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Cholangiocytes: cell transplantation.

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Abbreviations.

Aldehydedehydrogenase (ALDH); Cluster differentiation (CD); Epidermal growth factor (EGF); Epithelial cell adhesion molecule (EpCAM); Fibroblast growth factor (FGF); Good Manufacturing Practice (GMP); Granulocyte colony-stimulating factor (G-CSF); Hepatocyte growth factor (HGF); Human biliary tree stem cells (hBTSCs); Human hepatic stem/progenitor cells (hHpSCs); Hyaluronic acid (HA); Extracellular matrix (ECM); Hyaluronic Acid or Hyaluronan (HA); International Normalized Ratio (INR); Kruppel-like factor (KLF); Leucine-rich repeat containing G protein-coupled Receptor 5 (LGR5); Mesenchymal stem cells (MSCs); Model of End-stage Liver Disease (MELD); Nanog homeobox (NANOG); Non-alcoholic steato-hepatitis (NASH); Octamer-binding Transcription factor 4 (OCT4); Orthotopic liver transplantation (OLT); Pancreatic and duodenal homeobox 1 (PDX1); Peri-biliary glands (PBGs); Polyethylene glycol-diacrylate (PEG-DA); Quantitative Reverse-Transcription Polymerase Chain Reaction (RT-qPCR); Sal-like protein 4 (SALL4); Severe Combined Immunodeficiency (SCID); Sry-related HMG box (SOX).

Abstract

Background: Due to significant limitations to the access to orthotropic liver transplantation, cell therapies for liver diseases have gained large interest worldwide.

Scope of Review: to revise current literature dealing with cell therapy for liver diseases. We discussed the advantages and pitfalls of the different cell sources tested so far in clinical trials and the rationale underlying the potential benefits of transplantation of human biliary tree stem cells (hBTSCs).

Major Conclusions: transplantation of adult hepatocytes showed transient benefits but requires immune-suppression that is a major pitfall in patients with advanced liver diseases. Mesenchymal stem cells and hematopoietic stem cells transplanted into patients

with liver diseases are not able to replace resident hepatocytes but, rather they target autoimmune or inflammatory processes into the liver. Stem cells isolated from fetal or adult liver have been recently proposed as an alternative cell sources for advanced liver cirrhosis and metabolic liver disease. We demonstrated the presence of multipotent cells expressing a variety of endodermal stem cell markers in (peri)-biliary glands of bile ducts in fetal or adult human tissues, and in crypts of gallbladder epithelium. In the first cirrhotic patients treated in our center with biliary tree stem cell therapy we registered no adverse event but significant benefits.

General Significance: the biliary tree stem cell could represent the ideal cell source for the cell therapy of liver diseases.

Keywords: liver cell therapy, hepatic stem cells, biliary tree stem cells, hepatocytes, mesenchymal stem cells, grafting.

1. Introduction

Liver diseases represent a major public health problem affecting 5-15% of the inhabitants worldwide [1]. The final manifestation of chronic liver diseases is cirrhosis. When a successful etiologic approach is unavailable or has failed, or delayed, progressive, extensive fibrosis, with concurrently impaired hepatocyte regeneration, leads to irreversible cirrhosis and then liver failure. Currently, orthotopic liver transplantation (OLT) is the only therapy for acute and chronic liver diseases in the terminal stage. However, the number of donated livers is limited. In addition, OLT cannot be proposed for many cirrhotic patients, since at very advanced disease stages, surgical risks or other contraindications exist [2]. Post-surgery complications and rejection are still significant problems associated with organ transplantation. Other limitations of OLT relate to the high costs: typically, ~\$150,000 to \$180,000 for transplant and first-year medical follow-up [2]. These limitations drive interests to explore cell therapies aimed to sustain liver function, to stimulate liver regeneration and to reduce the scarring process.

2. Cell sources for cell therapy of liver diseases

Different strategies of cell therapy have been so far attempted and used not only for advanced cirrhosis but also for acute liver failure, fulminant hepatitis, inborn errors of metabolism, viral hepatitis, liver toxins, alcohol consumption, chronic cholestatic diseases, autoimmunity and metabolic disorders such as non-alcoholic steato-hepatitis (NASH) [3].

2.1 Adult Hepatocytes. The first strategies tried for cell therapies of liver diseases focused on adult hepatocyte transplantation which was first used for patients with liver diseases in the 1990s and has continued through for the last ~20 years [4]. The studies have not expanded into large clinical trials due to difficulties in obtaining high quality donor cells, limitations in the ability to cryopreserve hepatocytes, and the complications that can occur with transplantation of the hepatocytes into the liver [4]. These complications include

low engraftment (typically only 5% if transplanted via the portal vein), similar engraftment but with bleeding disorders if transplanted by direct injection, distribution to ectopic sites (most commonly the lungs), and formation of emboli that can be life threatening especially if they occur in the lungs [4]. Patients have been transplanted via the portal vein with isolated hepatocytes at concentrations from millions to billions. Immunosuppression was required in all cases. Although, it is a challenge to draw conclusions from the published reports, treatments resulted in improved liver functions, but the patient numbers in each trial are so small that they constitute only anecdotal evidence of effectiveness [4]. No trials have been done with sufficient numbers of patients to permit statistical analyses yielding Model of End-stage Liver Disease (MELD) or Child-Pugh scores, the standardized metrics used for liver condition. Therapeutic effects were observed to last from days to a few months and can provide bridging to a liver transplant [4]. Therapeutic benefits for congenital defects (e.g. Crigler-Najjar) [5-8] have lasted longer, typically for months and, in a few cases, up to a few years [8,9]. Leaders in the field have lobbied for alternate approaches but still retaining a focus on using adult hepatocytes [10], but the poor to mediocre results to date have diminished enthusiasm for this strategy.

