



## Synthesis and biological evaluation of a new vitamin D<sub>2</sub> analogue

Zoila Gándara<sup>a</sup>, Manuel Pérez<sup>a</sup>, Débora G. Salomón<sup>b</sup>, María J. Ferronato<sup>b</sup>, María E. Fermento<sup>b</sup>, Alejandro C. Curino<sup>b</sup>, María M. Facchinetti<sup>b</sup>, Generosa Gómez<sup>a</sup>, Yagamare Fall<sup>a,\*</sup>

<sup>a</sup>Departamento de Química Orgánica, Facultad de Química, Universidad de Vigo, 36200 Vigo, Spain

<sup>b</sup>Laboratorio de Biología del Cáncer, Instituto de Investigaciones Bioquímicas Bahía Blanca, Centro Científico Tecnológico Bahía Blanca, CONICET, Argentina

### ARTICLE INFO

#### Article history:

Received 12 June 2012

Revised 27 July 2012

Accepted 30 July 2012

Available online 7 August 2012

#### Keywords:

Vitamin D<sub>2</sub> analogues

Calcitriol

Cancer

Hypercalcemia

Cellular proliferation

Julia–Kocienski olefination

### ABSTRACT

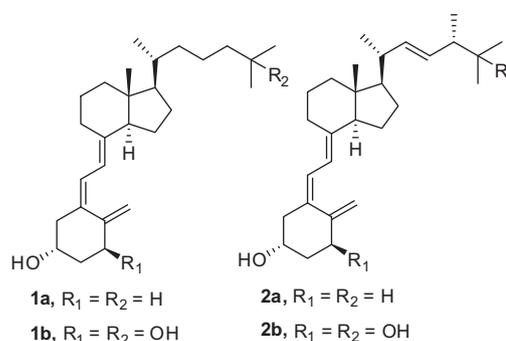
A new vitamin D<sub>2</sub> analogue was synthesized using the Julia–Kocienski olefination. It has antiproliferative effects on cell lines from squamous cell carcinomas of colon and head and neck, but is also as hypercalcaemic as calcitriol in vivo.

© 2012 Elsevier Ltd. All rights reserved.

1 $\alpha$ ,25-Dihydroxyvitamin D<sub>3</sub> (**1b**, calcitriol) (Fig. 1), the active hormone metabolized from vitamin D<sub>3</sub> (**1a**), acts as a regulator in calcium and phosphate homeostasis.<sup>1</sup> Apart from these classic activities, calcitriol has proved to inhibit cellular proliferation and to induce cellular differentiation.<sup>2</sup> However, therapeutic use of these latter effects of **1b** is hampered by the hypercalcaemic side effects of effective doses. This has stimulated a search for analogues that combine a relatively weak systemic effect on calcium metabolism with potent regulatory effects on cell differentiation and proliferation.

The metabolism and biological activity of the non-natural vitamin ergocalciferol (**2a**, vitamin D<sub>2</sub>) appear to parallel those of vitamin D<sub>3</sub>,<sup>3</sup> and it is in fact marketed as a treatment for refractory rickets. In particular, **2a** undergoes double hydroxylation to 1 $\alpha$ ,25-dihydroxyvitamin D<sub>2</sub> (**2b**),<sup>4</sup> which like **1b** induces cell differentiation and inhibits the proliferation of a number of tumour cell lines, including leukaemia cells<sup>5</sup> (in spite of which, very few syntheses of either 1 $\alpha$ ,25-(OH)<sub>2</sub>-D<sub>2</sub> or 25-(OH)-D<sub>2</sub> have been reported to date).<sup>6</sup>

The vitamin D<sub>2</sub> analogue paricalcitol has been shown to be less hypercalcaemic than calcitriol in vitro<sup>7</sup> and to have antiproliferative activity,<sup>8,9</sup> and in clinical trials it has proved to be partially effective against certain cancers.<sup>10,11</sup> These findings imply that the search for members of the vitamin D family with low hypercalcaemic activity and a high therapeutic index should not overlook



**Figure 1.** Structures of vitamin D<sub>3</sub> (**1a**) and vitamin D<sub>2</sub> (**2a**) and their metabolites **1b** and **2b**.

vitamin D<sub>2</sub> analogues. Here we describe the synthesis of vitamin D<sub>2</sub> analogue **3** (Fig. 2), the 22Z side chain of which was recently obtained serendipitously by means of a Julia–Kocienski olefination,<sup>12</sup> together with assays of its antiproliferative activities in various cancer cell lines and its hypercalcaemic effects in vivo.

