

## Journal Pre-proofs

### An Efficient Synthesis Tetrazole and Oxadiazole Analogues of Novel 2'-Deoxy-C-Nucleosides and their Antitumor Activity

Srishylam Penjarla, Subir Kumar Sabui, Dhande Sudhakar Reddy, Shyamapada Banerjee, Paidi Yella Reddy, Santhosh Penta, Yogesh S. Sanghvi

PII: S0960-894X(20)30723-X  
DOI: <https://doi.org/10.1016/j.bmcl.2020.127612>  
Reference: BMCL 127612



To appear in: *Bioorganic & Medicinal Chemistry Letters*

Received Date: 9 July 2020  
Revised Date: 7 September 2020  
Accepted Date: 9 October 2020

Please cite this article as: Penjarla, S., Kumar Sabui, S., Sudhakar Reddy, D., Banerjee, S., Yella Reddy, P., Penta, S., Sanghvi, Y.S., An Efficient Synthesis Tetrazole and Oxadiazole Analogues of Novel 2'-Deoxy-C-Nucleosides and their Antitumor Activity, *Bioorganic & Medicinal Chemistry Letters* (2020), doi: <https://doi.org/10.1016/j.bmcl.2020.127612>

This is a PDF file of an article that has undergone enhancements after acceptance, such as the addition of a cover page and metadata, and formatting for readability, but it is not yet the definitive version of record. This version will undergo additional copyediting, typesetting and review before it is published in its final form, but we are providing this version to give early visibility of the article. Please note that, during the production process, errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

## An Efficient Synthesis Tetrazole and Oxadiazole Analogues of Novel 2'-Deoxy-C-Nucleosides and their Antitumor Activity

Authors: Srishylam Penjarla<sup>a,b</sup>, Subir Kumar Sabui<sup>a</sup>, Dhande Sudhakar Reddy<sup>a</sup>, Shyamapada Banerjee<sup>a\*</sup>, Paidi Yella Reddy<sup>a</sup>, Santhosh Penta<sup>b\*</sup>, Yogesh S. Sanghvi<sup>a,c</sup>

Address: <sup>a</sup> Sapala Organics Pvt. Ltd, Plot Nos. 146B & 147 IDA Mallapur, Phase-II, Hyderabad-500076, Telangana, India; <sup>b</sup> Department of Chemistry, National Institute of Technology Raipur G.E. Road, Raipur, Chhattisgarh - 492010, India; <sup>c</sup> Rasayan Inc. 2802 Crystal Ridge Road, Encinitas, CA 92024-6615, U.S.A.

### Abstract:

Various tetrazole and oxadiazole C-nucleoside analogues were synthesized starting from pure  $\alpha$ - or  $\beta$ -glycosyl-cyanide. The synthesis of glycosyl-cyanide as key precursor was optimized on gram-scale to furnish crystalline starting material for the assembly of C-nucleosides. 1,2,4-Oxadiazole C-nucleosides were synthesized via two independent routes. First, both anomers of glycosyl-cyanide were transformed into tetrazole nucleosides followed by acylative rearrangement to furnish 1,2,4-oxadiazoles in high yields. Second, the glycosyl-cyanide was converted into an amidoxime which upon ring closure offered an alternative pathway for the assembly of 1,2,4-oxadiazoles in an efficient manner. These protocols offer an easy access to otherwise difficult to synthesize C-nucleosides in good yield and protecting group compatibility. These C-nucleosides were evaluated for their antitumor activity. This work paves a path for facile assembly of library of new chemical entities useful for drug discovery.

### Key Words:

C-Nucleoside, Cyano-sugar, Hoffer's Chloro-sugar, 1,2,4-oxadiazole, Anomers, Antitumor

### Introduction:

Unlike natural and synthetic N-nucleosides, C-nucleosides<sup>1,2</sup> are stable to enzymatic and acid-catalyzed hydrolysis of the glycosidic bond. Therefore, C-nucleosides offer a distinct advantage over the N-nucleosides for design of biologically active molecules. C-Nucleosides<sup>3,4,5</sup> have also attracted the interest of researchers looking for hydrogen-bond interactions alternative to those produced in the classical Watson-Crick model. Among well-known antitumor C-nucleosides, pyrazofurin,<sup>6</sup> showdomycin<sup>9</sup> and tiazofurin<sup>10</sup> are five-membered heterocyclic structures showing excellent biological activity (Figure- 1). Despite of their remarkable activity profile, lack of specificity and neurotoxicity prohibited the clinical progress of these nucleosides. More recently, remdesivir (GS-5734)<sup>7,8</sup> has shown promise for the treatment of COVID-19. Immucillins is yet another important class of C-nucleosides advancing into clinical trials as inhibitor of purine nucleoside phosphorylase. These observations have motivated us to revisit the study of C-nucleosides, with a particular interest of designing 2'-deoxyribose analogues of various five membered heterocycle and their antiviral and antitumor activity.

---

\* Corresponding author.

E-mail address: banerjee.s@sapalaorganics.com (S. Banerjee)

