



## Structure–activity relationship study of pyridazine derivatives as glutamate transporter EAAT2 activators

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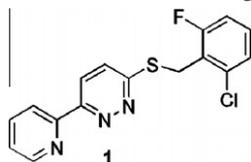
Excitotoxicity

### ABSTRACT

Excitatory amino acid transporter 2 (EAAT2) is the major glutamate transporter and functions to remove glutamate from synapses. A thiopyridazine derivative has been found to increase EAAT2 protein levels in astrocytes. A structure–activity relationship study revealed that several components of the molecule were required for activity, such as the thioether and pyridazine. Modification of the benzylthioether resulted in several derivatives (**7-13**, **7-15** and **7-17**) that enhanced EAAT2 levels by >6-fold at concentrations <5 μM after 24 h. In addition, one of the derivatives (**7-22**) enhanced EAAT2 levels 3.5–3.9-fold after 24 h with an EC<sub>50</sub> of 0.5 μM.

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Glutamate is a major neurotransmitter in the mammalian central nervous system (CNS) and essential for normal brain function including cognition, memory, and learning. However, the extracellular concentration of glutamate must remain below excitotoxic levels (~1 μM) to avoid overstimulation of glutamate receptors, leading to neuronal damage or death.<sup>1</sup> Excitotoxicity has been associated with multiple acute neurological conditions such as ischemic stroke, epilepsy, and trauma, as well as chronic adult-onset neurodegenerative disorders such as Alzheimer's disease and amyotrophic lateral sclerosis (ALS).<sup>2–6</sup> One potential approach to preventing excitotoxicity is to enhance glutamate reuptake. Excitatory amino acid transporter 2 (EAAT2) is the major glutamate transporter and functions to remove glutamate from synapses.<sup>7</sup> An increase in EAAT2 protein expression and function may provide a means to prevent insufficient glutamate reuptake and consequently reduce neuronal damage.



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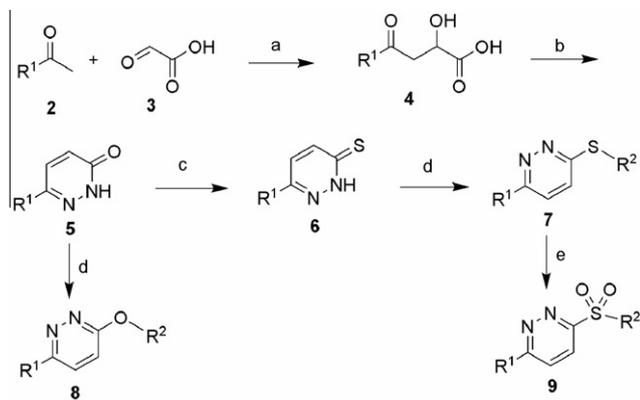
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In an effort to identify small molecules that can increase EAAT2 protein expression, a high-throughput screen of approximately 140,000 compounds was previously conducted in our laboratories using a cell-based enzyme-linked immunosorbent assay.<sup>8</sup> The hits identified from this screening provide starting points for further optimization in order to arrive at pharmacologically useful molecules for studying the role of EAAT2 in excitotoxicity induced neuronal injury and potentially as therapeutic agents.

The thiopyridazine **1** was confirmed to show a dose-dependent increase in EAAT2 protein levels after 24 h exposure. Herein, we report the structure–activity relationship (SAR) study of **1** for elevating EAAT2 protein levels.

Many of the pyridazine analogues utilized in the SAR study were prepared using the method depicted in *Scheme 1*. Treatment of ketone **2** with glyoxylic acid (**3**) and K<sub>2</sub>CO<sub>3</sub> gave **4**, which was used directly without purification. It was allowed to react with hydrazine in acetic acid at 100 °C to yield the desired pyridazinone **5** as an off-white solid after recrystallization from ethyl acetate.<sup>9</sup> Direct alkylation of **5** gave **8** in good yields. Intermediate **5** was also converted into pyridazinethione **6** in the presence of P<sub>2</sub>S<sub>5</sub> in pyridine at 120 °C.<sup>10</sup> Alkylation of **6** provided **7**, which could be subsequently oxidized to sulfone **9** with 3-chloroperoxybenzoic acid (m-CPBA) in CH<sub>2</sub>Cl<sub>2</sub>.<sup>11</sup>

