



## Review

## Peripheral analgesia: Hitting pain where it hurts

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## ABSTRACT

Pain is a complex biological phenomenon that encompasses intricate neurophysiological, behavioural, psychosocial and affective components. Protracted or chronic pain alerts an individual to a possible pathological abnormality and is the main reason why patients visit a primary care physician. Despite the pervasiveness of chronic pain in the population, the effectiveness of current pharmacological therapies remains woefully inadequate and prolonged treatment often leads to the development of undesirable side-effects. Since the vast majority of chronic pain originates in a specific tissue or group of tissues, it may be advantageous to target pain control in the periphery and thereby circumvent the known risks associated with non-specific systemic treatments. This review spotlights a number of promising targets for peripheral pain control including the transient receptor potential (TRP) family of neuronal ion channels, the family of proteinase activated receptors (PARs), cannabinoids, and opioids. A critical appraisal of these targets in preclinical models of disease is given and their suitability as future peripheral analgesics is discussed.

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As most of us can attest, pain is an unpleasant sensory experience which instils negative feelings and as such is something we generally try to avoid. Pain can either be short-lasting (acute) or more protracted (chronic). Acute pain is typically beneficial to the individual as it warns of actual or impending tissue damage allowing rapid, reflex evasive action to be carried out. This nociceptive pain arises in the periphery via direct activation of pain-sensing nerve terminals by noxious mechanical, thermal or chemical stimuli. Acute pain can usually be relieved by local administration of pharmacological agents (e.g. lidocaine, non-steroidal anti-inflammatory drugs) or non-pharmacological approaches (e.g. rubbing the affected area or application of ice). Chronic pain, on the other hand, tends to be a maladaptive response to some underlying pathology. Pain is considered chronic if it continues unabated beyond about 3 months and typically outlasts the normal healing response or continues in the absence of any observable tissue damage [1]. Chronic pain is the primary reason why people seek medical attention, yet the arsenal of available effective remedies is limited. Furthermore, all of the currently available analgesics have some degree of negative side-effect associated with them. The time has come to develop more effective, safe and consistent analgesics.

Firstly an overview of the classic pain pathway is provided. Most types of pain begin in the periphery with a noxious stimulus activating unspecialised free nerve endings associated with small diameter primary afferent neurones (Fig. 1). This physical stimulus is subsequently transduced into an electrochemical signal which is

transmitted from the periphery towards the central nervous system along slowly conducting primary afferent nerves. Upon entering the dorsal horn of the spinal cord, these impulses are transmitted via chemical synapses to second order neurones where they are subsequently conveyed along ascending tracts to higher centres in the brain. By relaying these impulses to specific areas of the brain such as the somatosensory cortex and the amygdala, the electrochemical signals are shaped into a psychophysical experience incorporating any protective motor reflexes and emotional responses. The critical details of this entire process are still inadequately understood and form the basis of a fascinating area of neuroscientific research.

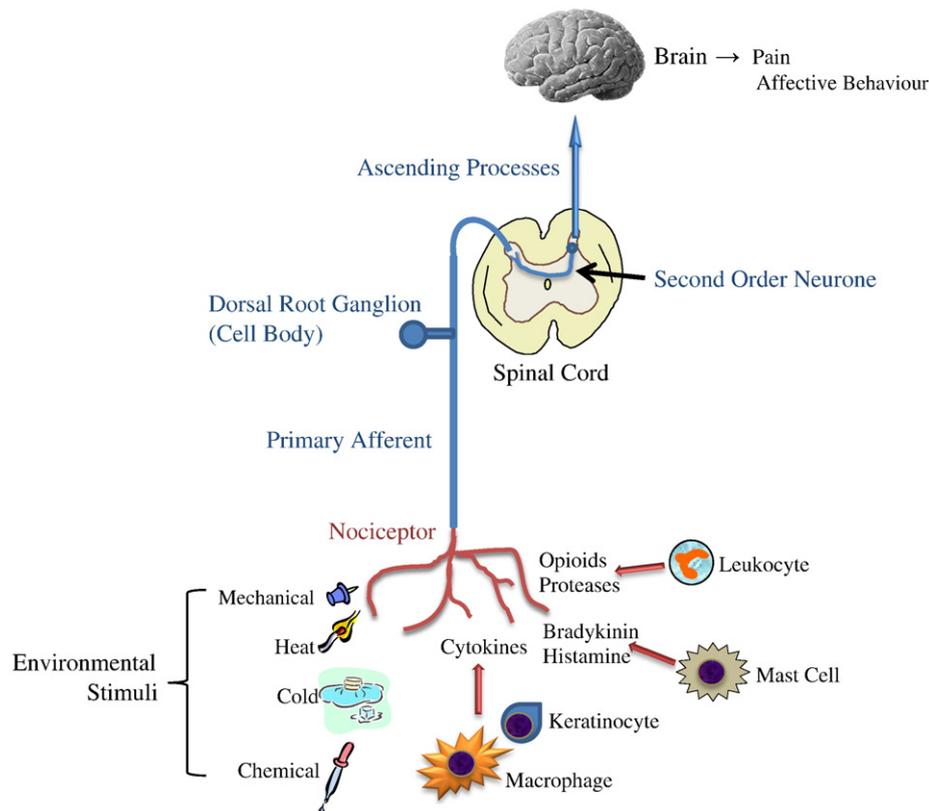
Current pharmacological agents attempt to tackle pain in both the central as well as the peripheral divisions of the pain pathway. Since centrally acting analgesics tend to produce numerous secondary unwanted side-effects (e.g. dysphoria, motor deficits and addiction) it stands to reason that peripherally acting drugs have the advantage of circumventing these problems by targeting pain at the source. This review will address these issues and highlight the palpable benefits of peripherally-directed analgesia. While the peripheral mediators covered here are by no means exhaustive, the intent is to highlight emerging and possibly contentious targets worthy of further discussion.

### 1. Transient receptor potential ion channels

In an era of molecular cloning technology, a large number of cation channels have emerged belonging to the transient receptor potential (TRP) family. Identification of these ion channels on polymodal nociceptors triggered intense interest in the pain research community, many of whom consider TRP channels to be central integrators of

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**Fig. 1.** The pain pathway from periphery to brain. Noxious environmental stimuli can activate peripheral nociceptors leading to the sensation of pain. Various cells residing in the vicinity of the nerve terminal can release mediators which either heighten or reduce the activation threshold of the nerve.

pain transduction and modulation. Indeed, TRP channels have been linked with all aspects of sensory physiology including mechanosensation, thermosensation, hearing, vision and taste. The latter sensory modality prompted a slew of molecular biology studies and at one point it felt like just about everything in the kitchen larder activated a TRP channel. It is plain to see why these prevalent and precocious ion channels have garnered favour as targets of interest by academia and the pharmaceutical industry alike. A summary of the most commonly studied TRP channels is shown in Table 1.

The channel which has received the greatest attention is the TRP-vanilloid 1 (TRPV1) channel. Cloned in 1997 [2], TRPV1 is known to be activated by protons, temperatures in excess of 42 °C, endovanilloids, cannabinoids, and a host of other chemical mediators known to be present in the “inflammatory brew”. The promiscuous nature of the TRPV1 channel renders it a central modulator of pro-algesic stimuli from disparate sensory modalities. Consequently, several small molecule TRPV1 antagonists are currently being assessed for potential analgesic properties in various painful diseases. In joints, intra-articular injection of the TRPV1 agonist capsaicin was found to cause synovial vasodilatation [3] which can be blocked by the selective TRPV1 antagonist SB366791 [4]. Thus, a role for TRPV1 in arthritic processes was established. This was later confirmed by the observation that the severity of inflammatory joint disease was significantly less in TRPV1 knockout mice compared to wild-types [5,6]. Pain behaviour studies found that the tactile allodynia associated with Freund’s complete adjuvant-induced arthritis was less in TRPV1 deficient mice compared to genetically normal controls [6,7]. Similarly, systemic treatment of osteoarthritic rats with the TRPV1 antagonist A-425619 reduced the weight bearing deficit associated with this disease [8].

In the gastrointestinal system, TRPV1 contributes to visceral hypersensitivity making it an intriguing target for intractable abdominal pain syndromes. Intestinal biopsies taken from subjects undergoing routine colonoscopy revealed that patients with inflam-

matory bowel syndrome had over three times as many TRPV1-positive nerve fibres compared to control subjects [9]. These additional TRPV1-positive afferents can then be sensitized by peripherally circulating inflammatory mediators released from enteric nerves, endothelial cells, and immunocytes. Data supporting a neuroimmune component of peripheral sensitization has recently been reported [10]. Supernatants of peripheral blood mononuclear

**Table 1**

Some of the commonly studied TRP channels in pain research and their activators/blockers.

TRP channel	Activator	Temperature sensitivity	Selective channel blocker
TRPV1	Protons Vanilloids Ethanol	43–52 °C	SB366791 Capsazepine
TRPV2	$\Delta^9$ -Tetrahydrocannabinol 2-aminoethoxydiphenyl borate Mechanical swelling	$\geq 52$ °C	None
TRPV3	Eugenol 2-aminoethoxydiphenyl borate Vanillin Camphor	32–39 °C	None
TRPV4	4 $\alpha$ -Phorbol 12,13-didecanoate Epoxyeicosatrienoic acids Mechanical	25–34 °C	HC-067047 RN-1734
TRPA1	Cinnamaldehyde Allicin Acrolein Formalin Mustard oil	17 °C	AP18 HC-030031
TRPM8	Menthol Eucalyptol Spearmint Icilin	18–24 °C	5-Benzyloxytryptamine

cells taken from patients with post-infectious inflammatory bowel syndrome caused increased firing of mouse visceral primary afferents. This observation suggests that during inflammatory bowel syndrome, immunocytes can locally release mediators that sensitize colonic nociceptors leading to intestinal pain.