2.2 Mesenchymal stem cells and hematopoietic stem cells. Transplantation of mesenchymal stem cells (MSCs) and hematopoietic stem cells into patients with liver disease, has resulted in large numbers of clinical trials throughout the world [11]. At present over 20 clinical trials have been published but with small numbers of patients (typically under 10). Thus, they are similar to the clinical trials of hepatocyte transplantation in providing anecdotal evidence or evidence with minimal possibility of statistically validated findings of the efficacy of the treatments. There are a small number with larger patient populations such as one reported by Peng et al [11] involving 53 patients who underwent a single transplantation with autologous MSCs by avascular route via the peripheral vasculature, the spleen or through the hepatic artery into the liver. The trials

comprised treatments with: a) unfractionated bone marrow or peripheral blood; b) used cytokines (e.g. Granulocyte colony-stimulating factor [G-CSF]) to mobilize cells in the bone marrow; c) used immune-selected cell populations (CD44⁺ cells, CD133⁺ cells) from bonemarrow; or d) used cultured MSCs or cultures treated with growth factors such as hepatocyte growth factor (HGF), epidermal growth factor (EGF) or fibroblast growth factor (FGF). Transplantations of any of these forms of MSCs or hematopoietic stem cells were found to be safe and significantly improved the quality of life and liver functions [3]. The patient response occurred within days to weeks, but long-term effects (more than a few months) were not observed. The conclusions are that effects are due to trophic and immune-modulatory factors. Finally, it was discovered a single specific molecule secreted by the MSCs, the Milk Fat Globule-EGF Factor 8, that protects against liver fibrosis in mice [12]. Andreone P. et al [13] performed a phase I study where 12 patients with end stage liver disease were reinfused with high purified CD133⁺ bone-marrow derived hematopoietic stem cells, after pretreatment with growth factors. No severe adverse events were registered, MELD worsened during mobilization in Child Turcotte-C patients. Improvement of liver function was observed 2 months after reinfusion (MELD 19.5 vs 16; $p = 0.045$), 5 patients underwent liver transplantation within 12 months from reinfusion and 2 died because of progressive liver failure. Again, the benefits were only transient and after a 6-month follow-up, liver function parameters returned to baseline. A recent multicenter phase-II open-label controlled trial of repeated autologous infusions of G-CSF mobilized CD133⁺ bone marrow stem cells in N. 27 patients with advanced cirrhosis (versus conservative management or treatment with G-CSF alone) found no impact on liver function nor on fibrosis and the trial was interrupted [14]. Therefore, also G-CSF and purified hematopoietic stem cells did not have an impact on liver function or fibrosis, according this large, powered, randomized controlled trial [14].

3. Human Biliary tree stem cells.

In a series of published studies [15-19] and summarized in recent reviews [20-22], we demonstrated the presence of cells expressing a variety of endodermal stem cell markers in (peri)-biliary glands (PBGs) of bile ducts in fetal or adult human tissues. We also found them in crypts of gallbladder epithelium [16]. The observations *in situ* in human biliary tree tissue have been complemented by demonstrations of culture selection of colonies of stem cells and that expand for months through self-renewal processes, and are multipotent as indicated by their ability to lineage restrict to hepatocytes, cholangiocytes or pancreatic islet cells depending on the precise conditions to which they are subjected. Isolation of major subpopulations of hBTSCs from tissues of all donor ages can be achieved by immunoselection for cells positive for expression of epithelial cell adhesion molecule (EpCAM). The hBTSCs constitute approximately 2-4% of the biliary tree cells with the percentages varying, depending on which part of the biliary tree is being analyzed. The richest sites are the large, intrahepatic bile ducts and the hepato-pancreatic common duct. The hBTSC subpopulations are all small (7-9 μm), approximately half the size of mature parenchymal cells; express various stem cell markers such as endodermal transcription factors (SOX9, SOX17, PDX1), pluripotency genes (OCT4, SOX2, NANOG, SALL4, KLF4/5), CD133 (prominin), CD44 (hyaluronan receptors), aldehyde dehydrogenase (ALDH), and hedgehog proteins and no markers of mature hepatic or pancreatic cells. They are also tolerant to ischemia [22]. Preliminary studies indicated that we can isolate viable and healthy hBTSCs from biliary tree tissue even 48 hours or more after cardiac arrest of the donor [23]. In addition, other preliminary studies suggest that the hBTSCs can be cryopreserved under serum-free, wholly defined conditions that we established and found effective for human hepatic stem cells [24].

In preclinical studies, we demonstrated that hBTSCs isolated from human gallbladder and transplanted in a model of liver cirrhosis yielded the formation of human hepatocytes and

cholangiocytes and, mostly important, the improvement of the liver function tests in the model [15].

