microenvironment, or “neurogenic niche”. Classic transplantation first demonstrated the contribution of extrinsic factors within the niche to neurogenesis (Shihabuddin et al., 2000; Suhonen et al., 1996), and many cellular components of the neurogenic niche have been identified. These include endothelial cells (Palmer et al., 2000; Shen et al., 2004, 2008; Tavazoie et al., 2008) which can release vascular-derived factors (Cao et al., 2004; Jin et al., 2002, 2003; Schanzer et al., 2004), astrocytes (Song et al., 2002a,b), which can release Wnt3, IL-1 beta and IL-6 (Barkho et al., 2006; Lie et al., 2005) and various mature neurons which via terminals and fibers of passage can release neurotransmitters, such as GABA and glutamate (Deisseroth et al., 2004; Tashiro et al., 2006; Ben-Ari, 2002; Ge et al., 2006; Liu et al., 2005; Tozuka et al., 2005). Scientists have begun to understand how neurogenesis is regulated by the factors released by these and other cellular components of the neurogenic niche. For example, GABA-mediated depolarization of Type 2 hippocampal progenitors leads to calcium influx and increased expression of the neurogenic bHLH transcription factor NeuroD1 (Tozuka et al., 2005) and the transcription factor cAMP response element-binding protein (CREB) (Jagasia et al., 2009) to promote maturation and survival of adult-born hippocampal neurons. While important progress is being made in identifying components of the

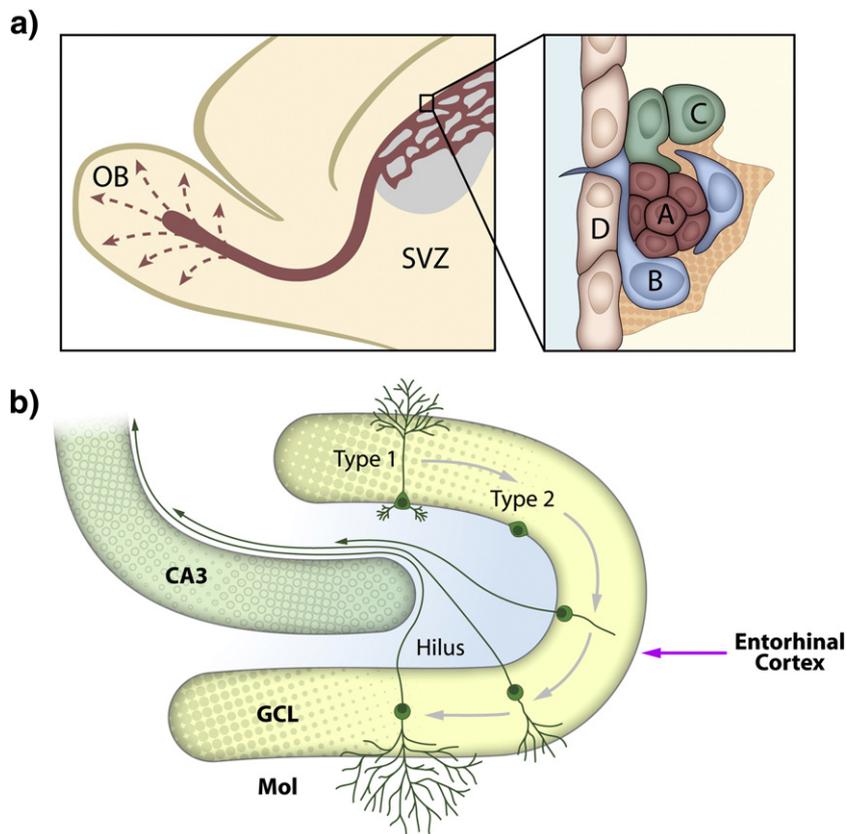


Fig. 1. Ongoing neurogenesis occurs in two discrete regions in the adult mammalian brain. (a) Progenitor cells (A–C) in the anterior subventricular zone (SVZ) lie adjacent to the ependymal cell (D) layer lining the lateral ventricles and interact with basal lamina extending from the vasculature. SVZ progenitors differentiate and migrate through the rostral migratory stream before they reach the olfactory bulb (OB) and integrate as granule neurons in the granule cell layer and as periglomerular neurons (not shown). (b) Type 1 and Type 2 progenitor cells in the subgranular zone (SGZ) proliferate and go through several stages of morphological and physiological changes as they differentiate into newborn neurons in the dentate gyrus of the hippocampus. Abbreviations are as follows: GCL, granule cell layer; Mol, molecular layer.

niches and understanding how signals in the niche regulate neurogenesis, the exact molecular mechanisms regarding how such diffusible factors signal to the NSC genome within the niche are unknown.

Here we review evidence that epigenetic modifications are in part responsible for maintenance and regulation of the process of adult hippocampal neurogenesis. This review is timely given several advances in the fields of epigenetics and adult neurogenesis. First, while overwhelming evidence supports the existence of adult-generated neurons and their functional integration (Zhao et al., 2006, 2008), more recent evidence indicates their importance in neuropsychiatric disorders (Eisch et al., 2008; Kempermann et al., 2008). In order to harness our knowledge of adult hippocampal neurogenesis for future translational applications that target these disorders, it is imperative to understand the molecular and genetic mechanisms that control the maintenance and regulation of adult neurogenesis. Second, evidence strongly supports the importance of epigenetic mechanisms in adult neurogenesis in regards to both cell-intrinsic (Lim et al., 2006) and cell-extrinsic (Ma et al., 2009) regulation. As reviewed here, the cell-intrinsic epigenetic regulation is reminiscent of classical studies on epigenetic regulation of differentiation and cell fate during embryonic and early postnatal development, while cell-extrinsic epigenetic regulation provides insight into the intriguing phenomena of “activity-dependent neurogenesis” (Deisseroth and Malenka, 2005; Deisseroth et al., 2004). Third, as indicated above, tools have been developed that allow dissection of intrinsic versus extrinsic regulators of adult-hippocampal neurogenesis (Jessberger et al., 2009b; Johnson et al., 2009). In addition, tools to study epigenetic modifications have become more standardized and thus more reliable for use; for example, high quality antibodies needed for approaches like chromatin immunoprecipitation are readily available. Therefore, we hope this review will highlight areas of research that need greater attention, and thus encourage applications of these new tools to further advance our understanding of the links among epigenetics, adult neurogenesis, and neuropsychiatric disorders.

Overview of epigenetic mechanisms

A major advance in our understanding of gene regulation was the discovery of transcription factors. Downstream of canonical intracellular signaling pathways, transcription factors bind DNA and activate or repress gene expression, opening enormous combinatorial options upon control of gene expression in regards to environmental or physiological stimuli. Epigenetic chromatin modification has emerged as an equally important discovery that, in working together with the action of transcription factors, allows fine-tuning and coordination of gene expression, thus provides even greater number of combinations and permutations to respond to stimuli. Our working definition of the term “epigenetics” refers to “changes above the genome”: heritable changes in patterns of gene expression that are not encoded in the primary DNA sequence itself, thus leading to new cellular phenotypes without a change in genotype (Riggs and Porter, 1996). As briefly described below, epigenetic mechanisms include histone modification, DNA methylation, and noncoding RNAs.

The most classically studied epigenetic modification that is also important for the present review is DNA methylation. Mammalian DNA can be covalently modified through methylation of the carbon at the fifth position on the pyrimidine ring of the cytosine residue, which is usually found at symmetrical CpG dinucleotides. DNA methylation is a major epigenetic modification for the establishment of parental-specific imprints during gametogenesis and gene silencing of the inactivated X-chromosome (Jaenisch and Bird, 2003). Cellular methyltransferases like Dnmt3a and Dnmt3b add methyl groups *de novo* to unmethylated DNA. Upon cell division, Dnmt1 preferentially recognizes hemimethylated DNA and methylates the unmethylated

strand, thus serving as a maintenance methyltransferase. Interestingly, both classes of methyltransferases have been shown to participate in various stages of neural fate and neurogenesis. During the initial specification of neurons and glia (Feng et al., 2005), as well as during later stages of neuronal maturation and function (Levenson et al., 2006), Dnmt3a and Dnmt3b are important. Dnmt1 is also very important in the brain and involved in JAK-STAT signaling to control the timing of when precursor cells switch from neurogenesis to gliogenesis during development (Fan et al., 2001, 2005; Namihira et al., 2009). With each round of cell division following DNA replication, passive demethylation of DNA occurs when maintenance methylases become inactivated. But what is known regarding active DNA demethylation? Plants use 5-methylcytosine glycosylases and the base excision repair pathway to remove excess cytosine methylation (Zhu, 2009). However, active DNA demethylation in mammals—though still controversial—is proposed to operate via several very different mechanisms from plants, but initiated by the same enzymes that are important in DNA methylation (Dnmt3a and Dnmt3b) (Gehring et al., 2009; Metivier et al., 2008; Ooi and Bestor, 2008). Clearly, more work is needed to elucidate the exact players and detailed mechanism of active DNA demethylation in mammals.

A second epigenetic modification of note is chromatin remodeling, which includes changes in histone modification. Chromatin is comprised of nucleosome repeats of 147 base pairs of DNA sequence physically wrapped around two copies of four distinct histone proteins: H2A, H2B, H3 and H4 (Luger and Richmond, 1998). One of the most exciting breakthroughs in chromatin biology this last decade is the discovery that the amino (N)-terminal tails of core histones are subject to a variety of covalent modifications or ‘marks’ such as acetylation, methylation, phosphorylation, and ubiquitylation. The DNA site- or domain-specific histone modifications (histone “code”) appear to exert powerful control over the activation or repression of the associated genes (Jenuwein and Allis, 2001). Some histone marks, like acetylation of lysine 9 and 14, di- or tri-methylation of lysine 4, and phosphorylation of serine 10 on histone H3, are signatures of actively expressed chromatin. Other marks, such as di- or trimethylation of lysine 9 or 27 on histone H3, are associated with silent chromatin domains. The histone code is established and maintained by dozens of chromatin-modifying enzymes such as histone acetyltransferases and deacetylases (HDACs) and histone methyltransferases and demethylases, which are targeted to specific chromatin loci, possibly through direct association with sequence-specific DNA binding proteins in large, multi-component complexes. Many of the components of these complexes—even the chromatin-modifying enzymes themselves—are signal responsive, resulting in a complex regulatory hierarchy for control of the genome. Notably, recent research suggests well-known neuronal transcription factors like ATF2 and CLOCK and CNS signaling molecules like nitric oxide have chromatin-modifying properties (Doi et al., 2006; Kawasaki et al., 2000; Nott et al., 2008), and likely more of such multi-purpose molecules will be identified in the future.