Having established TRPV1 as an integrator of peripheral pain processes, a number of small molecule TRPV1 antagonists rapidly emerged and touted as potential novel analgesics. Subsequent tests with small molecule TRPV1 antagonists, however, revealed that some of these pharmacological agents produced profound hyperthermia [11,12] and as such could not be used therapeutically. Later studies found that the primary site of TRPV1 antagonist-induced hyperthermia was the periphery and that targeting TRPV1 channels in the central nervous system may be more beneficial for pain control while avoiding thermoregulatory complications [13,14]. Some researchers claim to have discovered TRPV1 antagonists with differential pharmacology i.e. compounds which inhibit TRPV1-induced pain but have no effect on core body temperature control [15]. Since TRPV1 is activated by a large array of sensory stimuli and chemical agents, it remains to be seen whether it is possible to isolate the pain regulatory aspect of TRPV1 without evoking undesirable side-effects.

Another vanilloid channel receiving increasing attention is the TRPV4 channel. It is expressed on sensory free nerve endings [16] and is known to be activated by moderate temperatures (25–34 °C), mechanical stimuli [17], and chemical ligands such as 4 $\alpha$ -phorbol 12,13 didecanoate (4 $\alpha$ -PDD) and epoxyeicosatrienoic acids [18,19]. The identification of TRPV4 channels on cochlear hair cells and vibrissal Merkel cells gave the first indication that this particular TRP channel may be involved in mechanosensation [20]. Subsequent studies showing TRPV4 expression on A $\delta$  and C fibres in mouse skin highlighted the possibility that TRPV4 could be involved in peripheral mechanonociception [16]. In fact, TRPV4 knockout mice have a higher threshold to noxious mechanical stimuli compared to wildtype animals [21]. In the gut, TRPV4 is co-localised with inflammatory neuropeptides in small diameter sensory neurones and administration of a TRPV4 agonist greatly enhances visceral mechanosensation [22]. Furthermore, behavioural responses to noxious colonic distension were inhibited in animals lacking TRPV4 cation channels [22].

Moving from hot to cold, the first channel found to be responsive to lower temperatures was found to be expressed by a subset of TRPV1 expressing polymodal nociceptors and due to its large number of ankyrin repeating motifs was named TRPA1 [23,24]. TRPA1 channels are believed to be activated by moderate temperatures (approximately 17 °C) and their expression has been shown to be increased in dorsal root ganglia (DRG) ipsilateral to Freund's complete adjuvant-induced inflammation [25] or nerve injury [26]. Mice missing functional TRPA1 channels show reduced thermal and mechanical pain responses to intraplantar injection of bradykinin or allylthiocyanate [27,28]. Peripheral injection of the TRPA1 antagonist AP18 [(Z)-4-(4-chlorophenyl)-3-methylbutyl-3-en-2-oxime] significantly reduced mechanical hyperalgesia and cold allodynia in a model of chronic inflammation [29]. Another TRPA1 antagonist HC-030031 [(2-[1,3-dimethyl-2,6-dioxo-1,2,3,6-tetrahydro-7H-purin-7yl)-N-(4-isopropylphenyl)acetamide] dose-dependently reduced flinching in the formalin model of chemonociception [30] as well as mechanonociception in Freund's complete adjuvant model of inflammatory pain and in the spinal nerve ligation model of neuropathic pain [31]. More recently, mechanical hyperalgesia produced by either intra-plantar administration of tumour necrosis factor- $\alpha$  or intra-articular injection of Freund's complete adjuvant was attenuated by AP18 suggesting that TRPA1 channels are involved in inflammatory and arthritic pain [32].

Another ion channel known to be involved in cold sensation belongs to the melastatin family of TRP channels and was termed TRPM8 [33,34]. Triggered by temperatures in the range 18–24 °C, as well as other cooling compounds such as menthol, eucalyptol and icilin, TRPM8 is expressed by about 15% of small diameter DRG and

trigeminal neurones [33–35]. Being a non-selective cation channel, stimulation of TRPM8 channels leads to membrane depolarization and afferent nerve firing. Involvement of TRPM8 channels in cold hypersensitivity is not yet certain although the emerging evidence is highly suggestive of the involvement of TRPM8 in cold pain processing. In several neuropathic and inflammatory pain models, TRPM8 expression is increased and the channel offsets thermal as well as mechanical hypersensitivity although it is unclear whether this is achieved by TRPM8 activation or blockade [26,36,37]. Experiments examining acute inflammatory pain induced by intraplantar injection of Freund's complete adjuvant found that pain was inhibited by peripheral TRPM8 agonism [36]. Similarly, neuropathic pain elicited by chronic constriction injury to the sciatic nerve or peripheral nerve demyelination using lysolecithin could be ameliorated by topical cooling, menthol and icilin [36]. Katsura et al., however, found that antisense inactivation of central TRPM8 channels had no effect on L5 spinal nerve ligation-induced neuropathic pain [25]. The role of TRPM8 in pain modulation still requires further analysis; however, some of the controversy could be related to whether the agonists/antagonists are administered centrally versus peripherally leading to differential responses.

## 2. Proteinase activated receptors

Proteinases hydrolyse the peptide bonds in long chain proteins and are involved in multiple physiological processes such as tissue degradation, apoptosis, the blood clotting cascade and tissue remodelling. Further to this classic enzymatic activity, proteinases can also act as signalling molecules by triggering a small group of G-protein coupled receptors. There are currently four members of these proteinase activated receptors (PAR<sub>1–4</sub>) and they are expressed in a plethora of organs such as joints, skin, gut and lungs. Unlike other G-protein coupled receptors which rely on the binding of a specific chemical ligand to cause receptor stimulation, PARs are activated by a proteinase causing a change in receptor conformation. Tethered to the extracellular N-terminal loop of the receptor there is a short amino acid sequence which has the potential to stimulate the same receptor but is impeded from doing so by an amino acid cap. A locally released proteinase cleaves the receptor at a specific point thereby removing the cap and exposing the functional amino acid sequence which can now bind to its complementary domain on the extracellular N-terminal loop of the receptor rather like the hammer of a gun engaging with a cartridge. The stimulated receptor then initiates a series of second messenger cascades leading to intracellular signalling. Experimentally, it is possible to activate the PARs with a small peptide agonist which matches the tethered ligand sequence (see Table 2 for a list of the various PAR activating peptides). This approach has the benefit of selectively stimulating the PAR of interest without possible off target effects that would undoubtedly be initiated by addition of the proteinase.

With the exception of PAR<sub>3</sub>, all of the PARs have been implicated in the control of peripheral nociception. PAR<sub>1</sub> is primarily activated by thrombin and plays an important role in cardiovascular disease and inflammation [38]. Recent evidence has shown that PAR<sub>1</sub> is expressed by mouse DRG and treating these sensory neuronal cell bodies with a PAR<sub>1</sub> agonist leads to calcium mobilization [39]. Local injection of the PAR<sub>1</sub> agonist TFLR-NH<sub>2</sub> dose-dependently blocks hyperalgesia in both acute and chronic models of inflammation [39–41]. It is believed that PAR<sub>1</sub> achieves anti-nociception by causing the secondary release of endogenous opioids since PAR<sub>1</sub> responses are attenuated by the non-specific opioid antagonist naloxone [39]. Interestingly, PAR<sub>1</sub> knockout mice respond normally to noxious thermal and mechanical stimuli suggesting that PAR<sub>1</sub> is only active during inflammation and does not affect physiological pain [39].

PAR<sub>2</sub> has been localised on sensory nerves and is primarily activated by trypsin and mast cell tryptase resulting in the release of

**Table 2**  
Summary of PAR modulators and the role of PARs in pain.

PAR	Activating peptide	Inactive peptide	Antagonist	Role in pain
PAR1	TFLLR-NH <sub>2</sub>	FTLLR-NH <sub>2</sub>	RWJ-58259 SCH-205831 SCH-5303048	↓ Pain [36–38]
PAR2	SLIGRL-NH <sub>2</sub> 2-Furoyl- LIGRLO-NH <sub>2</sub>	LSIGRL-NH <sub>2</sub> 2-Furoyl- OLRGIL-NH <sub>2</sub>	ENMD-1068	↑ Pain [40–42]
PAR3	None	None	None	Unknown
PAR4	AYPGKF-NH <sub>2</sub>	YAPGKF-NH <sub>2</sub>	Pepducin p4Pal-10	↑ Pain joints [45,46] ↓ Pain skin, gut [49]

allogenic neuropeptides such as substance P and calcitonin gene-related peptide [42]. As such, PAR<sub>2</sub> is considered to be a pronociceptive receptor whose activation causes increased pain in joints [43], gut [44], and hindpaws [45]. The mechanism by which PAR<sub>2</sub> promotes pain appears to involve capsaicin-sensitive TRPV1 ion channels. Evidence supporting this concept comes from the fact that the PAR<sub>2</sub> agonist SLIGRL-NH<sub>2</sub> does not cause secondary hyperalgesia in TRPV1 knockout animals and PAR<sub>2</sub>-mediated nociception can be blocked by the TRPV1 receptor antagonist SB366791 [43]. Furthermore, Grant et al. found that TRPV4 ion channels are sensitised by PAR<sub>2</sub> agonists and PAR<sub>2</sub>-induced mechanical hyperalgesia is absent in TRPV4 knockout mice and attenuated by the TRPV4 antagonist 4 $\alpha$ PDD [46]. Mast cells have also been implicated in PAR<sub>2</sub> responses as the receptor has been localised on the surface of these cells and PAR<sub>2</sub> activation leads to mast cell degranulation [45,47]. The endogenous proteinases released during inflammation and which activate PAR<sub>2</sub> *in vivo* leading to pain have yet to be fully ascertained. Serine protease levels are known to be elevated in the synovial fluid of arthritis patients [48,49] and in the faeces from inflammatory bowel syndrome patients [50]. In the latter study, faecal supernatants taken from inflammatory bowel syndrome patients caused colono-rectal mechanical allodynia in wildtype mice which was less pronounced in PAR<sub>2</sub> knockout animals. This finding suggests that enzymatic mediators released during inflammatory bowel syndrome have the potential to sensitize visceral afferents leading to pain.

