



Review

Neuronal voltage-gated K⁺ (Kv) channels function in macromolecular complexes

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ARTICLE INFO

Article history:

Received 7 June 2010

Received in revised form 14 August 2010

Accepted 23 August 2010

Keywords:

Kv channels

Proteomics

Neuronal excitability

Macromolecular complex

Signaling

ABSTRACT

Considerable evidence indicates that native neuronal voltage-gated K⁺ (Kv) currents reflect the functioning of macromolecular Kv channel complexes, composed of pore-forming (α)-subunits, cytosolic and transmembrane accessory subunits, together with regulatory and scaffolding proteins. The individual components of these macromolecular complexes appear to influence the stability, the trafficking, the localization and/or the biophysical properties of the channels. Recent studies suggest that Kv channel accessory subunits subserve multiple roles in the generation of native neuronal Kv channels. Additional recent findings suggest that Kv channel accessory subunits can respond to changes in intracellular Ca²⁺ or metabolism and thereby integrate signaling pathways to regulate Kv channel expression and properties. Although studies in heterologous cells have provided important insights into the effects of accessory subunits on Kv channel expression/properties, it has become increasingly clear that experiments in neurons are required to define the physiological roles of Kv channel accessory and associated proteins. A number of technological and experimental hurdles remain that must be overcome in the design, execution and interpretation of experiments aimed at detailing the functional roles of accessory subunits and associated proteins in the generation of native neuronal Kv channels. With the increasing association of altered Kv channel functioning with neurological disorders, the potential impact of these efforts is clear.

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Voltage-gated K⁺ (Kv) currents are key regulators of neuronal membrane excitability, functioning to control resting membrane potentials [12], spontaneous firing rates [45], the back propagation (into dendrites) of action potentials [18], neurotransmitter release [20], synaptic integration [17,53], and even apoptosis [40]. Neurons typically express multiple types of Kv channels with distinct time- and voltage-dependent properties and subcellular distributions that differentially contribute to the regulation of firing properties and signal integration. Molecular cloning has revealed even greater potential for generating functional Kv channel diversity than was anticipated based on the physiology [10,45]. A large number of Kv channel pore-forming (α)-subunits and a variety of Kv channel accessory subunits have been identified [10,45], and studies in heterologous cells have revealed multiple, sometimes overlapping, effects of accessory subunits on the expression, distributions and properties of Kv α-subunit-encoded channels [25,32]. Electrophysiological and biochemical evidence indicates that native neuronal K⁺ currents reflect the functioning of macromolecular K⁺ channel complexes comprising four α-subunits, cytosolic and transmembrane accessory subunits, as well as regulatory and scaffolding proteins [25]. Recent proteomic studies, for example, demonstrated that native (mouse) brain Kv4 channel complexes contain many different types of acces-

sory and regulatory subunits [33], each of which may influence channel targeting, trafficking and/or biophysical properties. It has also become clear that the properties of Kv α-subunit-encoded channels and the modulatory effects of channel associated proteins vary in different cellular backgrounds. Thus, studies need to be carried out in neurons to understand the functions of Kv channel accessory subunits in the generation and regulation of native Kv channels. This review focuses on recent advances made in exploring the roles of Kv channel associated proteins in neurons with an emphasis on studies suggesting that Kv channel accessory subunits can serve as points of integration for signaling pathways, as well as link membrane excitability to other cellular processes.

Several recent studies, focused on examining the roles of K⁺ Channel Interacting Protein 3 (KChIP3), have highlighted the potential for Kv channel accessory subunits to serve as points of signal integration in the regulation of neuronal function [1,3,51]. KChIP3, like the other members of the Neuronal Ca²⁺ Sensor super family [5], binds Ca²⁺ through multiple EF-hand motifs [31,44]. KChIP3 also promotes the surface expression of heterologously expressed Kv4 channels [56] and regulates the functional expression of neuronal Kv4-encoded currents [29,39]. Exciting recent findings also demonstrate that KChIP3 regulates the voltage dependence of inactivation of Kv4-encoded channels in cerebellar stellate cells in response to Ca²⁺ entry through voltage-gated Ca²⁺ (Cav) channels [3]. In the presence of KChIP3 (but not the other KChIPs), Ca²⁺ entry through Cav3 channels induced a depolarizing shift in the voltage