The hBTSCs can be isolated from fetal or adult livers or from adult gallbladder. hBTSCs is an ideal source of multipotent endodermal stem cells with relevant self-renewal and differentiative capacities. Infact: First, tissues are largely available since fetal biliary trees can be easily obtained from the fetal liver given the large number of therapeutic abortions, postnatal biliary tree tissues are largely available since most of the biliary tree is routinely discarded from liver and pancreatic transplants and gallbladder tissue is readily available given large numbers of cholecystectomy done routinely. Second, the hBTSCs can be isolated easily from biliary tree of any donor age and survive and expand *ex vivo* under wholly defined and serum-free conditions that we established; v) hBTSCs from fetal tissue are non- or minimally immunogenic given their low or null expression of HLA class I and II [25]antigens and, therefore, can be transplanted without the need for immunosuppression; vi) hBTSCs, transplanted into experimental models of liver cirrhosis, are able to engraft, to differentiate into adult hepatocytes and cholangiocytes and to rescue liver function. The use of stem/progenitor cells has advantages over transplantation of mature hepatocytes. Firstly, stem/progenitor cells, capable of generating mature liver cells, can circumvent the shortage of hepatocytes. The hBTSCs are available from hepatic and biliary tree tissues of donors of all ages and can self-replicate under particular conditions indefinitely. Secondly, transplanting stem/progenitor cells may, in theory, yield better long-term repopulation and persistent metabolic activity due to the constant generation of newly formed mature parenchymal cells. Thirdly, stem/progenitor cells are not immunogenic [23,26] are more resistant than adult cells to ischemic damage, and are easier to cryopreserve [24]. All of these properties enable stem/progenitors to be a more robust product for clinical programs.

Khan et al. [27] isolated EpCAM⁺ cell from the fetal livers and administered these cells into 25 cirrhotic patients via hepatic artery and without immunosuppression. The procedure was safe and associated with significant improvement of liver function that lasted during the six-months follow up period. This procedure was proven to be safe, as assessed by ultrasound indicating persistence of echotexture, no focal lesions, and no abnormal changes in the size of the hepatic artery. Fetal liver-derived EpCAM⁺ cells (a mix of human Hepatic Stem Cells/hHpSCs and hepatoblasts) were marked with Tc99m-Hexamethylpropyleneamine Oxime and injected; most of the marked cells remained within the liver lobe injected. Also, most patients had grade 2 to grade 3 esophageal varices before the transplants, and the majority showed reduction in the varices grading. More recently, Pietrosi and co-workers [28] treated 9 patients affected by end-stage liver disease within transsplenic infusion of a total cell population obtained from the fetal liver and found that the procedure was safe and well tolerated with positive effects on clinical scores and on encephalopathy.

With all these premises, representing a strong rational basis for translating our preclinical research into a clinical setting, after having obtained the required authorization, two patients with advanced cirrhosis were treated with fetal hBTSCs transplantation via hepatic artery infusion (Figure 1) [29]. We injected immunoselected EpCAM-positive cells which also co-expressed Leucine-rich repeat containing G protein-coupled Receptor 5 (LGR5). In fetal livers, LGR5-positive cells were located in the ductal plate and in the epithelium of larger bile ducts [18]. In gallbladder and hepatic common duct, surface epithelial cells and bud of PBGs, were diffusely positive for LGR5 [17]. Cell products were characterized by Flow Cytometry (FC) for EpCAM and LGR5 before transplantation (Figure 1). Estimated cell viability by trypan blue exclusion was routinely higher than 95%. The immunomagnetic sorting resulted in an enrichment of EpCAM-positive cell population to 51% and contained cells with co-expression of EpCAM and LGR5 (48.4%). Immunosorted EpCAM-positive

cells were suspended in 10% glucose solution, at a concentration of 1 million cells per ml. The cell suspension was infused into the hepatic artery at an infusion velocity of 200 ml/h (Figure 1). The first patient received, via the right hepatic artery, 42 million viable EpCAM-sorted cells, showed an evident increasing trend of serum albumin, associated with a parallel gradual and constant decrease of International Normalized Ratio (INR). Before treatment, the patient received repeated hospitalizations, mostly for treatment of ascites through large volume paracentesis but, their frequency and duration were significantly shortened by the cell therapy (1-day vs 5-days hospital stay pre-treatment). The need of paracentesis was abolished during the follow-up, with no request after cell therapy. The patient observed the same pharmacological treatment during the whole period of observation, and did not receive albumin injection. Six months after receiving hBTSC transplantation, the Child-Pugh score decreased from C-12 to C-10 and the MELD score from 24 to 20, mainly due to improvement of coagulation and ascites.

The second patient is a Caucasian female affected by HCV-related cirrhosis. No side-effects related to the cell infusion were registered during the scheduled 6-month follow-up. Six months after the treatment, the patient displayed a consistent amelioration of liver function: Child-Pugh score improved from C-11 to B-8 and the MELD score from 21 to 16. After 12th month of follow-up the patient continued to maintain a state of compensated liver cirrhosis. Notably, this patient experienced a gradual and constant amelioration of albumin value and coagulation (INR) along the 12 months' follow-up. The patient did not receive intravenous albumin infusion. Exhaustive information on the different cell sources for cell therapy of liver diseases containing data on cell origin, number/size/characteristics of clinical trials, advantages/disadvantages, references, etc, have been included as supplementary tables (one for cell source) in an updated review published in Stem Cells by international experts including us and collaborators from Prof. Lola Reid and Prof. Luca Invernizzi groups (see ref. 20).

