



Contents lists available at ScienceDirect

Bioorganic & Medicinal Chemistry Letters

journal homepage: www.elsevier.com/locate/bmcl



Synthesis and properties of triazole-linked RNA

Daniel Mutisya^a, Chelliah Selvam^a, Scott D. Kennedy^b, Eriks Rozners^{a,*}

^a Department of Chemistry, Binghamton University, The State University of New York, Binghamton, NY 13902, USA

^b Department of Biochemistry and Biophysics, University of Rochester School of Medicine and Dentistry, Rochester, NY 14642, USA

ARTICLE INFO

Article history:

Received 1 March 2011

Revised 25 March 2011

Accepted 29 March 2011

Available online 5 April 2011

Keywords:

RNA

Modified nucleoside

Triazole internucleoside linkage

Huisgen dipolar cycloaddition

Click reaction

ABSTRACT

RNA oligonucleotides having triazole linkages between uridine and adenosine nucleosides have been prepared and studied using spectroscopic techniques. UV melting and CD showed that triazole strongly destabilized RNA duplex (7–14 °C per modification). NMR data suggested that, despite relative flexibility around the modified linkage, all base pairs were formed.

© 2011 Elsevier Ltd. All rights reserved.

RNA interference may become a promising alternative to conventional chemotherapy of genetic disorders, cancer, viral infections and other diseases for which the target mRNA can be identified.¹ However, for short interfering RNAs (siRNAs) to be efficient therapeutic agents they need to be chemically modified to optimize nuclease resistance, crossing of cell membranes, biodistribution and pharmacokinetics.² We are interested in developing RNA analogues that have the phosphate linkages replaced by nonionic and hydrophobic modifications, which we believe may improve the above properties of siRNAs. We have found that amide³ (3'-CH₂-CO-NH-5') and formacetal⁴ (3'-O-CH₂-O-5') linkages have surprisingly little effect on the global A-type structure, thermal stability and hydration of RNA and appear to be excellent mimics of phosphodiester linkages. Meanwhile, preparation of RNA bearing these modifications requires extensive synthetic effort, which hinders rapid testing and development of modified siRNAs. In a search for a nonionic modification that would be easy to synthesize both at the modified nucleoside and siRNA level, we became interested in triazole linkages that could be made via the Huisgen [3 + 2] dipolar cycloaddition (the 'click' reaction).^{5,6}

Von Matt et al.⁷ were the first to report synthesis and thermal stability of triazole-linked DNA, where the triazole was derived from 3'-azidothymidine. They found that the triazole modification of the DNA strand destabilized DNA–RNA heteroduplexes by about 1–2 °C per modification.^{7b} Recently, Isobe et al.⁸ synthesized triazole-linked dT₁₀, using Huisgen cycloaddition of 3'-azido and 5'-ethynyl thymidine. The triazole in Isobe's work was isomeric

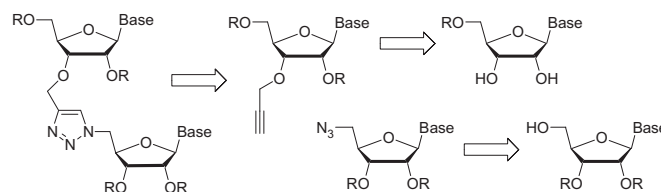


Figure 1. Retrosynthetic analysis of triazole-linked RNA.

to Von Matt's structure and led to an exceptionally stable duplex with complementary DNA.^{8a} Similar triazole linkage was later used by El-Sagheer and Brown to ligate synthetic DNA fragments and to amplify the resulting construct using PCR.⁹ Dondoni and co-workers¹⁰ also described preparation of triazole-linked DNA trinucleosides, but did not report on biophysical properties of these compounds.

Although triazole linkages prepared from 3'-azidothymidine have shown great promise in DNA, we expected that the presence of 2'-hydroxyl would complicate both preparation of 3'-azidouridine and synthesis of the corresponding triazole linked RNA. We envisioned that the easiest approach to triazole linked RNA would be a coupling of 3'-O-propargyl and 5'-azidoribonucleosides (Fig. 1).

Zerrouki and co-workers¹¹ incorporated the triazole linkage shown in Figure 1 in DNA but did not report on biophysical properties of the modified oligonucleotides. El-Sagheer and Brown used the 'click' reaction between 3'-O-propargyl-5-methyldeoxycytidine and 5'-azidouridine to incorporate a single triazole linkage at

* Corresponding author. Tel.: +1 607 777 2441; fax: +1 607 777 4478.