A series of additional analogues **14–17** was prepared using the methodology outlined in *Scheme 2*. Recently, 2-pyridyl *N*-methyliminodiacetic acid (MIDA) boronate (**10**) has been reported as an air-stable slow-release reagent with high cross-coupling efficiency even with heteroaryl chlorides.<sup>12</sup> Therefore, this material was used



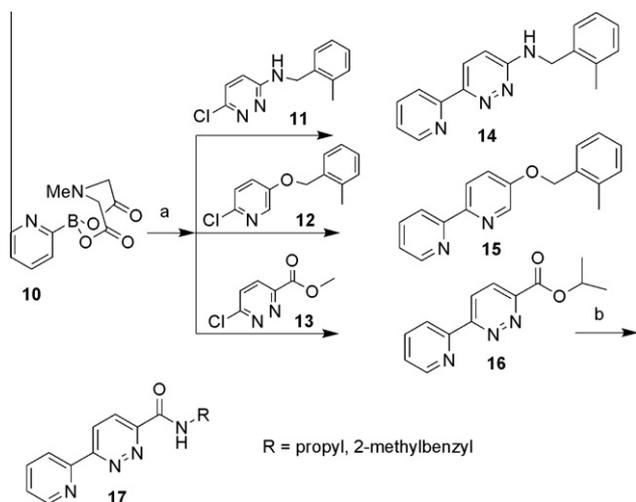
**Scheme 1.** Reagents and conditions: (a)  $K_2CO_3$ ,  $H_2O$ ; (b)  $NH_2NH_2$ ,  $AcOH$ ,  $100\text{ }^\circ C$  (20%); (c)  $P_2S_5$ , pyridine,  $120\text{ }^\circ C$  (80%); (d)  $R^2Br$ ,  $K_2CO_3$ ,  $DMF$  (90%); (e) *m*-CPBA,  $CH_2Cl_2$  (80%).

in palladium-mediated cross-couple reactions with heteroaryl chlorides **11–13** to obtain **14–16**, respectively. In the case of **13** transesterification occurred during the cross-coupling reaction to give the isopropyl ester **16**, which was then allowed to react with alkylamines in ethanol at  $85\text{ }^\circ C$  to generate amides **17**.

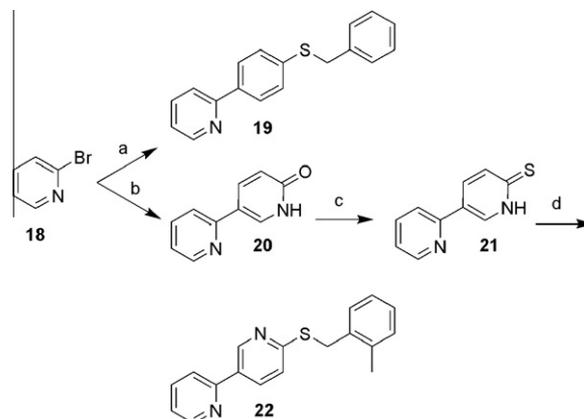
Finally, several additional analogues designed to evaluate replacement of the pyridazine in **1** with phenyl and pyridyl moieties were prepared using the methodology outlined in Scheme 3. Suzuki coupling of 2-bromopyridine (**18**) with various boronic acids afforded **19** and **20**.<sup>13</sup> Treatment of **20** with  $P_2S_5$  in pyridine at  $120\text{ }^\circ C$  provided **21** in 71% yield. Alkylation of **21** with 2-methylbenzyl bromide in the presence of  $K_2CO_3$  in  $DMF$  generated **22**.

All of the derivatives of **1** were initially evaluated in PA-EAAT2 cells<sup>3</sup> (a primary astrocyte line stably expressing EAAT2 mRNAs) following compound ( $10\text{ }\mu M$ ) incubation for 4 and 24 h before harvesting and measuring EAAT2 levels by Western blot analysis. The fold increases in EAAT2 protein levels relative to DMSO controls are reported (Tables 1–4).

