



## Synthesis of hydroxypyrrone- and hydroxythiopyrrone-based matrix metalloproteinase inhibitors: Developing a structure–activity relationship

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### ABSTRACT

The zinc(II)-dependent matrix metalloproteinases (MMPs) are associated with a variety of diseases. Development of inhibitors to modulate MMP activity has been an active area of investigation for therapeutic development. Hydroxypyrones and hydroxythiopyrones are alternative zinc-binding groups (ZBGs) that, when combined with peptidomimetic backbones, comprise a novel class of MMP inhibitors (MMPi). In this report, a series of hydroxypyrrone- and hydroxythiopyrrone-based MMPi with aryl backbones at the 2-, 5-, and 6-positions of the hydroxypyrrone ring have been synthesized. Synthetic routes for developing inhibitors with substituents at two of these positions (so-called double-handed inhibitors) are also explored. The MMP inhibition profiles and structure–activity relationship of synthesized hydroxypyrones and hydroxythiopyrones have been analyzed. The results here show that the ZBG, the position of the backbone on the ZBG, and the nature of the linker between the ZBG and backbone are critical for MMPi activities.

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Matrix metalloproteinases (MMPs) are a class of hydrolytic metalloenzymes involved in the degradation of the extracellular matrix (ECM).<sup>1–3</sup> At the catalytic center of MMPs is a conserved tris(histidine)-bound zinc(II) ion. The protein matrix surrounding the zinc center is comprised of a series of subsite pockets designated as S1', S2', S3', S1, S2, and S3 (Fig. 1). The different structures of the MMP subsites, and the amino acids comprising those subsites, lead to substrate selectivity for different MMP isoforms. MMPs are involved in tissue remodeling, wound healing, and growth. The misregulated activities of these enzymes are also implicated in a variety of diseases such as cancer, arthritis, atherosclerosis, and heart disease.<sup>1–3</sup> Thus, a number of investigations in both academia and industry have been carried out to develop MMP inhibitors (MMPi) as therapeutics to treat MMP-related diseases.<sup>1–4</sup> A typical MMPi has two parts (Fig. 1): a zinc-binding group (ZBG) able to chelate a zinc(II) ion thus blocking the access of the substrate to the catalytic center, and a peptidomimetic backbone that provides non-covalent interactions with the subsite pockets thus tuning inhibitor potency and selectivity. Often in an MMPi a linking group (L) can also be defined that connects the backbone substituent to the ZBG. Most reported MMPi employ a hydroxamic acid as the ZBG with at least part of the backbone directed toward the S1' pocket.<sup>1–4</sup> This strategy has been successful in providing potent MMPi, but hydroxamate inhibitors have limitations including *in vivo* hydrolysis and dose-limiting side effects. To

overcome the drawbacks of hydroxamic acids, new MMPi with alternative ZBGs have been explored.

Hydroxypyrones and hydroxythiopyrones have versatile metal coordination chemistry.<sup>5–10</sup> Among the most commonly studied hydroxypyrones are natural products maltol and kojic acid (Fig. 2), which are widely used as food and cosmetics additives, suggesting that they possess good biocompatibility.<sup>11</sup> Hydroxypyrones have been explored for the development of new MMPi. Tris(pyrazolyl)borate zinc(II) complexes, used to mimic the MMP catalytic zinc(II) center, show that hydroxypyrones and hydroxythiopyrones can bind to the zinc(II) ion in a bidentate fashion.<sup>12</sup> Maltol (hydroxypyrrone) and thiomaltol (hydroxythiopyrrone) are more effective ZBGs against MMP-3 (stromelysin) when compared to a simple hydroxamate ligand.<sup>13,14</sup> Using hydroxypyrrone as the ZBG, a series of potent and selective pyrone-based inhibitors of MMP-3 have been developed by attaching an aryl backbone to the 2-position of the pyrone ring.<sup>15,16</sup> Simple hydroxythiopyrones have shown much higher inhibition activity (30–60 fold) than corresponding hydroxypyrones.<sup>14</sup> This observation promoted us to explore the potential of a hydroxythiopyrrone ZBG for development of full length MMP inhibitors. Unlike hydroxamate terminal chelators, in which the inhibitor backbone can only be extended in one direction, the hydroxypyrrone ring has several positions (2-, 5-, and 6-) to attach backbones (Fig. 2). Hence, it is possible to develop novel hydroxypyrrone-based inhibitors with multiple backbones to interact with MMP pockets (Fig. 1). In this report, we describe the syntheses of hydroxythiopyrrone-based MMP inhibitors and their inhibition activities are compared with that

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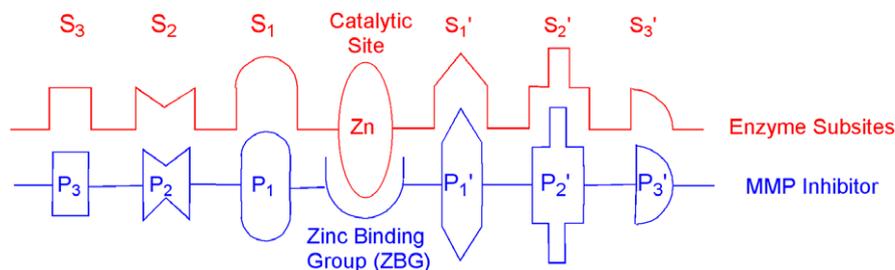


Figure 1. Schematic interactions between MMPs and MMP inhibitors.