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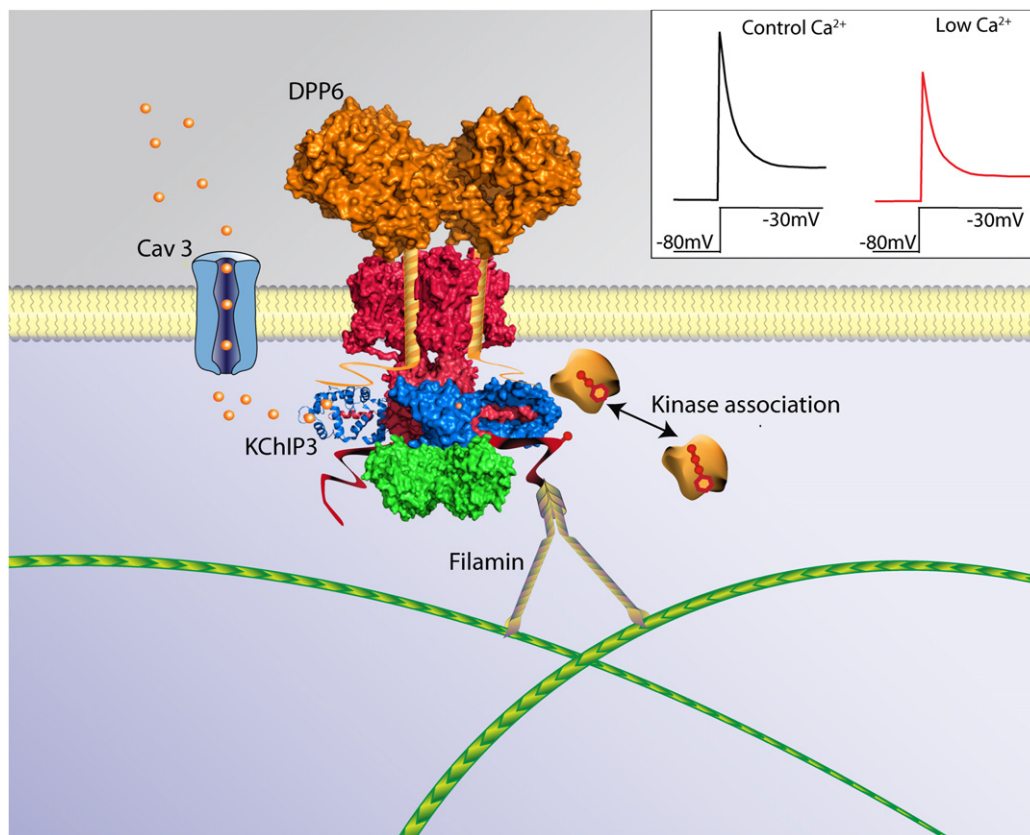


Fig. 1. Multiple components of Kv4 channel macromolecular protein complexes. Schematic representation of Kv4 channel α -subunits (red) bound to Kv β (green), DPP6 (orange) and KChIP3 (blue) accessory subunits. The channel complex is shown anchored to the actin cytoskeleton by filamin via the carboxyl-terminus of one of the Kv4 α -subunits. Kinases also associate with Kv4 channel complexes and phosphorylate α and/or accessory subunits, modifications that can affect channel stability, trafficking and/or properties. It has been reported that association of Kv4 α -subunits with KChIP3 accessory subunits is required for the modification of channel function by some kinases [51,52]. Cav3 channels can also associate with Kv4 channel complexes by binding to Kv4.2 [2]. In Kv4.2–Cav3 complexes identified in stellate neurons, KChIP3 acts as a Ca^{2+} sensor, and Ca^{2+} entry through Cav3 channels, shifts voltage dependence of Kv4.2 channel inactivation to more depolarized potentials [2,3]. Consistent with a role for Ca^{2+} entry through Cav3 channels in the regulation of Kv4.2 channels, reductions in extracellular Ca^{2+} concentration result in attenuation of Kv4.2 currents (inset) because more Kv4.2 channels are inactive at the holding potential when Ca^{2+} entry is reduced. Inset adapted with permission from Macmillan Publishers Ltd: Nature Neuroscience copyright 2010 [2].

dependence of Kv4 channel inactivation, thereby linking Kv4 channel availability to local changes in Ca^{2+} concentration [2] (Fig. 1, inset). Together, these observations suggest that KChIP3 may regulate functional Kv4-encoded current densities in neurons, both by controlling the numbers of Kv4 channels expressed at the cell surface and by dynamically regulating Kv4 channel availability in response to changes in Ca^{2+} entry (Fig. 2).

Additional experiments suggest that KChIP3 also regulates Kv4 channel modulation by kinases [52]. The association of G protein-coupled receptor kinases (GRK) with KChIP3, for example, is dependent on Ca^{2+} [51], suggesting that KChIP3 could function to integrate local changes in Ca^{2+} levels with kinase signaling pathways. In addition, the phosphorylation of Kv4 α -subunits has been suggested to be related to surface expression [19,54,61]. The possibility that KChIP3 acts as a point of signal integration to regulate the surface expression of Kv4 channels through multiple pathways is exciting, particularly given the importance of dynamic alterations in Kv4-encoded currents in modulating dendritic integration and synaptic potentiation [8,30]. Interestingly, it has also been reported that KChIPs regulate the surface expression and the functioning of voltage-gated Ca^{2+} channels [57], NMDA receptors [65] and other Kv channels [27], suggesting that these proteins are multifunctional, differentially regulating excitability in different cell types and/or subcellular compartments. Clearly, experiments in which the expression levels of the KChIPs are altered *in situ* are needed to explore these hypotheses directly.

Importantly, KChIP3 has also been demonstrated to function in other cellular processes and has been given additional names reflecting these roles: including regulating gene transcription (downstream regulatory element antagonist modulator or DREAM) [7], modulating the production of amyloid precursor protein by binding to presenilin (calsenilin) [6,28], and regulating gamma secretase activity and apoptosis [21,22]. The possibility that KChIP3 may help coordinate long-term changes, through regulating gene transcription, and short-term changes, in response to alterations in cellular activity (intracellular Ca^{2+} levels and phosphorylation state), in neuronal excitability is intriguing and warrants further investigation.