Replacement of the 2-pyridyl with 3-pyridyl (**7-1**, **7-5** and **7-8**), 4-pyridyl (**7-2**, **7-9** and **7-11**) or phenyl (**7-6**) resulted in equal or reduced activity (Table 1). Removing either one or both of the nitrogen atoms in the pyridazine resulted in loss of activity (Table 2). Collectively, these results suggested that both the 2-pyridyl and the pyridazine were required for enhancing EAAT2 protein expression.



**Scheme 2.** Reagents and conditions: (a)  $Pd_2(dba)_3$ , XPhos,  $Cu(OAc)_2$ ,  $K_2CO_3$ ,  $DMF/i-PrOH$  (4/1),  $100\text{ }^\circ C$ ; (b)  $H_2NR$ , ethanol,  $85\text{ }^\circ C$  (90%).



**Scheme 3.** Reagents and conditions: (a)  $Pd(PPh_3)_4$ ,  $Na_2CO_3$ ,  $CH_3CN/H_2O$  (1/1),  $75\text{ }^\circ C$ , 4-benzylthiophenylboronic acid (54%); (b)  $Pd(PPh_3)_4$ ,  $Na_2CO_3$ ,  $CH_3CN/H_2O$  (1/1),  $75\text{ }^\circ C$ , 6-hydroxypyridine-3-boronic acid pinacol ester (70%); (c)  $P_2S_5$ , pyridine,  $120\text{ }^\circ C$  (71%); (d) 2-methylbenzyl bromide,  $K_2CO_3$ ,  $DMF$  (76%).

**Table 1**

Effects of the modified pyridyl and benzyl substituent on EAAT2 protein levels

Compound	R <sup>1</sup>	R <sup>2</sup>	Fold increase for EAAT2 <sup>a</sup>	
			4 h	24 h
<b>1</b>	2-Py	2-Cl-6-F-Bn	na <sup>b</sup>	2.0 ± 0.8
<b>7-1</b>	3-Py	2-Cl-6-F-Bn	na	2.3 ± 0.8
<b>7-2</b>	4-Py	2-Cl-6-F-Bn	na	1.9 ± 0.9
<b>7-3</b>	4-Me-Ph	2-Cl-6-F-Bn	1.2 ± 0.2	2.3 ± 0.7
<b>7-4</b>	2-Py	2-Me-Bn	1.7 ± 0.1	3.5 ± 0.3
<b>7-5</b>	3-Py	2-Me-Bn	1.5 ± 0.4	na
<b>7-6</b>	Ph	2-Me-Bn	1.1 ± 0.1	2.0 ± 0.4
<b>7-7</b>	2-Py	2-Cl-Bn	2.1 ± 1.4	4.0 ± 0.3
<b>7-8</b>	3-Py	2-Cl-Bn	1.2 ± 0.4	1.2 ± 0.3
<b>7-9</b>	4-Py	2-Cl-Bn	1.5 ± 0.2	1.7 ± 0.3
<b>7-10</b>	2-Py	4-Me-Bn	2.1 ± 1.1	2.6 ± 0.4
<b>7-11</b>	4-Py	4-Me-Bn	1.3 ± 0.1	2.4 ± 0.6

<sup>a</sup> Compound concentration of  $10\text{ }\mu M$ .

<sup>b</sup> na = not active.

**Table 2**

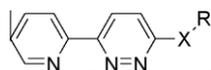
Effects of modifications of the pyridazine on EAAT2 protein levels

Compound	X	Y	Z	R	Fold increase for EAAT2 <sup>a</sup>	
					4 h	24 h
<b>7-4</b>	S	N	N	2-Me-Bn	1.7 ± 0.1	3.5 ± 0.3
<b>22</b>	S	C	N	2-Me-Bn	1.3 ± 0.6	na <sup>b</sup>
<b>8-1</b>	O	N	N	2-Me-Bn	2.0 ± 0.9	2.5 ± 0.5
<b>15</b>	O	N	C	2-Me-Bn	1.5 ± 0.3	na
<b>7-12</b>	S	N	N	Bn	1.3 ± 0.2	3.0 ± 0.4
<b>19</b>	S	C	C	Bn	1.2 ± 0.5	1.1 ± 0.4

<sup>a</sup> Compound concentration of  $10\text{ }\mu M$ .