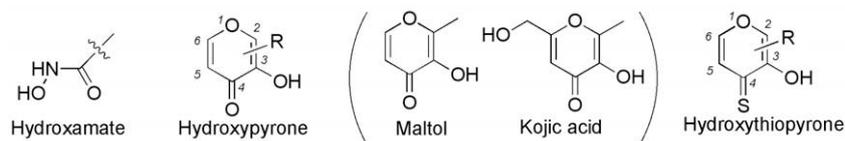


Figure 2. Structures of hydroxamate, hydroxypyrrone, and hydroxythiopyrrone chelators.

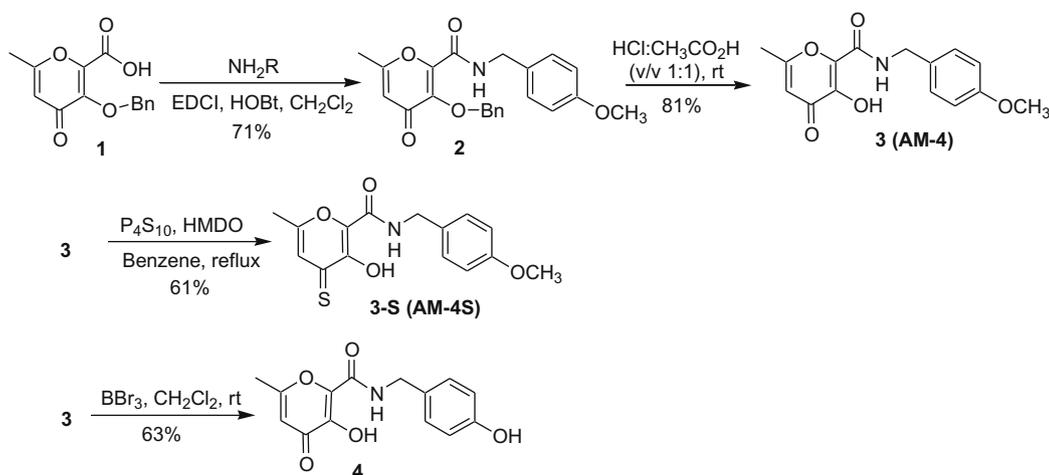
of corresponding hydroxypyrones. The synthetic schemes for developing double-handed hydroxypyrrone-based inhibitors have been explored. Hydroxypyrones and their hydroxythiopyrrone analogues have also been widely reported in other biomedical applications such as iron balance in anemia and iron overload disorder,<sup>7,8</sup> aluminium removal in Alzheimer's disease,<sup>9,18,19</sup> treatment of diabetes,<sup>20–23</sup> and contrast agents for medical imaging.<sup>24</sup> As such, the synthetic studies and activity analysis provided here should be a valuable reference for development of new hydroxypyrones and hydroxythiopyrones for a wide range of medicinal applications.

Previously, our laboratory has reported several potent and selective hydroxypyrrone MMPi with aryl backbones at the 2-position.<sup>15,16</sup> Thus, we first synthesized corresponding 2-backbone hydroxythiopyrones and evaluated their inhibition activities (Scheme 1).<sup>15,16</sup> Intermediate **1** was prepared according to a literature method.<sup>8</sup> A coupling reaction of compound **1** with 4-methoxybenzylamine in the presence of EDCI and HOBt provided amide **2** in 71% yield. It was found that coupling reagent 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide (EDCI) was preferable over dicyclohexylcarbodiimide (DCC), due to the difficulty in removing dicyclohexylurea (DCU) from reaction mixtures employing DCC. The benzyl protecting group on compound **2** was easily

removed by stirring in  $\text{CH}_3\text{CO}_2\text{H}/\text{HCl}$  (v/v 1:1) at room temperature overnight to give the previously reported hydroxypyrrone **AM-4** (referred to here as compound **3**) in 81% yield. The corresponding hydroxythiopyrrone **AM-4S** (**3-S**) was synthesized in 61% yield by thionation of compound **3** with  $\text{P}_4\text{S}_{10}$  in the presence of hexamethyldisiloxane (HMDO).<sup>25</sup> The methyl group on the aryl backbone of compound **3** was removed by reaction with  $\text{BBr}_3$  resulting in the new hydroxypyrrone **4** in 63% yield.