A number of recent studies also suggest that the inclusion of Kv β subunits in Kv channel complexes has broad ramifications for channel expression and functioning. Similar to KChIPs, Kv β subunits also appear to subserve multiple roles including: directly altering the time- and voltage-dependent properties of Kv currents [37,49], regulating the trafficking of Kv1 and Kv4 channels [14,15,64], and modulating changes in Kv currents in response to signaling pathways. Numerous kinases have been shown to phosphorylate Kv1 α -subunits, as well as Kv β subunits, and thereby modify the effects of Kv β subunits on Kv1-encoded currents [35]. Interestingly, Kv β subunits also reportedly associate with Kv4 α -subunits in brain [33], and coexpression of Kv β subunits with Kv4 α -subunits in heterologous cells alters the cell surface expression of Kv4-encoded channels [62,64]. Little is presently known, however, about the

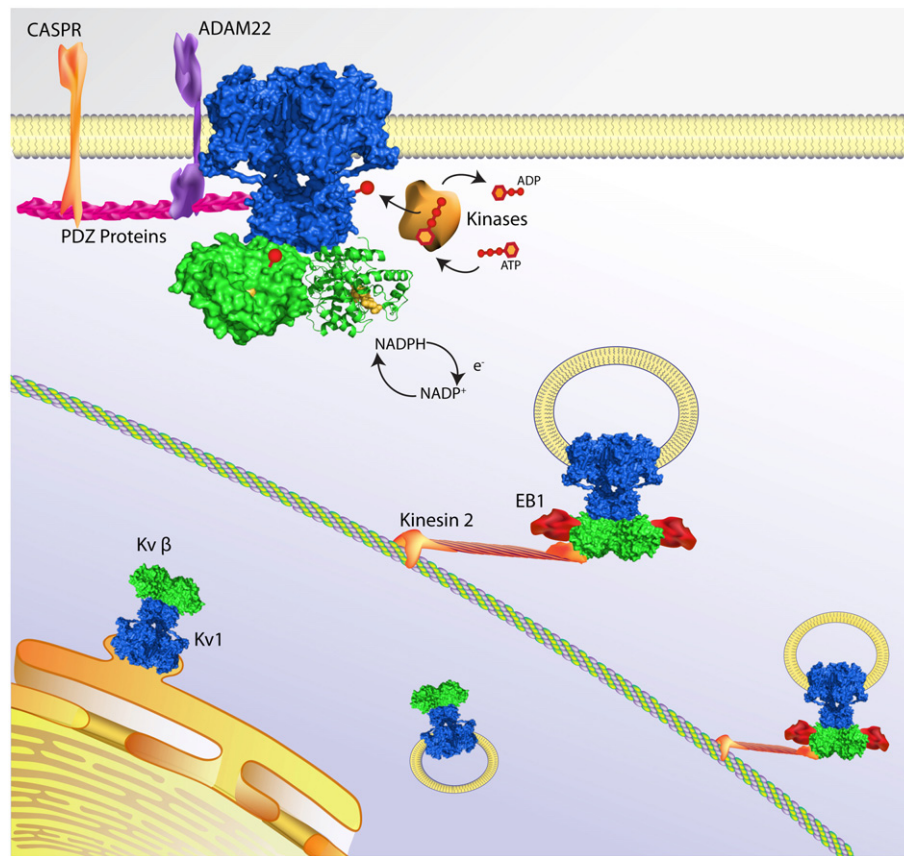


Fig. 2. Kv1 channel complexes targeted to axons. Schematic representation of the assembly and targeting of Kv1 channel macromolecular complexes. Previous studies have shown that Kv1 channel α -subunits (blue) associate with Kv β subunits (green) in the endoplasmic reticulum early during biosynthesis [55]. Through the binding of Kv β to EB-1 and Kinesin-2 channel complexes then are trafficked down the axon on microtubules. At nodes of Ranvier, Kv1-encoded channels associate with PDZ-domain containing proteins (e.g., PSD-95 and PSD-93) and with the transmembrane proteins, ADAM22 and CASPR. The functioning of Kv1-encoded channels can also be modulated by phosphorylation of both the Kv1 α and the Kv β subunit [35]. In addition, Kv β subunits can also regulate channel properties and functioning in response to changes in the oxidation state of the bound co-factor NADP $^{+}$, providing a potential link between cell metabolism and membrane excitability [42,63].

physiological role(s) of Kv β subunits in regulating the expression or the properties of native neuronal Kv4 channels.