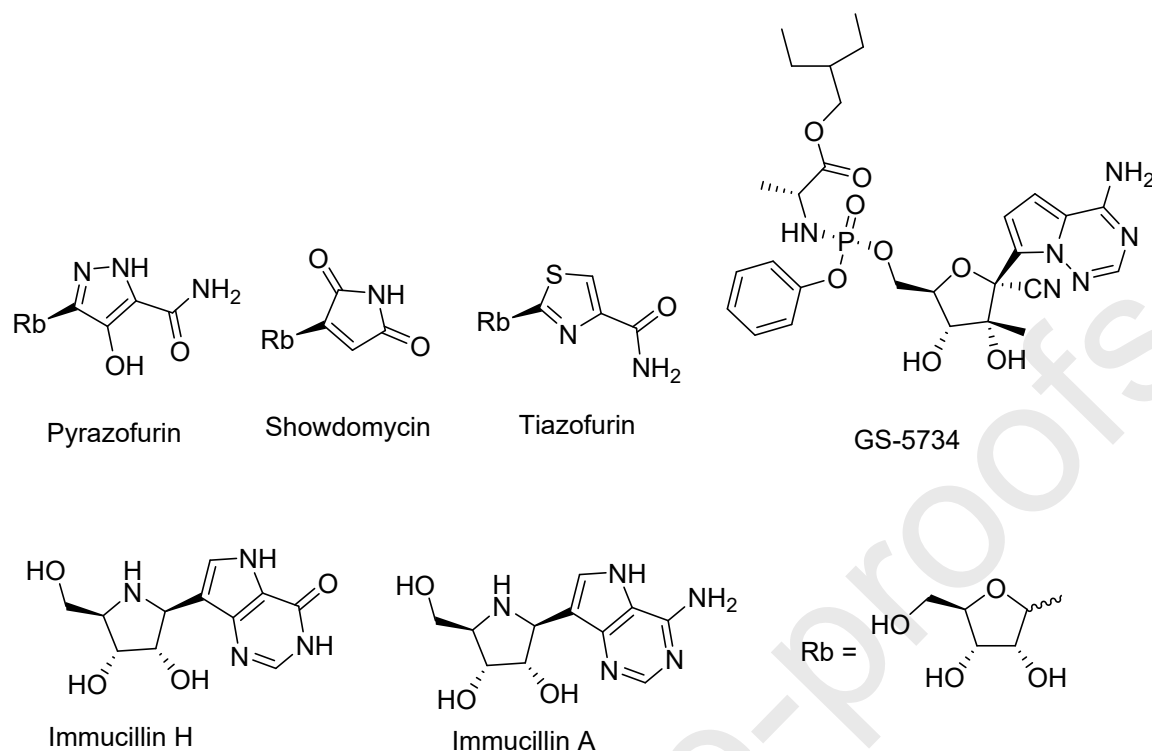


Figure -1

The presence of five-membered heterocyclic system is also a common feature in raltegravir,<sup>11</sup> an antiviral drug for treatment of HIV. Interestingly, the oxadiazole ring system is also present in ataluren<sup>12</sup> and zibotentan<sup>13</sup> used for the treatment of Duchenne muscular dystrophy (DMD) and prostate cancer, respectively (Figure -2). These facts and other reports on the promising biological activity of various regioisomeric oxadiazoles inspired us to synthesise and evaluate the biological activity of novel C-glycosides assembled from 2'-deoxyribose and oxadiazoles.

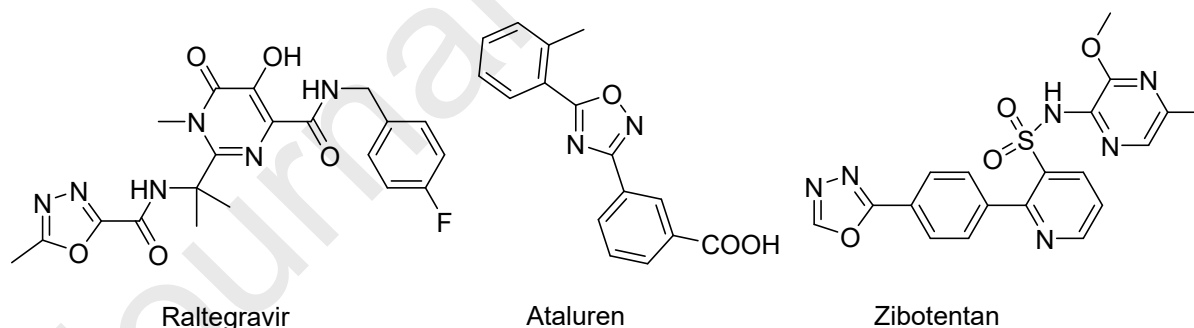


Figure-2

In particular, the oxadiazole ring<sup>14</sup> is an essential part of the pharmacophore favouring ligand binding, act as a flat aromatic linker to place substituents in the proper orientation and finally mimics as bioisoster of esters, amides, carbamates, and hydroxamic esters. The 1,2,3-oxadiazole ring is unstable. 1,2,4-Oxadiazole, 1,2,5-oxadiazole, and 1,3,4-oxadiazole are well known and appear in numerous marketed drugs. We have chosen 1,2,4-oxadiazole and 1,3,4-oxadiazole moiety for C-nucleoside synthesis. We envisioned the synthesis of a common building block by C-C bond formation at C1 and further integration of the heterocycle to assemble these C-nucleosides. We utilized the glycosyl cyanide as the key starting material which is obtained as mixture of  $\alpha/\beta$ -cyanide anomers from 1-chloro carbohydrate by reaction with trimethyl silyl cyanide in the presence of a Lewis acid as catalyst. These two anomers of glycosyl cyanide were transformed into novel C-nucleosides. The main advantage of this strategy resides in availability of stereochemically pure glycosyl cyanide that is transformed into C-nucleosides without anomerization at the C1 position. The C-nucleosides of both anomers of 2'-deoxyribose of tetrazoles and oxadiazoles synthesised are

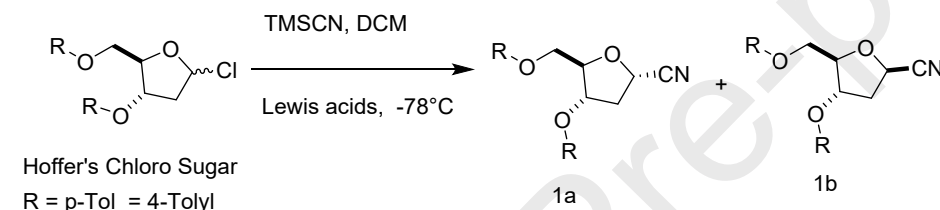
reported for the first time and the methodologies developed are general, which can be applied to construct other structurally diverse anomerically pure C-nucleosides.

### Result and Discussion:

Herein, we describe a modular synthesis approach allowing rapid assembly of C-nucleoside library in an efficient manner. Synthesis of a common building block by C-C bond formation at C1 and further integration of the heterocycle was the key feature of our approach. We have divided our study in three parts: (i) synthesis and separation of glycosyl cyanine anomers on multigram scale, (ii) synthesis of 1,2,4-oxadiazole C-nucleoside via amidoxime intermediate and (iii) synthesis of 1,3,4-oxadiazole via acylative rearrangement of tetrazole.