4. Strategies to ameliorate the engraftment efficiency of hBTSCs into the liver (see Box 1 and 2).

Liver parenchymal repopulation with exogenous cells is a prerequisite for a successful cell therapy. Cell translocation from sinusoids into liver plates requires disruption of the sinusoidal endothelium and progressive proliferation of the transplanted cells [30] (Box 1). BTSCs and hepatic stem/progenitor cells (HpSCs) have intrinsic proliferative advantage when transplanted in liver parenchyma with respect to mature hepatocytes [31] (Box 1). However, the main pitfall of liver stem cell therapy is the uncertain yield of transplanted cells effectively grafting in the target organ. Indeed, engraftment efficiencies of less than 5% were reported for stem cells when delivered by vascular routes into the livers of primates [32] or in the livers of humans when injected into the portal vein [33]. That's because the modulation of cell adhesion and engraftment are key issues in regenerative medicine (Box 1). To this regard, a lot of recent researches focus on strategies to improve grafting efficiency of transplanted cells (Box 2). Hyaluronic acid (HA) is one of the most utilized biomatrix in human medicine and it could be one of the best candidate to promote the acceptance of transplanted cells in the target organs (Box 2). In the last decades, different evidences showed how extracellular matrix (ECM) plays a fundamental dynamic role in regulating cell homeostasis. Among the main ECM components, HA plays a major role because of its biologic, rheological, viscoelastic and hygroscopic properties [34]. HA molecules are highly active in interacting each other by forming large polymers or in association with other molecules [35]. HA is expressed on cell surface of both normal and tumor cells, it is an important component of the stem cell niche involved in preserving the multipotent character of stem/progenitor cells, preventing their differentiation and modulating stem cell migration during embryonic development [36]. Indeed, the HA matrix supports cell adhesion, growth and differentiation, regulates cell trafficking and affects

various biologic processes including development and organogenesis, inflammation, wound healing, and tissue remodeling [34]. For this reason, many research groups used scaffold/gel with HA, alone or with other molecules [37-44]. Lozoya et al. [43] studied the grafting of hHpSC using hydrogel in several concentrations with thiol-modified HA derivatives, and polyethylene glycol-diacrylate (PEG-DA), as cross-link. Remarkably, they demonstrate that both the composition and mechanical properties of the microenvironment can regulate phenotypic changes. Based on these results obtained *in vitro*, Reid L. et al. [46] devised grafting strategies for transplantation of hHpSCs embedded into a mix of soluble signals and extracellular matrix biomaterials (hyaluronans, type III collagen, laminin) found in stem cell niches. Grafted cells remained localized to the livers, resulting in a larger bolus of engrafted cells in the host livers under quiescent conditions and with potential for more rapid expansion under injured liver conditions. By contrast, transplantation by direct injection or via a vascular route resulted in inefficient engraftment and cell spreading to ectopic sites (Box 2). Recently, we developed a fast and easy method to coat hBTSCs with HA and assessed the effects of HA-coating on cell properties *in vitro* and *in vivo* [47] (Box 2). The HA coating markedly improved the viability, colony formation, and population doubling of hBTSCs in primary cultures, and resulted in a higher expression of integrins that mediate cell attachment to matrix components. When HA-coated hBTSCs were transplanted via the spleen into the liver of immunocompromised mice, the engraftment efficiency increased to 11% with respect to 3% of uncoated cells (Box 2). Studies in distant organs showed minimal ectopic cell distribution without differences between HA-coated and uncoated hBTSCs and, specifically, cell seeding in the kidney was excluded. The danger of thrombi generation and ectopic distribution has been minimized, because the coating process did not result in large cell clusters. Moreover, the fact that HA is selectively and actively cleared by the liver could be an additive factor favoring the selective hepatic engraftment [48]. Finally, HA should create a

favorable microenvironment allowing a greater number of hBTSCs to engraft within the liver parenchyma, survive, proliferate, and enhance human albumin secretion. When, in previous studies, fibrin was tested, instead of the HA, a similar phenomenon was observed [32]. Previous studies highlighted varied HA trophic effects in different cell systems [49-54]. An especially advantageous aspect of using HAs to facilitate engraftment in condition of liver injury is that they are anti-inflammatory and entirely biocompatible. In prior studies with use of HAs for diverse forms of transplantations, they minimize fibrotic reactions and foster vascularization that facilitates engraftment [55]. HA is a natural candidate for stem cell grafting because it is abundant during embryogenesis, wound repair, and organ regeneration [45]. To this latter regards HA cell coating can be seen as a form of host conditioning, which, it is currently a target to monitor cell rejection in liver cell therapy in humans [56]. HA coating could improve outcomes of stem cell therapies of liver diseases and could be immediately translated into the clinic given that Good Manufactory Practice (GMP)-grade HAs are already available for clinical use. This molecule indeed is already approved for clinical use and has been tested in clinical trials [57].

5. Conclusions and perspectives

The improvement of regenerative medicine approaches for liver diseases requires the identification of sustainable and readily available cell sources (66). In keeping, other than mature hepatocytes and the different stem cells already tested in clinic, the possibility of reprogramming adult somatic cells to generate mature liver cells attracted (66). As far as biliary tissue, several groups have reported defined protocols to differentiate human induced pluripotent stem cells (iPSCs) in functionally biliary epithelium cells (67-71). This technology associated with dedicated tissue engineering strategies could lead to the autologous replacement of damaged biliary tree (67-71). However, the clinical application of reprogrammed cells raises concerns due to the possible uncontrolled tumorigenic

expansion within the recipient, particularly, in long life expectancy situations as in pediatric liver diseases or in chronic liver diseases (66). In this context, the small molecule-based reprogramming has raised attention due to its safety (66), and a recent paper by Wang et al. described a novel approach to generate human induced endodermal progenitor cells (hiEndoPCs) through the lineage reprogramming of gastrointestinal epithelial cells induced by a cocktail of defined small molecules in association with the support of tissue-specific mesenchymal feeders (72).