Given that the Kv β subunits have a high degree of structural homology to aldo-keto reductases, it is perhaps not surprising that Kv β 2 was crystallized bound to the co-factor nicotinamide adenine dinucleotide phosphate (NADP $^{+}$) [9,16]. The Kv β subunits appear positioned, therefore, to serve as sensors of the oxidative state of cells, suggesting a direct link to metabolism [63]. Mutations in the NADP $^{+}$ binding site have been reported to disrupt Kv β mediated alterations in the trafficking of Kv1 α -subunits [42]. In addition, changes in the oxidation state of bound NADP $^{+}$ were shown to result in changes in the expression/properties of heterologously expressed Kv1-encoded currents [26,63]. Interestingly, Kv β subunits reportedly confer sensitivity to oxygen concentration to heterologously expressed Kv4, but not Kv1-encoded currents [41]. Accumulating evidence also suggests that Kv β subunits may couple changes in cellular metabolism to the regulation of membrane excitability [58], although a direct link in neurons has yet to be demonstrated.

As highlighted by the results of several recent studies [15,24,36], a number of challenges exist to efforts focused on defining the physiological roles of Kv channel accessory subunits in the generation of native neuronal Kv channels. Recent proteomic studies, focused on Kv4 channel complexes isolated from brain, indicated that Kv4 α -subunits channels interact with multiple different proteins. Many of the proteins traditionally thought of as Kv channel accessory subunits, such as DPP6 and the KChIPs were identified, but sev-

eral additional, unexpected proteins, such as the GABA receptor subunit, Gabra-6, and the poorly understood metabotropic glutamate receptor, Gpr158, were also found to co-immunoprecipitate with the Kv4.2 protein from brain [33]. Consistent with findings from studies on Kv4.2, other Kv channels have been suggested to interact with scaffolding proteins like ADAM22 and PSD-95. Thus, it seems likely that native Kv channel macromolecular complexes contain numerous accessory and regulatory proteins. In efforts to determine the functional roles of the proteins associated with Kv channels, experiments utilizing heterologous cells will be important, but experiments in native cells will be required to understand fully how each Kv channel accessory subunit functions in the generation of native neuronal Kv channel complexes.

There are, however, a number of potential challenges to the design, execution and interpretation of experiments focused on exploring the functioning of Kv channel accessory subunits in neurons. Among the potential challenges is the fact that many families of Kv channel accessory subunits, including the KChIPs, Kv β s and Mink/MiRPs, have multiple members with properties (and functional effects) that are likely conserved throughout the family. The individual subunits may be able to compensate for the loss of other family members. The disruption of the expression of an individual accessory subunit, therefore, may not reveal defects in the expression or the targeting of α -subunits and/or in the properties of expressed Kv currents dramatic enough to be resolved. Recent studies focused on exploring the contributions of the various KChIPs to the expression of Kv4-encoded channels

in (mouse) cortical pyramidal neurons, in fact, revealed that these cells express KChIP2, 3 and 4 and that when the expression of an individual KChIP was disrupted, the protein expression levels of the remaining two KChIPs are up regulated and Kv4 currents were only modestly affected [39]. Simultaneous RNAi mediated knock-down of all three KChIPs, however, caused a marked reduction in Kv4-encoded current densities [39].

Functional compensation may also occur in other families of Kv channel accessory subunits. There are, for example, three members of the Kv β family in mammals [46]. Expression of mutant Kv β subunits has revealed a critical role for Kv β subunits in the trafficking of Kv1 channels [14]. In animals lacking Kv β 2, the most abundantly expressed β subunit [50], however, Kv1.1 and Kv1.2 α -subunits traffic normally. Mice lacking Kv β 2, however, display cold swim-induced tremors, suffer occasional seizures and have a reduced average lifespan [36], suggesting that the loss of Kv β 2 produces defects that are undetected in the molecular studies focused on examining the effects of Kv β 2 on the trafficking Kv1 channels.

An additional challenge to determining the functional roles of Kv channel associated proteins is the potential for these proteins to interact with multiple channel types and to participate in a number of cellular processes. This point is well illustrated with proteins like the AKAPs and filamins, which interact with specific types of ion channels [13,43], but are also scaffolding proteins generally involved in the formation of macromolecular protein complexes [43,47]. Several Kv channel accessory subunits are also involved in multiple cellular processes. Both KChIP3 and KChIP4, for example, were independently identified as binding partners for the presenilins and were termed calsenilin and calsenilin related protein, respectively, suggesting roles for both proteins in the regulation of gamma secretase activity [6,38]. Disruption of the expression of molecules like the filamins and the KChIPs, therefore, could potentially have numerous consequences for cellular functioning not directly related to Kv channels.

Mutations in K⁺ channel α -subunits (for example, Kv7.2 and Kv7.3 in epilepsy [48], and Kv1.1 in Episodic Ataxia Type 1 [4]) have been associated with neurological disorders/diseases. It is perhaps not surprising then that recent results from genetic studies in patient populations and studies using genetically altered mice suggest roles for altered expression and/or functioning of Kv channel accessory subunits in a number of different neurological disorders/diseases. Structural variants of both of the Kv channel accessory subunit genes, *DPP6* and *DPP10*, for example, were identified in a genome-wide association study as candidate genes in autism spectrum disorder [34]. Interestingly, the *DPP6* gene has also been associated with increased risk of progressive spinal muscular atrophy [60] and sporadic amyotrophic lateral sclerosis [11,59]. Mutations in another K⁺ channel associated protein, leucine-rich glioma-inactivated 1 (Lgi1), have been linked to multiple epilepsy syndromes [23]. As the functional roles of channel accessory/associated proteins are increasingly appreciated, it seems certain that alterations in the expression and/or properties of these proteins will be linked to more disease states. Clearly, carefully crafted experiments, combining molecular genetic, pharmacological, electrophysiological and biochemical approaches, will be needed to define the roles of Kv channel accessory and associated proteins in the functioning of native neuronal Kv channel macromolecular complexes and to further delineate the contributions of Kv channel accessory subunits to neuronal functioning under normal and pathologic conditions.