Recent studies have demonstrated PAR<sub>4</sub> expression in rat DRG [51] as well as in neural elements associated with joints [51,52] and the bladder [53]. Immunolocalisation and reverse transcription-polymerase chain reaction studies on rat cultured DRG found that PAR<sub>4</sub> is co-localised with the inflammatory neuropeptides substance P and calcitonin gene-related peptide [54]. The role of PAR<sub>4</sub> in nociceptive processing is still unclear with conflicting reports of hyperalgesia and analgesia. Peripheral administration of the PAR<sub>4</sub> agonist AYPGKF-NH<sub>2</sub> to knee joints, for example, results in increased mechanosensitivity of joint afferents and a reduction in pain threshold [51,52]. This pronociceptive effect of PAR<sub>4</sub> activation can be blocked by the bradykinin B<sub>2</sub> antagonist HOE140 but not by TRPV1 antagonism. In contrast, intraplantar and intracolonic injection of AYPGKF-NH<sub>2</sub> caused a reduction in nociception by deactivating the pain promoting effects of the TRPV4 channel [55]. Furthermore, fecal supernatants taken from patients suffering from ulcerative colitis produced visceral hyposensitivity in mice which was found to be due to the actions of the serine proteinase cathepsin G on PAR<sub>4</sub> [56]. Thus, the effect of PAR<sub>4</sub> activation on peripheral pain modulation appears to be organ specific and suggests the possible existence of PAR<sub>4</sub> subtypes (e.g. PAR<sub>4a</sub>, PAR<sub>4b</sub>, etc.) This hypothesis requires further investigation and characterisation.

### 3. Cannabinoids

Cannabinoids are highly lipophilic alkaloids derived from the hemp plant *Cannabis sativa*. At least 66 distinct chemicals have been

identified in *C. sativa* and these plant-borne agents are collectively called phytocannabinoids. There is a growing collection of man-made cannabinoids (synthetocannabinoids) which are based on the structure of their plant counterparts. Finally, it has been found that the body produces its own natural cannabinoids called endocannabinoids of which anandamide and 2-arachidonylglycerol are the most prominent.

Cannabinoids bind to two G protein-coupled receptor subtypes (CB<sub>1</sub> and CB<sub>2</sub>) with a third putative cannabinoid receptor (GPR55) having recently been described [57]. The CB<sub>1</sub> receptor was originally identified in the central nervous system [58] where it is believed to be the most abundant G protein-coupled receptor [58–60]. The CB<sub>2</sub> receptor was later identified on peripherally circulating macrophages and in the spleen [61]. Originally thought to be restricted to the periphery, CB<sub>2</sub> receptors have also been localised in the brainstem where they regulate emesis [62]. CB<sub>1</sub> receptors are present throughout the pain pathway including on peripheral neurones, spinal neurones and in pain processing areas of the brain [63–66]. Stimulation of neuronal CB<sub>1</sub> receptors by a cannabinoid ligand leads to hyperpolarisation and a decrease in calcium dependent neurotransmitter release. Since CB<sub>2</sub> receptors are relatively scarce in the central nervous system, they were originally thought to be an ideal target for pain control as selective CB<sub>2</sub> receptor agonists would not be able to produce the psychotropic effects commonly associated with CB<sub>1</sub> receptor activation. The reality has been less impressive with a number of CB<sub>2</sub> receptor agonists having low selectivity and producing multiple off-target responses. Some of the pharmacological tools used in cannabinoid research are listed in Table 3.

A plethora of cultures have used phytocannabinoids for millennia to treat a whole host of illnesses. Public acceptance of cannabinoids as a viable means of treating disease has been slow to catch on despite a significant body of evidence supporting a role for cannabinoids in the safe treatment of various chronic pain disorders. In joints, for example, the CB<sub>1</sub> receptor has been identified on nerve terminals innervating the synovium [67] and peripheral administration of the selective CB<sub>1</sub> receptor agonist arachidonyl-2-chloroethylamide (ACEA) dramatically reduced nociception in a rat model of osteoarthritis [68]. Local injection of the CB<sub>1</sub> receptor antagonist AM251 into arthritic joints led to an increase in nociception activity suggesting that endocannabinoids are released into the joint to help offset articular pain [68]. Other studies looking at models of inflammatory joint disease have found that both CB<sub>1</sub> and CB<sub>2</sub> receptors are involved in mediating articular analgesia [69–71]. Thus, there is a strong rationale for putting cannabis in our joints. The mechanism by which cannabinoids achieve anti-nociception in joints appears to involve TRPV1 ion channels [68,72,73]. During inflammation, TRPV1 channels

**Table 3**  
Overview of some of the commonly used cannabinoid reagents and their target receptor.

	Cannabinoid receptor		
	CB <sub>1</sub>	CB <sub>2</sub>	GPR55
Non-selective agonists	WIN55,212-2 CP55,940 HU210		N/A
Full agonists	Arachidonyl-2-chloroethylamide (ACEA) O1812 Methanandamide Arachidonyl-cyclopropylamide	AM1241 JWH015 JWH133 GW405833 HU308 L-759,633 L-759,656	O1602
Selective antagonists	AM251 AM281 SR141716A LY320135	AM630 SR144528	O1908

are upregulated [74,75] and TRPV1/ CB<sub>1</sub> receptor co-expression is enhanced [76]. In inflamed tissue, the TRPV1 channel exists in a phosphorylated state [77] rendering it more sensitive to endocannabinoids such as anandamide [78–80]. Endocannabinoids then bind directly to either TRPV1 channels or TRPV1/ CB<sub>1</sub> receptor heterodimers leading to channel deactivation and pain relief.

Cannabis has been used as an adjunct to chemotherapy in cancer patients due to its anti-emetic effects and palliation properties. A seminal study by Noyes et al. found that low dose Δ-9 tetrahydrocannabinol was equipotent to codeine in alleviating pain in cancer patients, although higher doses produced psychotropic side-effects such as ataxia, dizziness and blurred vision [81]. In pancreatic cancer patients, it was noted that peripheral CB<sub>1</sub> receptor density was inversely proportional to subjective pain scores suggesting that cannabinoid receptors may be involved in controlling cancer pain [82]. Animal studies also support a role for cannabinoids in reducing cancer pain. The non-selective synthetocannabinoids WIN55,212-2 and CP55,940 were found to reduce bone cancer pain and this effect could be blocked by a CB<sub>1</sub> receptor antagonist but not a CB<sub>2</sub> receptor blocker [83,84]. The cancer pain produced by intraplantar injection of human oral carcinoma cells could also be attenuated by peripheral injection of WIN55,212-2 or the CB<sub>2</sub> receptor agonist AM1241 [85].

Cannabinoids have also been shown to be analgesic in painful disorders of the gastrointestinal system. The writhing response elicited by intraluminal deposition of formic acid in mice could be alleviated by oral administration of either Δ-9 tetrahydrocannabinol or cannabinal [86]. Pain induced by colorectal distension could also be reduced by treating animals with a local injection of either WIN55,212-2 or the CB<sub>2</sub> receptor agonist JWH015 [87]. By administering the same cannabinoid agonists, these investigators were also able to reduce mechanical hyperalgesia in a model of colitis [87] confirming that peripherally administered cannabinoids may be viable analgesics for the treatment of visceral pain.

Damage to peripheral sensory nerves by either injury, altered metabolism or viral infection often results in neuropathic pain which may be episodic or constant. Neuropathic pain is often difficult to diagnose and hence treat since there is often no overt sign of tissue injury. Animal studies indicate that cannabinoids can alleviate neuropathic pain, offering a much needed alternative to the current first order treatment options of anti-convulsants and tricyclic antidepressants. In the rat spinal ligation model of neuropathic pain [88], CP55,940 was able to reverse tactile allodynia while WIN55,212-2 has been shown to reduce pain in the sciatic nerve chronic constriction model [89]. Unfortunately, these non-selective cannabinoid agonists were administered systemically in these studies rendering it impossible to know which cannabinoid receptor subtype was involved and where the site of analgesic action was taking place. Peripheral injection of WIN55,212-2 into the hindpaw was found to be anti-nociceptive in neuropathic pain rats with the effect being blocked by the CB<sub>1</sub> receptor antagonist SR141716A [90]. In contrast, spinal *cfos* expression is elevated in CB<sub>1</sub> receptor knockout mice compared to wild-type littermates [91] while CB<sub>1</sub> receptor deficiency has no effect on neuropathic pain responses [92]. In an ingenious

model in which peripheral CB<sub>1</sub> receptors were deleted and central CB<sub>1</sub> receptors were maintained, the analgesia produced by systemic injection of cannabinoids was reduced [93]. These data indicate that peripheral CB<sub>1</sub> receptors are critical for neuropathic pain control and suggest that peripherally-restricted CB<sub>1</sub> receptor agonists could be efficacious in the treatment of neuropathic pain.

CB<sub>2</sub> receptors have also been implicated in neuropathic pain control. The CB<sub>2</sub> receptor agonists AM1241 and JWH133 have been found to attenuate neuropathic pain in the spinal nerve ligation model [94,95]. Furthermore, neuropathic pain generated by sciatic nerve constriction was ameliorated by GW405833 [96,97], although it should be noted that extremely high doses of the compound were required to achieve anti-nociception and compound selectivity is equivocal.