For the synthesis of **3** we started from alcohol **4** (Scheme 1), which is readily obtained in large quantities from vitamin D<sub>2</sub> using the procedures originally described by Calverley<sup>13</sup> and later modified by Choudhry.<sup>14</sup> TPAP oxidation of **4** afforded aldehyde **5** in 95% yield, and Julia–Kocienski olefination of **5** with sulfone **6** gave a 65% yield of ester **7**, which upon reaction with ethyl magnesium iodide in ether at 0°C yielded alcohol **8**. Removal of the silyl protecting

\* Corresponding author.

E-mail address: [yagamare@uvigo.es](mailto:yagamare@uvigo.es) (Y. Fall).

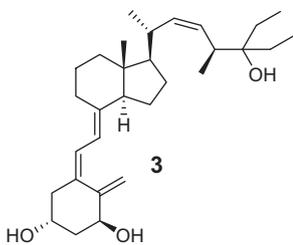
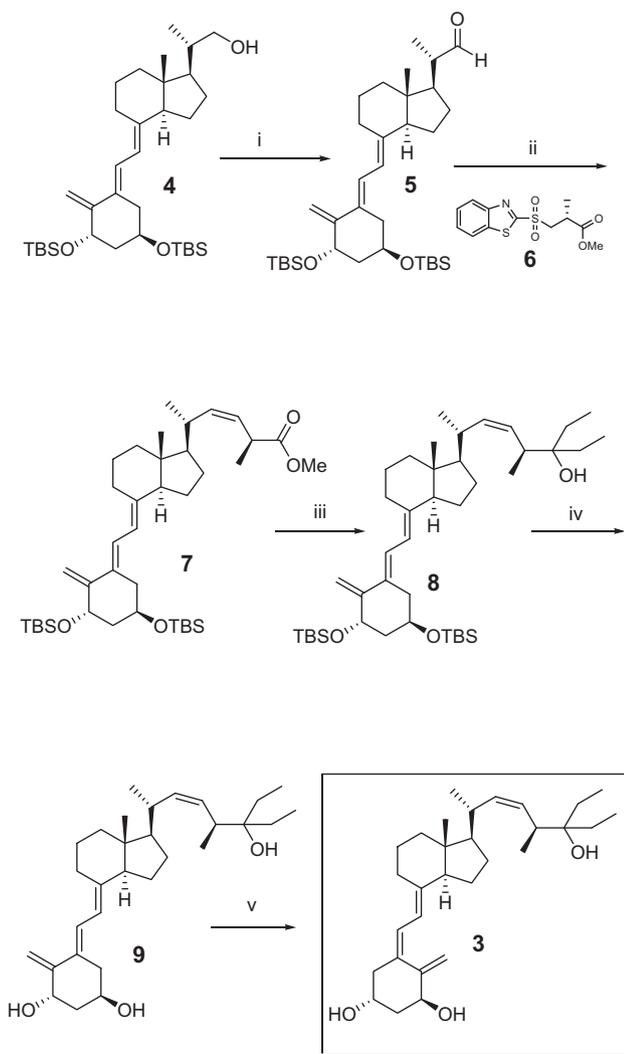


Figure 2. Structure of vitamin D<sub>2</sub> analogue **3**.

groups of **8** with TBAF in THF afforded a 98% yield of triol **9**, and photoisomerization of **9** using anthracene as sensitizer finally gave the target analogue **3** in 90% yield. The overall yield of this five-step synthetic sequence was an excellent 44%.

In order to test the biological activity of analogue **3**, we first analyzed the antiproliferative effects of this compound in several cancer cell lines. For this purpose we performed cell count following treatment of human squamous cell carcinoma (HN12), glioblastoma (T98G), mammary carcinoma (human T47D and murine LM3) and colorectal cancer (HCT116) cell lines with analogue **3**, calcitriol or vehicle. We performed 120 h dose-response analyses comparing