Acknowledgements

The authors thank Mr. Rick Wilson for technical assistance in the creation of the figures. We would also like to thank Dr. Yarimar Car-

rasquillo for many valuable discussions and for comments on the manuscript. In addition, the authors acknowledge the financial support provided by the National Institutes of Health (NS030676 and NS065295 to JMN); AJN was supported by an institutional training grant (T32-EY013360) from the National Eye Institute and NCF was supported by an institutional training grant (T32-HL007275) from the National Heart Lung and Blood Institute.

References

- [1] J.C. Alexander, C.M. McDermott, T. Tunur, V. Rands, C. Stelly, D. Karhson, M.R. Bowlby, W.F. An, J.D. Sweatt, L.A. Schrader, The role of calsenilin/DREAM/KChIP3 in contextual fear conditioning, *Learn. Mem.* 16 (2009) 167–177.
- [2] D. Anderson, W.H. Mehafeff, M. Iftinca, R. Rehak, J.D. Engbers, S. Hameed, G.W. Zamponi, R.W. Turner, Regulation of neuronal activity by Cav3-Kv4 channel signaling complexes, *Nat. Neurosci.* 13 (2010) 333–337.
- [3] D. Anderson, R. Rehak, S. Hameed, W.H. Mehafeff, G.W. Zamponi, R.W. Turner, Regulation of the Kv4.2 complex by Cav3.1 calcium channels, *Channels (Austin)* 4 (2010).
- [4] D.L. Browne, S.T. Gancher, J.G. Nutt, E.R. Brunt, E.A. Smith, P. Kramer, M. Litt, Episodic ataxia/myokymia syndrome is associated with point mutations in the human potassium channel gene, *KCNA1*, *Nat. Genet.* 8 (1994) 136–140.
- [5] R.D. Burgoyne, Neuronal calcium sensor proteins: generating diversity in neuronal Ca²⁺ signalling, *Nat. Rev. Neurosci.* 8 (2007) 182–193.
- [6] J.D. Buxbaum, E.K. Choi, Y. Luo, C. Lilliehook, A.C. Crowley, D.E. Merriam, W. Wasco, Calsenilin: a calcium-binding protein that interacts with the presenilins and regulates the levels of a presenilin fragment, *Nat. Med.* 4 (1998) 1177–1181.
- [7] A.M. Carrion, W.A. Link, F. Ledo, B. Mellstrom, J.R. Naranjo, DREAM is a Ca²⁺-regulated transcriptional repressor, *Nature* 398 (1999) 80–84.
- [8] X. Chen, L.L. Yuan, C. Zhao, S.G. Birnbaum, A. Frick, W.E. Jung, T.L. Schwarz, J.D. Sweatt, D. Johnston, Deletion of Kv4.2 gene eliminates dendritic A-type K⁺ current and enhances induction of long-term potentiation in hippocampal CA1 pyramidal neurons, *J. Neurosci.* 26 (2006) 12143–12151.
- [9] S.W. Chouinard, G.F. Wilson, A.K. Schlimgen, B. Ganetzky, A potassium channel beta subunit related to the aldo-keto reductase superfamily is encoded by the *Drosophila* hyperkinetic locus, *Proc. Natl. Acad. Sci. U.S.A.* 92 (1995) 6763–6767.
- [10] W.A. Coetzee, Y. Amarillo, J. Chiu, A. Chow, D. Lau, T. McCormack, H. Moreno, M.S. Nadal, A. Ozaita, D. Pountney, M. Saganich, E. Vega-Saenz de Miera, B. Rudy, Molecular diversity of K⁺ channels, *Ann. N.Y. Acad. Sci.* 868 (1999) 233–285.