The complexity of this system raises the obvious question: why are there so many modifications? One hypothesis is that specific modifications link with individual biological processes and read out as simple binary “on” or “off” states (Strahl and Allis, 2000; Turner, 2000). An alternate, more complicated, but perhaps more realistic scenario is that histone modifications set up a platform for recognition and binding of many other proteins to carry out a multitude of cellular functions (Schreiber and Bernstein, 2002).

Yet another layer of complexity on top of DNA methylation, histone modification, and transcription factor expression is the presence of regulatory noncoding RNAs. Transcribed from non-protein-coding regions, several classes of noncoding RNAs are expressed in a regulated manner and serve to “fine-tune” gene expression networks. For example, microRNAs (miRNAs) are 21–23 bp noncoding RNAs that bind 3′ untranslated regions of target mRNAs resulting in translational

repression or mRNA destabilization (Kosik, 2006). Other small noncoding RNAs include small-interfering RNAs (siRNAs), small nucleolar RNAs (snoRNAs), and piwi protein-interacting RNAs (piRNAs) (Mattick and Makunin, 2005). Recently, many small noncoding RNAs have been discovered to function in development and disease, including those of the mammalian brain (Cao et al., 2006; Mehler and Mattick, 2007).

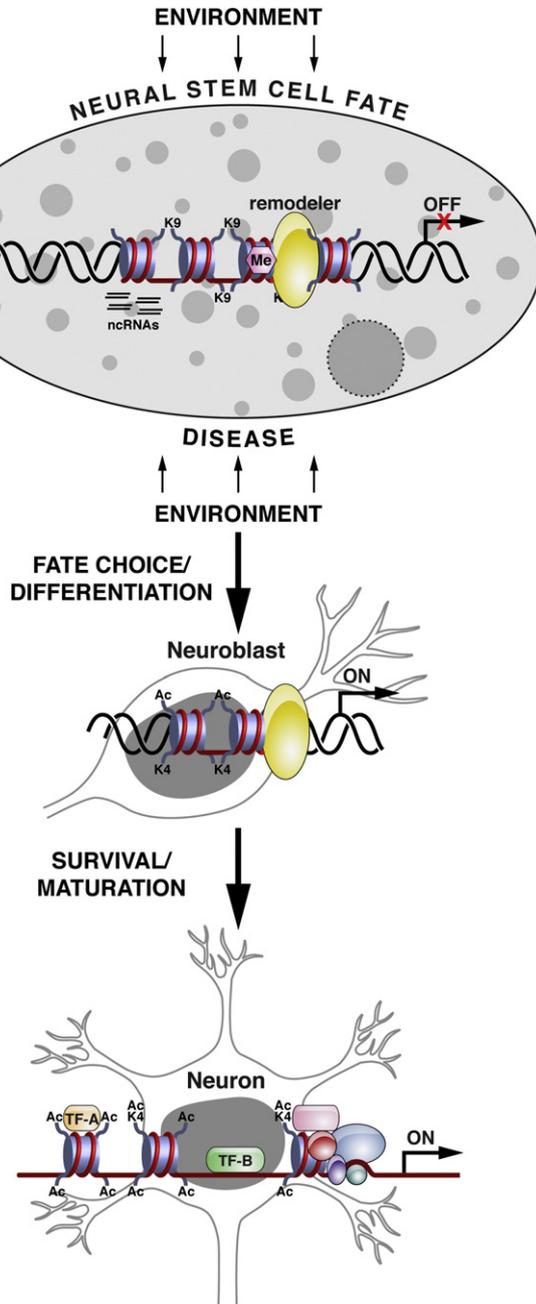


Fig. 2. Environmental factors signal to the neural stem cell genome to regulate cell fate decisions and neurogenesis. Sequence-specific transcription factors work in concert with the chromatin machinery to direct the neuronal lineage program within neural stem cells. In the stem cell state, repressive chromatin remodeling machinery maintains neuronal gene repression (OFF) through one set of histone modifications, such as histone H3 lysine K9 methylation and DNA cytosine methylation. Stimulation by environmental factors and/or stress signals (during disease) can induce adult neurogenesis and survival/maturation of newborn neurons by de-repressing or activating neuronal gene expression (ON) through the hyperacetylation and/or switch in histone modification to histone H3 lysine K4 methylation. The emergence of noncoding RNAs adds another layer of regulatory complexity to help fine-tune gene expression. Abbreviations are as follows: Me: methylation, Ac: acetylation, K: lysine, ncRNAs: noncoding RNAs, TF: transcription factor.

Below we expand on current aspects of the epigenetic regulation of adult neurogenesis. We begin with a review of classic and recent studies on epigenetic regulation of nervous system development, and then provide an overview of the progress made in understanding epigenetic mechanisms in regards to adult neurogenesis (Fig. 2). We conclude with a summary of the progress made in exploring the links among epigenetics, hippocampal neurogenesis, and neuropsychiatric disorders (Fig. 3).

Epigenetic regulation of nervous system development from mouse and *in vitro* models

The understanding of postnatal and adult neurogenesis to date has greatly benefited from studies of embryonic nervous system development. During CNS development, a diverse spectrum of neuronal and glial cell-types originates from multipotent neuroepithelial precursor cells lining the ventricles of the brain and spinal cord (Guillemot, 2007; Temple, 2001). Neuroepithelial cells differentiate into radial glia progenitors cells which further divide in a temporal fashion ('first neuron and then glia') to generate transit-amplifying progenitors to expand the population of neurons and glia that constitute the developing cortex (Okano and Temple, 2009). A number of cell-intrinsic factors have been reported to play a role in the switch from neurogenesis to gliogenesis, including proneural bHLH genes, Smad/CBP-p300 proteins, and nuclear hormone receptors (Schuurmans and Guillemot, 2002; Sun et al., 2001; Tomita et al., 2000) as well as chromatin remodeling and DNA methylation of glia-specific genes (Nakashima et al., 1999; Namihira et al., 2004; Takizawa et al., 2001). However, these studies still leave open the exact role of chromatin-based epigenetic mechanisms in early neural development.

A set of recent papers sheds light on this issue, implicating mammalian SWI/SNF-like ATP-dependent chromatin-remodeling complexes and demonstrating that a subunit composition change of the Brg- and Brahma- (Brm) associated factor-complexes (BAFs or mSWI/SNF) is critical for neuronal differentiation and dendritic development during early embryonic and postnatal development (Lessard et al., 2007; Wu et al., 2007). The transition of multipotent progenitors to postmitotic neurons involves an essential switch in ATP-dependent complexes: the BAF45a and BAF53a subunits are exchanged for the homologous BAF45b and BAF53b subunits. Knockdown of individual BAF subunits (BAF45a and/or 53a) affected the proliferation of E14.5 cortical progenitors *in vitro* and exogenous BAF45a was sufficient to drive progenitor proliferation *in vivo* (Lessard et al., 2007). Consistent with these findings, stem cell deletion of the core subunit Brg, which is associated with BAF45a and BAF53a, resulted in reduced cortical thickness and decreased proliferation of progenitors. Perhaps the most interesting observation was that the neuron-specific subunit BAF45b, a closely related subunit, could not substitute for BAF45a, suggesting a level of specificity that is necessary for the transition from proliferation of progenitors to the differentiation of neuronal subtypes.

Recently, an intriguing study came out regarding a microRNA-based mechanism that controls the essential BAF subunit composition switch. Specifically miR-9* and miR-124 appear necessary to repress BAF53a as neural progenitors differentiate into neurons (Yoo et al., 2009). Moreover, expression of neuron-restrictive silencer factor (NRSF, also known as repressor element-1 silencing transcription factor or REST; a known transcriptional repressor of miR-9* and miR-124 as well as many other neuronal genes) leads to de-repression of BAF53a in postmitotic neurons. These results highlight the intricate interplay between transcriptional regulators and regulatory noncoding RNAs, which will be discussed in a subsequent section.

In contrast to the downregulation of BAF subunits, CNS deletion of HDACs 1 and 2 during embryonic development resulted in increased proliferation of ventricular zone progenitors that similarly resulted in abrogated neuronal differentiation *in vivo* and *in vitro*

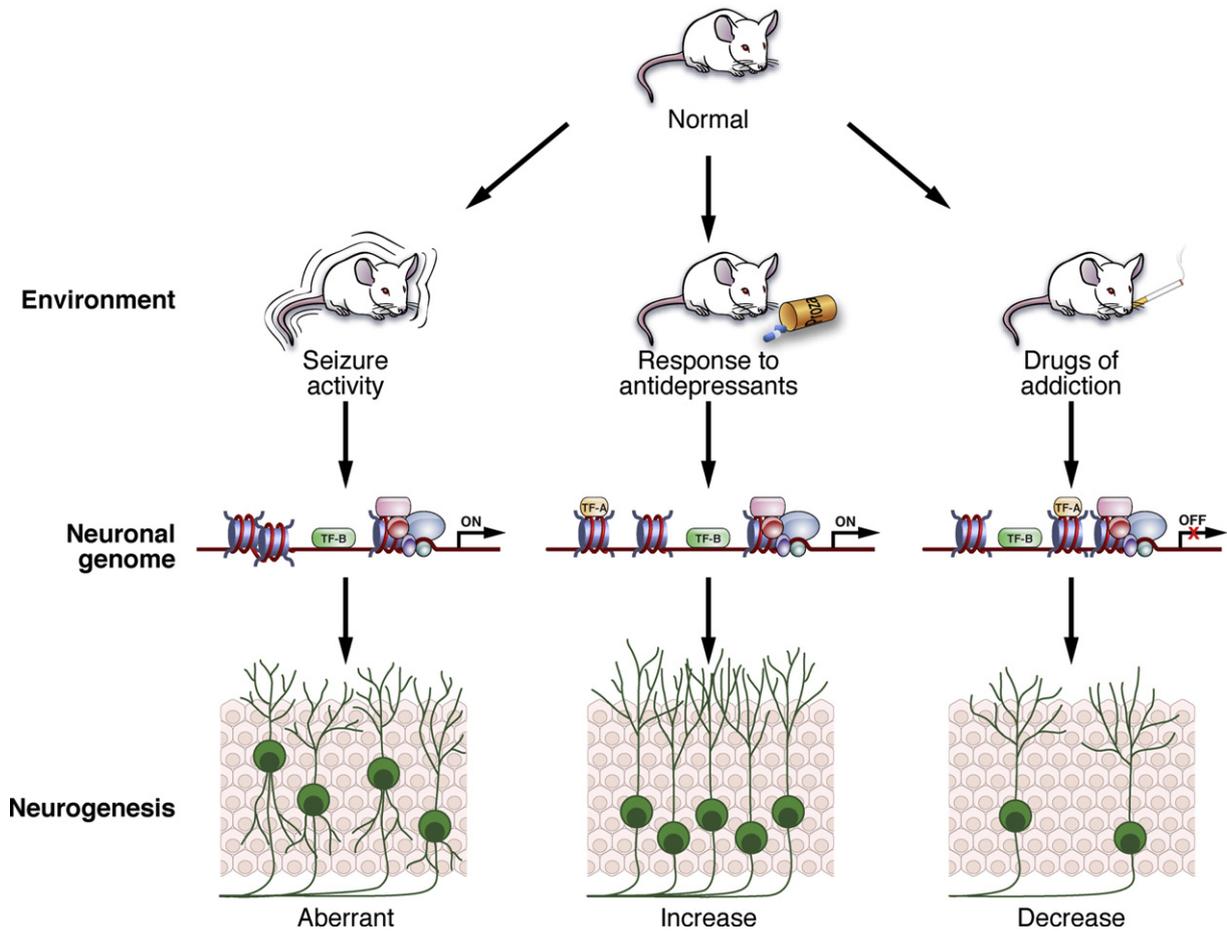


Fig. 3. Hypothetical relationship among animal models of psychiatric disorders, epigenetic regulation in SGZ stem cells, and altered adult hippocampal neurogenesis. Environmental/physiological stimuli, such as drug-induced seizure activity or chronic exposure to antidepressant agents or drugs of abuse, lead to complex neuroadaptations in discrete brain regions including altered neurogenesis. We hypothesize that these stimuli may also produce cell-intrinsic changes in chromatin remodeling that contributes to altered neurogenesis and ultimately altered neuronal genome structure. Recent advances in understanding epigenetic regulation and technical advances in determining and independently regulating chromatin modifications now make it possible to test these hypothetical relationships.