(i) Synthesis and separation of glycosyl cyanide anomers on multigram scale:

The glycosyl cyanide is one of the most important type of C-glycosyl intermediate, which is usually obtained as mixture of cyanide anomers from commercially available Hoffer's chloro sugar<sup>15</sup> by reaction with trimethyl silyl cyanide in the presence of a Lewis acid as catalyst (Scheme 1). Synthesis of 2'-deoxy glycosyl cyanine anomers (**1a** and **1b**) have been reported<sup>16</sup> only on small-scale. Since 2'-deoxy glycosyl cyanine anomers (**1a** and **1b**) are the key starting materials for our study, it was essential to optimize the yield and anomeric ratio with an ultimate objective of making it in hundred-gram quantity. Because chloro-sugar is devoid of neighbouring group participation, selective stereochemical outcome is challenging. Hence development of process which is robust and greener was undertaken. We screened various Lewis acids and solvents to improve yield and obtain better ratio of anomers in favour of  $\beta$ -selectivity.  $\beta$ -Anomer **1b** is desirable to produce C-nucleoside having resemblance to the naturally abundant 2'-deoxy-nucleosides.



Scheme-1: Synthesis of glycosyl cyanide

Entry	Lewis Acid	anomeric ratio ( $\beta/\alpha$ )	Isolated Yield (%)
1	SnCl <sub>4</sub>	3:1	93
2	BF <sub>3</sub> .OEt <sub>2</sub>	2:1	70
3	TMSOTf	0.59:1	60
4	CeCl <sub>3</sub>	No Reaction (Starting material remain intact)	-
5	ZnCl <sub>2</sub>	2:1	55
6	Mg (ClO <sub>4</sub> ) <sub>2</sub>	No Reaction (Insoluble in DCM)	-
7	FeCl <sub>3</sub>	5.7:1	70

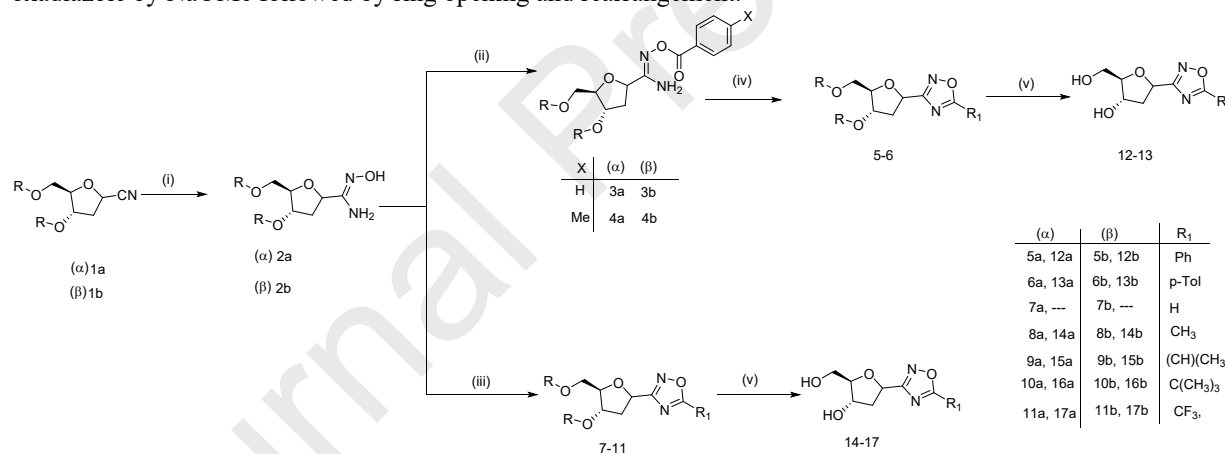
Table-1: Screening of Lewis acid for cyanation of Hoffer's chloro sugar in DCM at -78°C

Upon screening of various Lewis acids,  $\text{FeCl}_3$  afforded best  $\beta:\alpha = 5.7:1$  ratio in 70% yield (entry 7 Table 1). Next solvent screening using nitromethane, toluene, 1,2-dichloroethane, tetrahydrofuran, acetonitrile, 1,2-dimethoxyethane, acetone and dimethyl formamide failed to improve the  $\beta:\alpha$  ratio and yield compared to the reaction performed in dichloromethane. Considering desired  $\beta:\alpha$  ratio, yield (93%) and scalability,  $\text{SnCl}_4$  was chosen as preferred Lewis acid for the present study. It is important to note that low temperature is essential for  $\beta$ -selectivity and high yield. Further optimization effort is underway in our laboratory to find a robust process with non-toxic Lewis acid.

The two anomers of glycosyl cyanides were easily separated by silica gel column chromatography and anomeric configuration was established by  $^1\text{H}$  NMR experiments. The  $\text{SnCl}_4$  protocol (entry 1 Table 1) was scaled-up to furnish 366g of the pure  $\beta$ -anomer required for the transformation into various C-nucleosides containing five membered heterocycles. This route is the largest scale synthesis of glycosyl cyanide **1b** reported to date in high yield.

(ii) Synthesis of 1,2,4-oxadiazole C-nucleoside via amidoxime intermediate:

Nitrile functional group has served as an excellent handle to install several heterocyclic rings. Separately, both anomers of glycosyl cyanide (**1a** and **1b**) were converted into amidoxime (**2a** and **2b**) following Tiemann protocol<sup>17</sup> using hydroxylamine hydrochloride under basic condition. Reaction was performed with  $\text{NH}_2\text{OH}.\text{HCl}$  in presence of Hünig's base, instead of using  $\text{NH}_2\text{OH}.\text{HCl}$  and  $\text{Na}_2\text{CO}_3$  was reported by Adelfinskaya et al.<sup>18</sup> Excellent yields were obtained for both anomers. These amidoxime derivatives were then converted into 1,2,4-oxadiazoles derivatives (**5-6** and **7-11**) following two distinct protocols (Scheme 2). First, sequential synthesis of *O*-acylated amidoximes using acetyl chloride followed by cyclization to 1,2,4-oxadiazole ring (**5-6**) using alkaline DMSO solution. Whereas the second protocol involve direct cyclization of amidoximes<sup>19</sup> to 1,2,4-oxadiazoles<sup>20-27</sup> using orthoformate or acid anhydride in presence of  $\text{BF}_3.\text{Et}_2\text{O}$  as Lewis acid. The later protocol is shorter and offered higher yields compared to the first route. Deprotection of *p*-tolyl group was accomplished using  $\text{NaOMe}$  in methanol at room temperature in excellent yield except for **7a** and **7b**. Multiple product formation was observed on TLC for **7a** and **7b**. This phenomenon of multiple product formation can be attributed to the deprotonation<sup>28,29</sup> of acidic C5-H of 1,2,4-oxadiazole by  $\text{NaOMe}$  followed by ring opening and rearrangement.