- [11] R. Del Bo, S. Ghezzi, S. Corti, D. Santoro, A. Prella, M. Mancuso, G. Siciliano, C. Briani, L. Murri, N. Bresolin, G.P. Comi, DPP6 gene variability confers increased risk of developing sporadic amyotrophic lateral sclerosis in Italian patients, *J. Neurol. Neurosurg. Psychiatry* 79 (2008) 1085.
- [12] P. Enyedi, G. Czirkak, Molecular background of leak K⁺ currents: two-pore domain potassium channels, *Physiol. Rev.* 90 (2010) 559–605.
- [13] B. Gravante, A. Barbuti, R. Milanesi, I. Zappi, C. Viscomi, D. DiFrancesco, Interaction of the pacemaker channel HCN1 with filamin A, *J. Biol. Chem.* 279 (2004) 43847–43853.
- [14] C. Gu, Y.N. Jan, L.Y. Jan, A conserved domain in axonal targeting of Kv1 (Shaker) voltage-gated potassium channels, *Science* 301 (2003) 646–649.
- [15] C. Gu, W. Zhou, M.A. Puthenveedu, M. Xu, Y.N. Jan, L.Y. Jan, The microtubule plus-end tracking protein EB1 is required for Kv1 voltage-gated K⁺ channel axonal targeting, *Neuron* 52 (2006) 803–816.
- [16] J.M. Gulbis, S. Mann, R. MacKinnon, Structure of a voltage-dependent K⁺ channel beta subunit, *Cell* 97 (1999) 943–952.
- [17] A.T. Guldedge, B.M. Kampa, G.J. Stuart, Synaptic integration in dendritic trees, *J. Neurobiol.* 64 (2005) 75–90.
- [18] D.A. Hoffman, J.C. Magee, C.M. Colbert, D. Johnston, K⁺ channel regulation of signal propagation in dendrites of hippocampal pyramidal neurons, *Nature* 387 (1997) 869–875.
- [19] H.J. Hu, Y. Carrasquillo, F. Karim, W.E. Jung, J.M. Nerbonne, T.L. Schwarz, R.W.T. Gereau, The kv4.2 potassium channel subunit is required for pain plasticity, *Neuron* 50 (2006) 89–100.
- [20] T. Ishikawa, Y. Nakamura, N. Saitoh, W.B. Li, S. Iwasaki, T. Takahashi, Distinct roles of Kv1 and Kv3 potassium channels at the calyx of Held presynaptic terminal, *J. Neurosci.* 23 (2003) 10445–10453.
- [21] D.G. Jo, M.J. Kim, Y.H. Choi, I.K. Kim, Y.H. Song, H.N. Woo, C.W. Chung, Y.K. Jung, Pro-apoptotic function of calsenilin/DREAM/KChIP3, *FASEB J.* 15 (2001) 589–591.
- [22] D.G. Jo, J.Y. Lee, Y.M. Hong, S. Song, I. Mook-Jung, J.Y. Koh, Y.K. Jung, Induction of pro-apoptotic calsenilin/DREAM/KChIP3 in Alzheimer's disease and cultured neurons after amyloid-beta exposure, *J. Neurochem.* 88 (2004) 604–611.
- [23] S. Kalachnikov, O. Evgrafov, B. Ross, M. Winawer, C. Barker-Cummings, F. Martinelli Boneschi, C. Choi, P. Morozov, K. Das, E. Teplitskaya, A. Yu, E. Cayanis, G. Penchaszadeh, A.H. Kottmann, T.A. Pedley, W.A. Hauser, R. Ottman, T.C. Gilliam, Mutations in LGI1 cause autosomal-dominant partial epilepsy with auditory features, *Nat. Genet.* 30 (2002) 335–341.
- [24] J. Kim, M.S. Nadal, A.M. Clemens, M. Baron, S.C. Jung, Y. Misumi, B. Rudy, D.A. Hoffman, Kv4 accessory protein DPPX (DPP6) is a critical regulator of membrane excitability in hippocampal CA1 pyramidal neurons, *J. Neurophysiol.* 100 (2008) 1835–1847.