(Montgomery et al., 2009). However it remains to be determined whether BAF chromatin remodeling complexes and HDACs interact to control neuronal and glial lineage specific differentiation during CNS development.

In addition to traditional mouse models of developmental neurogenesis, studies utilizing *in vitro* stem cell systems—such as embryonic stem (ES) and induced pluripotent stem cells (iPSCs)—have emerged to address the question: to what extent do epigenetic mechanisms maintain cells in an undifferentiated or differentiated state? A major breakthrough came in 2006 when it was discovered that terminally differentiated somatic cells can be coaxed to adopt a pluripotent state via introduction of four transcription factors Oct4, Sox2, Klf4, and c-Myc (Takahashi and Yamanaka, 2006). These “reprogrammed” cells represent an immensely powerful tool for biomedical research and potentially allow every man, woman, and child to have his or her own matched stem cells for therapeutic use (Park et al., 2008; Takahashi et al., 2007). Moreover, the possibility of modeling neurological disorders like Parkinson’s disease *in vitro* where primary neuronal tissue is not available (Soldner et al., 2009; Wernig et al., 2008) make patient-specific iPSCs immediately valuable. A major obstacle to widespread use of iPSCs for neuroregenerative medicine is the knowledge gap regarding the basic mechanisms underlying direct reprogramming to pluripotency and subsequent neurogenesis from iPSCs. A slew of exciting recent papers (Hester et al., 2009; Kim et al., 2008, 2009a,b; Marchetto et al., 2009) showed that NSCs from human and mice could be

reprogrammed to pluripotency with the expression of a single transcription factor Oct4 and/or a combination of two factors in defined culture conditions.

These studies are consistent with the previous work of global chromatin modification in ES cells and support the hypothesis that NSCs represent an intermediate state between differentiated and pluripotent ES cells. Of great interest in this regard is a study by Bernstein et al. (2006). Using a combination of chromatin immunoprecipitation and tiling oligonucleotide arrays, they examined histone H3 lysine 4 and lysine 27 methylation patterns in ES cells across a subset of highly conserved noncoding elements (~2.5% of the genome), which are enriched for the 4 Hox clusters as well as transcription factor genes. Their study revealed a novel histone modification pattern termed “bivalent domains” consisting of smaller regions of lysine 4 methylation within larger regions of lysine 27 methylation within ES cells. Moreover, upon differentiation into neural lineages, bivalent domains appeared to resolve into regions selectively enriched for either lysine 27 or lysine 4 methylation. These resolved regions may provide an “epigenetic memory” for the maintenance of lineage-specific gene activation or repression. These studies also suggest that bivalent domains silence differentiation-specific genes in ES cells, but keep them poised for activation. Thus, uncovering the mechanisms that initiate and maintain bivalent domains in ES cells may shed light on the underlying mechanisms involved in reprogramming somatic cells, such as neurons, to a pluripotent state.

DNA methylation/demethylation and chromatin remodeling during adult neurogenesis

Compared to studies of embryonic and early postnatal neurogenesis, relatively little is known about cell-intrinsic epigenetic mechanisms that control adult neurogenesis. In fact, most of what is known about epigenetic modifications and adult neurogenesis comes from mouse models in which epigenetic mediators, like methyl-CpG binding protein-1 (MBD1), are constitutively deleted.

For example, one of the first studies of epigenetic regulation in adult hippocampal NSCs showed that MBD1-deficient mice had reduced neurogenesis and deficits in spatial learning and DG-specific LTP (Zhao et al., 2003), in addition to exhibiting enhanced susceptibility to depressive behaviors (Allan et al., 2008). Although there was no change in global methylation levels in hippocampal NSCs in MBD1 deficient mice, there was increased expression of the endogenous virus IAP and increased aneuploidy, supporting the important role that DNA methylation may play in maintaining genomic stability. MBDs can bind directly to methylated gene promoters and silence gene expression by blocking transcription factor binding and/or recruit HDACs to promote transcriptional repression (Klose and Bird, 2006). In a follow-up paper, MBD-1 was shown to directly regulate the methylation status of the FGF-2 promoter in adult NSCs, consistent with its intrinsic epigenetic effects (Li et al., 2008b). Future work is needed to determine whether MBD-1 actually plays a cell-intrinsic role *in vivo* or whether MBD-1 in other cells in the niche regulates neurogenesis (e.g. via cell-extrinsic mechanisms).

Another MBD, MeCP2, is highly expressed in mature neurons in the adult CNS. More recently, MeCP2 appears to be involved in suppressing the expression of glia-specific genes (like GFAP) in neurons (Kohyama et al., 2008). This finding is consistent with the importance of DNA methylation in the maintenance of neuronal identity and function which is critical for the maturation of new neurons in adult brain (Smrt et al., 2007). Interestingly, acute overexpression of MeCP2 in NSCs *in vitro* leads to enhanced neuronal differentiation (Tsujiyama et al., 2009), suggesting that MeCP2 regulates NSC function. In related work, the maintenance of hyperacetylation with HDAC inhibitors in adult hippocampal NSCs resulted in neuronal differentiation, and dominantly blocked glial differentiation (Hsieh et al., 2004). These studies lay the groundwork for additional *in vivo* experiments to determine whether MeCP2 has a cell-intrinsic vs. cell-extrinsic regulation of neurogenesis *in vivo*, what role specific HDACs play in neurogenesis *in vivo*, and what the precise relationship is between HDACs and MBDs during adult neurogenesis.

As mentioned above, electrochemical activity is emerging to be a potent trigger of adult neurogenesis. While the detailed molecular mechanisms underlying this 'activity-dependent neurogenesis' are still unknown, an exciting recent report attempted to address the question of how transient activation of mature neurons modulates adult neurogenesis. The authors focused on the functional role of the activity-induced gene *Gadd45b* (DNA-damage-inducible protein 45 beta) (Ma et al., 2009), which was previously implicated in 5-methylcytosine excision (Barreto et al., 2007; Jung et al., 2007; Tran et al., 2002). Mice deficient for *Gadd45b* display decreased proliferation after electroconvulsive treatment (ECT) or voluntary exercise. Moreover, *Gadd45b* is required for ECT-induced dendritic development of newborn neurons in DG. Interestingly, the mechanism by which *Gadd45b* regulates activity-dependent neurogenesis appears to be epigenetic in nature. While *Gadd45b* did not appear to affect global DNA demethylation after ECT, DNA from micro-dissected DG from knockout mice apparently lacked demethylation at the regulatory regions of two ECT-induced genes, FGF-1B and BDNF IX, suggesting that the role of *Gadd45b* is to promote epigenetic DNA demethylation at specific activity-induced target genes to control adult neurogenesis and dendritic development in a non-cell autonomous manner. The

precise nature of how, for example, BDNF from mature neurons signals to dividing cells in the neurogenic niche remains unclear. However, these studies importantly highlight that epigenetic mechanisms like DNA methylation and demethylation may be broadly employed for long-lasting modulation of plasticity after neuronal activity. In addition, recent studies of active DNA demethylation activity mediated by *Gadd45a* were reported to occur in mammalian cell lines as well as in *Xenopus* oocytes (Barreto et al., 2007), and in zebrafish embryos (Rai et al., 2008). However, in a further study, the functional role of *Gadd45a* in DNA demethylation was unsubstantiated (Jin et al., 2008), possibly due to species differences. This and work reviewed elsewhere (Wu and Sun, 2009) thus leave open the controversial role for *Gadd45a* as a DNA demethylase. In sum, this groundbreaking work by Ma et al. provides a tantalizing example of how cell-extrinsic epigenetic modifications (likely in mature neurons, and thus cell-extrinsic relative to NSCs) can lead to potent alterations in adult neurogenesis.

A final example of recent work on chromatin remodeling and adult neurogenesis focuses on proteins in the polycomb (PcG) and trithorax (TrxG) groups, which are required for establishing and maintaining cellular states during development and been the subject of intense study for decades (Gould, 1997). PcG and TrxG genes are organized into large multimeric complexes that regulate their target genes through a "ying-yang" fashion, most notably *Hox* (and other) promoters that control fate of individual body segments in *Drosophila*, by counterbalancing each other functionally and modulating chromatin structure. During SVZ neurogenesis, *Bmi-1*, a member of the Polycomb group of chromatin remodeling factors, is important for the self-renewal of embryonic and postnatal NSCs (Fasano et al., 2009; Molofsky et al., 2003). In the adult SVZ, Lim et al. (2006, 2009) showed that the histone methyltransferase (HMT) *Mll1* is expressed in the SVZ and olfactory bulb (OB) and is required for proliferation and neurogenesis of SVZ/OB NSCs. One downstream gene that failed to upregulate in *Mll1*-deficient SVZ neurospheres was *Dlx2*, a homeodomain-containing transcription factor. Interestingly, although *Mll1* is known to be a HMT for histone H3 lysine 4, the level of methylated lysine 4 did not change between wild type and *Mll1* KO cells. However, the level of tri-methylated lysine 27 on histone H3, usually associated with silent chromatin, was apparently higher in the absence of *Mll1*. Thus, the main function of *Mll1* in the regulation of *Dlx2* appears to be to recruit a histone H3 lysine 27 demethylase. *Mll1* is a member of the Trithorax family of proteins, which are known to antagonize Polycomb-mediated silencing (Ringrose and Paro, 2004). Future work will likely be aimed at uncovering the detailed mechanisms by which these chromatin-remodeling factors control adult neurogenesis, and dissecting whether the modifications are cell-intrinsic to NSCs and their immediate progeny or—as shown in the first part of this section—are rather cell-extrinsic and occur in postmitotic components in the neurogenic niche.