<sup>b</sup> na = not active.

Next, the sulfur linker was examined (Table 3). Replacing the sulfur with oxygen generally yielded less active derivatives. An

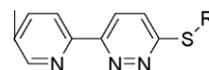
**Table 3**  
Effects of modifications of the sulfur linker on EAAT2 protein levels

Compound	X	R	Fold increase for EAAT2 <sup>a</sup>	
			4 h	24 h
<b>1</b>	S	2-Cl-6-F-Bn	na <sup>b</sup>	2.0 ± 0.8
<b>8-2</b>	O	2-Cl-6-F-Bn	1.1 ± 0.3	2.9 ± 0.4
<b>7-4</b>	S	2-Me-Bn	1.7 ± 0.1	3.5 ± 0.3
<b>8-1</b>	O	2-Me-Bn	2.0 ± 0.9	2.5 ± 0.5
<b>7-13</b>	S	2,6-Di-Me-Bn	na	6.5 ± 1.0
<b>8-3</b>	O	2,6-Di-Me-Bn	2.1 ± 1.2	3.2 ± 1.4
<b>7-14</b>	S	1-(2-Cl-6-F-phenyl)ethyl	1.7 ± 0.9	5.5 ± 1.0
<b>8-4</b>	O	1-(2-Cl-6-F-phenyl)ethyl	1.6 ± 0.2	3.0 ± 0.7
<b>7-15</b>	S	2-(2-Cl-6-F-phenylethyl)	1.9 ± 0.7	6.7 ± 1.5
<b>8-5</b>	O	2-(2-Cl-6-F-phenylethyl)	2.2 ± 0.4	3.1 ± 1.2
<b>7-16</b>	S	2,4-Di-F-Bn	2.4 ± 1.5	3.8 ± 0.8
<b>8-6</b>	O	2,4-Di-F-Bn	na	2.3 ± 0.6
<b>7-17</b>	S	2,4-Di-Me-Bn	1.9 ± 0.8	6.4 ± 1.0
<b>8-7</b>	O	2,4-Di-Me-Bn	na	2.0 ± 0.9
<b>6-1</b>	S	H	na	2.0 ± 0.8
<b>5-1</b>	O	H	1.5 ± 0.5	1.6 ± 1.0
<b>14</b>	NH	2-Me-Bn	1.7 ± 1.1	2.6 ± 0.5
<b>9-1</b>	SO <sub>2</sub>	2-Cl-6-F-Bn	1.4 ± 0.3	2.8 ± 0.4
<b>9-2</b>	SO <sub>2</sub>	2-Me-Bn	2.3 ± 0.4	1.1 ± 0.2
<b>9-3</b>	SO <sub>2</sub>	2-Cl-Bn	1.6 ± 0.3	na
<b>17-1</b>	CONH	<i>n</i> -Pr	na	1.2 ± 0.2
<b>17-2</b>	CONH	2-Me-Bn	na	1.4 ± 0.2

<sup>a</sup> Compound concentration of 10 μM.<sup>b</sup> na = not active.

analogue containing a NH linker (**14**) showed weaker activity. Likewise, oxidation of the sulfur to a sulfone (**9-1**, **9-2**, and **9-3**) was also detrimental. Finally, replacing the sulfur with an amide moiety (**17-1** and **17-2**) was not tolerated. Collectively, these results indicated that the sulfur linker was optimal.

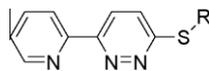
Finally, the benzyl group was examined (Table 4). Compared to **1**, 2-chloro, 3-chloro and 2,6-dichloro substitutions improved the potency by 2-fold (**7-7**, **7-18** and **7-22**), but 2,3-dichloro, 2,4-di-

