

Decreased glutamate transporter (*GLT-1*) expression in frontal cortex of rats with acute liver failure

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Abstract

It has been suggested that reduced astrocytic uptake of neuronally released glutamate contributes to the pathogenesis of hepatic encephalopathy in acute liver failure. In order to further address this issue, the recently cloned and sequenced astrocytic glutamate transporter GLT-1 was studied in brain preparations from rats with ischemic liver failure induced by portacaval anastomosis followed 24 h later by hepatic artery ligation and from appropriate sham-operated controls. *GLT-1* expression was studied using reverse transcriptase-polymerase chain reaction (RT-PCR). Expression of *GLT-1* transcript was significantly decreased in frontal cortex at coma stages of acute liver failure. Western blotting using a polyclonal antibody to *GLT-1* revealed a concomitant decrease in expression of transporter protein in the brains of rats with acute liver failure. Reduced capacity of astrocytes to reuptake neuronally released glutamate, resulting from a GLT-1 transporter deficit and the consequently compromised neuron-astrocytic trafficking of glutamate could contribute to the pathogenesis of hepatic encephalopathy and brain edema, two major complications of acute liver failure. © 1997 Elsevier Science Ireland Ltd.

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Acute liver failure resulting from viral infections or ingestion of toxic substances results in severe neurological impairment progressing through stupor and coma within hours or days. The pathophysiologic mechanisms responsible for hepatic encephalopathy (HE) in acute liver failure are unknown but deficits of neurotransmission rather than primary cerebral energy failure are currently considered to be implicated [1,3]. In this regard, there is increasing evidence to suggest that abnormalities of the glutamate neurotransmitter system are involved in the pathogenesis of HE in acute liver failure. Brain concentrations of glutamate are reduced in both human [13] and experimental [7,17] acute liver failure and recent studies using in vivo cerebral microdialysis in both rats [2,9] and rabbits [5] with acute (ischemic) liver failure have consistently revealed increased extracellular brain concentrations of glutamate. It was suggested that the increased extracellular concentrations of glu-

tamate in the brain of these animals was the consequence of decreased astrocytic uptake [9]. The major high affinity astrocytic transporter for glutamate, GLT-1, has recently been cloned and sequenced [12,14]. In order to directly assess glutamate uptake capacity in the brain in acute liver failure, GLT-1 gene and protein expression were measured in frontal cortex of rats with acute (ischemic) liver failure.

Male, Sprague-Dawley rats (175–200 g) were anesthetized with halothane and an end-to-side portacaval shunt (SH) was performed according to the guidelines of Lee and Fisher [6]. Sham-operated control rats (SM) were anesthetized and underwent a laparotomy and clamping of the portal vein. 24 h later, the same rats were reanesthetized with halothane and subjected to hepatic artery ligation (HAL) or laparotomy (SM). In this way, four groups of surgically operated animals were created with four rats in each group, namely acute liver failure (SH + HAL) and the three obligate control groups SH + SM, SM + HAL and SM + SM. After surgery, arterial blood glucose levels

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were monitored and supplemental glucose was administered intraperitoneally, as needed. Control animals received an equivalent volume of 0.9% saline. Body temperature was maintained at 37°C.

Approximately 10–12 h after surgery, animals lost their righting reflex followed 2–3 h later by loss of corneal reflex (coma). At coma stages of ischemic liver failure, rats in acute liver failure together with their appropriate controls were killed by decapitation and brains were removed and stored at –80°C until use.