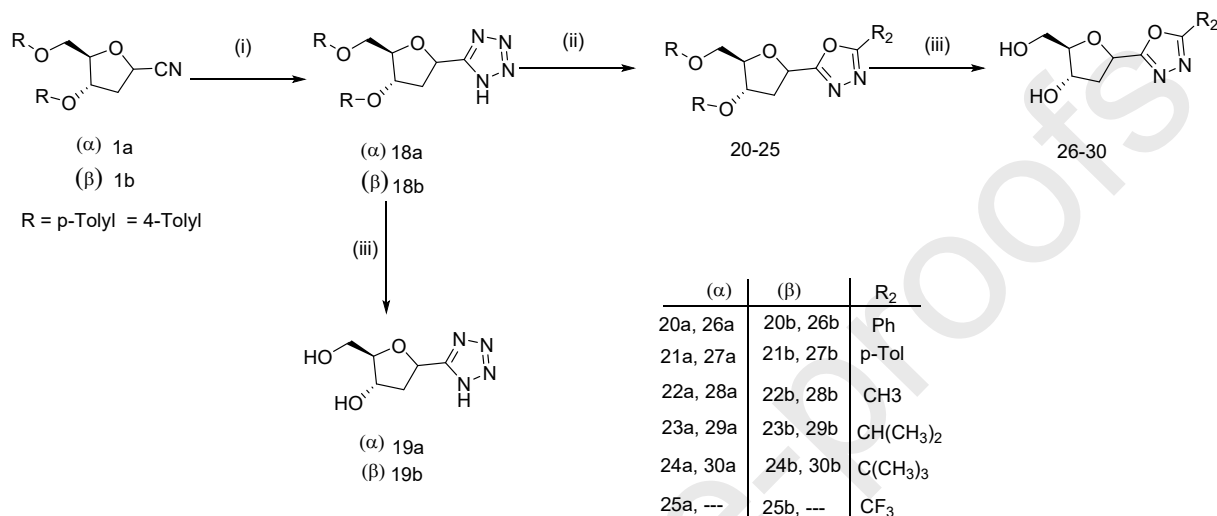


Scheme-2 (Reagents and Conditions): (i)  $\text{NH}_2\text{OH}.\text{HCl}$ , DIEA, EtOH, reflux, 1h (92-96%); (ii) Acyl chloride, 1,4-dioxane, RT, 16 h (73-97%); (iii) Trimethyl orthoformate,  $\text{BF}_3.\text{Et}_2\text{O}$  110°C, 3 h (or) Acid anhydride,  $\text{BF}_3.\text{Et}_2\text{O}$ , 110°C (or) Trifluoroacetic anhydride, DCM, RT, 5 h (80-98%); (iv) KOH, DMSO, RT, 6 h (72-98%); (v)  $\text{NaOMe}$ , DCM: MeOH (3:2), RT, 16h (48-97%).

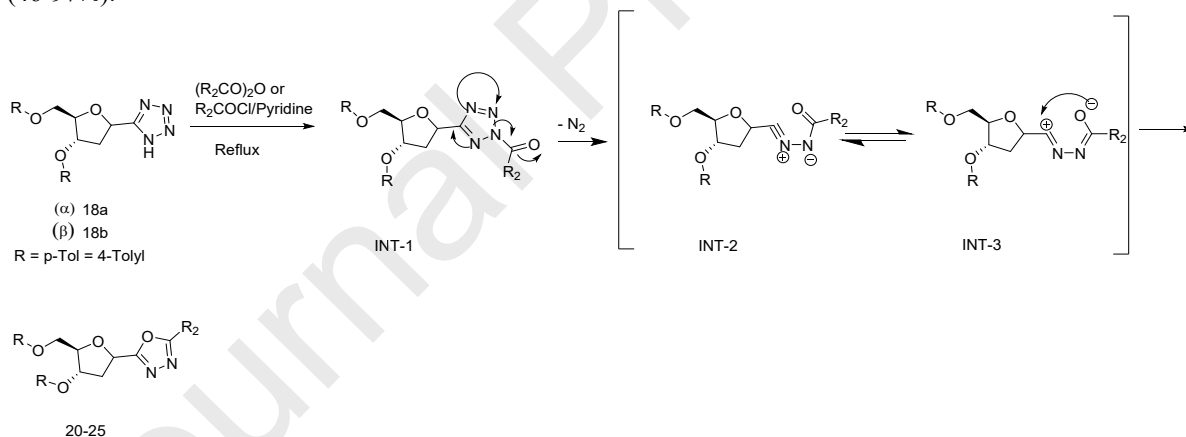
(iii) Synthesis of 1,3,4-oxadiazole via acylative rearrangement of tetrazole:

In 1994, Kobe et al.<sup>30</sup> synthesised 5- $\beta$ -D-ribofuranosyl-1*H*-tetrazole from allononitrile using  $\text{NaN}_3$  and  $\text{AlCl}_3$  in excellent yield. Our attempt to utilize same protocol starting with glycosyl cyanide **1a** or **1b** resulted in low yield of tetrazole derivative along with other unidentified products. Therefore, tetrazole derivatives<sup>31-33</sup> (**18a** and **18b**) were successfully synthesized in good yield from both anomers of glycosyl cyanide (**1a** and **1b**) using azide click reaction with copper and cupric sulphate in DMF at 120°C. Unprotected tetrazole nucleosides (**19a** and **19b**) were obtained by cleaving tolyl protecting group using sodium methoxide in methanol at room temperature (Scheme 3). The conversion of tetrazole to 1,3,4-oxadiazole derivatives<sup>34-38</sup> was achieved either by reacting tetrazole derivatives with carboxylic acid anhydride in presence of hydroquinone under reflux or by reacting with carboxylic acid chloride in pyridine.<sup>39-42</sup> The Deprotection of *p*-tolyl group was executed using  $\text{NaOMe}$  in methanol at room temperature in excellent yield. However, the deprotection protocol suffers from a drawback for C5-unsubstituted and C5-substitution with electron

withdrawing groups. In both cases, multiple product formation was observed due to the ring opening of the oxadiazole ring. This decomposition can be explained by the nucleophilic addition of NaOMe to C5-carbon and ring opening.<sup>43</sup> The postulated mechanism<sup>44</sup> of this conversion is illustrated in Scheme-4. 5-Substituted tetrazole undergoes N2-acylation upon treatment with acylation reagent due to steric bulk of 5-substitution. This unstable intermediate (INT-1) then ring opens via nitrogen extrusion and formation of *N*-acyl nitrilimine as putative intermediates (INT-2 and INT-3). These intermediates are then cyclized to form 1, 3, 4-oxadiazoles (**20-25**) in good yield. Structural elucidation of the new compounds described in this study was based on NMR and mass spectral data.