Waiting for protocols allowing a safe and effective clinical translation of iPSCs in regenerative medicine of liver, hBTSCs represent a ready cellular source. The Rationale of “hBTSC treatment of liver cirrhosis” is based on these key concepts: i) autochthonous hBTSCs are normally involved in liver repair but they are impaired in their regeneration potential and in their capacity to differentiate in mature liver cells, by the long-lasting inflammatory milieu characterizing liver cirrhosis; ii) in preclinical studies we demonstrated that isolated fetal hBTSCs are capable of generating *in vitro* mature liver cells and that; iii) hBTSCs, transplanted in a model of liver cirrhosis, yielded the formation of human liver cells (hepatocytes and cholangiocytes) and, mostly important, the improvement of the liver function tests in the model, iv) hBTSCs cells are not immunogenic and are more resistant than adult cells to ischemic damage; v) in the first two cirrhotic patients treated in our center, we registered no adverse event but significant benefits. With this premises, the expectancy is that hBTSCs transplantation in patients with advanced liver cirrhosis may yield long-term repopulation and persistent metabolic activity due to the constant generation of newly formed mature parenchymal cells and that this could finally result in improvement of clinically relevant liver functions.

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Conflicts of interest: V. C., G. C., E. G., and D. A. are inventors of an international patents related to hBTSCs and granted a sponsored research agreement from Vesta Therapeutics (Bethesda, MD) for further development of this patent.

REFERENCES

- [1] G. D'Amico, G. Garcia-Tsao, L. Pagliaro. Natural history and prognostic indicators of survival in cirrhosis: a systematic review of 118 studies. *J Hepatol*44 (2006) 217-231,
- [2] M.R. Lucey, N. Terrault, L. Ojo, J.E. Hay, J. Neuberger, E. Blumberg, L.W. Teperman. Long-term management of the successful adult liver transplant: 2012 practice guideline by the American Association for the Study of Liver Diseases and the American Society of Transplantation. *Liver Transpl*19 (2013) 3-26,
- [3] X. Qi., X. Guo, C. Su. Clinical Outcomes of the Transplantation of Stem Cells from Various Sources of for the Management of Liver Cirrhosis: A Systematic Review and Meta-Analysis. *Curr Stem Cell Res Ther*10 (2), (2015) 166-180.
- [4] M.C. Hansel, R. Gramignoli, K.J. Skvorak, K. Dorko, F. Marongiu, W. Blake, J. Davila, et al. The history and use of human hepatocytes for the treatment of liver diseases: the first 100 patients. *CurrProtocToxicol*62 (2014) 14 12 11-14 12 23,
- [5] A.A. Khan, N. Parveen, V.S. Mahaboob, A. Rajendraprasad, H.R. Ravindraprakash, J. Venkateswarlu, P. Rao, et al. Treatment of Crigler-Najjar Syndrome type 1 by hepatic progenitor cell transplantation: a simple procedure for management of hyperbilirubinemia. *Transplant Proc*40 (2008) 1148-1150,
- [6] P.A. Lysy, M. Najimi, X. Stephenne, A. Bourgois, F. Smets, E.M. Sokal. Liver cell transplantation for Crigler-Najjar syndrome type I: update and perspectives. *World J Gastroenterol*14 (2008) 3464-3470.
- [7] I.J. Fox, J.R. Chowdhury, S.S. Kaufman, T.C. Goertzen, N.R. Chowdhury, P.I. Warkentin, K. Dorko, et al. Treatment of the Crigler-Najjar syndrome type I with hepatocyte transplantation. *N Engl J Med* 338 (1998) 1422-1426,
- [8] J. Meyburg, F. Hoerster, J. Schmidt, J. Poeschl, G.F. Hoffmann, J.P. Schenk. Monitoring of intraportal liver cell application in children. *Cell Transplant* 19 (2010) 629-638,
- [9] J. Meyburg, A.M. Das, F. Hoerster, M. Lindner, H. Kriegbaum, G. Engelmann, J. Schmidt, et al. One liver for four children: first clinical series of liver cell transplantation for severe neonatal urea cycle defects. *Transplantation* 87 (2009) 636-641,
- [10] S.N. Bhatia, G.H. Underhill, K.S. Zaret, I.J. Fox, G.H. Underhill, K.S., Zaret, I.J. Fox. Cell and tissue engineering for liver disease. *SciTransl Med* 6 (2014) 245sr242,
- [11] L. Peng, D.Y. Xie, B.L. Lin, J. Liu, H.P. Zhu, C. Xie, Y.B. Zheng, Z.L. Gao. Autologous bone marrow mesenchymal stem cell transplantation in liver failure patients caused by hepatitis B: short-term and long-term outcomes. *Hepatology*54 (2011) 820-828,
- [12] S.Y An, Y.J. Jang, H.J. Lim, J. Han, J. Lee, G. Lee, J.Y. Park, et al. Milk Fat Globule-EGF Factor 8, Secreted by Mesenchymal Stem Cells, Protects Against Liver Fibrosis in Mice. *Gastroenterology*. (9) 2016. pii: S0016-5085(16)35463-4.
- [13] P. Andreone, L. Catani, C. Margini, L. Brodosi, S. Lorenzini, D. Sollazzo, B. Nicolini, R. et al. Reinfusion of highly purified CD133+ bone marrow-derived stem/progenitor cells in patients with end-stage liver disease: A phase I clinical trial. *Dig Liver Dis*. 47 (2015) 1059-1066.
- [14] P.N. Newsome, R. Fox, A. King, N. Than, J. Moore, C. Corbett, S. Townsend, et al. A multicentre, phase ii, open-label, randomised controlled trial of repeated autologous infusions of granulocyte colony stimulating factor mobilised cd133+ bone marrow stem cells in patients with cirrhosis. *Journal of Hepatology* 64 (2016) S183–S212.