E-mail address: erozners@binghamton.edu (E. Rozners).

the active site of a hammerhead ribozyme.¹² The modified ribozyme was fully functional suggesting that the triazole linkage in **Figure 1** may be well tolerated in RNA despite being two atoms longer than the phosphate. Our own recent results, suggesting that RNA may tolerate hydrophobic nonionic modifications better than DNA,^{3,4} inspired us to test the properties of triazole-linked RNA. We found that the triazole linkage strongly destabilized RNA double helix by about 7–14 °C per modification depending on the sequence and location of the modifications. Although satisfactory melting curves could be recorded for all model systems, NMR and CD spectra suggested that the double helical structure is relatively flexible.

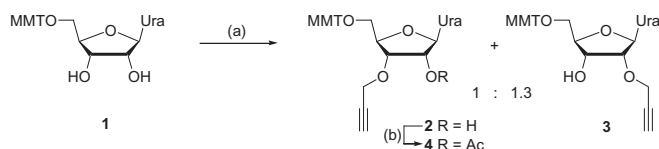
The key consideration in our design was minimizing the synthetic effort. While preparation of 5'-azidoribonucleosides was straightforward,¹³ synthesis of 3'-O-propargyl ribonucleosides involved alkylation of the 2',3'-cis-diol system, which we expected to proceed with low selectivity and require separation of isomeric products.¹⁴ However, we envisioned that the quick access of the alkyne coupling partner (only three to four steps from ribonucleosides) would offset the difficulties and low yields of the alkylation step.

Treatment of the known 5'-O-methoxytrityluridine **1** with dibutyltin oxide followed by propargyl bromide gave the expected mixture of 3'- and 2'-O-propargyluridine (**2** and **3**, in an approximate ratio of 1:1.3, respectively) from which the target compound was isolated in 28% yield (**Scheme 1**). To complete the synthesis of the alkyne coupling partner **4**, 2'-OH was protected with acetyl group.

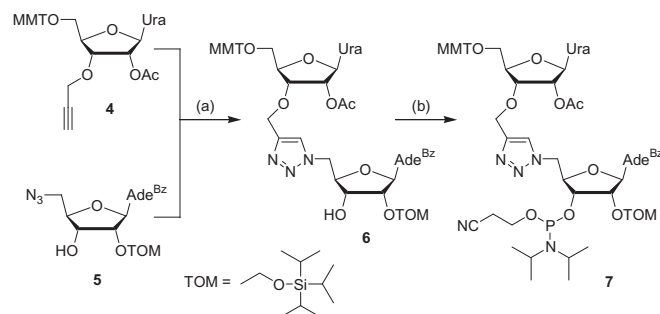
We recently developed synthesis of selectively protected 5'-azido-2'-O-TOM adenosine **5**,^{3a} which in the present study served well as the coupling partner for **4** in the Huisgen cycloaddition reaction to prepare the triazole linked r(U_{TR}A) dimer **6** (**Scheme 2**). The structure of the triazole linkage was confirmed using 2D NOESY NMR spectroscopy. In particular, the NOEs of adenosine H5', triazole aromatic H and 3'-OCH₂ of uridine confirmed the assigned structure of **6** (for details see **Supplementary data**, **Figs. S1 and S2**). Standard phosphoramidite synthesis gave dimer **7** suitable for solid phase synthesis (Expedite 8909) of RNA sequences bearing the triazole modification.

To evaluate the effect of triazole linkage on thermal stability of RNA we synthesized several self-complementary sequences (**Table 1**) that had performed well in our previous thermodynamic and structural studies.^{3a,4a} UV thermal melting (**Fig. S7**) was done as previously reported.^{3a,4a} Results in **Table 1** show that triazole linkage strongly destabilized RNA helices. The effect was more dramatic for **RNA3** and **RNA4** ($\Delta t_m > 10$ °C per modification) where the triazole linkages were placed opposite to each other in the double helix than for **RNA1** and **RNA2** (Δt_m ca. 7 °C per modification) where the triazoles were shifted relative to each other.