Scheme-3 (Reagents and Conditions): (i) NaN<sub>3</sub>, Cu, CuSO<sub>4</sub>, DMF, 120°C, 16 h (87-90%); (ii) (R<sub>2</sub>CO)<sub>2</sub>O, hydroquinone, reflux, 1 h (or) R<sub>2</sub>COCl, Pyridine, 90°C, 2 h (78-97%); (iii) NaOMe, DCM : MeOH (3:2), RT, 16 h (46-97%).



Scheme-4: Plausible reaction mechanism for 2-substituted 1,3,4-oxadiazole ring formation from **18a** or **18b**

### Biological Activity:

We tested a set of 12 C-nucleosides both α- and β-anomers for their in-vitro cytotoxicity activity in five tumor cell lines,<sup>45</sup> namely HeLa, MDA-MB-231 (breast cancer), PANC-1 (pancreatic cancer), PC3 (prostate cancer) and SK-OV-3 (ovarian cancer) using the MTT (3-(4,5-dimethyl thiazol-2-yl)-2,5-diphenyltetrazolium bromide) assay. Doxorubicin was used as a positive control to validate the MTT assay<sup>46</sup>. Majority of the compounds failed to exhibit significant cytotoxicity against five cell lines tested at 10 μmol concentration. The results are summarized in the graph shown below. Compounds **15b** and **28a** exhibited modest (9-11%) inhibition of breast cancer cell line MDA-MB-231. Whereas in case of SK-OV-3, we observed 10-14% inhibition exerted by five C-nucleosides (**19b**, **17b**, **12b**, **14b** & **14a**) (Figure 3).

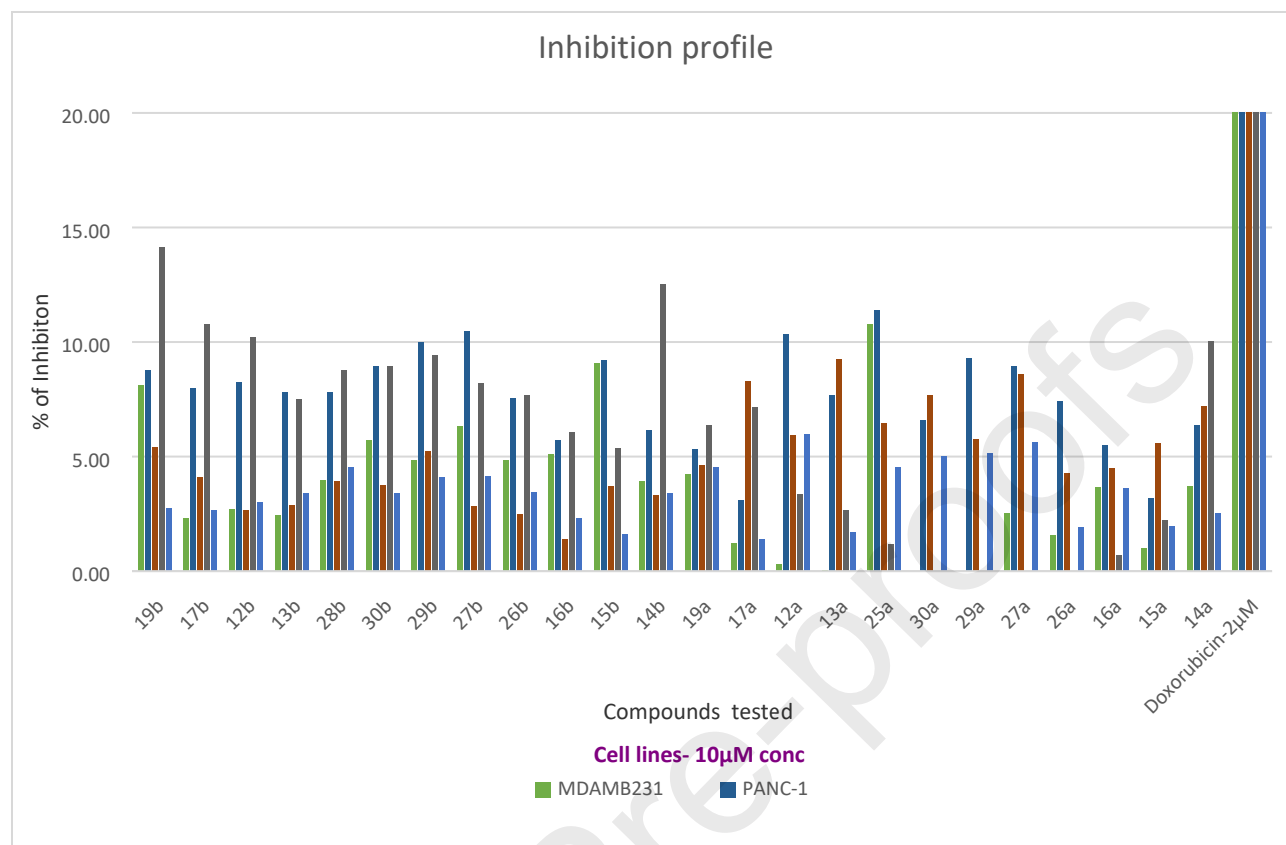


Figure 3: Inhibition profile for 12 C-nucleosides against 5 cell lines. Doxorubicin was used as a control which exhibited >90% inhibition at 2µM.