The inhibitory activities of synthesized hydroxypyrones and hydroxythiopyrones against MMPs were evaluated against human MMP catalytic domain using a fluorescent substrate assay.<sup>17</sup> MMP-1 (collagenase), MMP-2 (gelatinase A), and MMP-3 (stromelysin) are examples of MMPs with shallow, intermediate, and deep S1' pockets, respectively. For comparison of structural impact on the inhibitor activity, all inhibitors were evaluated at a concentration of 50  $\mu\text{M}$  against the aforementioned MMPs (Table 1).  $\text{IC}_{50}$  values were determined in cases where an inhibitor showed relatively strong inhibition against a given MMP.

Inhibition data comparison of 2-substituted thiopyrrone **3-S** and pyrrone **3** and **4** shows that hydroxythiopyrrone **3-S** led to higher inhibition than hydroxypyrones **3** and **4** for all MMPs tested at 50  $\mu\text{M}$  concentration. This outcome agrees with the earlier finding



Scheme 1. Synthetic scheme for 2-C backbone MMPi.

**Table 1**  
Percent inhibition at inhibitor concentrations of 50  $\mu$ M and select  $IC_{50}$  values (in parentheses)

Compound ID	Structure	MMP-1	MMP-2	MMP-3
3-S (AM-4S)		70 $\pm$ 1 (30)	94 $\pm$ 1 (19)	97 $\pm$ 1 (16.4 $\pm$ 2.3)
3 (AM-4)		29 $\pm$ 6	26 $\pm$ 2	93 $\pm$ 2 (2.4) <sup>a</sup>
4		66 $\pm$ 4	25 $\pm$ 4	94 $\pm$ 1 (3.7 $\pm$ 1.2)
AM-2S		–	–	(5.6) <sup>b</sup>
AM-2 <sup>a</sup>		(>50)	(9.3)	(0.24)
9a-S		62 $\pm$ 2 (44)	98 $\pm$ 1 (14.4)	91 $\pm$ 1 (13.3)
9a		12 $\pm$ 2	14 $\pm$ 3	44 $\pm$ 2
9b-S		31 $\pm$ 1	85 $\pm$ 2	91 $\pm$ 1 (13.9 $\pm$ 0.9)
9b		16 $\pm$ 4	11 $\pm$ 4	22 $\pm$ 4
14a-S <sup>c</sup>		52	94 (23)	45
14a		15 $\pm$ 4	21 $\pm$ 1	28 $\pm$ 1
14b-S		49 $\pm$ 5	69 $\pm$ 1 (30)	36 $\pm$ 1

Table 1 (continued)

Compound ID	Structure	MMP-1	MMP-2	MMP-3
14b		15 ± 1	33 ± 2	18 ± 1
15		17 ± 5	24 ± 5	32 ± 1
21		13 ± 1	18 ± 1	<1
22		18 ± 3	24 ± 4	32 ± 1
26		21 ± 4	15 ± 4	26 ± 2
30		6 ± 1	63 ± 1 (37.7 ± 1.2)	4 ± 1

<sup>a</sup> Data cited from Ref. 15.

<sup>b</sup> Data cited from Ref. 29.

<sup>c</sup> Data cited from Ref. 26.

that O,S donor ligands are more potent ZBGs than O,O ligands.<sup>13,14</sup> But it is interesting to note that thiopyrone **3-S** and **AM-2S** have poorer IC<sub>50</sub> values than corresponding pyrone **3** and **AM-2** (Table 1) against MMP-3.<sup>15,16</sup> X-ray structures of thiomaltol and maltol tris(pyrazolyl)borate zinc(II) complexes reveal that the hydroxythiopyrone and hydroxypyrene ligands coordinate to tris(pyrazolyl)borate zinc(II) in different geometries (Fig. 3),<sup>12</sup> suggesting that substituents at the same positions of thiopyrone and pyrone rings may interact with MMP active sites in different fashions. These findings promote us to investigate 5- and 6-substituted hydroxypyrones and hydroxythiopyrones.

The synthesis of inhibitors with backbones in the 6-C position was performed according to Scheme 2, resulting in compounds **9a**, **9b**, **9a-S**, and **9b-S**. The ring hydroxyl of kojic acid (**5**) was selectively protected with a benzyl group by treatment with BnBr in the presence of NaOH to give compound **6**. The hydroxymethyl group of **6** was oxidized to carboxylic acid **7** in 65% yield using Jones reagent. The aryl backbones were introduced by coupling reactions using EDCI/HOBt which afforded compounds **8a** and **8b** in