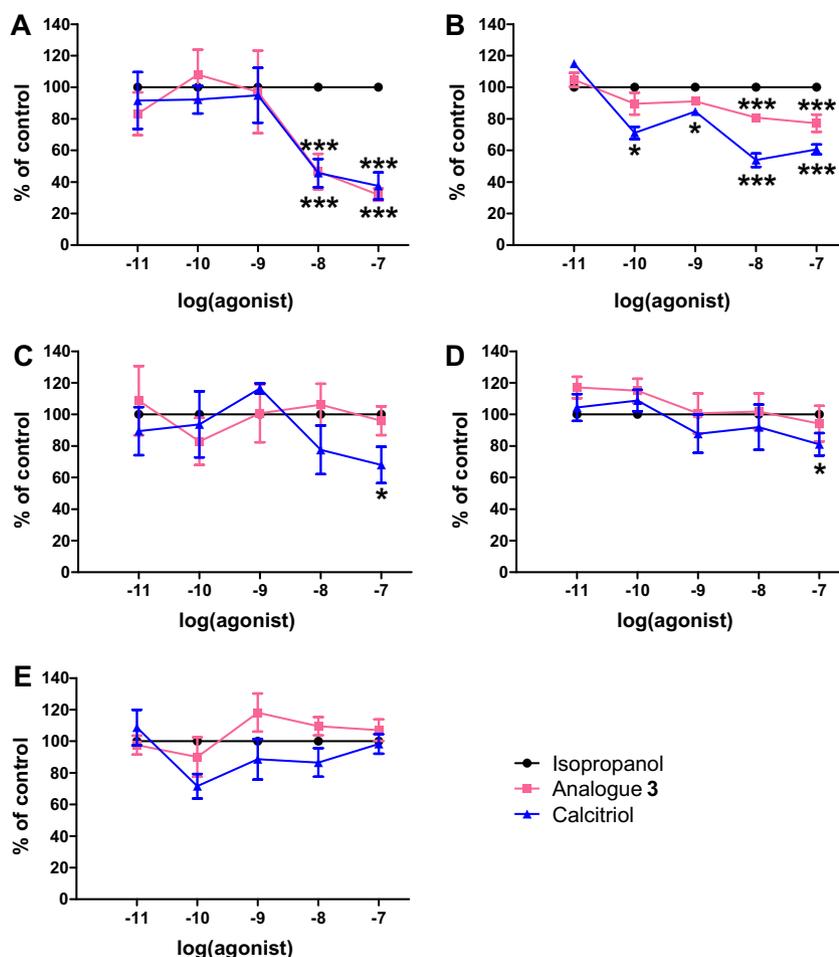
Scheme 1. Reagents and conditions: (i) TPAP, NMO, CH<sub>2</sub>Cl<sub>2</sub>, molecular sieves (95%); (ii) **6**, LiHMDS, THF, −78 °C (65%); (iii) EtMgI, Et<sub>2</sub>O, 0 °C (80%); (iv) TBAF, THF (98%); (v) anthracene, Et<sub>3</sub>N, hv, CH<sub>2</sub>Cl<sub>2</sub>, MeOH (90%).

the effects of analogue **3** with those elicited by the natural hormone calcitriol. As shown in Figure 3, we observed a significant decrease in cell count after treatment with analogue **3**, in the human squamous cell carcinoma (A) and the human colorectal cancer (B) cell lines. The antiproliferative effect exerted by analogue **3** in HN12 cells was similar to that elicited by calcitriol. The half maximal inhibitory concentration (IC<sub>50</sub>) was 5.05 nM and 4.68 nM for analogue **3** and calcitriol, respectively. In HCT116 cells, the IC<sub>50</sub> was 0.76 nM and 0.01 nM for analogue **3** and calcitriol, respectively. No effect on cell proliferation was observed in the mammary cancer (C and D) and the glioma (E) cell lines.

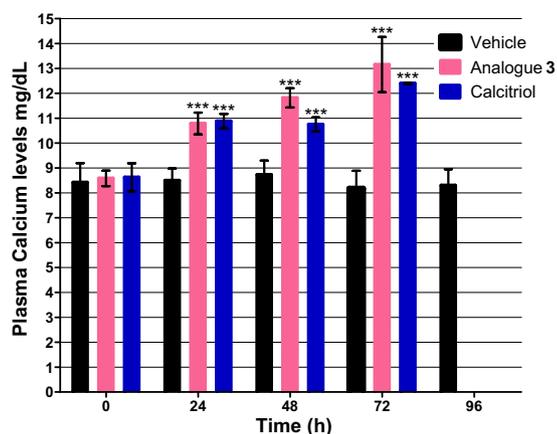
It is known that cancer cell lines display a range of sensitivities to the anti-proliferative effects of calcitriol and its derivatives; the reason for this is largely unknown and could result from defects in any component in the VDR signaling pathway including VDR and 24-hydroxylase (CYP24A1).<sup>15</sup> Calcitriol action is limited by its catabolism, which occurs mainly by CYP24A1 resulting in 1 $\alpha$ ,24,25 (OH)<sub>3</sub>-D<sub>3</sub>, a metabolite with substantially lower affinity for the vitamin D receptor VDR. Although this enzyme is located primarily in liver, it has been demonstrated to be expressed by many tissues.<sup>15</sup> Augmented expression of CYP24A1 has been shown to be detrimental to calcitriol antiproliferative effects. In prostate cancer cell lines, it has been demonstrated that enzyme expression was inversely correlated to the antiproliferative effects displayed by the cells.<sup>16</sup> In this regard, analogue **3** may be degraded by cells displaying high expression of CYP24A1, which could account for the lack of activity observed in some of the cell lines, similarly to what occurs with calcitriol treatment. It has also been reported that splice variants of the enzyme may have implications for the antitumorigenic effects of the hormone and analogues.<sup>17</sup>