#### Summary:

Various regio-isomeric five-membered oxadiazoles based 2'-deoxy-C-nucleosides were synthesized for the first time in good yield and high purity. All C-nucleosides were assembled from pure  $\alpha$ - or  $\beta$ -anomer of glycosyl cyanide. The synthesis of glycosyl cyanide as key starting material was established on large-scale and in excellent yield. The easy accessibility of glycosyl cyanide further allows its utility in design of therapeutic oligonucleotides.<sup>47</sup> The synthetic methodologies developed in this study are general and offer future scope to generate other nucleoside analogues for SAR study. Biological evaluation was carried out for synthesised compounds and shows reasonable cytotoxicity in five different tumor cell lines. Studies on antiviral activity of these compounds is in progress and it will be published elsewhere.

#### Declaration of Competing Interest:

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

#### Acknowledgement:

SS acknowledge the help and support of Director, NIT Raipur for allowing him to register and pursue his PhD. SS also thanks Mr. Madika Chandrakanth for his help in preparation of manuscript.

#### Supplementary Data:

Experimental data and product characterization is available as SI material.

#### References:

1. C-Nucleosides: Synthetic Strategies and Biological Applications, Jan Štambaský, Michal Hocek and Pavel Kočovský *Chem. Rev.*, **2009**, 109, 6729–6764, DOI: 10.1021/cr9002165
2. C-Nucleosides To Be Revisited, Erik De Clercq, *J. Med. Chem.*, **2016**, 59 (6), 2301–2311. DOI: 10.1021/acs.jmedchem.5b01157



3. Synthetic Methodologies for C-Nucleosides, Qinpei Wu, Claire Simons, *Synthesis*, 2004, 10, 1533–1553, DOI: 10.1055/s-2004-829106
4. Recent advances in synthetic approaches for medicinal chemistry of C-nucleosides, Kartik Temburnikar and Katherine L. Seley-Radtke, *Beilstein JOC* **2018**, 14, 772-785, DOI:10.3762/bjoc.14.65
5. Modular synthesis of new C-aryl-nucleosides and their anti-CML activity, Hamid Marzag, Marwa Zerhouni, Hamza Tachallait, Luc Demange, Guillaume Robert, Khalid Bougrin, Patrick Auberger, Rachid Benhida, *Bioorganic and Medicinal Chemistry Letter*, **2018**, 28, 1931-1936., DOI: 10.1016/j.bmcl.2018.03.063
6. Biochemistry and biological effects of the pyrazofurins (pyrazomycins): initial clinical trial. G E Gutowski, M J Sweeney, D C DeLong, R L Hamill, K Gerzon, R W Dyke. *Ann. N. Y. Acad. Sci.* **1975**, 255, 544-551, DOI: 10.1111/j.1749-6632.1975.tb29257.x
7. Discovery of the first C-nucleoside HCV polymerase inhibitor (GS-6620) with demonstrated antiviral response in HCV infected patients. Cho, A.; Zhang, L.; Xu, J.; Lee, R.; Butler, T.; Metobo, S.; Aktoudianakis, V.; Lew, W.; Ye, H.; Clarke, M.; Doerffler, E.; Byun, D.; Wang, T.; Babusis, D.; Carey, A. C.; German, P.; Sauer, D.; Zhong, W.; Rossi, S.; Fenaux, M.; McHutchison, J. G.; Perry, J.; Feng, J.; Ray, A. S.; Kim, C. U *J. Med. Chem.* **2014**, 57, 1812–1825, DOI: 10.1021/jm400201a
8. Current knowledge about the antivirals remdesivir (GS-5734) and GS441524 as therapeutic options for coronaviruses, E. Susan Amiriana, Julie K. Levy, *One Health*, 2020, 9, 100128, DOI: 10.1016/j.onehlt.2020.100128
9. Showdomycin, A New Nucleoside Antibiotic, S. Roy-Burman, P. Roy-Burman and D. W. Visser, *Cancer Research*, **1968**, 28, 1605-1610
10. Tiazofurin: A new antitumor agent, Peter J. O'Dwyer, D. Dale Shoemaker, Hiremagalur N. Jayaram, David G. Johns, David A. Cooney, Silvia Marsoni, Louis Malspeis, Jacqueline Plowman, J. Paul Davignon and Ruth D. Davis, *Investigational New Drugs*, **1984**, 2, 79-84, DOI: 10.1007/BF00173791
11. Raltegravir, Croxtall JD and Keam SJ *Drugs*, 2009, 69(8), 1059-1075. DOI:10.2165/00003495-200969080-00007
12. Ataluren stimulates ribosomal selection of near-cognate tRNAs to promote nonsense suppression. Roy B, Friesen, WJ, Tomizawa Y, Leszyk JD, Zhuo J, Johnson B, Dakka J, Trotta CR, Xue X, Mutyam V, Keeling KM, Mobley JA, Rowe SM, Bedwell DM, Welch EM, Jacobson A *Proceedings of the National Academy of Sciences of the United States of America*. **2016**, 113 (44): 12508–12513. DOI: 10.1073/pnas.1605336113
13. Phase III, randomized, placebo-controlled study of once-daily oral zibotentan (ZD4054) in patients with non-metastatic castration-resistant prostate cancer, K Miller, JW Moul, M Gleave, K Fizazi, JB Nelson, T Morris, FE Nathan, S McIntosh, K Pemberton and CS Higano, *Prostate Cancer and Prostatic Disease* **2013**, 16, 187–192, DOI:10.1038/pcan.2013.2
14. (a) Oxadiazoles in Medicinal Chemistry, Jonas Boström, Anders Hogner, Antonio Llinàs, Eric Wellner, and Alleyn T. Plowright, *J. Med. Chem.* **2012**, 55, 1817–1830, DOI: 10.1021/jm2013248 (b) C-(2-Deoxy-D-arabino-hex-1-enopyranosyl)-oxadiazoles: synthesis of possible isomers and their evaluation as glycogen phosphorylase inhibitors, Eva Bokor, Eszter Szennyés, Tibor Csúpsz, Nora Toth, Tibor Docsa, Pal Gergely, Laszlo Somsak, *Carbohydrate Research* **2015**, 412, 71-79. DOI: 10.1016/j.carres.2015.04.016
15.  $\alpha$ -Thymidin, Hoffer M, *Chem. Ber.* **1960**, 93, 2777-2781. DOI: 10.1002/cber.19600931204
16. Oligonucleotide labelling methods. 4. Direct labelling reagents with a novel, non-nucleoside, chirally defined 2-deoxy- $\beta$ -D-ribose backbone, Smith, Thomas H.; Kent, Mark A.; Muthini, Sylvester; Boone, Steven J.; Nelson, Paul S. *Nucleosides & Nucleotides*, **1996**, 15(10), 1581-1594, DOI: 10.1080/07328319608002458