- [15] V. Cardinale, Y. Wang, G. Carpino, C.B. Cui, M. Gatto, M. Rossi, P.B. Berloco, et al. Multipotent stem/progenitor cells in human biliary tree give rise to hepatocytes, cholangiocytes, and pancreatic islets. *Hepatology* 54 (2011) 2159-2172,
- [16] G. Carpino, V. Cardinale, R. Gentile, P. Onori, R. Semeraro, A. Franchitto, Y. Wang, et al. Evidence for multipotent endodermal stem/progenitor cell populations in human gallbladder. *J Hepatol* 60 (2014) 1194-1202,
- [17] G. Carpino, V. Cardinale, P. Onori, A. Franchitto, P.B. Berloco, M. Rossi, Y. Wang, et al. Biliary tree stem/progenitor cells in glands of extrahepatic and intrahepatic bile ducts: an anatomical in situ study yielding evidence of maturational lineages. *J Anat* 220 (2012) 186-199,
- [18] R. Semeraro, G. Carpino, V. Cardinale, P. Onori, R. Gentile, A. Cantafora, A. Franchitto, et al. Multipotent stem/progenitor cells in the human foetal biliary tree. *J Hepatol* 57 (2012) 987-994,
- [19] Y. Wang, G. Lanzoni, G. Carpino, C.B. Cui, J. Dominguez-Bendala, E. Wauthier, V. Cardinale, et al. Biliary tree stem cells, precursors to pancreatic committed progenitors: evidence for possible life-long pancreatic organogenesis. *Stem Cells* 31 (2013) 1966-1979,
- [20] G. Lanzoni, T. Oikawa, Y. Wang, C.B. Cui, G. Carpino, V. Cardinale, D. Gerber, et al. Concise review: clinical programs of stem cell therapies for liver and pancreas. *Stem Cells* 31 (2013) 2047-2060,
- [21] V. Cardinale, Y. Wang, G. Carpino, G. Mendel, G. Alpini, E. Gaudio, L.M. Reid, et al. The biliary tree—a reservoir of multipotent stem cells. *Nat Rev Gastroenterol Hepatol* (2012) 231-240,
- [22] R. Turner, O. Lozoya, Y. Wang, V. Cardinale, E. Gaudio, G. Alpini, G. Mendel, et al. Human hepatic stem cell and maturational liver lineage biology. *Hepatology* 53 (2011) 1035-1045,
- [23] E. Schmelzer, L. Zhang, A. Bruce, E. Wauthier, J. Ludlow, H.L. Yao, N. Moss, et al. Human hepatic stem cells from fetal and postnatal donors. *J Exp Med* 204 (2007) 1973-1987,
- [24] R. Turner, G. Mendel, E. Wauthier, C. Barbier, L.M. Reid. Hyaluronan-supplemented buffers preserve adhesion mechanisms facilitating cryopreservation of human hepatic stem/progenitor cells. *Cell Transplant* 21 (2012) 2257-2266,
- [25] M. Riccio, G. Carnevale, V. Cardinale, L. Gibellini, S. De Biasi, A. Pisciotta, G. Carpino, et al. The Fas/Fas ligand apoptosis pathway underlies immunomodulatory properties of human biliary tree stem/progenitor cells. *J Hepatol.* 61 (2014) 1097-1105.
- [26] J.H. Lee, H.J. Park, Y.A. Kim, D.H. Lee, J.K. Noh, C.H. Kwon, S.M. Jung, et al. Differentiation and major histocompatibility complex antigen expression in human liver-derived stem cells. *Transplant Proc* 44 (2012) 1113-1115,
- [27] A.A. Khan, N. Parveen, V.S. Mahaboob, A. Rajendraprasad, H.R. Ravindraprakash, J. Venkateswarlu, P. Rao, et al. Management of hyperbilirubinemia in biliary atresia by hepatic progenitor cell transplantation through hepatic artery: a case report. *Transplant Proc* 40 (2008) 1153-1155,
- [28] G. Pietrosi, G. Vizzini, J. Gerlach, C. Chinnici, A. Luca, G. Amico, M. D'Amato, et al. Phase I-II Matched Case-Control Study of Human Fetal Liver Cell Transplantation for Treatment of Chronic Liver Disease. *Cell Transplant*, (2014)
- [29] V. Cardinale, G. Carpino, R. Gentile, C. Napoletano, H. Rahmi, A. Franchitto, R. Semeraro, et al. Transplantation of human fetal biliary tree stem/progenitor cells into two patients with advanced liver cirrhosis. *BMC Gastroenterology*, 14 (2014).
- [30] M. Muraca. Evolving concepts in cell therapy of liver disease and current clinical perspectives. *Dig Liver Dis.* 43 (2011) :180-187.