GLT-1 expression was quantitated by RT-PCR using purified total RNA from frontal cortex of animals from the four treatment groups (Fig. 1). Total RNA was extracted using the Trizol reagent (Life Technologies Inc., Gaithersburg, MD, USA) according to the manufacturer's protocol. DNA contamination of the RNA samples was removed by incubation with RNase-free DNase. Quantification of expression by RT-PCR requires a linear ratio between cycle number and resulting DNA amount. Thus, conditions for expression studies for each gene were performed. Glyceraldehyde-3-phosphate dehydrogenase (*GAPDH*) was used as internal standard to evaluate RNA samples and to correct for loading variations. Optimal conditions were obtained after 22 amplification cycles for *GLT-1* and 17 for *GAPDH* using 0.5 µg of RNA per amplification. The monospecificity of the oligonucleotide primers was verified by computer analyses using the PRIMER program (Genetics Computer Group, Madison, WI). Reverse transcription of mRNA and PCR amplification of resulting cDNA molecules were performed in a one-step reaction. The reaction mixture in a final volume of 50 µl contained 0.5 µg of the total RNA, which was incubated in 10 mM Tris–HCl (pH 8.3), 50 mM KCl, 0.1% (w/v) BSA, 1 mM MgCl₂, 200 µM dNTPs, 10 µCi/ml [α -³²P]dCTP (3000 µCi/mmol), 320 U/ml of RNasin, 100 U/ml of AMV reverse transcriptase, 20 U/ml of Taq DNA polymerase and the primers either for *GLT-1* (1 µM of each forward and reverse, respectively) or

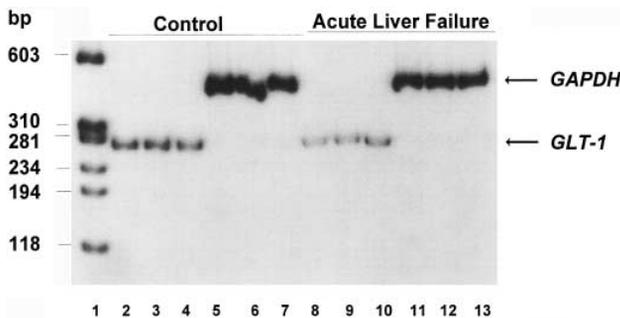


Fig. 1. Expression of the astrocytic glutamate transporter *GLT-1* and *GAPDH* in frontal cortex of rats with acute liver failure versus control (sham operation). Total RNA was extracted from the brain of 3 animals with acute liver failure at coma stages of encephalopathy or at comparable times following sham operations. Lane 1, molecular weight standards; lanes 2–7, *GLT-1* and *GAPDH* expression in sham-operated control rats; lanes 8–13, *GLT-1* and *GAPDH* expression in rats with acute liver failure.

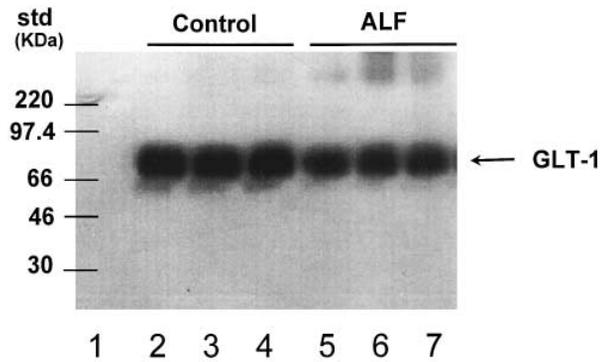


Fig. 2. *GLT-1* protein expression in frontal cortex of rats at coma stage of acute liver failure (ALF, lanes 5–7) and sham-operated controls (lanes 2–4). Lane 1, molecular weight markers.

GAPDH (0.5 µM of each). The mRNA was reverse transcribed at 50°C for 10 min followed by PCR including a denaturing step at 95°C for 30 s, primer annealing at 62°C for 1 min and DNA-elongation at 72°C for 1.5 min. Using these primer sets the amplification of *GLT-1*-mRNA resulted in a 285 bp fragment, whereas the *GAPDH*-product was 470 bp in length. The PCR products were separated on an 8% polyacrylamide gel which was then apposed to film for visualization. The amplified *GLT-1* and *GAPDH*-cDNA-fragments were subsequently excised from the gel and Cerenkov radiation was quantitated using a β -counter.