Noncoding RNAs and control of NSC fate

In addition to chromatin remodeling and neurogenesis, recent progress has been made in investigating the links between noncoding RNAs and neurogenesis. Noncoding RNAs play key roles in the modulation of transcriptional networks and appear to have important functions in CNS development and neurological disease as well (reviewed in Cao et al., 2006; Kosik and Krichevsky, 2005; Mehler, 2008; Mehler and Mattick, 2007). An interesting recent study profiled microRNA expression in developing and adult olfactory epithelium and found a dynamic expression of microRNAs, particularly miR-200 family members (Choi et al., 2008). Interestingly, inducible conditional deletion of *Dicer*, an enzyme required for the production of functional miRNAs (Bernstein et al., 2001), in olfactory progenitors during development resulted in defects in terminal differentiation of olfactory neurons as well as the maintenance of olfactory progenitor

cells. Moreover, a forebrain-specific deletion of Dicer displays smaller brains, abnormally large ventricles, increased cortical apoptosis and defects in hippocampal development (Davis et al., 2008) and conditional deletion of Dicer also results in defects in cerebellar Purkinje neurons (Schaefer et al., 2007), striatal neurons (Kim et al., 2007) and retinal degeneration (Damiani et al., 2008). Although these studies point to the global role of microRNAs in brain development, there are ~500 microRNA genes (and many mRNA targets per individual microRNA) identified in human and mice to date (Saini et al., 2007, 2008). Therefore, detailed studies of individual microRNAs are warranted to further establish the link between small noncoding RNAs and their contribution to neurological disorders. However, it is important to note that the work by Bernstein et al. is one of the few to identify a causal role for epigenetic modification intrinsic to neural progenitors in modification of neurogenesis *in vivo*.

One of the most fascinating stories regarding noncoding RNAs and neuronal differentiation is that of miR-124, the most abundant microRNA in the adult brain (Lagos-Quintana et al., 2002). During neurogenesis, miR-124 expression is undetectable or expressed at low levels in progenitor cells and is upregulated in differentiating and mature neurons (Deo et al., 2006). Initial studies demonstrated that overexpression of miR-124 in HeLa cells led to decreased expression of many non-neuronal genes to reflect a gene expression program more similar to that of neuronal cells (Lim et al., 2005). Another set of studies showed that the transcriptional repressor NRSF/REST can silence the expression of miRNA-124 and the downregulation of NRSF and increased levels of miR-124 and neuronal gene expression is permissive for neuronal differentiation in mouse embryonal carcinoma cells (Conaco et al., 2006). However, inhibiting miR-124 does not affect neuronal differentiation, at least in chick neural tube (Cao et al., 2007). A regulatory noncoding RNA/NRSF transcriptional circuit has been described previously, with the discovery of a double-stranded small RNA (smRNA) that acts to convert NRSF from a repressor to an activator in adult hippocampal NSCs during neurogenesis (Kuwabara et al., 2004).

To add to the controversial role of miR-124 in the brain, Makeyev et al. (2007) found that miR-124 targets PTBP1, a polypyrimidine tract binding protein and a repressor of neuronal splicing. Thus as neurons differentiate, miR-124 reduces PTBP1 mRNA levels, which results in a switch from general to neuron-specific alternative splicing. MiR-124 is also critical during adult neurogenesis in SVZ stem cells (Cheng et al., 2009). Blocking miR-124 maintains SVZ stem cells as dividing precursors at the transient amplifying stage and ectopic expression of miR-124 in earlier stages results in precocious neuronal differentiation *in vitro* and *in vivo*. Interestingly, miR-124 targets the SRY-box transcription factor Sox9, as well as the Dlx2 transcription factor and the Notch ligand Jag1, and functional experiments indicate that Sox9 protein levels need to be tightly regulated by miR-124 for the transition of SVZ progenitors to neurons.

In addition to microRNAs, another subset of noncoding RNAs is long, polyadenylated noncoding RNAs (lncRNAs). lncRNAs may act cooperatively and recruit protein partners to regulate gene expression (Shamovsky and Nudler, 2006). The discovery of an embryonic brain noncoding RNA called Evf2, a lncRNA target of sonic hedgehog signaling and cooperates with Dlx homeodomain proteins in NSCs (Feng et al., 2006) is consistent with noncoding RNAs having transcription-regulating activity. Recently, it was shown that Evf2 controls the balance of positive (Dlx) and negative (Mecp2) transcriptional factor recruitment to regulate Gad1 and early GABAergic interneuron development reinforcing the notion that epigenetic regulation is critical in the developing embryo which may impact mental disorders (Bond et al., 2009).

These above studies highlight the fundamental role that epigenetic mechanisms have in the regulation of adult neurogenesis under basal conditions and suggest that they may also have important effects in the context of neurological disease. In the next section, we present the

current knowledge regarding the links between hippocampal neurogenesis and the pathophysiology of neuropsychiatric disorders and examine the question of whether epigenetic mechanisms underlying hippocampal neurogenesis might contribute to neuropsychiatric diseases.

Adult hippocampal neurogenesis and neuropsychiatric diseases

Strong evidence shows that adult-generated neurons are incorporated into hippocampal circuitry (Fig. 1b) and the hippocampus itself is clearly involved in myriad neuropsychiatric disorders (Kobayashi, 2009; Sapolsky, 2000). Thus it is perhaps not surprising that adult-generated hippocampal neurons themselves have been implicated in the pathophysiology of disorders as diverse as depression, addiction, schizophrenia, epilepsy, Alzheimer's disease and even autism. As several excellent recent reviews have highlighted the links between adult neurogenesis and neuropsychiatric disorders (e.g. Danzer, 2008; Eisch et al., 2008; Kuruba et al., 2009; Parent et al., 2007; Perera et al., 2008; Scharfman and Hen, 2007), only a few critical points will be reiterated here.

First, animal models of neuropsychiatric disorders are in general marked by decreased or abnormal hippocampal neurogenesis. For example, animals chronically exposed to stress (a predisposing event in depression) or to drugs like nicotine, opiates, ethanol, or psychostimulants have fewer Type 2 hippocampal progenitors, which in general leads to decreased hippocampal neurogenesis (Fig. 3). Interestingly, while pharmacologically induced seizure activity leads to increased neurogenesis, the resulting neurons are abnormal and have dramatically aberrant migration and dendritic processes (Fig. 3). Such work with animal models suggests that similar changes would be seen in the brains of humans diagnosed with neuropsychiatric disorders. Indeed, tissue from epileptic patients reveals both increases and abnormalities in neurogenesis (Geha et al., 2009; Liu et al., 2007, 2008b) and tissue from schizophrenics reveals decreased neurogenesis (Reif et al., 2006), in accordance with what might be expected based on work in animal models of these disorders. While tissue from depressed patients does not have increased indices of neurogenesis relative to tissue from control patients (Boldrini et al., 2009; Reif et al., 2006), antidepressants appear to enhance proliferation in humans and non-human primates (Boldrini et al., 2009; Perera et al., 2007), which is consistent with laboratory animal work. Data from patients diagnosed with Alzheimer's disease are more challenging to interpret, as both animal models and human studies show both increased and decreased indices of neurogenesis (Donovan et al., 2006; He and Shen, 2009; Jin et al., 2004a,b, 2008a; Yu et al., 2009b; Zhang et al., 2007). Other disorders, like addiction, have yet to be thoroughly assessed for their impact on human neurogenesis. Clearly, more work is needed on human neurogenesis in relation to neuropsychiatric disorders. However, major challenges exist to performing human studies in a meaningful manner, such as the limited availability of proven markers appropriate for human neurogenesis and for consideration of the many stages of neurogenesis, and other general complexities that accompany human post-mortem studies (as reviewed in DeCarolis and Eisch, 2010). Thus more studies—and more technical advances, such as imaging neurogenesis in the human brain (Manganas et al., 2007)—are needed to clarify whether indeed neuropsychiatric disorders are linked to decreased or abnormal neurogenesis as is the case in most animal models of these disorders.

A second notable link is that decreased or abnormal hippocampal neurogenesis is strongly correlated to deficits in hippocampal structure and function in animal models of these disorders. This is reviewed extensively elsewhere (Abrous et al., 2005; Leuner and Gould; Ming and Song, 2005; Zhao et al., 2008), but the overview is worth mentioning. In general, stimuli or manipulations that improve performance on cognitive tasks (like exercise or the opportunity to

learn a spatial task) increase neurogenesis, while stimuli or manipulations that diminish performance on behavioral testing on cognitive tasks (like stress, age, drugs of abuse, or as is relevant for this review, HDAC inhibitors (Umka et al., 2009)) decrease neurogenesis. These correlative links suggest a functional importance for hippocampal neurogenesis in key aspects of cognition, and this is supportive by numerous publications in which neurogenesis is inducibly ablated (e.g. Clelland et al., 2009; Deng et al., 2009; Dupret et al., 2008; Garthe et al., 2009; Hernandez-Rabaza et al., 2009; Imayoshi et al., 2008; Jessberger et al., 2009a; Ko et al., 2009). Of great interest for this review, suppression of hippocampal neurogenesis in mice blocks behavioral responses in antidepressant-sensitive tests (Santarelli et al., 2003), is anxiogenic (Revest et al., 2009), and confers vulnerability in an animal model of cocaine addiction (Noonan et al., 2010). However, a role for new neurons in cognition and mental health remains controversial (e.g. DeCarolis and Eisch, 2010; Leuner and Gould; Sapolsky, 2004), as ablation of neurogenesis does not always diminish cognition, result in anxiety or lead to a depressive phenotype, and new neurons are not always needed for antidepressant efficacy (e.g. David et al., 2009; Hernandez-Rabaza et al., 2009; Holick et al., 2008; Jaholkowski et al., 2009; Ko et al., 2009; Singer et al., 2009; Surget et al., 2008). These discrepancies emphasize the need for more studies, which are further prompted by the intriguing support from human imaging studies (Manganas et al., 2007) that NSCs may be important for cognitive function. Thus more work is needed to fully clarify the relationship between the regulation of adult neurogenesis and the pathophysiology or treatment of many neuropsychiatric disorders.