To gain insights into how the triazole linkage affected the structure of modified RNA we compared the circular dichroism (CD) spectra of **RNA1–RNA4** with the spectra of the corresponding unmodified RNAs (**Fig. 2**). **RNA2** gave the expected CD spectrum (**Fig. 2A**) similar to the unmodified RNA. Despite the similar t_m and melting curves (**Fig. S7**) **RNA1** gave almost no CD signal



Scheme 1. Propargylation of 5'-O-methoxytrityluridine. Steps: (a) Bu₄NBr, Bu₂SnO, DMF 100 °C, 2 h, then propargyl bromide, 45 °C, 24 h, 28%; (b) acetic anhydride, pyridine, rt, 5 h, 98%.



Scheme 2. Synthesis of triazole-linked r(U_{TR}A) phosphoramidite. Steps: (a) CuSO₄, sodium ascorbate, THF/ethanol/water, 60 °C, 12 h, 79%; (b) iPr₂NEt, CIP(OCH₂CH₂CN)N(iPr)₂, CH₂Cl₂, rt, 6 h, 47%.

Table 1
Thermal stability of triazole modified RNA

Entry	Sequence	t_m TR ^a	t_m RNA ^b	Δt_m
RNA1	GCGUAU _{TR} ACGC	39.5	53.7	−7.1
RNA2	GCGU _{TR} AUACGC	38.7	53.7	−7.5
RNA3	GUGU _{TR} ACAC	16.0	40.8	−12.4
RNA4	GCGU _{TR} ACGC	29.5	58.3	−14.4

^a t_m (°C) of triazole-modified RNA, TR denotes position of the triazole linkage.

^b t_m (°C) of unmodified RNA control.

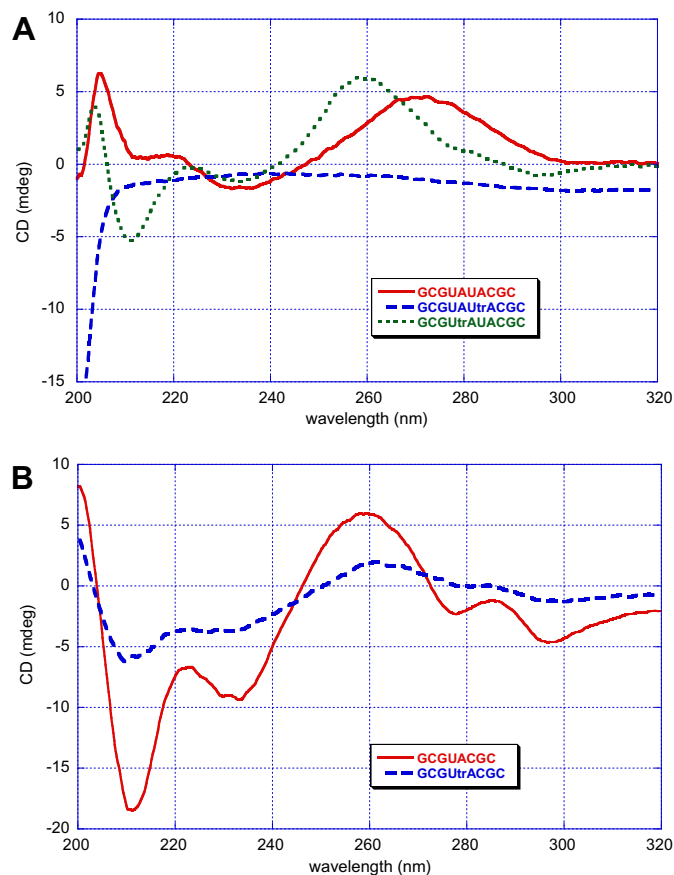


Figure 2. Comparison of CD spectra of triazole-linked RNAs (broken lines) and the unmodified RNA controls (solid lines). Recorded at **A** 20 °C, **B** 0 °C.