For the study of *GLT-1* protein, brain tissue was homogenized in 50 mM Tris–HCl (pH 7.4), 300 mM NaCl, 0.1% Tween (Fig. 2). After centrifugation (10 000 \times g, 15 min) the supernatant was separated by a denaturing SDS-PAGE (7.5%) as previously described [14]. Blotting was performed with the semi-dry transfer cell from Bio-Rad at 15 V (1 A) for 45–60 min using ECL-nitrocellulose and Whatmann paper (3MM) each presoaked in transfer buffer (48 mM Tris (pH 8.3), 39 mM glycine, 0.037% SDS, 20% methanol) for at least 15 min. After the transfer the membrane was briefly rinsed in 1 \times PBS/0.1% Tween-20 and air dried. For immunological protein detection the membrane was incubated in blocking buffer (10% horse serum, 5% nonfat dried milk, 1% BSA, 1 \times PBS/0.1% Tween-20) for 1 h followed by incubations with the antisera against *GLT-1* and secondary antibodies (Amersham: 1:4000 diluted anti-rabbit IgG horseradish peroxidase-coupled) each in blocking buffer for 1 h. Subsequent to each incubation with the primary and secondary antibody, blots were washed three times (1 \times 20 min and 2 \times 5 min) with 1 \times PBS/0.1% Tween-20. For the detection of specific antibody binding the membranes were treated in accordance with the Amersham ECL-Kit instructions and apposed to photosensitive ECL-hybrid film for 5–60 s. Signal intensities were measured with a densitometer.

Results of these studies reveal significant reductions in *GLT-1* protein (by 31%, $P < 0.05$) and in *GLT-1* mRNA (by 60%, $P < 0.01$) in frontal cortex of rats with ischemic liver failure at coma stages of encephalopathy compared to SM + SM, SM + HAL or SH + SM control groups of ani-

mals (Figs. 1 and 2). These findings add to a growing body of evidence suggesting that acute liver failure results in altered glutamatergic synaptic regulation in brain. Previous studies using in vivo cerebral microdialysis demonstrated increased extracellular glutamate concentrations in frontal cortex [9] as a function of the deterioration of neurological status and of arterial ammonia concentrations in the same animal model of acute liver failure as that used in the present study. Such increases could result directly from diminished capacity for astrocytic uptake of glutamate as a consequence of decreased expression of GLT-1. Reduced expression of GLT-1 and the consequently increased extracellular brain glutamate concentrations also offers a possible explanation for the observed decrease in densities (down regulation) of AMPA/kainate binding sites also described in this model of acute liver failure [10]. Diminished uptake of neuronally released glutamate by the astrocyte as a consequence of decreased expression of GLT-1 would therefore constitute further evidence of impaired neuron-astrocytic trafficking of glutamate, a phenomenon which has previously been proposed to contribute to the pathogenesis of hepatic encephalopathy based on neurochemical studies [4] and by ^{13}C -NMR spectroscopy [16] in portacaval-shunted rats.

Impaired removal of glutamate from brain extracellular space due to decreased *GLT-1* expression could also be implicated in the pathogenesis of cytotoxic brain edema, a major complication of acute liver failure in both humans and in the experimental animal model used in the present study [17]. Exposure of cultured astrocytes to glutamate leads to significant cell swelling [15]. Finally, it is conceivable that, under certain circumstances, reduction in the capacity for glutamate uptake by astrocytes as a result of decreased GLT-1 expression could impair ammonia removal by brain. Glutamine synthetase, the enzyme primarily responsible for ammonia detoxification by brain has a predominantly astrocytic localization [11] and a recent study provides evidence to suggest that the metabolic fate of glutamate in astrocytes (i.e. conversion to glutamine versus oxidation in the tricarboxylic acid cycle) is regulated by extracellular glutamate concentrations [8].

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