- [25] H.C. Lai, L.Y. Jan, The distribution and targeting of neuronal voltage-gated ion channels, *Nat. Rev. Neurosci.* 7 (2006) 548–562.
- [26] T. Leicher, R. Bähring, D. Isbrandt, O. Pongs, Coexpression of the KCNA3B gene product with Kv1.5 leads to a novel A-type potassium channel, *J. Biol. Chem.* 273 (1998) 35095–35101.
- [27] H. Li, W. Guo, R.L. Mellor, J.M. Nerbonne, KChIP2 modulates the cell surface expression of Kv 1.5-encoded K(+) channels, *J. Mol. Cell. Cardiol.* 39 (2005) 121–132.
- [28] C. Lilliehook, O. Bozdagi, J. Yao, M. Gomez-Ramirez, N.F. Zaidi, W. Wasco, S. Gandy, A.C. Santucci, V. Haroutunian, G.W. Huntley, J.D. Buxbaum, Altered Abeta formation and long-term potentiation in a calenilin knock-out, *J. Neurosci.* 23 (2003) 9097–9106.
- [29] B. Liss, O. Franz, S. Sewing, R. Bruns, H. Neuhoff, J. Roeper, Tuning pacemaker frequency of individual dopaminergic neurons by Kv4.3L and KChIP3.1 transcription, *EMBO J.* 20 (2001) 5715–5724.
- [30] A. Losonczy, J.K. Makara, J.C. Magee, Compartmentalized dendritic plasticity and input feature storage in neurons, *Nature* 452 (2008) 436–441.
- [31] J.D. Lusin, M. Vanarotti, C. Li, A. Valiveti, J.B. Ames, NMR structure of DREAM: implications for Ca(2+)-dependent DNA binding and protein dimerization, *Biochemistry* 47 (2008) 2252–2264.
- [32] J. Maffie, B. Rudy, Weighing the evidence for a ternary protein complex mediating A-type K⁺ currents in neurons, *J. Physiol.* 586 (2008) 5609–5623.
- [33] C. Marionneau, R.D. LeDuc, H.W. Rohrs, A.J. Link, R.R. Townsend, J.M. Nerbonne, Proteomic analyses of native brain Kv4.2 channel complexes, *Channels (Austin)* 3 (2009) 284–294.
- [34] C.R. Marshall, A. Noor, J.B. Vincent, A.C. Lionel, L. Feuk, J. Skaug, M. Shago, R. Moessner, D. Pinto, Y. Ren, B. Thiruvahindrapuram, A. Fiebig, S. Schreiber, J. Friedman, C.E. Ketelaars, Y.J. Vos, C. Ficicioglu, S. Kirkpatrick, R. Nicolson, L. Sloman, A. Summers, C.A. Gibbons, A. Teebi, D. Chitayat, R. Weksberg, A. Thompson, C. Vardy, V. Crosbie, S. Luscombe, R. Baatjes, L. Zwaigenbaum, W. Roberts, B. Fernandez, P. Szatmari, S.W. Scherer, Structural variation of chromosomes in autism spectrum disorder, *Am. J. Hum. Genet.* 82 (2008) 477–488.
- [35] J.R. Martens, Y.G. Kwak, M.M. Tamkun, Modulation of Kv channel alpha/beta subunit interactions, *Trends Cardiovasc. Med.* 9 (1999) 253–258.
- [36] K. McCormack, J.X. Connor, L. Zhou, L.L. Ho, B. Ganetzky, S.Y. Chiu, A. Messing, Genetic analysis of the mammalian K⁺ channel beta subunit Kvbeta 2 (Kcnab2), *J. Biol. Chem.* 277 (2002) 13219–13228.
- [37] M.J. Morales, J.O. Wee, S. Wang, H.C. Strauss, R.L. Rasmusson, The N-terminal domain of a K⁺ channel beta subunit increases the rate of C-type inactivation from the cytoplasmic side of the channel, *Proc. Natl. Acad. Sci. U.S.A.* 93 (1996) 15119–15123.
- [38] Y. Morohashi, N. Hatano, S. Ohya, R. Takikawa, T. Watabiki, N. Takasugi, Y. Imaizumi, T. Tomita, T. Iwatsubo, Molecular cloning and characterization of CALP/KChIP4, a novel EF-hand protein interacting with presenilin 2 and voltage-gated potassium channel subunit Kv4, *J. Biol. Chem.* 277 (2002) 14965–14975.
- [39] A.J. Norris, N.C. Foeger, J.M. Nerbonne, Interdependent roles for accessory KChIP2, KChIP3 and KChIP4 subunits in the generation of Kv4-encoded I_A channels in cortical pyramidal neurons, *J. Neurosci.*, in press.
- [40] S. Pal, K.A. Hartnett, J.M. Nerbonne, E.S. Levitan, E. Aizenman, Mediation of neuronal apoptosis by Kv2.1-encoded potassium channels, *J. Neurosci.* 23 (2003) 4798–4802.
- [41] M.T. Perez-Garcia, J.R. Lopez-Lopez, C. Gonzalez, Kvbeta1.2 subunit coexpression in HEK293 cells confers O₂ sensitivity to kv4.2 but not to Shaker channels, *J. Gen. Physiol.* 113 (1999) 897–907.
- [42] R. Peri, B.A. Wible, A.M. Brown, Mutations in the Kv beta 2 binding site for NADPH and their effects on Kv1.4, *J. Biol. Chem.* 276 (2001) 738–741.
- [43] K. Petrecca, D.M. Miller, A. Shrier, Localization and enhanced current density of the Kv4.2 potassium channel by interaction with the actin-binding protein filamin, *J. Neurosci.* 20 (2000) 8736–8744.
- [44] M. Pioletti, F. Findeisen, G.L. Hura, D.L. Minor Jr., Three-dimensional structure of the KChIP1-Kv4.3 T1 complex reveals a cross-shaped octamer, *Nat. Struct. Mol. Biol.* 13 (2006) 987–995.
- [45] O. Pongs, Voltage-gated potassium channels: from hyperexcitability to excitement, *FEBS Lett.* 452 (1999) 31–35.