As already stated, vitamin D and many of its analogues induce hypercalcemia, a fact that limits the therapeutic efficacy of these compounds for cancer treatment. Therefore, analyses of the calcemic effects need to be performed for each novel analogue synthesized. Because of its significant *in vitro* antiproliferative activity in human squamous cell carcinoma and in colorectal carcinoma, analogue **3** was then evaluated for its calcemic effects *in vivo*. Previous pharmacokinetic studies performed in normal mice indicated that calcitriol at 0.125  $\mu$ g/mouse (approximately 5  $\mu$ g/kg body weight) results in a C<sub>max</sub> > 10.0 ng/mL and AUC > 40.0 ng h/mL<sup>18</sup>, exceeding the concentration needed for calcitriol anti-tumor activity *in vitro*.<sup>19</sup> Other studies using either calcitriol or vitamin D analogues at doses lower than 5  $\mu$ g/kg body weight showed a reduction in tumor burden in animal models of cancer.<sup>20–22</sup> Therefore we chose the dose of 5  $\mu$ g/kg body weight that have antitumor effects *in vivo* as a starting concentration for the studies of calcemia, with the idea of increasing the doses in escalating experiments, provided no toxic effects were observed. Mice were divided into three groups ( $n = 5$ /group) and given a daily intraperitoneal injection of calcitriol, analogue **3** or vehicle at 5  $\mu$ g/kg body weight for 4 days. Blood was collected prior to dose administration, then at 24, 48, 72 and 96 h post-treatment. The health condition of the animals was recorded. Plasma calcium levels were measured by reading the absorbance of metallochromic indicator arsenazo III, as previously described.<sup>23</sup> As observed in Figure 4, analogue **3** induced hypercalcemia starting at 24 h of compound delivery, similar to that elicited by calcitriol. Moreover, mice treated with analogue **3** or calcitriol died after 4 days of treatment. Together with the high calcium plasma levels we observed, in the calcitriol- treated mice and the analogue **3**-treated ones, the appearance of conjunctivitis, chills and thirst which are symptoms and signs that have been associated to hypercalcemia.<sup>24</sup> Additionally, an increase in haematocrit was observed (50 at 72 hours; normal levels range from 39 to 47<sup>25</sup>), which is a sign of intoxication.<sup>26</sup>

Vitamin D<sub>2</sub> analogues such as paricalcitol have been shown to be less calcemic than calcitriol in *in vitro* studies<sup>7</sup> and have been



**Figure 3.** Dose-response effects of analogue 3 on cellular survival and its comparison with calcitriol. (A) Human head and neck squamous cell carcinoma HN12. (B) Human colorectal carcinoma HCT116. (C) Murine mammary adenocarcinoma LM3. (D) Human mammary adenocarcinoma T47D and (E) Human glioma T98G. Cells were exposed to the indicated doses of vehicle (isopropanol), analogue 3 or calcitriol over a total time of 120 h. Cellular survival was expressed as percentage of the vehicle. Data points represent means  $\pm$  SD from five replicates; \* $p \leq 0,05$  and \*\*\* $p \leq 0001$ , with respect to vehicle. The experiments were repeated at least three times for each cell line.