#### 4. Opioids

Lauded for their therapeutic benefits, infamous for their euphoric effects, opioids are the archetypical pain killer. Opioids are a group of alkaloids derived from the poppy plant *Papaver somniferum*. The earliest description of poppies being actively cultivated as a crop was in c. 3400 BC by the Sumerians in Mesopotamia who referred to the plant as Hul Gil or “Joy Plant”. For millennia opioids have been used medically to treat health problems as diverse as dysmenorrhoea, cough, teething, asthma and arthritis. The most familiar opioid is morphine which was first isolated by the pharmacologist Friedrich Sertürner in 1806. Named after Morpheus the Greek god of dreams, morphine is one of the most widely used analgesics and is still a first line treatment for most chronic pain sufferers.

The opioids act on four distinct G-protein-coupled receptors viz. the δ-opioid receptor (DOR), κ-opioid receptor (KOR), μ-opioid receptor (MOR), and the nociceptin/orphanin FQ receptor (NOP). Stereospecific binding assays originally discovered that opioid receptors were located in the central nervous system [98–100] although the identification of the individual opioid receptor subtypes would not be determined for another 3 years. Martin et al. [101] found that morphine and ketacyclozine acted via the MOR and KOR respectively while Hughes et al. determined that the endogenous opioids Met- and Leu-enkephalin bound to a receptor in the vas deferens and called it the DOR [102]. In 1995, a novel opioid peptide with significant homology to dynorphin A was discovered by two independent laboratories [103,104]. One group called the peptide orphanin FQ since it did not bind to any known receptor subtype, while the other group referred to the opioid as nociceptin based on its ability to modulate pain processing in mammals. Based on these two independent findings, the peptide was referred to the rather clumsy concatenation of nociceptin/orphanin FQ.

Following on from their identification in the central nervous system, opioid receptors were found to be expressed in the periphery [105]. Functional studies showed that prostaglandin-induced hyperalgesia in the rat hindpaw could be ameliorated by locally applied low dose morphine thereby establishing a role for opioids in peripheral pain control [106]. This effect was confined to the treated hindpaw since contralateral morphine injection was unable to offset ipsilateral

**Table 4**

Overview of some of the commonly used opioid agonists/antagonists and their target receptor.

	Opioid receptor			
	DOR	KOR	MOR	NOP
Selective agonists	DPDPE [DAla <sup>2</sup> ]deltorphin-I or II SNC80	U69593 CI977 Salvinorin A	DAMGO Endomorphin-1 or -2 Sufentanil	N/OFQ, Ro646198 UFP112
Selective antagonists	Navirindole	Nor-binaltorphimine GNTI	CTOP	J113397 SB612111 UFP101

hyperalgesia. Similarly, the MOR agonist fentanyl reduced the pain produced by bilateral intraplantar carrageenan injection, but only in the paw in which the opioid was administered [107]. Systemic treatment with low dose fentanyl had no effect on paw withdrawal threshold confirming a peripheral site of action of the opioid. In addition to MOR-mediated analgesia, peripheral administration of DOR, KOR, and NOP agonists can also modulate inflammatory pain. Table 4 highlights some of the commonly used opioids. The selective KOR agonist U50488 reduced nociceptor spontaneous activity in acutely inflamed cat knee joints [108] as well as reducing pain behaviour in Freund's complete adjuvant inflamed paws [109]. Confirmation that U50488 was producing antihyperalgesia via KORs was indicated by the blocking effects of the KOR antagonist nor-BNI [110] but not the MOR antagonist CTAP [111] nor the DOR antagonist ICI174864 [112]. The synthetic DOR agonist DPDPE has been found to be antihyperalgesic when injected locally into the inflamed rat hindpaw [109] and when administered around the primary tumour site in an animal model of bone cancer pain [113]. Some of the original studies looking at a peripheral site of action for nociceptin/orphanin FQ found that the opioid acted on mast cells and caused secondary release of substance P from nerve terminals leading to peripheral sensitization and pain [114–117]. Curiously, during inflammation nociceptin/orphanin FQ desensitized nerve fibres suggesting that the peptide has a dual role in pain modulation depending upon the inflammatory status of the tissue [118]. This anti-nociceptive effect of nociceptin/orphanin FQ has been demonstrated in models of arthritis [118], colitis [119], capsaicin-induced neurogenic inflammation [120], as well as in response to noxious heat [121] and formalin-induced hyperalgesia [122]. Thus, it appears that under normal conditions peripherally administered nociceptin/orphanin FQ is pro-nociceptive but switches to having anti-nociceptive properties in tissues where there is pre-existing pain.

A number of reports describe an augmentation in the analgesic capacity of opioids in various inflamed tissues [123–126]. The main explanation proffered to account for this phenomenon is that during inflammation there is believed to be an increased synthesis and peripheral axonal transport of opioid receptors resulting in an upregulation of receptors in the inflamed tissue [127–129]. Since endogenous opioid expression is also increased following inflammation [130,131], it begs the question as to why these tissues are still painful. Closer inspection of the preceding studies reveals that opioid receptor expression was only assessed in the acute phase of inflammation (*i.e.* up to four days after induction) and overlooks the more clinically relevant chronic phase of the inflammatory process. In a chronic model of arthritis, it was found that peripheral MORs are actually downregulated leading to a loss in opioid anti-nociception [132]. The decrease in receptor number is thought to be due to an overaccumulation of endogenous ligand in the chronically inflamed tissue leading to internalisation of membrane-bound MOR [130]. This hypothesis would explain the inability of endogenous opioids to offset chronic pain as well as accounting for the poor analgesia reported by patients receiving intra-articular  $\mu$ -opioid therapy [133–135]. Recovery strategies aimed at preserving opioid receptor number during chronic inflammation could improve endogenous opioid analgesia and improve the efficacy of peripherally restricted opioid agonists.

## 5. Summary

At first glance, the pain pathway appears to be a straightforward line: pain signals originate in the periphery, travel centrally to the spinal cord and thence to the brain. This overly simplistic notion is, however, deceptive and belies complex neuroanatomical circuitry and neurophysiological processing of which we still only have a rudimentary understanding. Spinal and supraspinal mechanisms augment, diminish and refine the nociceptive signal shaping it into the emotional and psychophysical experience we call pain. Central

plasticity changes are thought to be responsible for the chronicity of long-term pain, but are these higher centres truly necessary for the ultimate treatment of nociceptive pain? Management of neuropathic pain is of course entirely different since the source of pain appears to be in the various divisions of the nervous system itself. However, for the majority of chronic pain, targeting nociceptive activity in the periphery should enable us to directly control pain generation at the source and thereby limit input to the central nervous system. It could be argued, however, that by the time a patient seeks medical assistance and ultimately receives appropriate pain management, the central nervous system has undergone plasticity changes and is now driving the pain. But surely the opposite must also hold true. A prolonged and targeted reduction in nociceptor activity by pharmacological means and/or physical therapy would allow the central nervous system to undergo structural reorganisation back to normal thereby returning the patient to a pain free state. As we have seen, controlling nociceptor activity is a difficult task with a multitude of chemical mediators and neurophysiological processes being involved. Adding to the complexity is that peripheral targets can change their sensitivity and even phenotype depending upon the severity of tissue pathology, concentration of agonist and crossreactivity of other inflammatory mediators. Nevertheless, by tackling pain in the periphery it should be possible to provide symptomatic relief to millions of chronic pain sufferers while avoiding the hazardous side-effects of pervasive systemic treatments.