17. Effect of Hydroxylamine on Nitriles. Tiemann, T. *Chem. Ber.* **1884**, 17, 126–129 DOI: 10.1002/cber.18840170230
18. Synthesis and structural analysis of oxadiazole carboxamide deoxyribonucleoside analogs, Olga Adelfinskaya, Weidong Wu, V. Jo Davisson, and Donald E. Bergstrom, *Nucleosides, Nucleotides, and Nucleic Acids*, **2005**, 24:1919–1945, DOI: 10.1080/15257770500269267
19. Ethionamide Boosters. 2. Combining Bioisosteric Replacement and Structure-Based Drug Design To Solve Pharmacokinetic Issues in a Series of Potent 1,2,4-Oxadiazole EthR Inhibitors, Marion Flipo, Matthieu Desroses, Nathalie Lecat-Guillet, Baptiste Villemagne, Nicolas Blondiaux, Florence Leroux, Catherine Piveteau, Vanessa Mathys, Marie-Pierre Flament, Juergen Siepmann, Vincent Villeret, Alexandre Wohlkönig, René Wintjens, Sameh H. Soror, Thierry Christophe, Hee Kyoung Jeon, Camille Loch, Priscille Brodin, Benoit Déprez, Alain R. Baulard and Nicolas Willand, *J. Med. Chem.* **2012**, 55, 1, 68–83, DOI: 10.1021/jm200825u
20. Synthesis and characterization of novel bioactive 1,2,4-oxadiazole natural product analogs bearing the N-phenylmaleimide and N-phenylsuccinimide moieties, Catalin V. Maftai, Elena Fodor, Peter G. Jones, M. Heiko Franz, Gerhard Kelter, Heiner Fiebig and Ion Neda, *Beilstein J. Org. Chem.* **2013**, 9, 2202–2215. DOI:10.3762/bjoc.9.259
21. (a) 1,2,4-oxadiazole nucleus with versatile biological applications, Mohammad Arshad, Taqi Ahmed Khan, Meraj Alam Khan, *International Journal of Pharma Sciences and Research*, 2014, 5(7), 303–316 (b) Synthesis and Antiviral Activity of 3-( $\beta$ -D-Ribofuranosyl)-1,2,4-oxadiazole -5-carboxamide Ram Pratap and V. N. Yarovenko, *Nucleosides, Nucleotides and Nucleic Acids*, **2000**, 19:5–6, 845–849, DOI: 10.1080/15257770008033026 (c) A novel synthesis of 1,2,4-oxadiazoles and isoxazoles, Arif Kivrak, Metin Zora, *Tetrahedron*, **2014**, 70, 817–831, DOI: 10.1016/j.tet.2013.12.043, (d) The new era of 1,2,4-oxadiazoles, Andrea Pace and Paola Pierro, *Org. Biomol. Chem.*, **2009**, 7, 4337–4348, DOI:10.1039/B908937C
22. A Straightforward and High-Yielding Synthesis of 1,2,4-Oxadiazoles from Chiral N-Protected  $\alpha$ -Amino Acids and Amidoximes in Acetone-Water: An Eco-Friendly Approach, Andre C. Sauer, Lucas Wolf, Natalia Quoos, Mariele B. Rodrigues, Antonio L. Braga, Oscar E. D. Rodrigues, and Luciano Dornelles, *Journal of Chemistry*, Volume **2019**, Article ID 8589325, 9 pages. DOI: 10.1155/2019/8589325
23. In Search of Glycogen Phosphorylase Inhibitors: 5-Substituted 3-C-Glucopyranosyl- 1,2,4-oxadiazoles from  $\beta$ -D-Glucopyranosyl Cyanides upon Cyclization of O-Acylamidoxime Intermediates, Mahmoud Benlifa, Sébastien Vidal, Bernard Fenet, Moncef Msaddek, Peter G. Goekjian, Jean-Pierre Praly, Attila Brunyánszki, Tibor Docsa, and Pál Gergely, *Eur. J. Org. Chem.* **2006**, 4242–4256. DOI: 10.1002/ejoc.200600073
24. A new one-pot synthesis of 1,2,4-oxadiazoles from aryl nitriles, hydroxylamine and crotonoyl chloride, Masoumeh Zakeri, Majid M Heravi and Ebrahim Abouzari-Loft, *J. Chem. Sci.* **2013**, 125, (4), 731–735 DOI: 10.1007/s12039-013-0426-6
25. PTSA-ZnCl<sub>2</sub>: An Efficient Catalyst for the Synthesis of 1,2,4-Oxadiazoles from Amidoximes and Organic Nitriles, John Kallikat Augustine, Vani Akabote, Shrivats Ganapati Hegde, and Padma Alagarsamy, *J. Org. Chem.* **2009**, 74, 5640–5643, DOI: 10.1021/jo900818h
26. 3-Glucosylated 5-amino-1,2,4-oxadiazoles: synthesis and evaluation as glycogen phosphorylase inhibitors, Marion Donnier-Maréchal, David Goyard, Vincent Folliard, Tibor Docsa, Pal Gergely, Jean-Pierre Praly and Sébastien Vidal, *Beilstein J. Org. Chem.* **2015**, 11, 499–503, DOI:10.3762/bjoc.11.56
27. Synthesis and structure–activity relationships of C-glycosylated oxadiazoles as inhibitors of glycogen phosphorylase, Marietta Tóth, Sándor Kun, Éva Bokor, Mahmoud Benlifa, Gaylord Tallec, Sébastien Vidal, Tibor Docsa, Pál Gergely, László Somsák, Jean-Pierre Praly, *Bioorganic & Medicinal Chemistry* **2009**, 17, 4773–4785, DOI:10.1016/j.bmc.2009.04.036