**Figure 4.** Plasma calcium levels in mice in response to daily intraperitoneal injections of isopropanol (vehicle), analogue 3 or calcitriol during a period of 4 days. Animals were injected with 5  $\mu$ g/kg body weight of compounds, and plasma calcium was measured before the injection (0 h, basal levels) and at 24, 48, 72 and 96 h. Normal levels range from 8.8 to 10.4 mg/dL.<sup>27</sup> Values for calcitriol and analogue 3 at 96 h were not available because animals died following 3 days of treatment due to hypercalcemia. Values are means  $\pm$  SDs from five animals in each group. The experiment was repeated two times. \*\*\* $p \leq 0.0001$ , with respect to vehicle.

demonstrated to have antiproliferative activity.<sup>8,9</sup> Clinical trials showed a partial response of paricalcitol in cancer<sup>10,11</sup> and therefore new vitamin D<sub>2</sub> analogues should be assayed for antitumoral activity with the aim to identify one with low hypercalcemic effects and high therapeutic index. The study of the biologic effects of analogue 3 showed that although it exerts antiproliferative effects in colon and head and neck squamous cell carcinomas, it displays similar hypercalcemic effects to those elicited by calcitriol.

#### Acknowledgments

This work was supported financially by the Spanish Ministry of Foreign Affairs and Cooperation (PCI A/030052/10) and the Xunta de Galicia (INCITE845B-2010/020 and INCITE08PXIB314255PR) and the ANPCyT from Argentina. The work of the NMR and MS divisions of the research support services of the University of Vigo (CACTI) is also gratefully acknowledged. Zoila Gándara and Manuel Pérez thank the Xunta de Galicia for an Angeles Alvariño contract.

#### References and notes

1. Norman, A. W. *Vitamin D, the Calcium Homeostatic Steroid Hormone*; Academic Press: New York, 1979.
2. (a) For general reviews of vitamin D chemistry and biology, see: Vitamin D: Chemistry, Biology and clinical Application of the Steroid Hormone; Norman, A. W.; Bouillon, R.; Thomasset, M. Eds., Vitamin D Workshop: Riverside, CA,