## References

- [1] D.C. Turk, A. Okifuji, Pain terms and taxonomies of pain, in: S.M. Fishman, J.C. Ballantyne, J.P. Rathmell (Eds.), *Bonica's Management of Pain*, Lippincott Williams & Wilkins, Philadelphia, PA, USA, 2009, pp. 14–23.
- [2] M.J. Caterina, M.A. Schumacher, M. Tominaga, T.A. Rosen, J.D. Levine, D. Julius, The capsaicin receptor: a heat-activated ion channel in the pain pathway, *Nature* 389 (1997) 816–824.
- [3] S.M. Karimian, J.J. McDougall, W.R. Ferrell, Neuropeptidergic and autonomic control of the vasculature of the rat knee joint revealed by laser Doppler perfusion imaging, *Exp. Physiol.* 80 (1995) 341–348.
- [4] A. Varga, J. Nemeth, A. Szabo, J.J. McDougall, C. Zhang, K. Elekes, E. Pinter, J. Szolcsanyi, Z. Helyes, Effects of the novel TRPV1 receptor antagonist SB366791 *in vitro* and *in vivo* in the rat, *Neurosci. Lett.* 385 (2005) 137–142.
- [5] N.J. Barton, D.S. McQueen, D. Thomson, S.D. Gaudie, A.W. Wilson, D.M. Salter, I.P. Chessell, Attenuation of experimental arthritis in TRPV1R knockout mice, *Exp. Mol. Pathol.* 81 (2006) 166–170.
- [6] A. Szabo, Z. Helyes, K. Sandor, A. Bite, E. Pinter, J. Nemeth, A. Banvolgyi, K. Bolcskei, K. Elekes, J. Szolcsanyi, Role of transient receptor potential vanilloid 1 receptors in adjuvant-induced chronic arthritis: *in vivo* study using gene-deficient mice, *J. Pharmacol. Exp. Ther.* 314 (2005) 111–119.
- [7] J. Keeble, F. Russell, B. Curtis, A. Starr, E. Pinter, S.D. Brain, Involvement of transient receptor potential vanilloid 1 in the vascular and hyperalgesic components of joint inflammation, *Arthritis Rheum.* 52 (2005) 3248–3256.
- [8] P. Honore, C.T. Wismer, J. Mikusa, C.Z. Zhu, C. Zhong, D.M. Gauvin, A. Gontsyan, R. El Kouhen, C.H. Lee, K. Marsh, J.P. Sullivan, C.R. Faltynek, M.F. Jarvis, A-425619 [1-isoquinolin-5-yl-3-(4-trifluoromethyl-benzyl)-urea], a novel transient receptor potential type V1 receptor antagonist, relieves pathophysiological pain associated with inflammation and tissue injury in rats, *J. Pharmacol. Exp. Ther.* 314 (2005) 410–421.
- [9] A. Akbar, Y. Yiangou, P. Facer, J.R. Walters, P. Anand, S. Ghosh, Increased capsaicin receptor TRPV1-expressing sensory fibres in irritable bowel syndrome and their correlation with abdominal pain, *Gut* 57 (2008) 923–929.
- [10] P.A. Hughes, S.M. Brierley, C.M. Martin, T. Liebrechts, J. Persson, B. Adam, G. Holtmann, L.A. Blackshaw, TRPV1-expressing sensory fibres and IBS: links with immune function, *Gut* 58 (2009) 465–466.
- [11] N.R. Gavva, A.W. Bannon, S. Surapaneni, D.N. Hovland Jr., S.G. Lehto, A. Gore, T. Juan, H. Deng, B. Han, L. Klionsky, R. Kuang, A. Le, R. Tamir, J. Wang, B. Youngblood, D. Zhu, M.H. Norman, E. Magal, J.J. Treanor, J.C. Louis, The vanilloid receptor TRPV1 is tonically activated *in vivo* and involved in body temperature regulation, *J. Neurosci.* 27 (2007) 3366–3374.
- [12] D.M. Swanson, A.E. Dubin, C. Shah, N. Nasser, L. Chang, S.L. Dax, M. Jetter, J.G. Breitenbucher, C. Liu, C. Mazur, B. Lord, L. Gonzales, K. Hoey, M. Rizzolio, M. Bogenstaetter, E.E. Codd, D.H. Lee, S.P. Zhang, S.R. Chaplan, N.I. Carruthers, Identification and biological evaluation of 4-(3-trifluoromethylpyridin-2-yl)piperazine-1-carboxylic acid (5-trifluoromethylpyridin-2-yl)amide, a high affinity TRPV1 (VR1) vanilloid receptor antagonist, *J. Med. Chem.* 48 (2005) 1857–1872.
- [13] A.A. Steiner, V.F. Turek, M.C. Almeida, J.J. Burmeister, D.L. Oliveira, J.L. Roberts, A.W. Bannon, M.H. Norman, J.C. Louis, J.J. Treanor, N.R. Gavva, A.A. Romanovsky, Nonthermal activation of transient receptor potential vanilloid-1 channels in