A third and final link between neurogenesis and neuropsychiatric disorders worth mentioning is that treatments that ameliorate the alterations in behavior in animal models or in humans typically enhance or normalize neurogenesis. The first example of this was with antidepressants, where Duman and colleagues showed that chronic, but not acute, exposure to several different classes of antidepressants enhanced hippocampal neurogenesis (Malberg et al., 2000). In fact, all major pharmacological and non-pharmacological treatments for depression enhance proliferation and/or neurogenesis in laboratory animals, including electroconvulsive shock (ECS), monoamine oxidase inhibitors (MAOIs) like tranylcypromine, serotonin selective reuptake inhibitors (SSRIs) like fluoxetine, norepinephrine (NE) selective reuptake inhibitors like reboxetine, (all shown in Malberg et al., 2000), NMDA antagonists like memantine (Jin et al., 2006; Namba et al., 2009), tricyclic antidepressants like imipramine (Sairanen et al., 2005), transcranial magnetic stimulation (Arias-Carrion et al., 2004), and exercise (e.g. van Praag et al., 1999). Intriguingly, some antidepressants themselves do not statistically enhance neurogenesis but rather block stress-induced decreases in proliferation or neurogenesis (Czeh et al., 2001; Liu et al., 2008a). After this surge in research on antidepressants, medications for other neuropsychiatric disorders have similarly been found to normalize neurogenesis. For example, some antiepileptic drugs block seizure-induced changes in neurogenesis in animal models (e.g. Chen et al., 2009a).

Epigenetics, adult neurogenesis and neuropsychiatric diseases

As stated above, there are notable links between neuropsychiatric disorders and hippocampal neurogenesis. This leads us to the main point of this review: might epigenetics play a role in mediating hippocampal neurogenesis and perhaps contribute to neuropsychiatric disorders? Certainly epigenetic modifications in non-hippocampal brain regions are linked to neuropsychiatric disorders, including depression (Castren et al., 2007; Renthal et al., 2007; Renthal and Nestler, 2008) and schizophrenia (Sharma, 2005). In addition, a wealth of information is known now about epigenetic modifications with regards to hippocampal function and dysfunction (Alarcon et al., 2004; Crepaldi and Riccio, 2009; Sweatt, 2009; Tsankova et al., 2006). Thus it

is timely to turn the spotlight on what is known about the relationship among epigenetics, neurogenesis and neuropsychiatric disorders (also see reviews in Newton and Duman, 2006).

Prior to reviewing what is specifically known about the links among epigenetics, neurogenesis, and neuropsychiatric disorders, it is important to state that there is still great complexity—and some understandable confusion—around the progress in understanding these links and advancing the promise of utilizing epigenetic regulation to modify neurogenesis and perhaps treat or prevent neuropsychiatric disorders. To help guide future research on these important topics, here we suggest two ways of thinking about the relationship among epigenetics, adult neurogenesis, and neuropsychiatric disorders. The first is that perhaps there are cell-intrinsic epigenetic mechanisms that underlie altered neurogenesis in animal models of these disorders or even in human disease. An alternative—and complementary way—of thinking about the relationship among epigenetics, adult neurogenesis, and neuropsychiatric disorders is that perhaps epigenetic mechanisms in mature cellular components in the neurogenic niche—or cell-extrinsic mechanisms—contribute to altered signaling in the niche, which then modulates adult hippocampal neurogenesis indirectly. Distinguishing between these two possibilities is important since it clarifies the challenges ahead.

However, distinguishing between these two possibilities is also extremely challenging. As mentioned throughout this review, constitutive knockout mice are frequently the focus of epigenetic studies, and the pharmaceutical agents typically used to manipulate epigenetic mechanisms are given systemically (e.g. intraperitoneal sodium butyrate) or at best intra-hippocampally, thus influencing all cells in the hippocampus. For example, mice with constitutive mutations in or deletions of epigenetic-relevant genes, like CBP, MBD, Mecp2, HDAC1/2, or NRSF, present robust behavioral and cognitive deficits that are associated with numerous neuropsychiatric disorders (Adachi et al., 2009; Allan et al., 2008; Amir et al., 1999; Guan et al., 2009; Lepagnol-Bestel et al., 2007; Zhao et al., 2003). Intriguingly, mice deficient in these and other relevant genes, such as NRSF/REST, MeCP2, Gadd45b or MRG15 (a component of HAT and HDAC complexes) also have abnormal neurogenesis under basal or stimulated conditions (e.g. Ballas et al., 2005; Chen et al., 2009b; Ma et al., 2009; Smrt et al., 2007; Zhao et al., 2003). However, it is not clear if these results are a function of mutations in mature neurons leading to altered neurogenesis or to the mutations also causing cell-intrinsic changes in neurogenesis. As another example, exposure to antidepressants or drugs of abuse alter epigenetic-relevant molecules in the hippocampus (e.g. Cassel et al., 2006; Tsankova et al., 2004, 2006) similar to actions in other brain regions (e.g. Renthal et al., 2007; Renthal and Nestler, 2008), perhaps driving the changes in neurogenesis seen after antidepressants and cocaine (Malberg et al., 2000; Yamaguchi et al., 2004). However, it is unclear whether epigenetic changes occur within NSCs and their progeny to mediate changes in neurogenesis, or whether the changes occur within nonneurogenic components of the niche. Essentially, these published studies are elegant and excellent for what they reveal about epigenetic mechanisms in hippocampal neurons and thus the relationship between the hippocampus and neuropsychiatric disorders. However, few mechanistic conclusions can be drawn from these about how cell-intrinsic or cell-extrinsic epigenetic modifications regulate neurogenesis and adult-generated neuron involvement in neuropsychiatric disorders.

There are a few lines of research, however, that have utilized a clever combination of *in vivo* cell-specific targeting and *in vitro* approaches, and whose results support our hypothesis that epigenetic changes in mature neurons or cell-intrinsic epigenetic changes in NSCs and their progeny contribute to altered neurogenesis and are linked to neuropsychiatric disorders. For example, valproic acid (VPA) is a clinically effective mood stabilizer that drives hippocampal neurogenesis (Manji et al., 2000) partially via its positive influence on

ERK signaling (Hao et al., 2004). However, VPA also is an HDAC inhibitor, and it drives neuronal differentiation *in vitro* in part through upregulation of the proneural transcription factor NeuroD1 (Hsieh et al., 2004). Intriguingly, VPA's HDAC inhibition blocks kainic acid-induced seizures and prevents the resulting abnormal neurogenesis and cognitive deficits (Jessberger et al., 2007), strongly suggesting a role for HDACs in seizure-induced behavioral problems. More recent work has emphasized a role for VPA in induction of proneural factors (Yu et al., 2009a) and the importance of these factors, like NeuroD1, in driving neurogenesis (Gao et al., 2009). As VPA also can decrease memory function and neurogenesis *in vivo* (Umka et al., 2009), the question of whether and how HDAC inhibition alters neurogenesis will await the utilization of modern transgenic mice and viral-mediated gene transfer to answer these more mechanistic questions (Johnson et al., 2009).

Another example of a line of research that support our hypothesis that epigenetic changes in mature neurons or cell-intrinsic epigenetic changes in NSCs and their progeny contribute to altered neurogenesis and are linked to neuropsychiatric disorders involves the gene *Disc1* (disrupted-in-schizophrenia-1 gene). *Disc1* is notable since the clinical link was discovered prior to its epigenetic involvement. Initially linked to schizophrenia (Millar et al., 2000), *Disc1* was subsequently found to be important in neurite outgrowth and other fundamental neuronal functions (Miyoshi et al., 2003; Morris et al., 2003). Seminal work showing the importance of *Disc1* in adult neurogenesis (Duan et al., 2007) was followed by recent work that *Disc1* interacts with the GSK3 β / β -catenin pathway to regulate adult neurogenesis (Mao et al., 2009). This is exceptionally intriguing since the GSK3 β / β -catenin pathway is considered a common target for many neuropsychiatric disorders (Wada, 2009), including its involvement in the regulation of histone modifications that are hallmarks of epigenetic mechanisms (Mosimann et al., 2009).

Several other papers that were discussed above (Allan et al., 2008; Ma et al., 2009; Zhao et al., 2003) are other excellent examples of how altered epigenetic signaling in mature neurons might be linked to adult neurogenesis and altered behavior relevant to neuropsychiatric disorders. Taken together, these studies point to a set of hypothetical epigenetic changes that may occur within NSCs in the adult SGZ after seizure activity, antidepressant administration, or exposure to drugs of abuse (Fig. 3), and that are the focus of current studies within many laboratories. Clearly, more work is needed utilizing cell-specific transgenic and viral-mediated protein expression in order to fully understand the relationship among epigenetics, neurogenesis, and neuropsychiatric disorders.

Conclusions and future directions

In this review, we hope to convey that many types of epigenetic mechanisms are interrelated—DNA methylation and demethylation, histone modifications, noncoding RNAs—to regulate gene expression in adult neural stem/progenitor cells which crossover to diverse neuropsychiatric conditions. While these epigenetic approaches to explore the links between adult neurogenesis and neurological disease remain vastly promising, there are still unanswered questions that need to be addressed in future research towards the development of potential therapeutics. Do the recently identified epigenetic mechanisms that function to regulate embryonic neurogenesis also function in adult neurogenesis? What other CNS signaling molecules have chromatin-modifying properties? As human studies with *in vivo* imaging of NSCs become more commonplace, and as our ability to study neurogenesis in human tissues improve, it will be important to aggressively test whether, as in animal models of these disorders, strong links exist between hippocampal function and neurogenesis. Importantly for this review, it will be critical to assess how epigenetic mechanisms in human tissue contribute to regulation of neurogenesis and whether, as we hypothesize, chromatin remodeling is in fact a

promising target for future treatment avenues. Finally, it will further be critical to establish whether treatments that target the epigenetic state of mature hippocampal neurons have untoward or additional unappreciated effect on the adult-generated neurons that reside nearby.