**Table 4**  
Effects of the benzyl group on EAAT2 protein levels

Compound	R	Fold increase for EAAT2 <sup>a</sup>	
		4 h	24 h
<b>1</b>	2-Cl-6-F-Bn	na <sup>b</sup>	2.0 ± 0.8
<b>7-7</b>	2-Cl-Bn	2.1 ± 1.4	4.0 ± 0.3
<b>7-18</b>	3-Cl-Bn	2.1 ± 0.3	4.1 ± 0.5
<b>7-19</b>	2,3-Di-Cl-Bn	na	2.4 ± 0.4
<b>7-20</b>	2,5-Di-Cl-Bn	1.4 ± 0.1	2.5 ± 0.4
<b>7-21</b>	2,4-Di-Cl-Bn	1.8 ± 0.5	na
<b>7-22</b>	2,6-Di-Cl-Bn	1.5 ± 0.8	3.9 ± 0.4
<b>7-23</b>	2-F-Bn	2.2 ± 0.1	3.1 ± 0.2
<b>7-24</b>	4-F-Bn	2.5 ± 0.6	2.6 ± 0.3
<b>7-16</b>	2,4-Di-F-Bn	2.4 ± 1.5	3.8 ± 0.8
<b>7-25</b>	2,6-Di-F-Bn	2.1 ± 1.4	2.2 ± 0.7
<b>7-26</b>	2,4,6-Tri-F-Bn	1.6 ± 1.0	2.4 ± 0.6
<b>7-4</b>	2-Me-Bn	1.7 ± 0.1	3.5 ± 0.3
<b>7-27</b>	3-Me-Bn	2.0 ± 0.2	2.8 ± 0.5
<b>7-28</b>	4-Me-Bn	2.1 ± 1.1	2.6 ± 0.4
<b>7-29</b>	2,4-Di-Me-Ph	1.2 ± 0.3	2.2 ± 0.3
<b>7-17</b>	2,4-Di-Me-Bn	1.9 ± 0.8	6.4 ± 1.0
<b>7-13</b>	2,6-Di-Me-Bn	na	6.5 ± 1.0
<b>7-30</b>	2,4,6-Tri-Me-Bn	1.7 ± 0.3	4.7 ± 0.5
<b>7-15</b>	2-(2-Cl-6-F-phenylethyl)	1.9 ± 0.7	6.7 ± 1.5
<b>7-31</b>	2-(2-Cl-phenylethyl)	1.7 ± 0.2	4.8 ± 0.6
<b>7-32</b>	2-(2-Me-phenylethyl)	1.2 ± 0.3	2.8 ± 0.4
<b>7-33</b>	1-(1,2,3,4-Tetrahydronaphthalenyl)	na	5.3 ± 1.1
<b>7-14</b>	1-(2-Cl-6-F-phenyl)ethyl	1.7 ± 0.9	5.5 ± 1.0
<b>7-34</b>	2-Cl-4-F-Ph	1.3 ± 0.1	2.5 ± 0.7
<b>7-35</b>	2-Cl-4-F-Bn	1.6 ± 0.6	1.7 ± 0.4
<b>7-12</b>	Bn	1.3 ± 0.2	3.0 ± 0.5
<b>7-36</b>	Et	1.6 ± 0.7	1.9 ± 0.9
<b>6-1</b>	H	na	2.0 ± 0.8

<sup>a</sup> Compound concentration of 10 μM.<sup>b</sup> na = not active.

chloro and 2,5-dichloro analogues (**7-19**, **7-21** and **7-20**) were equivalent to **1**. In the case of fluorine substitutes, only the 2-fluoro