- abdominal viscera tonically inhibits autonomic cold-defense effectors, *J. Neurosci.* 27 (2007) 7459–7468.
- [14] N. Tamayo, H. Liao, M.M. Stec, X. Wang, P. Chakrabarti, D. Retz, E.M. Doherty, S. Surapaneni, R. Tamir, A.W. Bannon, N.R. Gavva, M.H. Norman, Design and synthesis of peripherally restricted transient receptor potential vanilloid 1 (TRPV1) antagonists, *J. Med. Chem.* 51 (2008) 2744–2757.
- [15] S.G. Lehto, R. Tamir, H. Deng, L. Klionsky, R. Kuang, A. Le, D. Lee, J.C. Louis, E. Magal, B. H. Manning, J. Rubino, S. Surapaneni, N. Tamayo, T. Wang, J. Wang, W. Wang, B. Youngblood, M. Zhang, D. Zhu, M.H. Norman, N.R. Gavva, Antihyperalgesic effects of (R, E)-N-(2-hydroxy-2, 3-dihydro-1H-inden-4-yl)-3-(2-(piperidin-1-yl)-4-(tri fluoromethyl)phenyl)-acrylamide (AMG8562), a novel transient receptor potential vanilloid type 1 modulator that does not cause hyperthermia in rats, *J. Pharmacol. Exp. Ther.* 326 (2008) 218–229.
- [16] M. Suzuki, Y. Watanabe, Y. Oyama, A. Mizuno, E. Kusano, A. Hirao, S. Ookawara, Localization of mechanosensitive channel TRPV4 in mouse skin, *Neurosci. Lett.* 353 (2003) 189–192.
- [17] H. Mutai, S. Heller, Vertebrate and invertebrate TRPV-like mechanoreceptors, *Cell Calcium* 33 (2003) 471–478.
- [18] J. Vriens, G. Owsianik, B. Fisslthaler, M. Suzuki, A. Janssens, T. Voets, C. Morisseau, B.D. Hammock, I. Fleming, R. Busse, B. Nilius, Modulation of the Ca<sub>2</sub> permeable cation channel TRPV4 by cytochrome P450 epoxygenases in vascular endothelium, *Circ. Res.* 97 (2005) 908–915.
- [19] H. Watanabe, J. Vriens, J. Prenen, G. Droogmans, T. Voets, B. Nilius, Anandamide and arachidonic acid use epoxyeicosatrienoic acids to activate TRPV4 channels, *Nature* 424 (2003) 434–438.
- [20] W. Liedtke, Y. Choe, M.A. Marti-Renom, A.M. Bell, C.S. Denis, A. Sali, A.J. Hudspeth, J.M. Friedman, S. Heller, Vanilloid receptor-related osmotically activated channel (VR-OAC), a candidate vertebrate osmoreceptor, *Cell* 103 (2000) 525–535.
- [21] M. Suzuki, A. Mizuno, K. Kodaira, M. Imai, Impaired pressure sensation in mice lacking TRPV4, *J. Biol. Chem.* 278 (2003) 22664–22668.
- [22] S.M. Brierley, A.J. Page, P.A. Hughes, B. Adam, T. Liebrechts, N.J. Cooper, G. Holtmann, W. Liedtke, L.A. Blackshaw, Selective role for TRPV4 ion channels in visceral sensory pathways, *Gastroenterology* 134 (2008) 2059–2069.
- [23] D. Jaquemar, T. Schenker, B. Truebe, An ankyrin-like protein with transmembrane domains is specifically lost after oncogenic transformation of human fibroblasts, *J. Biol. Chem.* 274 (1999) 7325–7333.
- [24] G.M. Story, A.M. Peier, A.J. Reeve, S.R. Eid, J. Mosbacher, T.R. Hricik, T.J. Earley, A.C. Hergarden, D.A. Andersson, S.W. Hwang, P. McIntyre, T. Jegla, S. Bevan, A. Patapoutian, ANKTM1, a TRP-like channel expressed in nociceptive neurons, is activated by cold temperatures, *Cell* 112 (2003) 819–829.
- [25] H. Katsura, K. Obata, T. Mizushima, H. Yamanaka, K. Kobayashi, Y. Dai, T. Fukuoka, A. Tokunaga, M. Sakagami, K. Noguchi, Antisense knock down of TRPA1, but not TRPM8, alleviates cold hyperalgesia after spinal nerve ligation in rats, *Exp. Neurol.* 200 (2006) 112–123.
- [26] J. Frederick, M.E. Buck, D.J. Matson, D.N. Corbridge, Increased TRPA1, TRPM8, and TRPV2 expression in dorsal root ganglia by nerve injury, *Biochem. Biophys. Res. Commun.* 358 (2007) 1058–1064.
- [27] D.M. Bautista, S.E. Jordt, T. Nikai, P.R. Tsuruda, A.J. Read, J. Poblete, E.N. Yamoah, A.I. Basbaum, D. Julius, TRPA1 mediates the inflammatory actions of environmental irritants and proalgesic agents, *Cell* 124 (2006) 1269–1282.
- [28] K.Y. Kwan, A.J. Allchorne, M.A. Vollrath, A.P. Christensen, D.S. Zhang, C.J. Woolf, D.P. Corey, TRPA1 contributes to cold, mechanical, and chemical nociception but is not essential for hair-cell transduction, *Neuron* 50 (2006) 277–289.
- [29] L. Klionsky, R. Tamir, B. Gao, W. Wang, D.C. Immke, N. Nishimura, N.R. Gavva, Species-specific pharmacology of trichloro(sulfanyl)ethyl benzamides as transient receptor potential ankyrin 1 (TRPA1) antagonists, *Mol. Pain* 3 (2007) 39.
- [30] C.R. McNamara, J. Mandel-Brehm, D.M. Bautista, J. Siemans, K.L. Deranian, M. Zhao, N.J. Hayward, J.A. Chong, D. Julius, M.M. Moran, C.M. Fanger, TRPA1 mediates formalin-induced pain, *Proc. Natl. Acad. Sci. USA* 104 (2007) 13525–13530.
- [31] S.R. Eid, E.D. Crown, E.L. Moore, H.A. Liang, K.C. Choong, S. Dima, D.A. Henze, S.A. Kane, M.O. Urban, HC-030031, a TRPA1 selective antagonist, attenuates inflammatory- and neuropathy-induced mechanical hypersensitivity, *Mol. Pain* 4 (2008) 48.
- [32] E.S. Fernandes, F.A. Russell, D. Spina, J.J. McDougall, R. Graepel, C. Genry, A.A. Staniland, J.E. Keeble, M. Malcangio, S. Bevan, S.D. Brain, A distinct role for TRPA1, in addition to TRPV1, in mechanical hypernociception induced by TNF $\alpha$  and in CFA-induced monoarthritis, *Arthritis Rheum.* (in press), doi:10.1002/art.30150.
- [33] D.D. McKemy, W.M. Neuhauser, D. Julius, Identification of a cold receptor reveals a general role for TRP channels in thermosensation, *Nature* 416 (2002) 52–58.
- [34] A.M. Peier, A. Moqrich, A.C. Hergarden, A.J. Reeve, D.A. Andersson, G.M. Story, T.J. Earley, I. Dragoni, P. McIntyre, S. Bevan, A. Patapoutian, A TRP channel that senses cold stimuli and menthol, *Cell* 108 (2002) 705–715.
- [35] D.M. Bautista, J. Siemans, J.M. Glazer, P.R. Tsuruda, A.I. Basbaum, C.L. Stucky, S.E. Jordt, D. Julius, The menthol receptor TRPM8 is the principal detector of environmental cold, *Nature* 448 (2007) 204–208.
- [36] C.J. Proudfoot, E.M. Garry, D.F. Cottrell, R. Rosie, H. Anderson, D.C. Robertson, S.M. Fleetwood-Walker, R. Mitchell, Analgesia mediated by the TRPM8 cold receptor in chronic neuropathic pain, *Curr. Biol.* 16 (2006) 1591–1605.
- [37] H. Xing, M. Chen, J. Ling, W. Tan, J.G. Gu, TRPM8 mechanism of cold allodynia after chronic nerve injury, *J. Neurosci.* 27 (2007) 13680–13690.
- [38] O. Dery, C.U. Corvera, M. Steinhoff, N.W. Bunnett, Proteinase-activated receptors: novel mechanisms of signaling by serine proteases, *Am. J. Physiol. Cell Physiol.* 274 (1998) C1429–C1452.
- [39] L. Martin, C. Auge, J. Boue, M.C. Buresi, K. Chapman, S. Asfaha, P. Andrade-Gordon, M. Steinhoff, N. Cenac, G. Dietrich, N. Vergnolle, Thrombin receptor: an endogenous inhibitor of inflammatory pain, activating opioid pathways, *Pain* 146 (2009) 121–129.
- [40] S. Asfaha, V. Brussee, K. Chapman, D.W. Zochodne, N. Vergnolle, Proteinase-activated receptor-1 agonists attenuate nociception in response to noxious stimuli, *Br. J. Pharmacol.* 135 (2002) 1101–1106.
- [41] A. Kawabata, N. Kawao, R. Kuroda, A. Tanaka, C. Shimada, The PAR-1-activating peptide attenuates carrageenan-induced hyperalgesia in rats, *Peptides* 23 (2002) 1181–1183.
- [42] M. Steinhoff, N. Vergnolle, S.H. Young, M. Tognetto, S. Amadesi, H.S. Ennes, M. Trevisani, M.D. Hollenberg, J.L. Wallace, G.H. Caughey, S.E. Mitchell, L.M. Williams, P. Geppetti, E.A. Mayer, N.W. Bunnett, Agonists of proteinase-activated receptor 2 induce inflammation by a neurogenic mechanism, *Nat. Med.* 6 (2000) 151–158.
- [43] Z. Helyes, K. Sandor, E. Borbely, V. Tekus, E. Pinter, K. Elekes, D.M. Toth, J. Szolcsanyi, J.J. McDougall, Involvement of transient receptor potential vanilloid 1 receptors in protease-activated receptor-2-induced joint inflammation and nociception, *Eur. J. Pain* 14 (2010) 351–358.
- [44] A.M. Coelho, N. Vergnolle, B. Guiard, J. Fioramonti, L. Bueno, Proteinases and proteinase-activated receptor 2: a possible role to promote visceral hyperalgesia in rats, *Gastroenterology* 122 (2002) 1035–1047.
- [45] N. Vergnolle, M.D. Hollenberg, K.A. Sharkey, J.L. Wallace, Characterization of the inflammatory response to proteinase-activated receptor-2 (PAR2)-activating peptides in the rat paw, *Br. J. Pharmacol.* 127 (1999) 1083–1090.
- [46] A.D. Grant, G.S. Cottrell, S. Amadesi, M. Trevisani, P. Nicoletti, S. Materazzi, C. Altier, N. Cenac, G.W. Zamponi, F. Bautista-Cruz, C.B. Lopez, E.K. Joseph, J.D. Levine, W. Liedtke, S. Vanner, N. Vergnolle, P. Geppetti, N.W. Bunnett, Protease-activated receptor 2 sensitizes the transient receptor potential vanilloid 4 ion channel to cause mechanical hyperalgesia in mice, *J. Physiol.* 578 (2007) 715–733.
- [47] M.R. D'Andrea, C.J. Rogahn, P. Andrade-Gordon, Localization of protease-activated receptors-1 and -2 in human mast cells: indications for an amplified mast cell degranulation cascade, *Biotech. Histochem.* 75 (2000) 85–90.
- [48] J. Martel-Pelletier, J.M. Cloutier, J.P. Pelletier, Neutral proteases in human osteoarthritic synovium, *Arthritis Rheum.* 29 (1986) 1112–1121.
- [49] S. Nakano, T. Ikata, I. Kinoshita, J. Kanematsu, S. Yasuoka, Characteristics of the protease activity in synovial fluid from patients with rheumatoid arthritis and osteoarthritis, *Clin. Exp. Rheumatol.* 17 (1999) 161–170.
- [50] K. Gecse, R. Roka, L. Ferrier, M. Leveque, H. Eutamene, C. Cartier, A. Ait-Belgnaoui, A. Rosztoczy, F. Izbeki, J. Fioramonti, T. Wittmann, L. Bueno, Increased faecal serine protease activity in diarrhoeic IBS patients: a colonic luminal factor impairing colonic permeability and sensitivity, *Gut* 57 (2008) 591–599.
- [51] F.A. Russell, V.E. Veldhoen, D. Tchitchkan, J.J. McDougall, Proteinase-activated receptor-4 (PAR4) activation leads to sensitization of rat joint primary afferents via a bradykinin B2 receptor-dependent mechanism, *J. Neurophysiol.* 103 (2010) 155–163.
- [52] J.J. McDougall, C. Zhang, L. Cellars, E. Joubert, C.M. Dixon, N. Vergnolle, Triggering of proteinase-activated receptor 4 leads to joint pain and inflammation in mice, *Arthritis Rheum.* 60 (2009) 728–737.
- [53] M.R. D'Andrea, M.R. Saban, N.B. Nguyen, P. Andrade-Gordon, R. Saban, Expression of protease-activated receptor-1, -2, -3, and -4 in control and experimentally inflamed mouse bladder, *Am. J. Pathol.* 162 (2003) 907–923.
- [54] S. Asfaha, N. Cenac, S. Houle, C. Altier, M.D. Papez, C. Nguyen, M. Steinhoff, K. Chapman, G.W. Zamponi, N. Vergnolle, Protease-activated receptor-4: a novel mechanism of inflammatory pain modulation, *Br. J. Pharmacol.* 150 (2007) 176–185.
- [55] C. Auge, D. Balz-Hara, M. Steinhoff, N. Vergnolle, N. Cenac, Protease-activated receptor-4 (PAR 4): a role as inhibitor of visceral pain and hypersensitivity, *Neurogastroenterol. Motil.* 21 (2009) 1189–e1107.
- [56] A. Annahazi, K. Gecse, M. Dabek, A. Ait-Belgnaoui, A. Rosztoczy, R. Roka, T. Molnar, V. Theodorou, T. Wittmann, L. Bueno, H. Eutamene, Fecal proteases from diarrheic-IBS and ulcerative colitis patients exert opposite effect on visceral sensitivity in mice, *Pain* 144 (2009) 209–217.
- [57] E. Ryberg, N. Larsson, S. Sjogren, S. Hjorth, N.O. Hermansson, J. Leonova, T. Elebring, K. Nilsson, T. Drmota, P.J. Greasley, The orphan receptor GPR55 is a novel cannabinoid receptor, *Br. J. Pharmacol.* 152 (2007) 1092–1101.
- [58] L.A. Matsuda, S.J. Lolait, M.J. Brownstein, A.C. Young, T.I. Bonner, Structure of a cannabinoid receptor and functional expression of the cloned cDNA, *Nature* 346 (1990) 561–564.
- [59] W.A. Devane, F.A. Dysarz III, M.R. Johnson, L.S. Melvin, A.C. Howlett, Determination and characterization of a cannabinoid receptor in rat brain, *Mol. Pharmacol.* 34 (1988) 605–613.
- [60] D. Piomelli, The molecular logic of endocannabinoid signalling, *Nat. Rev. Neurosci.* 4 (2003) 873–884.
- [61] S. Munro, K.L. Thomas, M. Abu-Shaar, Molecular characterization of a peripheral receptor for cannabinoids, *Nature* 365 (1993) 61–65.
- [62] M.D. Van Sickle, M. Duncan, P.J. Kingsley, A. Mouhate, P. Urbani, K. Mackie, N. Stella, A. Makriyannis, D. Piomelli, J.S. Davison, L.J. Marnett, V. Di Marzo, Q.J. Pittman, K.D. Patel, K.A. Sharkey, Identification and functional characterization of brainstem cannabinoid CB2 receptors, *Science* 310 (2005) 329–332.
- [63] A.G. Hohmann, Spinal and peripheral mechanisms of cannabinoid antinociception: behavioral, neurophysiological and neuroanatomical perspectives, *Chem. Phys. Lipids* 121 (2002) 173–190.
- [64] R.G. Pertwee, Evidence for the presence of CB1 cannabinoid receptors on peripheral neurones and for the existence of neuronal non-CB1 cannabinoid receptors, *Life Sci.* 65 (1999) 597–605.
- [65] R.G. Pertwee, Cannabinoid receptors and pain, *Prog. Neurobiol.* 63 (2001) 569–611.