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References

- Abrous, D.N., et al., 2005. Adult neurogenesis: from precursors to network and physiology. *Physiol. Rev.* 85, 523–569.
- Adachi, M., et al., 2009. MeCP2-mediated transcription repression in the basolateral amygdala may underlie heightened anxiety in a mouse model of Rett syndrome. *J. Neurosci.* 29, 4218–4227.
- Alarcon, J.M., et al., 2004. Chromatin acetylation, memory, and LTP are impaired in CBP+/- mice: a model for the cognitive deficit in Rubinstein-Taybi syndrome and its amelioration. *Neuron* 42, 947–959.
- Allan, A.M., et al., 2008. The loss of methyl-CpG binding protein 1 leads to autism-like behavioral deficits. *Hum. Mol. Genet.* 17, 2047–2057.
- Amir, R.E., et al., 1999. Rett syndrome is caused by mutations in X-linked MECP2, encoding methyl-CpG-binding protein 2. *Nat. Genet.* 23, 185–188.
- Arias-Carrion, O., et al., 2004. Neurogenesis in the subventricular zone following transcranial magnetic field stimulation and nigrostriatal lesions. *J. Neurosci. Res.* 78, 16–28.
- Ballas, N., et al., 2005. REST and its corepressors mediate plasticity of neuronal gene chromatin throughout neurogenesis. *Cell* 121, 645–657.
- Barkho, B.Z., et al., 2006. Identification of astrocyte-expressed factors that modulate neural stem/progenitor cell differentiation. *Stem Cells Dev.* 15, 407–421.
- Barreto, G., et al., 2007. Gadd45a promotes epigenetic gene activation by repair-mediated DNA demethylation. *Nature* 445, 671–675.
- Ben-Ari, Y., 2002. Excitatory actions of GABA during development: the nature of the nurture. *Nat. Rev. Neurosci.* 3, 728–739.
- Bernstein, B.E., et al., 2006. A bivalent chromatin structure marks key developmental genes in embryonic stem cells. *Cell* 125, 315–326.
- Bernstein, E., et al., 2001. Role for a bidentate ribonuclease in the initiation step of RNA interference. *Nature* 409, 363–366.
- Boldrini, M., et al., 2009. Antidepressants increase neural progenitor cells in the human hippocampus. *Neuropsychopharmacology* 34, 2376–2389.
- Bond, A.M., et al., 2009. Balanced gene regulation by an embryonic brain ncRNA is critical for adult hippocampal GABA circuitry. *Nat. Neurosci.* 12, 1020–1027.
- Cao, L., et al., 2004. VEGF links hippocampal activity with neurogenesis, learning and memory. *Nat. Genet.* 36, 827–835.
- Cao, X., et al., 2006. Noncoding RNAs in the mammalian central nervous system. *Annu. Rev. Neurosci.* 29, 77–103.
- Cao, X., et al., 2007. A functional study of miR-124 in the developing neural tube. *Genes Dev.* 21, 531–536.
- Cassel, S., et al., 2006. Fluoxetine and cocaine induce the epigenetic factors MeCP2 and MBD1 in adult rat brain. *Mol. Pharmacol.* 70, 487–492.
- Castren, E., et al., 2007. Role of neurotrophic factors in depression. *Curr. Opin. Pharmacol.* 7, 18–21.
- Chen, J., et al., 2009a. Effects of lamotrigine and topiramate on hippocampal neurogenesis in experimental temporal-lobe epilepsy. *Brain Res.* 1313, 270–282.
- Chen, M., et al., 2009b. MRG15, a component of HAT and HDAC complexes, is essential for proliferation and differentiation of neural precursor cells. *J. Neurosci. Res.* 87, 1522–1531.
- Cheng, L.C., et al., 2009. miR-124 regulates adult neurogenesis in the subventricular zone stem cell niche. *Nat. Neurosci.* 12, 399–408.
- Choi, P.S., et al., 2008. Members of the miRNA-200 family regulate olfactory neurogenesis. *Neuron* 57, 41–55.
- Clelland, C.D., et al., 2009. A functional role for adult hippocampal neurogenesis in spatial pattern separation. *Science* 325, 210–213.
- Conaco, C., et al., 2006. Reciprocal actions of REST and a microRNA promote neuronal identity. *Proc. Natl. Acad. Sci. U. S. A.* 103, 2422–2427.
- Crepaldi, L., Riccio, A., 2009. Chromatin learns to behave. *Epigenetics* 4, 23–26.
- Czeh, B., et al., 2001. Stress-induced changes in cerebral metabolites, hippocampal volume, and cell proliferation are prevented by antidepressant treatment with tianeptine. *Proc. Natl. Acad. Sci. U. S. A.* 98, 12796–12801.
- Damiani, D., et al., 2008. Dicer inactivation leads to progressive functional and structural degeneration of the mouse retina. *J. Neurosci.* 28, 4878–4887.
- Danzer, S.C., 2008. Postnatal and adult neurogenesis in the development of human disease. *Neuroscientist* 14, 446–458.

- David, D.J., et al., 2009. Neurogenesis-dependent and -independent effects of fluoxetine in an animal model of anxiety/depression. *Neuron* 62, 479–493.
- Davis, T.H., et al., 2008. Conditional loss of Dicer disrupts cellular and tissue morphogenesis in the cortex and hippocampus. *J. Neurosci.* 28, 4322–4330.
- DeCarolis, N. A., Eisch, A. J., 2010. Hippocampal neurogenesis as a target for the treatment of mental illness: a critical evaluation. *Neuropharmacology* 58, 884–893.
- Deisseroth, K., Malenka, R.C., 2005. GABA excitation in the adult brain: a mechanism for excitation–neurogenesis coupling. *Neuron* 47, 775–777.
- Deisseroth, K., et al., 2004. Excitation–neurogenesis coupling in adult neural stem/progenitor cells. *Neuron* 42, 535–552.
- Deng, W., et al., 2009. Adult-born hippocampal dentate granule cells undergoing maturation modulate learning and memory in the brain. *J. Neurosci.* 29, 13532–13542.
- Deo, M., et al., 2006. Detection of mammalian microRNA expression by in situ hybridization with RNA oligonucleotides. *Dev. Dyn.* 235, 2538–2548.
- Doi, M., et al., 2006. Circadian regulator CLOCK is a histone acetyltransferase. *Cell* 125, 497–508.
- Donovan, M.H., et al., 2006. Decreased adult hippocampal neurogenesis in the PDAPP mouse model of Alzheimer's disease. *J. Comp. Neurol.* 495, 70–83.
- Duan, X., et al., 2007. Disrupted-in-schizophrenia 1 regulates integration of newly generated neurons in the adult brain. *Cell* 130, 1146–1158.
- Dupret, D., et al., 2008. Spatial relational memory requires hippocampal adult neurogenesis. *PLoS ONE* e1959, 3.
- Eisch, A.J., et al., 2008. Adult neurogenesis, mental health, and mental illness: hope or hype? *J. Neurosci.* 28, 11785–11791.
- Fan, G., et al., 2001. DNA hypomethylation perturbs the function and survival of CNS neurons in postnatal animals. *J. Neurosci.* 21, 788–797.
- Fan, G., et al., 2005. DNA methylation controls the timing of astroglialogenesis through regulation of JAK-STAT signaling. *Development* 132, 3345–3356.
- Fasano, C.A., et al., 2009. Bmi-1 cooperates with Foxg1 to maintain neural stem cell self-renewal in the forebrain. *Genes Dev.* 23, 561–574.
- Feng, J., et al., 2006. The Evf-2 noncoding RNA is transcribed from the Dlx-5/6 ultraconserved region and functions as a Dlx-2 transcriptional coactivator. *Genes Dev.* 20, 1470–1484.
- Feng, J., et al., 2005. Dynamic expression of de novo DNA methyltransferases Dnmt3a and Dnmt3b in the central nervous system. *J. Neurosci. Res.* 79, 734–746.
- Gao, Z., et al., 2009. Neurod1 is essential for the survival and maturation of adult-born neurons. *Nat. Neurosci.* 12, 1090–1092.
- Garthe, A., et al., 2009. Adult-generated hippocampal neurons allow the flexible use of spatially precise learning strategies. *PLoS ONE* e5464, 4.
- Ge, S., et al., 2006. GABA regulates synaptic integration of newly generated neurons in the adult brain. *Nature* 439, 589–593.
- Geha, S., et al., 2009. NG2+/Olig2+ cells are the major cycle-related cell population of the adult human normal brain. *Brain Pathol.* 20, 399–411.
- Gehring, M., et al., 2009. DNA demethylation by DNA repair. *Trends Genet.* 25, 82–90.
- Gould, A., 1997. Functions of mammalian Polycomb group and trithorax group related genes. *Curr. Opin. Genet. Dev.* 7, 488–494.
- Guan, J.S., et al., 2009. HDAC2 negatively regulates memory formation and synaptic plasticity. *Nature* 459, 55–60.
- Guillemot, F., 2007. Cell fate specification in the mammalian telencephalon. *Prog. Neurobiol.* 83, 37–52.
- Hao, Y., et al., 2004. Mood stabilizer valproate promotes ERK pathway-dependent cortical neuronal growth and neurogenesis. *J. Neurosci.* 24, 6590–6599.
- He, P., Shen, Y., 2009. Interruption of beta-catenin signaling reduces neurogenesis in Alzheimer's disease. *J. Neurosci.* 29, 6545–6557.
- Hernandez-Rabaza, V., et al., 2009. Inhibition of adult hippocampal neurogenesis disrupts contextual learning but spares spatial working memory, long-term conditional rule retention and spatial reversal. *Neuroscience* 159, 59–68.
- Hester, M.E., et al., 2009. Two factor reprogramming of human neural stem cells into pluripotency. *PLoS One* e7044, 4.
- Holick, K.A., et al., 2008. Behavioral effects of chronic fluoxetine in BALB/cj mice do not require adult hippocampal neurogenesis or the serotonin 1A receptor. *Neuropsychopharmacology* 33, 406–417.
- Hsieh, J., et al., 2004. Histone deacetylase inhibition-mediated neuronal differentiation of multipotent adult neural progenitor cells. *Proc. Natl. Acad. Sci. U. S. A.* 101, 16659–16664.
- Imayoshi, I., et al., 2008. Roles of continuous neurogenesis in the structural and functional integrity of the adult forebrain. *Nat. Neurosci.* 11, 1153–1161.
- Imayoshi, I., et al., 2009. Continuous neurogenesis in the adult brain. *Dev. Growth Differ.* 51, 379–386.
- Jaenisch, R., Bird, A., 2003. Epigenetic regulation of gene expression: how the genome integrates intrinsic and environmental signals. *Nat. Genet.* 33 (Suppl.), 245–254.
- Jagasia, R., et al., 2009. GABA-cAMP response element-binding protein signaling regulates maturation and survival of newly generated neurons in the adult hippocampus. *J. Neurosci.* 29, 7966–7977.
- Jaholkowski, P., et al., 2009. New hippocampal neurons are not obligatory for memory formation; cyclin D2 knockout mice with no adult brain neurogenesis show learning. *Learn. Mem.* 16, 439–451.
- Jenuwein, T., Allis, C.D., 2001. Translating the histone code. *Science* 293, 1074–1080.
- Jessberger, S., et al., 2007. Epigenetic modulation of seizure-induced neurogenesis and cognitive decline. *J. Neurosci.* 27, 5967–5975.
- Jessberger, S., et al., 2009a. Dentate gyrus-specific knockdown of adult neurogenesis impairs spatial and object recognition memory in adult rats. *Learn. Mem.* 16, 147–154.
- Jessberger, S., et al., 2009b. Making a neuron: Cdk5 in embryonic and adult neurogenesis. *Trends Neurosci.* 32, 575–582.
- Jun, K., et al., 2002. Vascular endothelial growth factor (VEGF) stimulates neurogenesis in vitro and in vivo. *Proc. Natl. Acad. Sci. U. S. A.* 99, 11946–11950.
- Jun, K., et al., 2003. Cerebral neurogenesis is induced by intranasal administration of growth factors. *Ann. Neurol.* 53, 405–409.
- Jun, K., et al., 2004a. Enhanced neurogenesis in Alzheimer's disease transgenic (PDGF-APP^{Sw,Ind}) mice. *Proc. Natl. Acad. Sci. U. S. A.* 101, 13363–13367.
- Jun, K., et al., 2004b. Increased hippocampal neurogenesis in Alzheimer's disease. *Proc. Natl. Acad. Sci. U. S. A.* 101, 343–347.
- Jun, K., et al., 2006. Alzheimer's disease drugs promote neurogenesis. *Brain Res.* 1085, 183–188.
- Jun, S.G., et al., 2008. GADD45A does not promote DNA demethylation. *PLoS Genet.* 4, e1000013.
- Johnson, M.A., et al., 2009. Cell-intrinsic signals that regulate adult neurogenesis in vivo: insights from inducible approaches. *BMB Rep.* 42, 245–259.
- Jung, H.J., et al., 2007. Base excision DNA repair defect in Gadd45a-deficient cells. *Oncogene* 26, 7517–7525.
- Kawasaki, H., et al., 2000. ATF-2 has intrinsic histone acetyltransferase activity which is modulated by phosphorylation. *Nature* 405, 195–200.
- Kempermann, G., et al., 2008. The contribution of failing adult hippocampal neurogenesis to psychiatric disorders. *Curr. Opin. Psychiatry* 21, 290–295.
- Kim, J., et al., 2007. A microRNA feedback circuit in midbrain dopamine neurons. *Science* 317, 1220–1224.
- Kim, J.B., et al., 2008. Pluripotent stem cells induced from adult neural stem cells by reprogramming with two factors. *Nature* 454, 646–650.
- Kim, J.B., et al., 2009a. Direct reprogramming of human neural stem cells by OCT4. *Nature* 461, 649–653.
- Kim, J.B., et al., 2009b. Oct4-induced pluripotency in adult neural stem cells. *Cell* 136, 411–419.
- Klose, R.J., Bird, A.P., 2006. Genomic DNA methylation: the mark and its mediators. *Trends Biochem. Sci.* 31, 89–97.
- Ko, H.G., et al., 2009. Effect of ablated hippocampal neurogenesis on the formation and extinction of contextual fear memory. *Mol. Brain* 2, 1.
- Kobayashi, K., 2009. Targeting the hippocampal mossy fiber synapse for the treatment of psychiatric disorders. *Mol. Neurobiol.* 39, 24–36.
- Kohyama, J., et al., 2008. Epigenetic regulation of neural cell differentiation plasticity in the adult mammalian brain. *Proc. Natl. Acad. Sci. U. S. A.* 105, 18012–18017.
- Kosik, K.S., 2006. The neuronal microRNA system. *Nat. Rev. Neurosci.* 7, 911–920.
- Kosik, K.S., Krichevsky, A.M., 2005. The elegance of the microRNAs: a neuronal perspective. *Neuron* 47, 779–782.
- Kuruba, R., et al., 2009. Hippocampal neurogenesis and neural stem cells in temporal lobe epilepsy. *Epilepsy Behav.* 14 (Suppl 1), 65–73.
- Kuwabara, T., et al., 2004. A small modulatory dsRNA specifies the fate of adult neural stem cells. *Cell* 116, 779–793.
- Lagos-Quintana, M., et al., 2002. Identification of tissue-specific microRNAs from mouse. *Curr. Biol.* 12, 735–739.
- Lepagnol-Bestel, A.M., et al., 2007. Nr5f silencing induces molecular and subcellular changes linked to neuronal plasticity. *Neuroreport* 18, 441–446.
- Lessard, J., et al., 2007. An essential switch in subunit composition of a chromatin remodeling complex during neural development. *Neuron* 55, 201–215.
- Leuner, B., Gould, E., 2010. Structural plasticity and hippocampal function. *Annu Rev Psychol.* 61, 111–140 C1–3.
- Levenson, J.M., et al., 2006. Evidence that DNA (cytosine-5) methyltransferase regulates synaptic plasticity in the hippocampus. *J. Biol. Chem.* 281, 15763–15773.
- Li, B., et al., 2008a. Failure of neuronal maturation in Alzheimer disease dentate gyrus. *J. Neuropathol. Exp. Neurol.* 67, 78–84.
- Li, X., et al., 2008b. Epigenetic regulation of the stem cell mitogen Fgf-2 by Mbd1 in adult neural stem/progenitor cells. *J. Biol. Chem.* 283, 27644–27652.
- Lie, D.C., et al., 2005. Wnt signalling regulates adult hippocampal neurogenesis. *Nature* 437, 1370–1375.
- Lim, L.P., et al., 2005. Microarray analysis shows that some microRNAs downregulate large numbers of target mRNAs. *Nature* 433, 769–773.
- Lim, D.A., et al., 2006. In vivo transcriptional profile analysis reveals RNA splicing and chromatin remodeling as prominent processes for adult neurogenesis. *Mol. Cell. Neurosci.* 31, 131–148.
- Lim, D.A., et al., 2009. Chromatin remodelling factor Mll1 is essential for neurogenesis from postnatal neural stem cells. *Nature* 458, 529–533.
- Liu, X., et al., 2005. Nonsynaptic GABA signaling in postnatal subventricular zone controls proliferation of GFAP-expressing progenitors. *Nat. Neurosci.* 8, 1179–1187.
- Liu, Y.W., et al., 2007. Adult neurogenesis in mesial temporal lobe epilepsy: a review of recent animal and human studies. *Curr. Pharm. Biotechnol.* 8, 187–194.
- Liu, Q., et al., 2008a. Repeated clomipramine treatment reversed the inhibition of cell proliferation in adult hippocampus induced by chronic unpredictable stress. *Pharmacogenomics* 9, 375–383.
- Liu, Y.W., et al., 2008b. Doublecortin expression in the normal and epileptic adult human brain. *Eur. J. Neurosci.* 28, 2254–2265.
- Luger, K., Richmond, T.J., 1998. The histone tails of the nucleosome. *Curr. Opin. Genet. Dev.* 8, 140–146.
- Ma, D.K., et al., 2009. Neuronal activity-induced Gadd45b promotes epigenetic DNA demethylation and adult neurogenesis. *Science* 323, 1074–1077.
- Makeyev, E.V., et al., 2007. The MicroRNA miR-124 promotes neuronal differentiation by triggering brain-specific alternative pre-mRNA splicing. *Mol. Cell* 27, 435–448.
- Malberg, J.E., et al., 2000. Chronic antidepressant treatment increases neurogenesis in adult rat hippocampus. *J. Neurosci.* 20, 9104–9110.
- Mangana, L.N., et al., 2007. Magnetic resonance spectroscopy identifies neural progenitor cells in the live human brain. *Science* 318, 980–985.