28. Rearrangements of 1,2,4-oxadiazole: "one ring to rule them all", Antonio Palumbo Piccionello, Andrea Pace, Silvestre Buscemi, *Chemistry of Heterocyclic Compounds* **2017**, 53(9), 936–947, DOI: 10.1007/s10593-017-2154-1
29. (a) Recent Advances in the Chemistry of 1,2,4-Oxadiazoles, Andrea Pace, Silvestre Buscemi, Antonio Palumbo Piccionello, Ivana Pibiri, *Advances in Heterocyclic Chemistry*, **2015**, Volume 116, Pages 85-136, DOI: 10.1016/bs.aihch.2015.05.001
30. Preparation and Utility of 5- $\beta$ -D-Ribofuranosyl-1H-tetrazole as a Key Synthon for C-Nucleoside Synthesis, J. Kobe, M. Prhac, M. Hohnjec & L. B. Townsend, *Nucleosides and Nucleotides*, **1994**, 13:10, 2209-2244, DOI: 10.1080/15257779408013218
31. Five-Membered Heterocycles with Four Heteroatoms: Tetrazoles, Ulhas Bhatt, In book: *Modern Heterocyclic Chemistry*, pp.1401-1430, Chapter-15, DOI: 10.1002/chin.201201256
32. Comproportionation based Cu(I) catalyzed [3+2] cycloaddition of nitriles and sodium azide, Yakambram B, Srinivasulu K Uday kumar N, Jaya Shree A, Rakeshwar Bandichhor, *Chemistry & Biology Interface*, **2015**, 5, 1, 51-62
33. (a) 1,3,4-Oxadiazoles by Ugi-Tetrazole and Huisgen Reaction, Qian W, Kumchok C. M, Markella K, Svitlana V. S, and Dömling A. *Org. Lett.* **2019**, 21, 7320–7323. DOI: 10.1021/acs.orglett.9b02614 (b) High-Temperature Continuous Flow Synthesis of 1,3,4-Oxadiazoles via N-Acylation of 5-Substituted Tetrazoles. Reichart, B.; Kappe, C. O. *Tetrahedron Lett.* **2012**, 53, 952–955. DOI: 10.1016/j.tetlet.2011.12.043
34. 1, 3, 4-oxadiazole nucleus with versatile pharmacological applications: A Review. Arshad M, *Int J Pharm Sci Res* **2014**; 5(4): 1124-37. DOI: 10.13040/IJPSR.0975-8232.5(4).1124-37
35. 1,2,4- and 1,3,4-Oxadiazoles as Scaffolds in the Development of Antiparasitic Agents, Paulo Pitasse-Santos, Vitor Sueth-Santiago and Marco E. F. Lima, *J. Braz. Chem. Soc.*, **2018**, Vol. 29, No. 3, 435-456, DOI: 10.21577/0103-5053.20170208
36. Selected nucleos(t)ide-based prescribed drugs and their multi-target activity Gabriela Pastuch-Gawolek, Danuta Gillner, Ewelina Król, Krzysztof Walczak, Ilona Wandzik, *European Journal of Pharmacology*, **2019**, 865, 172747, DOI: 10.1016/j.ejphar.2019.172747
37. Synthesis and Antiviral Activity of 3-( $\beta$ -D-Ribofuranosyl)-1,2,4-oxadiazole-5-carboxamide Ram Pratap and V. N. Yarovenko, *Nucleosides, Nucleotides and Nucleic Acids*, **2000**, 19:5-6, 845-849, DOI: 10.1080/15257770008033026
38. Xylo-C-nucleosides with a pyrrolo[2,1-f][1,2,4]triazin-4-amine heterocyclic base: Synthesis and antiproliferative properties Peng Nie, Elisabetta Groaz, Dirk Daelemans, Piet Herdewijn, *Bioorganic & Medicinal Chemistry Letters*, **2019**, 29 (12), 1450-1453, DOI: 10.1016/j.bmcl.2019.04.023
39. High-temperature continuous flow synthesis of 1,3,4-oxadiazoles via N-acylation of 5-substituted tetrazoles, Benedikt Reichart, C. Oliver Kappe, *Tetrahedron Letters*, **2012**, 53, 952-955, DOI:10.1016/j.tetlet.2011.12.043
40. Acylierung 5-substituierter Tetrazole zu 1.3.4-Oxdiazolen, Huisgen, R.; Sauer, J.; Sturm, H. J. *Angew. Chem.* **1958**, 70, 272–273; DOI: 10.1002/ange.19580700918
41. Zur acylierung von 5-aryl-tetrazolen; ein duplikationsverfahren zur darstellung von polyarylen, Sauer, J.; Huisgen, R.; Sturm, H. J. *Tetrahedron* **1960**, 11, 241–251; DOI: 10.1016/S0040-4020(01)93173-4
42. Synthesis and Functionalization of 5-Substituted Tetrazoles, Jaroslav Roh, Katerina Vávrová and Alexandr Hrabálek, *Eur. J. Org. Chem.* **2012**, 6101–6118; DOI: 10.1002/ejoc.201200469

43. A base-induced ring-opening process of 2-substituted-1,3,4-oxadiazoles for the generation of nitriles at room temperature, Guo-ping Lu and Ya-mei Lin, *Journal of Chemical Research* **2014**, 38, 371-374, DOI: 10.3184/174751914X14007780679741
44. New Polynuclear Nonfused Bis(1,3,4-Oxadiazole) Systems, Yagoub Mansoori, and Raana Sarvari, *J. Mex. Chem. Soc.* **2014**, 58(2), 205-210
45. Anti-Cancer Activity of Derivatives of 1,3,4-Oxadiazole, Teresa Glomb, Karolina Szymankiewicz and Piotr S'wiatek, *Molecules* **2018**, 23, 3361; DOI:10.3390/molecules23123361
46. Selective Cytotoxicity of Goniothalamine against Hepatoblastoma HepG2 Cells, Mothanna Al-Qubaisi, Rosli Rozita, Swee-Keong Yeap, Abdul-Rahman Omar, Abdul-Manaf Ali, and Noorjahan B. Alitheer, *Molecules* **2011**, 16, 2944-2959; doi:10.3390/molecules16042944
47. The impact of an extended nucleobase-2' - deoxyribose linker in the biophysical and biological properties of oligonucleotides, Alejandro Carnero, Sonia P ´erez-Rentero, Adele Alagia, Anna Avin˜o, Yogesh S. Sanghvi, Susana Fernandez, Miguel Ferrero and Ramon Eritja, *RSC Adv.*, **2017**, 7, 9579–9586, DOI: 10.1039/c6ra26852h