73–74% yield. The removal of the benzyl protecting groups from compound **8a** and **8b** was conducted in mixed-solvents HCl/CH<sub>3</sub>CO<sub>2</sub>H/CF<sub>3</sub>CO<sub>2</sub>H (v/v 10:10:1) at 60 °C, providing compounds **9a** and **9b** in 76–80% yield. In contrast to the benzyl group deprotection of compound **2** (Scheme 1), the benzyl group in **8a** and **8b** survived in CH<sub>3</sub>CO<sub>2</sub>H/HCl (v/v 1:1) at room temperature, but addition of CF<sub>3</sub>CO<sub>2</sub>H with increased temperature promoted its removal. This observation indicates that the substitution pattern on the pyrone can significantly change the electronic properties of the 3-benzyloxy group on the ring. Thionation of **9a** and **9b** by P<sub>4</sub>S<sub>10</sub> in the presence of HMDO provided **9a-S** and **9b-S** in 61–68% yield.<sup>25</sup>

Syntheses of 5-C backbone inhibitors **14a**, **14b**, **14a-S**, and **14b-S** followed a scheme recently developed in our laboratory (Scheme 3),<sup>26</sup> with some modifications. In the intramolecular condensation of activated bromo ester **10** to produce 3,3-diethoxy-pyran-4-one **11**, previously we used 2 equiv each of ethyl β-keto ester and NaH in order to make a homogeneous anion solution. Ethyl β-keto ester has an R<sub>f</sub> value similar to that of product **11** and is difficult to separate in large-scale preparations. In the improved procedure,

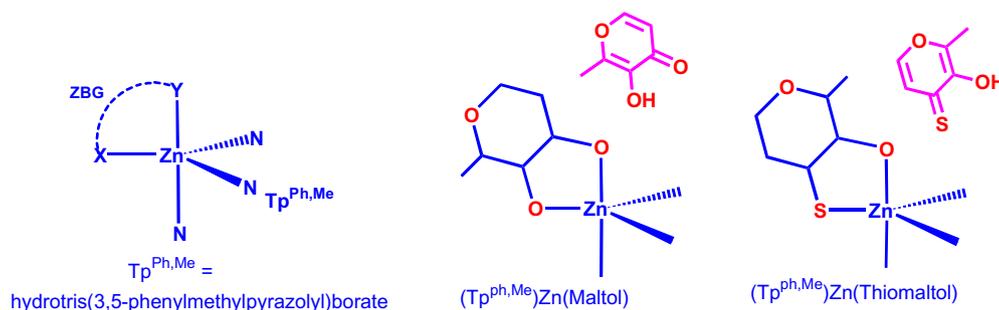
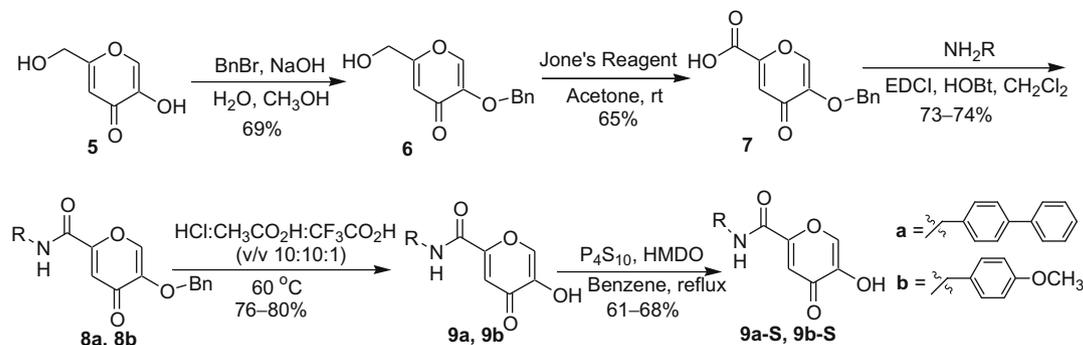


Figure 3. Geometries of maltol- and thiomaltol-tris(pyrazolyl)borate zinc(II) complexes.