- [66] K. Tsou, S. Brown, M.C. Sanudo-Pena, K. Mackie, J.M. Walker, Immunohistochemical distribution of cannabinoid CB1 receptors in the rat central nervous system, *Neuroscience* 83 (1998) 393–411.
- [67] J.J. McDougall, Cannabinoids and pain control in the periphery, in: B.E. Cairns (Ed.), *Peripheral Receptor Targets for Analgesia*, John Wiley & Sons Inc., Hoboken, NJ, 2009, pp. 325–345.
- [68] N. Schuelert, J.J. McDougall, Cannabinoid-mediated antinociception is enhanced in rat osteoarthritic knees, *Arthritis Rheum.* 58 (2008) 145–153.
- [69] M.L. Cox, V.L. Haller, S.P. Welch, The antinociceptive effect of delta9-tetrahydrocannabinol in the arthritic rat involves the CB(2) cannabinoid receptor, *Eur. J. Pharmacol.* 570 (2007) 50–56.
- [70] T. Croci, E. Zarini, Effect of the cannabinoid CB(1) receptor antagonist rimonabant on nociceptive responses and adjuvant-induced arthritis in obese and lean rats, *Br. J. Pharmacol.* 150 (2007) 559–566.
- [71] F.L. Smith, K. Fujimori, J. Lowe, S.P. Welch, Characterization of delta9-tetrahydrocannabinol and anandamide antinociception in nonarthritic and arthritic rats, *Pharmacol. Biochem. Behav.* 60 (1998) 183–191.
- [72] C.L. Baker, J.J. McDougall, The cannabinomimetic arachidonyl-2-chloroethylamide (ACEA) acts on capsaicin-sensitive TRPV1 receptors but not cannabinoid receptors in rat joints, *Br. J. Pharmacol.* 142 (2004) 1361–1367.
- [73] J.J. McDougall, V. Yu, J. Thomson, *In vivo* effects of CB2 receptor-selective cannabinoids on the vasculature of normal and arthritic rat knee joints, *Br. J. Pharmacol.* 153 (2008) 358–366.
- [74] L.J. Hudson, S. Bevan, G. Wotherspoon, C. Gentry, A. Fox, J. Winter, VR1 protein expression increases in undamaged DRG neurons after partial nerve injury, *Eur. J. Neurosci.* 13 (2001) 2105–2114.
- [75] Y. Yiangou, P. Facer, N.H. Dyer, C.L. Chan, C. Knowles, N.S. Williams, P. Anand, Vanilloid receptor 1 immunoreactivity in inflamed human bowel, *Lancet* 357 (2001) 1338–1339.
- [76] F. Amaya, G. Shimosato, Y. Kawasaki, S. Hashimoto, Y. Tanaka, R.R. Ji, M. Tanaka, Induction of CB1 cannabinoid receptor by inflammation in primary afferent neurons facilitates antihyperalgesic effect of peripheral CB1 agonist, *Pain* 124 (2006) 175–183.
- [77] P. Cesare, L.V. Dekker, A. Sardini, P.J. Parker, P.A. McNaughton, Specific involvement of PKC-epsilon in sensitization of the neuronal response to painful heat, *Neuron* 23 (1999) 617–624.
- [78] V. Di Marzo, P.M. Blumberg, A. Szallasi, Endovanilloid signaling in pain, *Curr. Opin. Neurobiol.* 12 (2002) 372–379.
- [79] S. Hong, J.W. Wiley, Early painful diabetic neuropathy is associated with differential changes in the expression and function of vanilloid receptor 1, *J. Biol. Chem.* 280 (2005) 618–627.
- [80] L.S. Premkumar, G.P. Ahern, Induction of vanilloid receptor channel activity by protein kinase C, *Nature* 408 (2000) 985–990.
- [81] R. Noyes Jr., S.F. Brunk, D.A. Avery, A.C. Canter, The analgesic properties of delta-9-tetrahydrocannabinol and codeine, *Clin. Pharmacol. Ther.* 18 (1975) 84–89.
- [82] C.W. Michalski, T. Laukert, D. Sauliunaite, P. Pacher, F. Bergmann, N. Agarwal, Y. Su, T. Giese, N.A. Giese, S. Batkai, H. Friess, R. Kuner, Cannabinoids ameliorate pain and reduce disease pathology in cerulein-induced acute pancreatitis, *Gastroenterology* 132 (2007) 1968–1978.
- [83] D.T. Hamamoto, S. Giridharagopalan, D.A. Simone, Acute and chronic administration of the cannabinoid receptor agonist CP 55, 940 attenuates tumor-evoked hyperalgesia, *Eur. J. Pharmacol.* 558 (2007) 73–87.
- [84] L.J. Kehl, D.T. Hamamoto, P.W. Wacnik, D.L. Croft, B.D. Norsted, G.L. Wilcox, D.A. Simone, A cannabinoid agonist differentially attenuates deep tissue hyperalgesia in animal models of cancer and inflammatory muscle pain, *Pain* 103 (2003) 175–186.
- [85] A.V. Guerrero, P. Quang, N. Dekker, R.C. Jordan, B.L. Schmidt, Peripheral cannabinoids attenuate carcinoma-induced nociception in mice, *Neurosci. Lett.* 433 (2008) 77–81.
- [86] P.J. Welburn, G.A. Starmer, G.B. Chesher, D.M. Jackson, Effect of cannabinoids on the abdominal constriction response in mice: within cannabinoid interactions, *Psychopharmacologia* 46 (1976) 83–85.
- [87] M. Sanson, L. Bueno, J. Fioramonti, Involvement of cannabinoid receptors in inflammatory hypersensitivity to colonic distension in rats, *Neurogastroenterol. Motil.* 18 (2006) 949–956.
- [88] D.A. Scott, C.E. Wright, J.A. Angus, Evidence that CB-1 and CB-2 cannabinoid receptors mediate antinociception in neuropathic pain in the rat, *Pain* 109 (2004) 124–131.
- [89] U. Herzberg, E. Eliav, G.J. Bennett, I.J. Kopin, The analgesic effects of R(+)-WIN 55, 212–2 mesylate, a high affinity cannabinoid agonist, in a rat model of neuropathic pain, *Neurosci. Lett.* 221 (1997) 157–160.
- [90] A. Fox, A. Kessingland, C. Gentry, K. McNair, S. Patel, L. Urban, I. James, The role of central and peripheral cannabinoid1 receptors in the antihyperalgesic activity of cannabinoids in a model of neuropathic pain, *Pain* 92 (2001) 91–100.
- [91] O. Pol, P. Murtra, L. Caracuel, O. Valverde, M.M. Puig, R. Maldonado, Expression of opioid receptors and c-fos in CB1 knockout mice exposed to neuropathic pain, *Neuropharmacology* 50 (2006) 123–132.
- [92] A. Castane, E. Celerier, M. Martin, C. Ledent, M. Parmentier, R. Maldonado, O. Valverde, Development and expression of neuropathic pain in CB1 knockout mice, *Neuropharmacology* 50 (2006) 111–122.
- [93] N. Agarwal, P. Pacher, I. Tegeder, F. Amaya, C.E. Constantin, G.J. Brenner, T. Rubino, C.W. Michalski, G. Marsicano, K. Monory, K. Mackie, C. Mariani, S. Batkai, D. Parolaro, M.J. Fischer, P. Reeh, G. Kunos, M. Kress, B. Lutz, C.J. Woolf, R. Kuner, Cannabinoids mediate analgesia largely via peripheral type 1 cannabinoid receptors in nociceptors, *Nat. Neurosci.* 10 (2007) 870–879.
- [94] S.J. Elmes, M.D. Jhaveri, D. Smart, D.A. Kendall, V. Chapman, Cannabinoid CB2 receptor activation inhibits mechanically evoked responses of wide dynamic range dorsal horn neurons in naive rats and in rat models of inflammatory and neuropathic pain, *Eur. J. Neurosci.* 20 (2004) 2311–2320.
- [95] M.M. Ibrahim, H. Deng, A. Zvonok, D.A. Cockayne, J. Kwan, H.P. Mata, T.W. Vanderah, J. Lai, F. Porreca, A. Makriyannis, T.P. Malan Jr., Activation of CB2 cannabinoid receptors by AM1241 inhibits experimental neuropathic pain: pain inhibition by receptors not present in the CNS, *Proc. Natl Acad. Sci. USA* 100 (2003) 10529–10533.
- [96] K.J. Valenzano, L. Tafesse, G. Lee, J.E. Harrison, J.M. Boulet, S.L. Gottshall, L. Mark, M.S. Pearson, W. Miller, S. Shan, L. Rabadi, Y. Rotshteyn, S.M. Chaffer, P.I. Turchin, D.A. Elsemore, M. Toth, L. Koetzner, G.T. Whiteside, Pharmacological and pharmacokinetic characterization of the cannabinoid receptor 2 agonist, GW405833, utilizing rodent models of acute and chronic pain, anxiety, ataxia and catalepsy, *Neuropharmacology* 48 (2005) 658–672.
- [97] G.T. Whiteside, S.L. Gottshall, J.M. Boulet, S.M. Chaffer, J.E. Harrison, M.S. Pearson, P.I. Turchin, L. Mark, A.E. Garrison, K.J. Valenzano, A role for cannabinoid receptors, but not endogenous opioids, in the antinociceptive activity of the CB2-selective agonist, GW405833, *Eur. J. Pharmacol.* 528 (2005) 65–72.
- [98] C.B. Pert, S.H. Snyder, Opiate receptor: demonstration in nervous tissue, *Science* 179 (1973) 1011–1014.
- [99] E.J. Simon, J.M. Hiller, I. Edelman, Stereospecific binding of the potent narcotic analgesic (3H) Etorphine to rat-brain homogenate, *Proc. Natl Acad. Sci. USA* 70 (1973) 1947–1949.
- [100] L. Terenius, Characteristics of the "receptor" for narcotic analgesics in synaptic plasma membrane fraction from rat brain, *Acta Pharmacol. Toxicol. (Copenh.)* 33 (1973) 377–384.
- [101] W.R. Martin, C.G. Eades, J.A. Thompson, R.E. Huppler, P.E. Gilbert, The effects of morphine- and nalorphine- like drugs in the nondependent and morphine-dependent chronic spinal dog, *J. Pharmacol. Exp. Ther.* 197 (1976) 517–532.
- [102] J. Hughes, T.W. Smith, H.W. Kosterlitz, L.A. Fothergill, B.A. Morgan, H.R. Morris, Identification of two related pentapeptides from the brain with potent opiate agonist activity, *Nature* 258 (1975) 577–580.
- [103] J.C. Meunier, C. Mollereau, L. Toll, C. Suaudeau, C. Moisand, P. Alvinerie, J.L. Butour, J.C. Guillemot, P. Ferrara, B. Monsarrat, H. Mazarquil, G. Vassart, M. Parmentier, J. Costentin, Isolation and structure of the endogenous agonist of opioid receptor- like ORL1 receptor, *Nature* 377 (1995) 532–535.
- [104] R.K. Reinscheid, H.P. Nothacker, A. Bourson, A. Ardati, R.A. Henningsen, J.R. Bunzow, D.K. Grandy, H. Langen, F.J. Monsma Jr., O. Civelli, Orphanin FQ: a neuropeptide that activates an opioidlike G protein- coupled receptor, *Science* 270 (1995) 792–794.
- [105] M. Wuster, R. Schulz, A. Herz, Multiple opiate receptors in peripheral tissue preparations, *Biochem. Pharmacol.* 30 (1981) 1883–1887.
- [106] S.H. Ferreira, M. Nakamura II, Prostaglandin hyperalgesia: the peripheral analgesic activity of morphine, enkephalins and opioid antagonists, *Prostaglandins* 18 (1979) 191–200.
- [107] J.L. Joris, R. Dubner, K.M. Hargreaves, Opioid analgesia at peripheral sites: a target for opioids released during stress and inflammation? *Anaesth. Analg.* 66 (1987) 1277–1281.
- [108] N.J. Russell, H.G. Schaible, R.F. Schmidt, Opiates inhibit the discharges of fine afferent units from inflamed knee joint of the cat, *Neurosci. Lett.* 76 (1987) 107–112.
- [109] C. Stein, M.J. Millan, T.S. Shippenberg, Peripheral opioid receptors mediating antinociception in inflammation: evidence for mu, delta, and kappa receptors, *J. Pharmacol. Exp. Ther.* 248 (1989) 1269–1275.
- [110] A.E. Takemori, B.Y. Ho, J.S. Naeseth, P.S. Portoghesse, Nor-binaltorphimine, a highly selective kappa-opioid antagonist in analgesic and receptor binding assays, *J. Pharmacol. Exp. Ther.* 246 (1988) 255–258.
- [111] T.H. Kramer, J.E. Shook, W. Kazmierski, E.A. Ayres, W.S. Wire, V.J. Hruby, T.F. Burks, Novel peptidic mu opioid antagonists: pharmacologic characterization *in vitro* and *in vivo*, *J. Pharmacol. Exp. Ther.* 249 (1989) 544–551.
- [112] R. Cotton, M.G. Giles, L. Miller, J.S. Shaw, D. Timms, ICI 174864: a highly selective antagonist for the opioid delta-receptor, *Eur. J. Pharmacol.* 97 (1984) 331–332.
- [113] A. Baamonde, A. Lastra, L. Juarez, V. Garcia, A. Hidalgo, L. Menendez, Effects of the local administration of selective mu-, delta- and kappa-opioid receptor agonists on osteosarcoma-induced hyperalgesia, *Naunyn-Schmiedeberg's Arch. Pharmacol.* 372 (2005) 213–219.
- [114] J.J. McDougall, S.E.M. Larson, Nociceptin/orphanin FQ evokes knee joint pain in rats via a mast cell independent mechanism, *Neurosci. Lett.* 398 (2006) 135–138.
- [115] J.J. McDougall, U. Hanesch, M. Pawlak, R.F. Schmidt, Participation of NK1 receptors in nociceptin-induced modulation of rat knee joint mechanosensitivity, *Exp. Brain Res.* 137 (2001) 249–253.
- [116] J.J. McDougall, M. Pawlak, U. Hanesch, R.F. Schmidt, Peripheral modulation of rat knee joint afferent mechanosensitivity by nociceptin/orphanin FQ, *Neurosci. Lett.* 288 (2000) 123–126.
- [117] M. Inoue, M. Kobayashi, S. Kozaki, A. Zimmer, H. Ueda, Nociceptin/orphanin FQ-induced nociceptive responses through substance P release from peripheral nerve endings in mice, *Proc. Natl Acad. Sci.* 95 (1998) 10949–10953.
- [118] J.J. McDougall, M. Pawlak, U. Hanesch, R.F. Schmidt, Peripheral modulation of rat knee joint afferent mechanosensitivity by nociceptin/orphanin FQ, *Neurosci. Lett.* 288 (2000) 123–126.
- [119] S. Agostini, H. Eutamene, M. Broccardo, G. Improta, C. Petrella, V. Theodorou, L. Bueno, Peripheral anti-nociceptive effect of nociceptin/orphanin FQ in inflammation and stress-induced colonic hyperalgesia in rats, *Pain* 141 (2009) 292–299.