- 1997.; (b) Feldman, D.; Glorieux, F. H.; Pike, J. W. *Vitamin D*; Academic Press: San Diego, CA, 1997; (c) Pardo, R.; Santelli, M. *Bull. Soc. Chim. Fr.* **1985**, 98; (d) Dai, H.; Posner, G. H. *Synthesis* **1994**, 1383; (e) Zhu, G.-D.; Okamura, W. H. *Chem. Rev.* **1877**, 1995, 95; (f) Posner, G. H.; Kahraman, M. *Eur. J. Org. Chem.* **2003**, 3889.
3. Jones, G.; Schnoes, H. K.; DeLuca, H. F. *Biochemistry* **1975**, 14, 1250.
4. (a) Drescher, D.; DeLuca, H. F.; Imrie, H. H. *Arch. Biochem. Biophys.* **1969**, 130, 657; (b) Norman, A. W.; Roth, J.; Orci, L. *Endocr. Rev.* **1982**, 3, 331; (c) Sjoden, G.; Smith, C.; Lindgren, U.; DeLuca, H. F. *Proc. Soc. Exp. Biol. Med.* **1985**, 178, 432; (d) Reddy, G. S.; Tserng, K. *Biochemistry* **1986**, 25, 5328; (e) Koszewski, N. J.; Reinhardt, T. A.; Napoli, J. L.; Beitz, D. C.; Horst, R. L. *Biochemistry* **1988**, 27, 5785, and referencies therein.
5. (a) Reinhardt, T. A.; Ramberg, C. F.; Horst, R. L. *Arch. Biochem. Biophys.* **1989**, 273, 64; (b) Norman, A. W.; Koeffler, H. P.; Bishop, J. E.; Collins, E. D.; Sergeev, I.; Zhou, L. X.; Nemer, I.; Zhou, J.; Henry, H. L.; Okamura, W. H. In *Vitamin D: Gene Regulation, Structure function Analysis and Clinical Application*; Norman, A. W., Bouillon, R., Thomasset, M., Eds.; Walter de Gruyter: Berlin, 1991; p 146.
6. (a) Morzycki, J. W.; Schnoes, H. K.; DeLuca, H. F. *J. Org. Chem.* **1984**, 49, 2148; (b) Baggiolini, E. G.; Iacobelli, J. A.; Hennessy, B. M.; Batcho, A. D.; Sereno, J. F.; Uskokovic, M. R. *J. Org. Chem.* **1986**, 51, 3098; (c) Wilson, S. R.; Davey, A. E.; Guazzaroni, M. E. *J. Org. Chem.* **2007**, 1992, 57; (d) Granja, J. R.; Castedo, L.; Mouriño, A. *J. Org. Chem.* **1993**, 58, 124; (e) Torneiro, M.; Fall, Y.; Castedo, L.; Mouriño, A. *J. Org. Chem.* **1997**, 62, 6344. and referencies therein; (f) Yamada, S.; Shiraishi, M.; Ohmori, M.; Takayama, H. *Tetrahedron Lett.* **1984**, 25, 3347.
7. Trump, D. L.; Deeb, K. K.; Johnson, C. S. *Cancer J.* **2010**, 1, 1.
8. Chen, T. C.; Schwartz, G. G.; Burnstein, K. L.; Lokeshwar, B. L.; Holick, M. F. *Clin. Cancer Res.* **2000**, 6, 901.
9. Schwartz, G. G.; Eads, D.; Naczki, C.; Northrup, S.; Chen, T.; Koumenis, C. *Cancer Biol. Ther.* **2008**, 7, 430.
10. Schwartz, G. G.; Hall, M. C.; Stindt, D.; Patton, S.; Lovato, J.; Torti, F. M. *Clin. Cancer Res.* **2005**, 11, 8680.
11. Liu, G.; Oettel, K.; Ripple, G.; Staab, M. J.; Horvath, D.; Alberti, D.; Arzoomanian, R.; Marnocha, R.; Bruskevitz, R.; Mazess, R.; Bishop, C.; Bhattacharya, A.; Bailey, H.; Wilding, G. *Clin. Cancer Res.* **2002**, 8(9), 2820.
12. Gándara, Z.; Pérez, M.; Pérez-García, X.; Gómez, G.; Fall, Y. *Tetrahedron Lett.* **2009**, 50, 4874.
13. Calverley, M. J. *Tetrahedron* **1987**, 43, 4609.
14. Choudhry, S. C.; Belica, P. S.; Coffen, D. L.; Focella, A.; Maehr, H.; Manchand, P. S.; Serico, L.; Yang, R. T. *J. Org. Chem.* **1993**, 58, 1496.
15. Trump, D. L.; Deeb, K. K.; Johnson, C. S. *Cancer J.* **2010**, 16, 1.
16. Moreno, J.; Krishnan, A. V.; Feldman, D. J. *Steroid Biochem. Mol. Biol.* **2005**, 97, 31.
17. Horváth, H. C.; Khabir, Z.; Nittke, T.; Gruber, S.; Speer, G.; Manhardt, T.; Bonner, E.; Kallay, E. *J. Steroid Biochem. Mol. Biol.* **2010**, 121(1–2), 76.
18. Muindi, J. R.; Modzelewski, R. A.; Peng, Y.; Trump, D. L.; Johnson, C. S. *Oncology* **2004**, 66(1), 62.
19. Muindi, J. R.; Yu, W. D.; Ma, Y.; Engler, K. L.; Kong, R. X.; Trump, D. L.; Johnson, C. S. *Endocrinology* **2010**, 151, 4301.
20. Kumagai, T.; O'Kelly, J.; Said, J. W.; Koeffler, H. P. *J. Natl. Cancer Inst.* **2003**, 95(12), 896.
21. Light, B. W.; Yu, W. D.; McElwain, M. C.; Russell, D. M.; Trump, D. L.; Johnson, C. S. *Cancer Res.* **1997**, 57(17), 3759.
22. Prudencio, J.; Akutsu, N.; Benlimame, N.; Wang, T.; Bastien, Y.; Lin, R.; Black, M. J.; Alaoui-Jamali, M. A.; White, J. H. *J. Natl. Cancer Inst.* **2001**, 93(10), 745.
23. Salomón, D. G.; Grioli, S. M.; Buschiazio, M.; Mascaró, E.; Vitale, C.; Radivoy, G.; Pérez, M.; Fall, Y.; Mesri, E. A.; Curino, A. C.; Facchinetti, M. M. *ACS Med. Chem. Lett.* **2011**, 2(7), 503.
24. Hathcock, J. N.; Shao, A.; Vieth, R.; Heaney, R. *Am. J. Clin. Nutr.* **2007**, 85, 6.
25. Windberger, U.; Bartholovitsch, A.; Plasenzotti, R.; Korak, K. J.; Heinze, G. *Exp. Physiol.* **2003**, 88(3), 431.
26. OECD. Repeated Dose 28-day Oral Toxicity Study in Rodents, guideline 407, the OECD guideline for testing of chemical. 1995.
27. Spina, C.; Tangpricha, V.; Yao, M.; Zhou, W.; Wolfe, M. M.; Maehr, H.; Uskokovic, M.; Adorini, L.; Holick, M. F. *J. Steroid Biochem. Mol. Biol.* **2005**, 97 (1–2), 111.