- [31] B. Gridelli, G. Vizzini, G. Pietrosi, A. Luca, M. Spada, S. Gruttadauria, D. Cintorino, et al. Efficient human fetal liver cell isolation protocol based on vascular perfusion for liver cell-based therapy and case report on cell transplantation. *B1Liver Transpl.* 18 (2012) 226-237.
- [32] A. Weber, M.T. Groyer-Picard, D. Franco, I. Dagher. Hepatocyte Transplantation in Animal Models. *Liver Transplantation.* 15 (2009) 7-14.
- [33] A.A. Khan, M.V. Shaik, N. Parveen, A. Rajendraprasad, M.A. Aleem, M.A. Habeeb, M. Aejaaz, et al. Human fetal liver-derived stem cell transplantation as supportive modality in the management of end-stage decompensated liver cirrhosis. *Cell Transplant.* 19 (2010) 409-418.
- [34] B.V. Nusgens. Hyaluronic acids and extracellular matrix: a primitive molecule?. *Ann DermatolVenereol.* 137 (2010) S3-8.
- [35] J.A. Burdick, G.D. Prestwich. Hyaluronic acid hydrogels for biomedical applications. *Adv Mater.* 25 (2011) 41-56. doi: 10.1002/adma.201003963. Epub 2011 Mar 10.
- [36] B.P. Toole. Hyaluronan in morphogenesis. *Semin Cell Dev Biol.* 12 (2001) 79-87.
- [37] A.E. Postlethwaite, J.M. Seyer, A.H. Kang. Chemotactic attraction of human fibroblast to type I, II and III collagens and collagen derived peptides. *ProcNatlAcad Sci U S A.* 75 (1978) 871-875.
- [38] J.L. Ifkovits, E. Tous, M. Minakawa, M. Morita, J.D. Robb, K.J. Koomalsingh, J.H. 3rdGorman, et al. Injectabel hydrogel properties influence infarct expansion and extent of postinfarction left ventricular remodeling in an ovine model. *Proc Natl AcadSci U S A.* (22) 2010 11507-11512.
- [39] S.J. Yoon, Y.H. Fang, C.H. Lim, B.S. Kim, H.S. Son, Y. Park, K. Sun. Regeneration of ischemic heart using hyaluronic acid-based injectable hydrogel. *J Biomed Mater Res B ApplBiomater* 91 (2009) 163-171.
- [40] L. Almany, D. Seliktar. Biosynthetic hydrogel scaffolds made from fibrinogen and polyethylene glycol for 3D cell cultures. *Biomaterials* 26 (2005) 2467-2477.
- [41] J.L. Drury, D.J. Mooney. Hydrogels for tissue engineering: scaffold design variables and applications. *Biomaterials* 24 (2003) 4337-4351.
- [42] G.D. Prestwich, Hyaluronic acid-based clinical biomaterials derived for cell and molecule delivery in regenerative medicine. *J Control Release* 155 (2011) 193-199.
- [43] G.D. Prestwich, T. Ghaly, P. Brudnicki, B. Ratliff, M.S. Goligorsky. Bioartificial stem cell niches: Engineering a regenerative microenvironment. *regenerative Nephrology First Edition*, (2011) 245-256.
- [44] C.Y. Chang, A.T. Chan, P.A. Armstrong, H.C. Luo, T. Higuchi, I.A. Strehin, S. Vakrou, et al. Hyaluronic acid-human blood hydrogels for stem cell transplantation. *Biomaterials* 33 (2012) 8026-8033.
- [45] O.A. Lozoya, E. Wauthier, R.A. Turner, C. Barbier, G.D. Prestwich, F. Guilak, R. Superfine, et al. Regulation of hepatic stem/progenitor phenotype by microenvironment stiffness in hydrogel models of the human liver cell stem niche. *Biomaterials.* 30 (2010) 7389-73402.
- [46] R.A. Turner, E. Wauthier, O. Lozoya, R. McClelland, J.E. Bowsher, C. Barbier, G. Prestwich, et al. Successful transplantation of human hepatic stem cells with restricted localization to liver using hyaluronan grafts. *Hepatology.* 57 (2013) 775-784.
- [47] L. Nevi, G. Carpino, D. Costantini, V. Cardinale, O. Riccioni, S. Di Matteo, F. Melandro, et al. Hyaluronan coating improves liver engraftment of transplanted human biliary tree stem/progenitor cells. *Stem Cell Research & Therapy*

- [48] M. Gudowska, E. Gruszevska, A. Panasiuk, B. Cylwik, R. Flisiak, M. Swiderska, M. Szmikowski, et al. Hyaluronic acid concentration in liver diseases. *ClinExp Med.* 16 (2016) 523–528.
- [49] Y.L. Ni, Z.R. Tang, W.X. Cao, H. Lin, Y.J. Fan, L.K. Guo, X. Zhang. Tough and elastic hydrogel of hyaluronic acid and chondroitin sulfate as potential cell scaffold materials. *Int J BiolMacromol.* 74 (2015) 367–375.
- [50] Y.M. Yan, X.S. Zuo, D.Y. Wei. Concise Review: Emerging role of CD44 in cancer stem cells: a promising biomarker and therapeutic target. *Stem Cells Transl Med.* 4 (2015) 1033–1043.
- [51] J. Lesley, V.C. Hascall, M. Tammi, R. Hyman. Hyaluronan binding by cell surface CD44. *J Biol Chem.* 275 (2000) 26967–26975.
- [52] D.S. Schmidt, P. Klingbeil, M. Schnolzer, M. Zoller. CD44 variant isoforms associate with tetraspanins and EpCAM. *Exp Cell Res.* 297 (2004) 329–347.
- [53] X.Z. Shu, S. Ahmad, Y.C. Liu, G.D. Prestwich. Synthesis and evaluation of injectable, in situ crosslinkable synthetic extracellular matrices for tissue engineering. *J Biomed Mater Res A.* 79 (2006) 902–912 .
- [54] M.J. Kujawa, D.A. Carrino, A.I. Caplan. Substrate-bonded hyaluronic-acid exhibits a size-dependent stimulation of chondrogenic differentiation of stage-24 limb mesenchymal cells in culture. *Dev Biol.* 114 (1986) 519–528.
- [55] G.D. Prestwich, I.E. Erickson, T.I. Zarembinski, M. West, W.P. Tew. The translational imperative: making cell therapy simple and effective. *Acta Biomater.* 8 (2012) 4200–4207. doi: 10.1016/j.actbio.2012.06.043. Epub 2012 Jul 7.
- [56] K.A. Soltys, K. Setoyama, E.N. Tafaleng, A. SotoGutiérrez, J. Fong, K. Fukumitsu, T. Nishikawa, et al. Host conditioning and rejection monitoring in hepatocyte transplantation in humans. *J Hepatol.* 24 (2016) pii: S0168-8278(16)30750-4. doi: 10.1016/j.jhep.2016.12.017.
- [57] R.B. De Santana, C.M. De Santana. Human intrabony defect regeneration with rhFGF-2 and hyaluronic acid—a randomized controlled clinical trial. *J Clin Periodontol.* 42 (2015) 658–665.
- [58] Haridass D, Yuan Q, Becker PD, Cantz T, Iken M, Rothe M, et al. Repopulation efficiencies of adult hepatocytes, fetal liver progenitor cells, and embryonic stem cell-derived hepatic cells in albumin-promoter-enhancer urokinase-type plasminogen activator mice. *Am J Pathol.* 175 (2009) 1483–1492.
- [59] Jorns C, Nowak G, Nemeth A, Zemack H, Mörk LM, Johansson H, et al. De Novo Donor-Specific HLA Antibody Formation in Two Patients With Crigler-Najjar Syndrome Type I Following HumanHepatocyte Transplantation With Partial Hepatectomy Preconditioning. *Am J Transplant.* 16 (2016) 1021–1030.
- [60] Dagher I, Boudechiche L, Branger J, Coulomb-Lhermine A, Parouchev A, Sentilhes L, et al. Efficient hepatocyte engraftment in a nonhuman primate model after partial portal vein embolization. *Transplantation.* 82 (2006) 1067–1073.
- [61] Dagher I, Nguyen TH, Groyer-Picard MT, Lainas P, Mainot S, Guettier C, et al. Efficient hepatocyte engraftment and long-term transgene expression after reversible portal embolization in nonhuman primates. *Hepatology.* 49 (2009) 950–959.
- [62] Wang Y, Cui CB, Yamauchi M, Miguez P, Roach M, Malavarca R, et al. Lineage restriction of human hepatic stem cells to mature fates is made efficient by tissue-specific biomatrix scaffolds. *Hepatology.* 53 (2011) 293–305.
- [63] Uygun BE, Yarmush ML, Uygun K. Application of whole-organ tissue engineering in hepatology. *Nat Rev Gastroenterol Hepatol.* 9 (2012): 738–744.