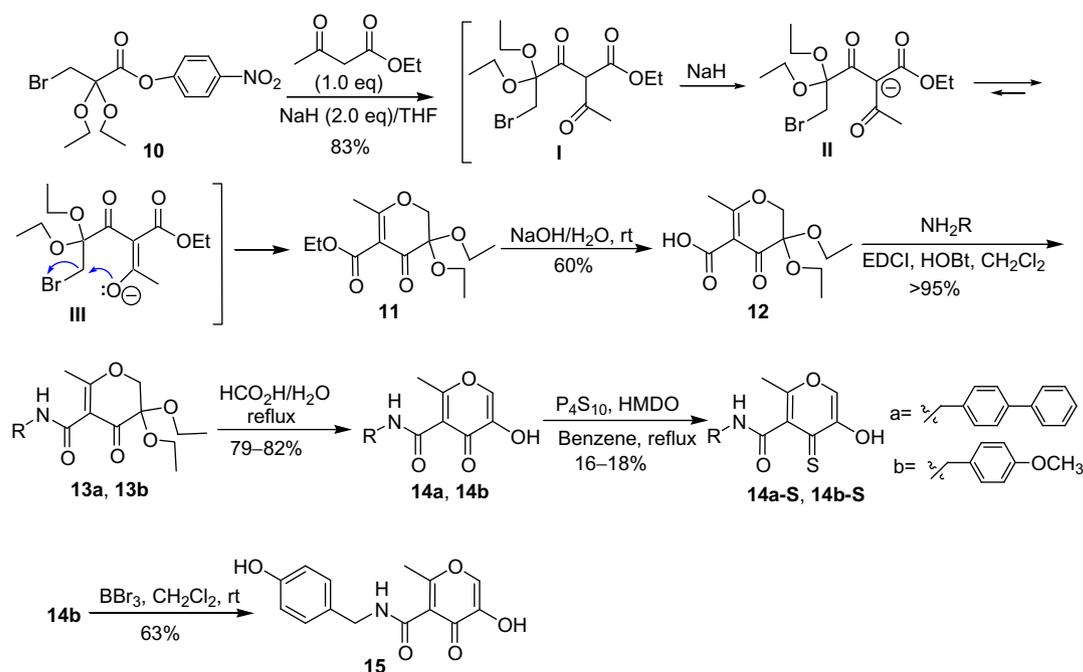
**Scheme 2.** Synthetic scheme for 6-C backbone MMPi.

1.0 equiv of ethyl  $\beta$ -keto ester and 2.0 equiv of NaH were employed. The reaction was first stirred at room temperature for 1 h to form intermediate **I** then refluxed to generate product **11**. In the preparation of amide **13a** and **13b**, EDCI/HOBT was employed as the coupling system instead of DCC/DMAP or DCC/NHS to avoid DCU contamination during product purification. After coupling of the backbone, the remaining synthetic procedures were performed as previously reported.<sup>26</sup> In contrast to the high yields (60–70%) in formation of the hydroxythiopyrones **3-S**, **9a-S**, and **9b-S** from the corresponding hydroxypyrones, thionation of **14a** and **14b** provided **14a-S** and **14b-S** in relatively low yields (16–18%).<sup>25</sup> The low yields in this transformation were tentatively explained by a steric effect at the keto position of **14a** and **14b** which arose from the neighboring 3-hydroxy and 5-amido groups. Demethylation of **14b** by  $\text{BBr}_3$  affords **15** in 63% yield.

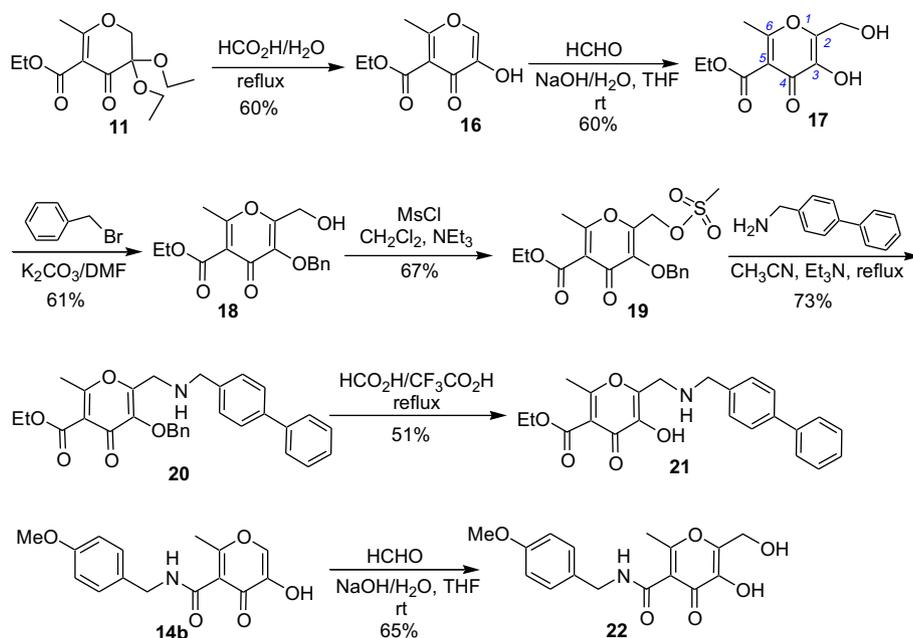
In vitro evaluation of 5- and 6-substituted inhibitors shows that hydroxythiopyrones such as **9a-S**, **9b-S**, **14a-S**, and **14b-S** were consistently more potent than their hydroxypyrene counterparts **9a**, **9b**, **14a**, and **14b** for all MMPs tested. For hydroxypyrene inhibitors, only inhibitors with backbones at the 2-position (e.g., **3**, **4**, and **AM-2**) were selective against MMP-3 over MMP-1 and MMP-2; and all 5- and 6-backbone hydroxypyrones **9a-b**, **14a-b**, and **15** were overall less potent for all MMPs and generally lacked isoform selectivity. The high potency and selectivity of these 2-C