- [46] O. Pongs, J.R. Schwarz, Ancillary subunits associated with voltage-dependent K⁺ channels, *Physiol. Rev.* 90 (2010) 755–796.
- [47] G.M. Popowicz, M. Schleicher, A.A. Noegel, T.A. Holak, Filamins: promiscuous organizers of the cytoskeleton, *Trends Biochem. Sci.* 31 (2006) 411–419.
- [48] C.A. Reid, S.F. Berkovic, S. Petrou, Mechanisms of human inherited epilepsies, *Prog. Neurobiol.* 87 (2009) 41–57.
- [49] J. Rettig, S.H. Heinemann, F. Wunder, C. Lorra, D.N. Parcej, J.O. Dolly, O. Pongs, Inactivation properties of voltage-gated K⁺ channels altered by presence of beta-subunit, *Nature* 369 (1994) 289–294.
- [50] K.J. Rhodes, M.M. Monaghan, N.X. Barrezaeta, S. Nawoschik, Z. Bekele-Arcuri, M.F. Matos, K. Nakahira, L.E. Schechter, J.S. Trimmer, Voltage-gated K⁺ channel beta subunits: expression and distribution of Kv beta 1 and Kv beta 2 in adult rat brain, *J. Neurosci.* 16 (1996) 4846–4860.
- [51] A. Ruiz-Gomez, B. Mellstrom, D. Tornero, E. Morato, M. Savignac, H. Holguin, K. Aurrekoetxea, P. Gonzalez, C. Gonzalez-Garcia, V. Cena, F. Mayor Jr., J.R. Naranjo, G protein-coupled receptor kinase 2-mediated phosphorylation of downstream regulatory element antagonist modulator regulates membrane trafficking of Kv4.2 potassium channel, *J. Biol. Chem.* 282 (2007) 1205–1215.
- [52] L.A. Schrader, A.E. Anderson, A. Mayne, P.J. Pfaffinger, J.D. Sweatt, PKA modulation of Kv4.2-encoded A-type potassium channels requires formation of a supramolecular complex, *J. Neurosci.* 22 (2002) 10123–10133.
- [53] L.A. Schrader, A.E. Anderson, A.W. Varga, M. Levy, J.D. Sweatt, The other half of Hebb: K⁺ channels and the regulation of neuronal excitability in the hippocampus, *Mol. Neurobiol.* 25 (2002) 51–66.
- [54] L.A. Schrader, Y. Ren, F. Cheng, D. Bui, J.D. Sweatt, A.E. Anderson, Kv4.2 is a locus for PKC and ERK/MAPK cross-talk, *Biochem. J.* 417 (2009) 705–715.
- [55] G. Shi, K. Nakahira, S. Hammond, K.J. Rhodes, L.E. Schechter, J.S. Trimmer, Beta subunits promote K⁺ channel surface expression through effects early in biosynthesis, *Neuron* 16 (1996) 843–852.
- [56] R. Shibata, H. Misonou, C.R. Campomanes, A.E. Anderson, L.A. Schrader, L.C. Doliveira, K.I. Carroll, J.D. Sweatt, K.J. Rhodes, J.S. Trimmer, A fundamental role for KChIPs in determining the molecular properties and trafficking of Kv4.2 potassium channels, *J. Biol. Chem.* 278 (2003) 36445–36454.
- [57] M.B. Thomsen, C. Wang, N. Ozgen, H.G. Wang, M.R. Rosen, G.S. Pitt, Accessory subunit KChIP2 modulates the cardiac L-type calcium current, *Circ. Res.* 104 (2009) 1382–1389.
- [58] A. Ueda, C.F. Wu, Effects of hyperkinetic, a beta subunit of Shaker voltage-dependent K⁺ channels, on the oxidation state of presynaptic nerve terminals, *J. Neurogenet.* 22 (2008) 1–13.
- [59] M.A. van Es, P.W. van Vught, H.M. Blauw, L. Franke, C.G. Saris, L. Van den Bosch, S.W. de Jong, V. de Jong, F. Baas, R. van't Slot, R. Lemmens, H.J. Schelhaas, A. Birve, K. Slegers, C. Van Broeckhoven, J.C. Schymick, B.J. Traynor, J.H. Wokke, C. Wijmenga, W. Robberecht, P.M. Andersen, J.H. Veldink, R.A. Ophoff, L.H. van den Berg, Genetic variation in DPP6 is associated with susceptibility to amyotrophic lateral sclerosis, *Nat. Genet.* 40 (2008) 29–31.
- [60] M.A. van Es, P.W. van Vught, G. van Kempen, H.M. Blauw, J.H. Veldink, L.H. van den Berg, Dpp6 is associated with susceptibility to progressive spinal muscular atrophy, *Neurology* 72 (2009) 1184–1185.
- [61] A.W. Varga, L.L. Yuan, A.E. Anderson, L.A. Schrader, G.Y. Wu, J.R. Gatchel, D. Johnston, J.D. Sweatt, Calcium-calmodulin-dependent kinase II modulates Kv4.2 channel expression and upregulates neuronal A-type potassium currents, *J. Neurosci.* 24 (2004) 3643–3654.
- [62] L. Wang, K. Takimoto, E.S. Levitan, Differential association of the auxiliary subunit Kvbeta2 with Kv1.4 and Kv4.3 K⁺ channels, *FEBS Lett.* 547 (2003) 162–164.
- [63] J. Weng, Y. Cao, N. Moss, M. Zhou, Modulation of voltage-dependent Shaker family potassium channels by an aldo-keto reductase, *J. Biol. Chem.* 281 (2006) 15194–15200.
- [64] E.K. Yang, M.R. Alvira, E.S. Levitan, K. Takimoto, Kvbeta subunits increase expression of Kv4.3 channels by interacting with their C termini, *J. Biol. Chem.* 276 (2001) 4839–4844.
- [65] Y. Zhang, P. Su, P. Liang, T. Liu, X. Liu, X.Y. Liu, B. Zhang, T. Han, Y.B. Zhu, D.M. Yin, J. Li, Z. Zhou, K.W. Wang, Y. Wang, The DREAM protein negatively regulates the NMDA receptor through interaction with the NR1 subunit, *J. Neurosci.* 30 (2010) 7575–7586.