- Manji, H.K., et al., 2000. Clinical and preclinical evidence for the neurotrophic effects of mood stabilizers: implications for the pathophysiology and treatment of manic-depressive illness. *Biol. Psychiatry* 48, 740–754.
- Mao, Y., et al., 2009. Disrupted in schizophrenia 1 regulates neuronal progenitor proliferation via modulation of GSK3 β /beta-catenin signaling. *Cell* 136, 1017–1031.
- Marchetto, M.C., et al., 2009. Transcriptional signature and memory retention of human-induced pluripotent stem cells. *PLoS One* e7076, 4.
- Mattick, J.S., Makunin, I.V., 2005. Small regulatory RNAs in mammals. *Hum. Mol. Genet.* 14 (Spec No 1), R121–R132.
- Mehler, M.F., 2008. Epigenetic principles and mechanisms underlying nervous system functions in health and disease. *Prog. Neurobiol.* 86, 305–341.
- Mehler, M.F., Mattick, J.S., 2007. Noncoding RNAs and RNA editing in brain development, functional diversification, and neurological disease. *Physiol. Rev.* 87, 799–823.
- Metivier, R., et al., 2008. Cyclical DNA methylation of a transcriptionally active promoter. *Nature* 452, 45–50.
- Millar, J.K., et al., 2000. Disruption of two novel genes by a translocation co-segregating with schizophrenia. *Hum. Mol. Genet.* 9, 1415–1423.
- Ming, G.L., Song, H., 2005. Adult neurogenesis in the mammalian central nervous system. *Annu. Rev. Neurosci.* 28, 223–250.
- Miyoshi, K., et al., 2003. Disrupted-in-schizophrenia 1, a candidate gene for schizophrenia, participates in neurite outgrowth. *Mol. Psychiatry* 8, 685–694.
- Molofsky, A.V., et al., 2003. Bmi-1 dependence distinguishes neural stem cell self-renewal from progenitor proliferation. *Nature* 425, 962–967.
- Montgomery, R.L., et al., 2009. Histone deacetylases 1 and 2 control the progression of neural precursors to neurons during brain development. *Proc. Natl. Acad. Sci. U. S. A.* 106, 7876–7881.
- Morris, J.A., et al., 2003. DISC1 (disrupted-in-schizophrenia 1) is a centrosome-associated protein that interacts with MAP1A, MIPT3, ATF4/5 and NUDEL: regulation and loss of interaction with mutation. *Hum. Mol. Genet.* 12, 1591–1608.
- Morrison, S.J., Spradling, A.C., 2008. Stem cells and niches: mechanisms that promote stem cell maintenance throughout life. *Cell* 132, 598–611.
- Mosimann, C., et al., 2009. Beta-catenin hits chromatin: regulation of Wnt target gene activation. *Nat. Rev. Mol. Cell Biol.* 10, 276–286.
- Nakashima, K., et al., 1999. Synergistic signaling in fetal brain by STAT3–Smad1 complex bridged by p300. *Science* 284, 479–482.
- Namba, T., et al., 2009. The Alzheimer's disease drug memantine increases the number of radial glia-like progenitor cells in adult hippocampus. *Glia* 57, 1082–1090.
- Namihira, M., et al., 2004. Developmental stage dependent regulation of DNA methylation and chromatin modification in a immature astrocyte specific gene promoter. *FEBS Lett.* 572, 184–188.
- Namihira, M., et al., 2009. Committed neuronal precursors confer astrocytic potential on residual neural precursor cells. *Dev. Cell* 16, 245–255.
- Newton, S.S., Duman, R.S., 2006. Chromatin remodeling: a novel mechanism of psychotropic drug action. *Mol. Pharmacol.* 70, 440–443.
- Noonan, M. A., et al., 2010. Reduction of adult hippocampal neurogenesis confers vulnerability in an animal model of cocaine addiction. *J. Neurosci.* 30, 304–315.
- Nott, A., et al., 2008. S-Nitrosylation of histone deacetylase 2 induces chromatin remodelling in neurons. *Nature* 455, 411–415.
- Okano, H., Temple, S., 2009. Cell types to order: temporal specification of CNS stem cells. *Curr. Opin. Neurobiol.* 19, 112–119.
- Ooi, S.K., Bestor, T.H., 2008. The colorful history of active DNA demethylation. *Cell* 133, 1145–1148.
- Palmer, T.D., et al., 2000. Vascular niche for adult hippocampal neurogenesis. *J. Comp. Neurol.* 425, 479–494.
- Parent, J.M., et al., 2007. Is neurogenesis reparative after status epilepticus? *Epilepsia* 48 (Suppl 8), 69–71.
- Park, I.H., et al., 2008. Reprogramming of human somatic cells to pluripotency with defined factors. *Nature* 451, 141–146.
- Perera, T.D., et al., 2007. Antidepressant-induced neurogenesis in the hippocampus of adult nonhuman primates. *J. Neurosci.* 27, 4894–4901.
- Perera, T.D., et al., 2008. Cognitive role of neurogenesis in depression and antidepressant treatment. *Neuroscientist* 14, 326–338.
- Rai, K., et al., 2008. DNA demethylation in zebrafish involves the coupling of a deaminase, a glycosylase, and gadd45. *Cell* 135, 1201–1212.
- Reif, A., et al., 2006. Neural stem cell proliferation is decreased in schizophrenia, but not in depression. *Mol. Psychiatry* 11, 514–522.
- Renthal, W., et al., 2007. Histone deacetylase 5 epigenetically controls behavioral adaptations to chronic emotional stimuli. *Neuron* 56, 517–529.
- Renthal, W., Nestler, E.J., 2008. Epigenetic mechanisms in drug addiction. *Trends Mol. Med.* 14, 341–350.
- Revest, J.M., et al., 2009. Adult hippocampal neurogenesis is involved in anxiety-related behaviors. *Mol. Psychiatry* 14, 959–967.
- Riggs, A.D., Porter, T.N., 1996. Overview of Epigenetic Mechanisms. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York.
- Ringrose, L., Paro, R., 2004. Epigenetic regulation of cellular memory by the Polycomb and Trithorax group proteins. *Annu. Rev. Genet.* 38, 413–443.
- Saini, H.K., et al., 2007. Genomic analysis of human microRNA transcripts. *Proc. Natl. Acad. Sci. U. S. A.* 104, 17719–17724.
- Saini, H.K., et al., 2008. Annotation of mammalian primary microRNAs. *BMC Genomics* 9, 564.
- Sairanen, M., et al., 2005. Brain-derived neurotrophic factor and antidepressant drugs have different but coordinated effects on neuronal turnover, proliferation, and survival in the adult dentate gyrus. *J. Neurosci.* 25, 1089–1094.
- Santarelli, L., et al., 2003. Requirement of hippocampal neurogenesis for the behavioral effects of antidepressants. *Science* 301, 805–809.
- Sapolsky, R.M., 2000. Glucocorticoids and hippocampal atrophy in neuropsychiatric disorders. *Arch. Gen. Psychiatry* 57, 925–935.
- Sapolsky, R.M., 2004. Is impaired neurogenesis relevant to the affective symptoms of depression? *Biol. Psychiatry* 56, 137–139.
- Schaefer, A., et al., 2007. Cerebellar neurodegeneration in the absence of microRNAs. *J. Exp. Med.* 204, 1553–1558.
- Schanzer, A., et al., 2004. Direct stimulation of adult neural stem cells in vitro and neurogenesis in vivo by vascular endothelial growth factor. *Brain Pathol.* 14, 237–248.
- Scharfman, H.E., Hen, R., 2007. Neuroscience. Is more neurogenesis always better? *Science* 315, 336–338.
- Schreiber, S.L., Bernstein, B.E., 2002. Signaling network model of chromatin. *Cell* 111, 771–778.
- Schuermans, C., Guillemot, F., 2002. Molecular mechanisms underlying cell fate specification in the developing telencephalon. *Curr. Opin. Neurobiol.* 12, 26–34.
- Shamoviy, I., Nudler, E., 2006. Gene control by large noncoding RNAs. *Sci. STKE* 2006, pe40.
- Sharma, R.P., 2005. Schizophrenia, epigenetics and ligand-activated nuclear receptors: a framework for chromatin therapeutics. *Schizophr. Res.* 72, 79–90.
- Shen, Q., et al., 2004. Endothelial cells stimulate self-renewal and expand neurogenesis of neural stem cells. *Science* 304, 1338–1340.
- Shen, Q., et al., 2008. Adult SVZ stem cells lie in a vascular niche: a quantitative analysis of niche cell–cell interactions. *Cell Stem Cell* 3, 289–300.
- Shihabuddin, L.S., et al., 2000. Adult spinal cord stem cells generate neurons after transplantation in the adult dentate gyrus. *J. Neurosci.* 20, 8727–8735.
- Singer, B.H., et al., 2009. Conditional ablation and recovery of forebrain neurogenesis in the mouse. *J. Comp. Neurol.* 514, 567–582.
- Smrt, R.D., et al., 2007. Mecp2 deficiency leads to delayed maturation and altered gene expression in hippocampal neurons. *Neurobiol. Dis.* 27, 77–89.
- Soldner, F., et al., 2009. Parkinson's disease patient-derived induced pluripotent stem cells free of viral reprogramming factors. *Cell* 136, 964–977.
- Song, H., et al., 2002a. Astroglia induce neurogenesis from adult neural stem cells. *Nature* 417, 39–44.
- Song, H.J., et al., 2002b. Neural stem cells from adult hippocampus develop essential properties of functional CNS neurons. *Nat. Neurosci.* 5, 438–445.
- Strahl, B.D., Allis, C.D., 2000. The language of covalent histone modifications. *Nature* 403, 41–45.
- Suh, H., et al., 2007. In vivo fate analysis reveals the multipotent and self-renewal capacities of Sox2+ neural stem cells in the adult hippocampus. *Cell Stem Cell* 1, 515–528.
- Suhonen, J.O., et al., 1996. Differentiation of adult hippocampus-derived progenitors into olfactory neurons in vivo. *Nature* 383, 624–627.
- Sun, Y., et al., 2001. Neurogenin promotes neurogenesis and inhibits glial differentiation by independent mechanisms. *Cell* 104, 365–376.
- Surget, A., et al., 2008. Drug-dependent requirement of hippocampal neurogenesis in a model of depression and of antidepressant reversal. *Biol. Psychiatry* 64, 293–301.
- Sweatt, J.D., 2009. Experience-dependent epigenetic modifications in the central nervous system. *Biol. Psychiatry* 65, 191–197.
- Takahashi, K., Yamanaka, S., 2006. Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. *Cell* 126, 663–676.
- Takahashi, K., et al., 2007. Induction of pluripotent stem cells from adult human fibroblasts by defined factors. *Cell* 131, 861–872.
- Takizawa, T., et al., 2001. DNA methylation is a critical cell-intrinsic determinant of astrocyte differentiation in the fetal brain. *Dev. Cell* 1, 749–758.
- Tashiro, A., et al., 2006. NMDA-receptor-mediated, cell-specific integration of new neurons in adult dentate gyrus. *Nature* 442, 929–933.
- Tavazoie, M., et al., 2008. A specialized vascular niche for adult neural stem cells. *Cell Stem Cell* 3, 279–288.
- Temple, S., 2001. The development of neural stem cells. *Nature* 414, 112–117.
- Tomita, K., et al., 2000. Mammalian achaete-scute and atonal homologs regulate neuronal versus glial fate determination in the central nervous system. *EMBO J.* 19, 5460–5472.
- Tozuka, Y., et al., 2005. GABAergic excitation promotes neuronal differentiation in adult hippocampal progenitor cells. *Neuron* 47, 803–815.
- Tran, H., et al., 2002. DNA repair pathway stimulated by the forkhead transcription factor FOXO3a through the Gadd45 protein. *Science* 296, 530–534.
- Tsankova, N.M., et al., 2004. Histone modifications at gene promoter regions in rat hippocampus after acute and chronic electroconvulsive seizures. *J. Neurosci.* 24, 5603–5610.
- Tsankova, N.M., et al., 2006. Sustained hippocampal chromatin regulation in a mouse model of depression and antidepressant action. *Nat. Neurosci.* 9, 519–525.
- Tsujimura, K., et al., 2009. Neuronal differentiation of neural precursor cells is promoted by the methyl-CpG-binding protein MeCP2. *Exp. Neurol.* 219, 104–111.
- Turner, B.M., 2000. Histone acetylation and an epigenetic code. *Bioessays* 22, 836–845.
- Umka, J., et al., 2009. Valproic acid reduces spatial working memory and cell proliferation in the hippocampus. *Neuroscience* 166, 15–22.
- van Praag, H., et al., 1999. Running enhances neurogenesis, learning, and long-term potentiation in mice. *Proc. Natl. Acad. Sci. U. S. A.* 96, 13427–13431.
- Wada, A., 2009. Lithium and neuropsychiatric therapeutics: neuroplasticity via glycogen synthase kinase-3 β , beta-catenin, and neurotrophin cascades. *J. Pharmacol. Cell.* 110, 14–28.
- Wernig, M., et al., 2008. Neurons derived from reprogrammed fibroblasts functionally integrate into the fetal brain and improve symptoms of rats with Parkinson's disease. *Proc. Natl. Acad. Sci. U. S. A.* 105, 5856–5861.
- Wu, H., Sun, Y.E., 2009. Reversing DNA methylation: new insights from neuronal activity-induced Gadd45b in adult neurogenesis. *Sci. Signal.* 2, pe17.