backbone hydroxypyrene inhibitors for MMP-3 is consistent with the favorable orientation of the 2-substituent of the hydroxypyrene toward the MMP-3 S1' pocket.<sup>15</sup> The low inhibition of MMP-1 by the 2-C backbone inhibitors confirms the incompatibility between the bulky aryl backbones and the smaller S1' pocket in this MMP. **9a** showed higher inhibition against MMP-3 over other MMPs, while **9a-S** showed comparable inhibition for MMP-2 and MMP-3; compound **14a** was not potent and lacked selectivity for all MMPs, but **14a-S** showed some preference against MMP-2. These findings further highlight the interplay between the ZBG structure and the position of the backbone on the ZBG can be optimized to improve both potency and selectivity.

Most reported MMPi interact with MMPs at the primed side of the active site (i.e., right-handed inhibitors). Inhibitors with backbones designed to interact with both the primed and unprimed pockets, so-called 'double-handed' inhibitors, may have higher selectivity and potency (Fig. 1).<sup>3,27,28</sup> Hydroxypyrones and hydroxythiopyrones have several positions (2-, 5-, and 6-) for derivatization (Fig. 2), and thus are ideal for development of MMPi with multiple backbones. In Scheme 4, a structural extension at both sides of the hydroxypyrene chelator was achieved. Intermediate **16** was obtained in 60% yield by refluxing compound **11** in formic acid in the presence of water. A hydroxymethyl group was introduced at the 2-position of compound **16** by reaction with



**Scheme 3.** Synthetic scheme for 5-C backbone MMPi.



**Scheme 4.** Synthetic scheme for double-handed (2-C, 5-C) MMPi intermediates.

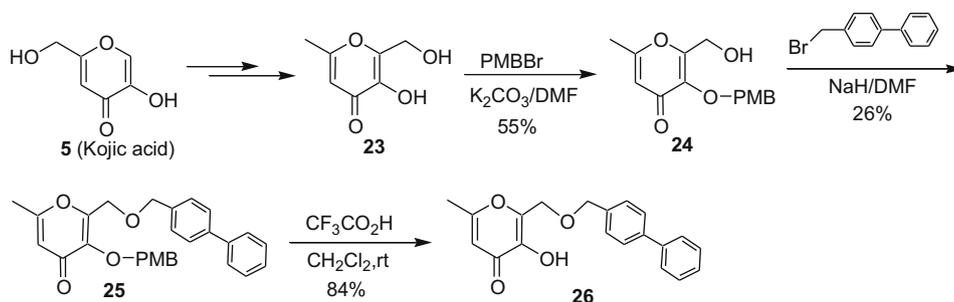
formaldehyde in the presence of NaOH aqueous solution. This hydroxymethylation reaction takes advantage of the ring hydroxyl of **16** resulting in compound **17** in 60% yield. Compared with the natural products kojic acid and maltol (Fig. 2), synthon **17** is highly versatile and provides synthetic handles for further functional extension at both sides of the hydroxypyronone chelator. The structure of **17** was unambiguously assigned by determining a single X-ray crystal structure of the compound complexed with iron (see Supplementary data). The ring hydroxyl was selectively benzylated using benzyl bromide in the presence of  $K_2CO_3$  providing intermediate **18** in 61% yield. Mesylation of **18** provided sulfonic acid ester **19** in 67% yield. The 4-biphenylmethyl backbone was introduced by refluxing mesylate **19** with 4-biphenylmethylamine in  $CH_3CN$  resulting in compound **20** in 73% yield. The benzyl group was removed by refluxing compound **20** in  $HCO_2H/CF_3CO_2H$  mixed solvents, which afforded compound **21** in 51% yield. As the 5-ester and 6-methyl groups provide attachment points for further functionalization of compound **21**, it is possible to synthesize novel hydroxypyronone inhibitors with multiple backbones to interact with MMP active sites. Similarly, compound **14b**, with an amido backbone at the 5-position, could also be hydroxymethylated at the 2-position generating compound **22** and thus providing a second route for introducing a new backbone at this position.