(**Fig. 2A**). A similar result was obtained with **RNA3** (**Fig. S8**). **RNA4** gave CD spectrum similar to but less intense than the unmodified control (**Fig. 2B**). Overall, the CD data suggested that

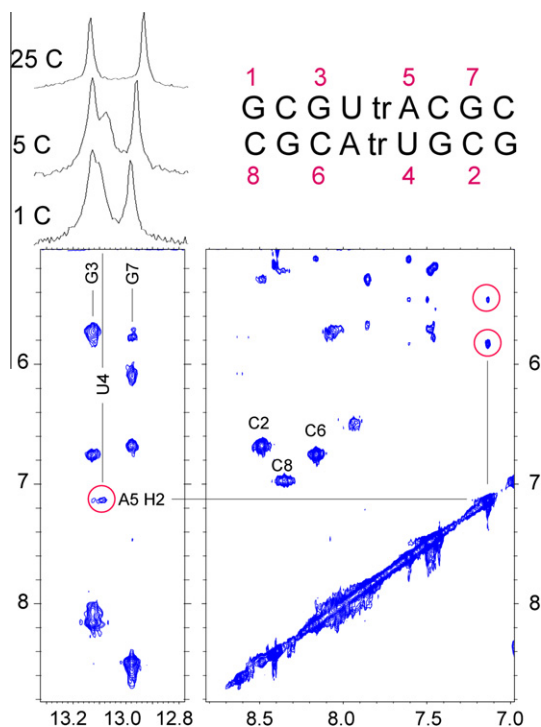


Figure 3. NMR spectra and 2D water-NOESY spectrum of **RNA4** duplex (0.1 mM). Upper left: 1D spectra of the imino proton region of the duplex (upper right with numbering) showing that U4 imino is only observed at low temperature. Bottom: 2D water-NOESY acquired at 1 °C with mixing time of 150 ms. Red circles indicate A5 H2 NOEs to U4 imino, C6 H1', and cross-strand A5 H1'. Cytosine intramolecular amino-amino cross-peaks typical in Watson-Crick GC pairs are labeled, including C8 in the terminal base pair for which no G imino is observed. Strong cross-peaks from G3 and G7 iminos to C6 and C2 aminos, respectively, indicate GC Watson-Crick pairs.

triazole modification made RNA duplexes more flexible and that **RNA1** and **RNA3** might be significantly disordered even at temperatures well below the t_m obtained by UV melting (20 and 0 °C, respectively).

To gain insight into base pair formation in the triazole modified RNA we studied **RNA4** using NMR techniques.¹⁵ Spectra in 95% H₂O/5% D₂O exhibit three clear imino proton signals (Fig. 3, top), one of which is clear only at low temperature where exchange of the imino proton with water is sufficiently slowed.

Strong NOEs to cytosine amino protons allow assignment of the two temperature-stable imino protons to G3 and G7 in GC base pairs. An NOE to the H2 proton of the lone adenine, despite the high rate of exchange with water, identifies the third imino peak as belonging to uracil in an AU base pair. Further evidence of this AU pair is contained in typical NOEs from adenine H2 to the cross-strand adenine H1' and to the intrastrand H1' of C6. Finally, the chemical shift of the adenine H2 proton (7.14 ppm) is nearly identical to that observed for the amide-modified and the unmodified RNA in our previous study (7.08 ppm).^{3a} While no imino proton signal was observed for the terminal guanosine (G1), formation of the closing GC base pair was suggested by the chemical shifts of C8 amino protons. One of these was shifted greater than 1 ppm downfield relative to the other, suggesting involvement in a hydrogen bond of one proton but not the other. This pattern, similar to that observed for C2 and C6 amino protons, is typical of Watson-Crick GC pairs. Thus, the NMR evidence indicates formation of a duplex that includes the central AU pair, although

that pair may be slightly distorted or dynamic allowing some water access. The relative flexibility of the central AU pair is consistent with reduced intensity of the CD signal for **RNA4** (Fig. 2B).

In summary, we found that the triazole 3'-O-CH₂-C=CH-N-5' linkage strongly destabilized RNA double helix (Δt_m -7 to -14 °C per modification) and is not a good mimic of the phosphate linkage in RNA. Our results on the triazole linkage that has four bonds (3'-O-CH₂-C=CH-N-5') instead of the two bonds long phosphate (3'-O-P(O)₂-O-5') linkage are in contrast to results reported by Isobe et al.⁸ who found that the two bonds long triazole (3'-N-CH=CH-5') linkage strongly stabilized DNA. The most likely explanation for these differences is that the significantly longer linkage adopts a conformation that is less than optimal for RNA double helix and causes the flexibility of the UA base pairs as suggested by the NMR and CD spectra of **RNA4**. Depending on the sequence the duplex may become disordered, as suggested by the lack of CD signal for **RNA1** and **RNA3**. The striking observation that modified duplexes with similar melting curves gave different CD spectra underlines the need for careful and comprehensive biophysical studies before conclusions can be made about the properties of a nucleic acid modification.