**Table 5**  
Fold increase and EC<sub>50</sub> values of EAAT2 protein levels of selected analogues<sup>a</sup>

Compound	R	Fold increase for EAAT2				EC <sub>50</sub> (μM)	
		1 μM, 24 h		10 μM, 24 h		β-actin <sup>b</sup>	GAPDH <sup>c</sup>
		β-actin <sup>b</sup>	GAPDH <sup>c</sup>	β-actin <sup>b</sup>	GAPDH <sup>c</sup>		
<b>7-4</b>	2-Me-Bn	1.0 ± 0.8	1.0 ± 0.3	3.5 ± 0.3	3.1 ± 0.2	2.6 ± 0.8	1.8 ± 0.3
<b>7-7</b>	2-Cl-Bn	1.1 ± 0.5	1.6 ± 0.4	4.0 ± 0.3	4.5 ± 0.6	3.3 ± 0.5	3.2 ± 0.5
<b>7-13</b>	2,6-Di-Me-Bn	2.0 ± 0.2	2.0 ± 0.4	6.5 ± 1.0	6.4 ± 0.9	2.6 ± 0.2	2.4 ± 0.4
<b>7-14</b>	1-(2-Cl-6-F-phenyl)ethyl	1.5 ± 0.5	1.8 ± 0.4	5.5 ± 1.0	5.2 ± 1.2	3.9 ± 0.5	2.4 ± 0.4
<b>7-17</b>	2,4-Di-Me-Bn	2.0 ± 0.3	2.2 ± 0.6	6.4 ± 1.0	6.0 ± 0.9	2.7 ± 0.3	1.8 ± 0.6
<b>7-22</b>	2,6-Di-Cl-Bn	3.0 ± 0.1	3.5 ± 0.0	3.9 ± 0.4	3.5 ± 0.5	0.5 ± 0.1	0.5 ± 0.0
<b>7-16</b>	2,4-Di-F-Bn	1.0 ± 0.2	1.0 ± 0.5	3.8 ± 0.8	3.7 ± 0.6	2.6 ± 0.2	2.6 ± 0.5
<b>7-30</b>	2,4,6-Tri-Me-Bn	1.5 ± 0.4	1.6 ± 0.4	4.7 ± 0.5	4.8 ± 0.6	2.4 ± 0.4	2.1 ± 0.4
<b>7-15</b>	2-(2-Cl-6-F-phenylethyl)	1.2 ± 0.3	1.2 ± 0.2	6.7 ± 1.5	5.3 ± 1.2	3.3 ± 0.3	3.3 ± 0.2
<b>7-31</b>	2-(2-Cl-phenylethyl)	1.0 ± 0.7	1.0 ± 0.1	4.8 ± 0.6	4.5 ± 0.6	4.7 ± 0.8	5.1 ± 0.1
<b>7-33</b>	1-(1,2,3,4-Tetrahydronaphthalenyl)	2.4 ± 0.3	2.8 ± 0.4	5.3 ± 1.1	4.5 ± 0.6	1.9 ± 0.3	1.7 ± 0.4
<b>7-18</b>	3-Cl-Bn	4.0 ± 0.4	4.2 ± 0.1	4.1 ± 0.5	3.8 ± 0.5	1.7 ± 0.4	0.5 ± 0.1

<sup>a</sup> Data represent the mean values of four separate experiments.<sup>b</sup> Data standardized by β-actin.<sup>c</sup> Data standardized by GAPDH.

and 2,4-difluoro derivatives (**7-23** and **7-16**) demonstrated improved activity. 4-Fluoro, 2,6-difluoro, and 2,4,6-trifluoro analogues (**7-24**, **7-25**, and **2-26**) did not result in significant improvement. Replacement of the halogens with methyls (**7-4**, **7-13**, **7-17**, and **7-30**) gave increased potency. Compounds containing 2-methyl substituted benzyl groups significantly increased EAAT2 protein level, where as the 3- or 4-methylbenzyl derivatives (**7-27** and **7-28**) were less potent. Increasing the tether length between the phenyl and the sulfur to an ethylene generally yielded more potent analogues (**7-15** and **7-31** vs **1** and **7-7**, respectively) with one noted exception (compare **7-32** to **7-4**). However, truncating the linker resulting in a diarylthioether decreased potency (**7-29** vs **7-17**). Interestingly, adding substitutes on the carbon linker (**7-14** and **7-33**) increased activity about 3-fold. Finally, other changes gave compounds with essentially the same potency as **1** (**7-12**, **7-36**, and **6-1**).

In order to further characterize twelve of the most active analogues from the initial assessment, they were evaluated in a 6-point dose–response assay (0.1, 0.3, 1.0, 3.3, 10.0, and 30.0  $\mu\text{M}$ ). The fold increases at 1 and 10  $\mu\text{M}$ , as well as the  $\text{EC}_{50}$  values are summarized in Table 5. In addition, these values were determined relative to two unrelated proteins (e.g.,  $\beta$ -actin and GAPDH). Several compounds (**7-13**, **7-17**, **7-31** and **7-18**) were able to increase EAAT2 levels >6-fold at 10  $\mu\text{M}$  compared to  $\beta$ -actin or GAPDH, but with  $\text{EC}_{50}$  values in the 2.6–3.3  $\mu\text{M}$  range. Analog **7-22** demonstrated the lowest  $\text{EC}_{50}$  of 0.5  $\mu\text{M}$  with a maximum increase of EAAT2 levels of 3- to 4-fold.