- Wu, J.I., et al., 2007. Regulation of dendritic development by neuron-specific chromatin remodeling complexes. *Neuron* 56, 94–108.
- Yamaguchi, M., et al., 2004. Repetitive cocaine administration decreases neurogenesis in adult rat hippocampus. *Ann. N. Y. Acad. Sci.* 1025, 351–362.
- Yoo, A.S., et al., 2009. MicroRNA-mediated switching of chromatin-remodelling complexes in neural development. *Nature* 460, 642–646.
- Yu, I.T., et al., 2009a. Valproic acid promotes neuronal differentiation by induction of proneural factors in association with H4 acetylation. *Neuropharmacology* 56, 473–480.
- Yu, Y., et al., 2009b. Increased hippocampal neurogenesis in the progressive stage of Alzheimer's disease phenotype in an APP/PS1 double transgenic mouse model. *Hippocampus* 19, 1247–1253.
- Zhao, X., et al., 2003. Mice lacking methyl-CpG binding protein 1 have deficits in adult neurogenesis and hippocampal function. *Proc. Natl. Acad. Sci. U. S. A.* 100, 6777–6782.
- Zhao, C., et al., 2006. Distinct morphological stages of dentate granule neuron maturation in the adult mouse hippocampus. *J. Neurosci.* 26, 3–11.
- Zhang, C., et al., 2007. Long-lasting impairment in hippocampal neurogenesis associated with amyloid deposition in a knock-in mouse model of familial Alzheimer's disease. *Exp. Neurol.* 204, 77–87.
- Zhao, C., et al., 2008. Mechanisms and functional implications of adult neurogenesis. *Cell* 132, 645–660.
- Zhu, J.K., 2009. Active DNA demethylation mediated by DNA glycosylases. *Annu. Rev. Genet.* 43, 143–166.