Acknowledgments

We thank National Institutes of Health (R01 GM071461) for financial support of this research and Dr. Pradeep S. Pallan (Vanderbilt University) for help with MALDI-TOF MS.

Supplementary data

Supplementary data (experimental procedures, copies of HPLC traces, UV melting curves, CD and NMR spectra) associated with this article can be found, in the online version, at doi:10.1016/j.bmcl.2011.03.111.

References and notes

- Watts, J. K.; Corey, D. R. *Bioorg. Med. Chem. Lett.* **2010**, *20*, 3203.
- (a) Watts, J. K.; Deleavey, G. F.; Damha, M. J. *Drug Discovery Today* **2008**, *13*, 842; (b) Corey, D. R. *J. Clin. Invest.* **2007**, *117*, 3615; (c) Rozners, E. *Curr. Org. Chem.* **2006**, *10*, 675; (d) Manoharan, M. *Curr. Opin. Chem. Biol.* **2004**, *8*, 570.
- (a) Selvam, C.; Thomas, S.; Abbott, J.; Kennedy, S. D.; Rozners, E. *Angew. Chem., Int. Ed.* **2011**, *50*, 2068; (b) Rozners, E.; Katkevica, D.; Bizdena, E.; Strömberg, R. *J. Am. Chem. Soc.* **2003**, *125*, 12125.
- (a) Kolarovic, A.; Schweizer, E.; Greene, E.; Gironde, M.; Pallan, P. S.; Egli, M.; Rozners, E. *J. Am. Chem. Soc.* **2009**, *131*, 14932; (b) Rozners, E.; Katkevica, D.; Strömberg, R. *Chem. Biol. Chem.* **2007**, *8*, 537; (c) Rozners, E.; Strömberg, R. *J. Org. Chem.* **1997**, *62*, 1846.
- (a) Rostovtsev, V. V.; Green, L. G.; Fokin, V. V.; Sharpless, K. B. *Angew. Chem., Int. Ed.* **2002**, *41*, 2596; (b) Tornøe, C. W.; Christensen, C.; Meldal, M. *J. Org. Chem.* **2002**, *67*, 3057.
- For a recent review, see: El-Sagheer, A. H.; Brown, T. *Chem. Soc. Rev.* **2010**, *39*, 1388.
- (a) Von Matt, P.; Lochmann, T.; Altmann, K.-H. *Bioorg. Med. Chem. Lett.* **1997**, *7*, 1549; (b) Von Matt, P.; Altmann, K.-H. *Bioorg. Med. Chem. Lett.* **1997**, *7*, 1553.
- (a) Isobe, H.; Fujino, T.; Yamazaki, N.; Guillot-Nieckowski, M.; Nakamura, E. *Org. Lett.* **2008**, *10*, 3729; (b) Fujino, T.; Yamazaki, N.; Isobe, H. *Tetrahedron Lett.* **2009**, *50*, 4101.
- El-Sagheer, A. H.; Brown, T. *J. Am. Chem. Soc.* **2009**, *131*, 3958.
- Nuzzi, A.; Massi, A.; Dondoni, A. *QSAR Comb. Sci.* **2007**, *1191*.
- (a) Lucas, R.; Zerrouki, R.; Granet, R.; Krausz, P.; Champavier, Y. *Tetrahedron* **2008**, *64*, 5467; (b) Lucas, R.; Neto, V.; Hadj Bouazza, A.; Zerrouki, R.; Granet, R.; Krausz, P.; Champavier, Y. *Tetrahedron Lett.* **2008**, *49*, 1004; (c) Lucas, R.; Elchinger, P. H.; Faugeras, P. A.; Zerrouki, R. *Nucleosides Nucleotides Nucleic Acids* **2010**, *29*, 168.
- El-Sagheer, A. H.; Brown, T. *Proc. Natl. Acad. Sci. U.S.A.* **2010**, *107*, 15329.
- Yamamoto, I.; Sekine, M.; Hata, T. *J. Chem. Soc., Perkin Trans. 1* **1980**, 306.
- Srivastava, S.C.; Raza, S.K. U.S. Patent US 5 744 595, 1998.
- Hammond, N. B.; Tolbert, B. S.; Kierzek, R.; Turner, D. H.; Kennedy, S. D. *Biochemistry* **2010**, *49*, 5817.