- [120] M.C. Ko, N.N. Naughton, J.R. Traynor, M.S. Song, J.H. Woods, K.C. Rice, A.T. McKnight, Orphanin FQ inhibits capsaicin-induced thermal nociception in monkeys by activation of peripheral ORL1 receptors, *Br. J. Pharmacol.* 135 (2002) 943–950.
- [121] Y.A. Kolesnikov, G.W. Pasternak, Peripheral orphanin FQ/nociceptin analgesia in the mouse, *Life Sci.* 64 (1999) 2021–2028.
- [122] M. Ambriz-Tututi, H.I. Rocha-Gonzalez, G. Castaneda-Corral, C.I. Araiza-Saldana, N.L. Caram-Salas, S.L. Cruz, V. Granados-Soto, Role of opioid receptors in the reduction of formalin-induced secondary allodynia and hyperalgesia in rats, *Eur. J. Pharmacol.* 619 (2009) 25–32.
- [123] J.N. Sengupta, A. Snider, X. Su, G.F. Gebhart, Effects of kappa opioids in the inflamed rat colon, *Pain* 79 (1999) 175–185.
- [124] W. Binder, H. Machelska, S. Mousa, T. Schmitt, P.J. Riviere, J.L. Junien, C. Stein, M. Schafer, Analgesic and antiinflammatory effects of two novel kappa-opioid peptides, *Anesthesiology* 94 (2001) 1034–1044.
- [125] S. Furst, P. Riba, T. Friedmann, J. Timar, M. Al-Khrasani, I. Obara, W. Makuch, M. Spetea, J. Schutz, R. Przewlocki, B. Przewlocka, H. Schmidhammer, Peripheral versus central antinociceptive actions of 6-amino acid-substituted derivatives of 14-O-methylxymorphone in acute and inflammatory pain in the rat, *J. Pharmacol. Exp. Ther.* 312 (2005) 609–618.
- [126] S. Nunez, J.S. Lee, Y. Zhang, G. Bai, J.Y. Ro, Role of peripheral mu-opioid receptors in inflammatory orofacial muscle pain, *Neuroscience* 146 (2007) 1346–1354.
- [127] R.R. Ji, Q. Zhang, P.Y. Law, H.H. Low, R. Elde, T. Hokfelt, Expression of mu-, delta-, and kappa-opioid receptor-like immunoreactivities in rat dorsal root ganglia after carrageenan-induced inflammation, *J. Neurosci.* 15 (1995) 8156–8166.
- [128] W. Puehler, C. Zollner, A. Brack, M.A. Shaqura, H. Krause, M. Schafer, C. Stein, Rapid upregulation of mu opioid receptor mRNA in dorsal root ganglia in response to peripheral inflammation depends on neuronal conduction, *Neuroscience* 129 (2004) 473–479.
- [129] C. Zöllner, M.A. Shaqura, C.P. Bopaiah, S. Mousa, C. Stein, M. Schafer, Painful inflammation-induced increase in mu-opioid receptor binding and G-protein coupling in primary afferent neurons, *Mol. Pharmacol.* 64 (2003) 202–210.
- [130] J.J. McDougall, A.K. Barin, C.M. McDougall, Loss of vasomotor responsiveness to the mu-opioid receptor ligand endomorphin-1 in adjuvant monoarthritic rat knee joints, *Am. J. Physiol. Reg. Integr. Comp. Physiol.* 286 (2004) R634–R641.
- [131] S.A. Mousa, H. Machelska, M. Schafer, C. Stein, Immunohistochemical localization of endomorphin-1 and endomorphin-2 in immune cells and spinal cord in a model of inflammatory pain, *J. Neuroimmunol.* 126 (2002) 5–15.
- [132] Z. Li, D. Proud, C. Zhang, S. Wiehler, J.J. McDougall, Chronic arthritis downregulates peripheral mu-opioid receptor expression with concomitant loss of endomorphin-1 anti-nociception, *Arthritis Rheum.* 52 (2005) 3210–3219.
- [133] L.A. Rosseland, A. Stubhaug, F. Grevbo, O. Reikeras, H. Breivik, Effective pain relief from intra-articular saline with or without morphine 2 mg in patients with moderate-to-severe pain after knee arthroscopy: a randomized, double-blind controlled clinical study, *Acta Anaesthesiol. Scand.* 47 (2003) 732–738.
- [134] P.A. Ruwe, I. Klein, C.L. Shields, The effect of intraarticular injection of morphine and bupivacaine on postarthroscopic pain control, *Am. J. Sports Med.* 23 (1995) 59–64.
- [135] I.J. Wrench, P. Taylor, G.J. Hobbs, Lack of efficacy of intra-articular opioids for analgesia after day-case arthroscopy, *Anaesthesia* 51 (1996) 920–922.