The simple double-handed intermediates **21** and **22** were not potent against the MMPs screened. Comparing the structure of **21** with potent inhibitor **AM-2**, which uses an amido linkage to at-

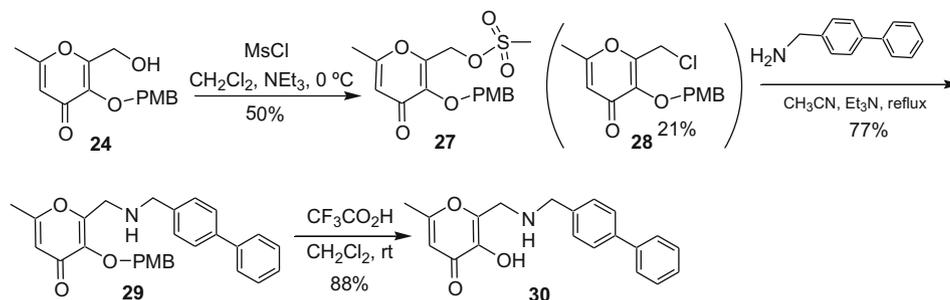
tach the backbone, we reason that the backbone linkage might be another important factor governing the interaction between an inhibitor and MMPs. Thus two analogous compounds of **AM-2**, compounds **26** and **30** with ether and amino linkages, respectively, were synthesized to investigate the impact of the linker on inhibitor potency.

The synthesis of compound **26** is shown in Scheme 5. Intermediate **23** was prepared from kojic acid (**5**) following a reported method.<sup>8</sup> The hydroxyl group on the pyrone ring was selectively protected with a PMB (*p*-methoxybenzyl) group using PMBBr to give compound **24**. The introduction of a 4-biphenylmethyl backbone was attained by treatment of compound **24** with 4-bromomethyl-biphenyl in the presence of NaH that led to the formation of compound **25**. The selective removal of the PMB group was achieved by treatment of compound **25** with  $CF_3CO_2H$  at room temperature in  $CH_2Cl_2$  providing inhibitor **26** in 84% yield. The use of a benzyl (BnBr) group instead of a PMB group to protect the pyrone hydroxyl group was also examined. It was found that the benzyl protecting group was not cleaved with  $CF_3CO_2H$  at room temperature. The benzyl group could be removed by refluxing in  $CH_3CO_2H/CF_3CO_2H$  (v/v 1:1) or hydrogenation in the presence of Pd/C, but under such conditions the 4-biphenylmethyl backbone was also partially removed, resulting in a low yield and difficulty in purifying product **26**.

The preparation of compound **30** is shown in Scheme 6. Treatment of intermediate **24** with methanesulfonyl chloride at 0 °C



**Scheme 5.** Synthetic scheme for compound **26**.



Scheme 6. Synthetic scheme for compound 30.

provided the mesylate ester **27** in 50% yield. The major side product in this transformation was the corresponding chloride **28** (21% yield). The 4-biphenylmethyl backbone was introduced by refluxing of **27** with 4-biphenylmethylamine in  $\text{CH}_3\text{CN}$  in the presence of triethylamine, resulting in compound **29**. Again, the PMB protecting group was easily removed by treatment with  $\text{CF}_3\text{CO}_2\text{H}$  at room temperature to give compound **30**. In the preparation of **30**, a benzyl protecting group was tested instead of the PMB protecting group, but again the 4-biphenylmethyl backbone was partially removed, as was found in the preparation of compound **26**.

The inhibition data for compounds **AM-2**, **26**, and **30** reveals the importance of the backbone linkage. The three inhibitors have the same ZBG and 4-biphenylmethyl backbone, but each has a different linker, namely amido (**AM-2**), ether (**26**), and amino (**30**). Compounds **26** and **30** are much less potent than **AM-2** against MMP-3, which may be partially explained by favorable hydrogen bonding between the amido carbonyl group of **AM-2** and the amino acid L164 in the MMP.<sup>15</sup> In addition, the ether linkage of **26** and the amino linkage of **30** are electron-donating, while the amido linkage of **AM-2** is electron-withdrawing. The differing electronic effects of linkers at the 2-position leads to significant differences in the  $\text{pK}_a$  values of the hydroxypyronone chelator,<sup>30</sup> with the amide compound being more acidic than either the ether- or amino-linked compound (unpublished results). These findings suggest that the linking group between the backbone and the ZBG can significantly influence the efficacy of an inhibitor through both direct (e.g., H-bonding) and inductive (e.g., ligand acidity) effects that must be carefully considered for successful MMPi design.

In summary, diverse arrays of hydroxypyronone and hydroxythiopyronone derivatives with different substitution patterns were synthesized and their inhibitory activity against three MMPs were examined. Our results suggest that hydroxypyronones and hydroxythiopyronones have different conformations with the same backbones leading to different selectivity and potency against MMPs. Our findings also show that an amide linkage between the chelator and the backbone is also essential for potent inhibition in this system. These synthetic approaches for manipulating hydroxypyronones and the structure–activity relationship information obtained from these MMPi should provide guidance for future design and optimization of potent and selective hydroxypyronone-based MMPi with multiple interactions with MMPs.

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## Supplementary data

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.bmcl.2009.02.044.