In summary, a series of pyridazine derivatives were synthesized and evaluated for increasing EAAT2 protein levels. Several compounds were found to significantly increase EAAT2 protein levels (>6-fold), such as **7-13**, **7-15** and **7-17**. Derivative **7-22** increased EAAT2 protein levels to a lower extent (3.5–3.9-fold), but with a lower  $\text{EC}_{50}$  value (0.5  $\mu\text{M}$ ). These compounds will provide useful tools for further assessing the role of glutamate excitotoxicity in cellular systems and potentially in animal models of acute and chronic neurodegeneration. In addition, these probe molecules

may also be beneficial in determining the biological mechanisms for regulating EAAT2 levels.

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### Supplementary data

Supplementary data associated with this article can be found, in the online version, at [doi:10.1016/j.bmcl.2011.08.009](https://doi.org/10.1016/j.bmcl.2011.08.009).

### References and notes

- Sheldon, A. L.; Robinson, M. B. *Neurochem. Int.* **2007**, *51*, 333.
- Guo, H.; Lai, L.; Butchbath, M. E. R.; Stockinger, M. P.; Shan, X.; Bishop, G. A.; Lin, C. L. G. *Hum. Mol. Genet.* **2003**, *12*, 2519.
- Tian, G.; Lai, L.; Guo, H.; Lin, Y.; Butchbach, M.; Chang, Y.; Lin, C.-L. *J. Biol. Chem.* **2007**, *282*, 1727.
- Hazell, A. *Neurochem. Int.* **2007**, *50*, 941.
- Seifert, G.; Carmignoto, G.; Steinhäuser, C. *Brain. Res. Rev.* **2010**, *63*, 212.
- Tian, G.; Kong, Q.; Lai, L.; Ray-Chaudhury, A.; Lin, C.-L. *J. Neurochem.* **2010**, *113*, 978.
- Lin, Y.; Tian, G.; Roman, K.; Handy, C.; Travers, J. B.; Lin, C. L. G.; Stephens, R. L., Jr. *Am. J. Physiol. Gastrointest Liver Physiol.* **2009**, *296*, 129.
- Colton, C. K.; Kong, Q.; Lai, L.; Zhu, M. X.; Seyb, K. I.; Cuny, G. D.; Xian, J.; Glicksman, M. A.; Lin, C. L. G. *J. Biomol. Screen.* **2010**, *15*, 653.
- Coates, W. J.; McKillop, A. *Synthesis* **1993**, 334–342.
- Arakawa, K.; Miyasaka, T.; Satoh, K. *Chem. Pharm. Bull.* **1977**, *25*, 299.
- Mylari, B. L.; Armento, S. J.; Beebe, D. A.; Conn, E. L.; Coutcher, J. B.; Dina, M. S.; O’Gorman, M. T.; Linhares, M. C.; Martin, W. H.; Oates, P. J.; Tess, D. A.; Withbroe, G. J.; Zembrowsky, W. J. *J. Med. Chem.* **2005**, *48*, 6326.
- Knapp, D. M.; Gillis, E. P.; Burke, M. D. *J. Am. Chem. Soc.* **2009**, *131*, 6961.
- Ghiron, C.; Haydar, S. N.; Aschmies, S.; Bothmann, H.; Castaldo, C.; Cocconcelli, G.; Comery, T. A.; Di, L.; Dunlop, J.; Lock, T.; Kramer, A.; Kowal, D.; Jow, F.; Grauer, S.; Harrison, B.; Rosa, S.; Maccari, L.; Marquis, K.; Micco, I.; Nencini, A.; Quinn, J.; Robichaud, A.; Roncarati, R.; Scali, C.; terstappen, G.; Turlizzi, E.; Valacchi, M.; Varrone, M.; Zalaletti, R.; Zanelli, U. *J. Med. Chem.* **2010**, *53*, 4379.