## References and notes

- Puerta, D. T.; Cohen, S. M. *Curr. Top. Med. Chem.* **2004**, *4*, 1551.
- Skiles, J. W.; Gonnella, N. C.; Jeng, A. Y. *Curr. Med. Chem.* **2004**, *11*, 2911.
- Whittaker, M. F.; Floyd, C. D.; Brown, P.; Gearing, A. J. H. *Chem. Rev.* **1999**, *99*, 2735.
- Matziari, M.; Dive, V.; Yiotakis, A. *Med. Res. Rev.* **2007**, *27*, 528.
- Ahmet, M. T.; Frampton, C. S.; Silver, J. *Dalton Trans.* **1988**, 1159.
- Ellis, B. L.; Duhme, A. K.; Hider, R. C.; Hossain, M. B.; Rizvi, S.; van der Helm, D. J. *Med. Chem.* **1996**, *39*, 3659.
- Liu, Z. D.; Hider, R. C. *Med. Res. Rev.* **2002**, *22*, 26.
- Liu, Z. D.; Piyamongkol, S.; Liu, D. Y.; Khodr, H. H.; Lu, S. L.; Hider, R. C. *Bioorg. Med. Chem.* **2001**, *9*, 563.
- Santos, M. A. *Coord. Chem. Rev.* **2002**, *228*, 187.
- Thompson, K. H.; Barta, C. A.; Orvig, C. *Chem. Soc. Rev.* **2006**, *35*, 545.
- Bentley, R. *Nat. Prod. Rep.* **2006**, *23*, 1046.
- Puerta, D. T.; Cohen, S. M. *Inorg. Chem.* **2003**, *42*, 3423.
- Puerta, D. T.; Griffin, M. O.; Lewis, J. A.; Romero-Perez, D.; Garcia, R.; Villarreal, F. J.; Cohen, S. M. *J. Biol. Inorg. Chem.* **2006**, *11*, 131.
- Puerta, D. T.; Lewis, J. A.; Cohen, S. M. *J. Am. Chem. Soc.* **2004**, *126*, 8388.
- Puerta, D. T.; Mongan, J.; Tran, B. L.; McCammon, J. A.; Cohen, S. M. *J. Am. Chem. Soc.* **2005**, *127*, 14148.
- Agrawal, A.; Romero-Perez, D.; Jacobsen, J. A.; Villarreal, F. J.; Cohen, S. M. *ChemMedChem* **2008**, *3*, 812.
- Knight, C. G.; Willenbrock, F.; Murphy, G. *FEBS Lett.* **1992**, *296*, 263.
- Finnegan, M. M.; Lutz, T. G.; Nelson, W. O.; Smith, A.; Orvig, C. *Inorg. Chem.* **1987**, *26*, 2171.
- Finnegan, M. M.; Rettig, S. J.; Orvig, C. *J. Am. Chem. Soc.* **1986**, *108*, 5033.
- McNeill, J. H.; Yuen, V. G.; Hoveyda, H. R.; Orvig, C. *J. Med. Chem.* **1992**, *35*, 1489.
- Saatchi, K.; Thompson, K. H.; Patrick, B. O.; Pink, M.; Yuen, V. G.; McNeill, J. H.; Orvig, C. *Inorg. Chem.* **2005**, *44*, 2689.
- Song, B.; Saatchi, K.; Rawji, G. H.; Orvig, C. *Inorg. Chim. Acta* **2002**, *339*, 393.
- Thompson, K. H.; Liboiron, B. D.; Sun, Y.; Bellman, K. D.; Setyawati, I. A.; Patrick, B. O.; Karunaratne, V.; Rawji, G.; Wheeler, J.; Sutton, K.; Bhanot, S.; Cassidy, C.; McNeill, J. H.; Yuen, V. G.; Orvig, C. *J. Biol. Inorg. Chem.* **2003**, *8*, 66.
- Puerta, D. T.; Botta, M.; Jocher, C. J.; Werner, E. J.; Avedano, S.; Raymond, K. N.; Cohen, S. M. *J. Am. Chem. Soc.* **2006**, *128*, 2222.
- Curphey, T. J. *J. Org. Chem.* **2002**, *67*, 6461.
- Yan, Y. L.; Cohen, S. M. *Org. Lett.* **2007**, *9*, 2517.
- Cuniassé, P.; Devel, L.; Makaritis, A.; Beau, F.; Georgiadis, D.; Matziari, A.; Yiotakis, A.; Dive, V. *Biochimie* **2005**, *87*, 393.
- Matziari, M.; Beau, F.; Cuniassé, P.; Dive, V.; Yiotakis, A. *J. Med. Chem.* **2004**, *47*, 325.
- Lewis, J. A.; Mongan, J.; McCammon, J. A.; Cohen, S. M. *ChemMedChem* **2006**, *1*, 694.
- Gordon, A. E. V.; Xu, J.; Raymond, K. N.; Durbin, P. *Chem. Rev.* **2003**, *103*, 4207.