- [64] Joshi M, B Patil P, He Z, Holgersson J, Olausson M, Sumitran-Holgersson S. Fetal liver-derived mesenchymal stromal cells augment engraftment of transplanted hepatocytes. *Cytotherapy*. 14 (2012) 657-669
- [65] Mitry RR, Jitraruch S, Iansante V, Dhawan A. Alginate Encapsulation of Human Hepatocytes and Assessment of Microbeads. *Methods Mol Biol*. 2017;1506:273-281.
- [66] Carpino G, Gaudio E. Cell sources for regenerative medicine of the liver and endoderm organs: strategies and perspectives. *Stem Cell Investig*. 3 (2016) 91.
- [67] Sampaziotis F, Cardoso de Brito M, Madrigal P, Bertero A, Saeb-Parsy K, Soares FA, Schrumpf E, Melum E, Karlsen TH, Bradley JA, Gelson WT, Davies S, Baker A, Kaser A, Alexander GJ, Hannan NR, Vallier L. Cholangiocytes derived from human induced pluripotent stem cells for disease modeling and drug validation. *Nat Biotechnol*. 2015 Aug;33(8):845-52.
- [68] Sampaziotis F, Cardoso de Brito M, Madrigal P, Bertero A, Saeb-Parsy K, Soares FA, Schrumpf E, Melum E, Karlsen TH, Bradley JA, Gelson WT, Davies S, Baker A, Kaser A, Alexander GJ, Hannan NR, Vallier L. Cholangiocytes derived from human induced pluripotent stem cells for disease modeling and drug validation. *Nat Biotechnol*. 2015 Aug;33(8):845-52.;
- [69] Ogawa M, Ogawa S, Bear CE, Ahmadi S, Chin S, Li B, Grompe M, Keller G, Kamath BM, Ghanekar A. Directed differentiation of cholangiocytes from human pluripotent stem cells. *Nat Biotechnol*. 2015 Aug;33(8):853-61. doi: 10.1038/nbt.3294. Epub 2015 Jul 13.
- [70] Sampaziotis F, de Brito MC, Geti I, Bertero A, Hannan NR, Vallier L. Directed differentiation of human induced pluripotent stem cells into functional cholangiocyte-like cells. *Nat Protoc*. 2017 Apr;12(4):814-827;
- [71] Ghanekar A, Kamath BM. Cholangiocytes derived from induced pluripotent stem cells for disease modeling. *Curr Opin Gastroenterol*. 2016 May;32(3):210-5.
- [72] Wang Y, Qin J, Wang S, Zhang W, Duan J, Zhang J, et al. Conversion of Human Gastric Epithelial Cells to Multipotent Endodermal Progenitors using Defined Small Molecules. *Cell Stem Cell*. 19 (2016) 449-461.

Legend to **Figure 1. Human biliary tree stem/progenitor therapy protocol.** The isolation protocol involves either the biliary tree from livers of therapeutic abortions or the extrahepatic biliary tree from donors which are digested mechanically and enzymatically to obtain a cell suspension which is filtered and then subjected to cell immune sorting using EpCAM microbeads. The EpCAM sorted cells underwent immediately to identity test by flow cytometry analysis and microbiological tests before sudden injection into the hepatic artery of patients which have been admitted to the transplant unit few hours in advance. TO: therapeutic abortion. EpCAM: epithelial adhesion molecule.

Human biliary tree stem/progenitor therapy protocol

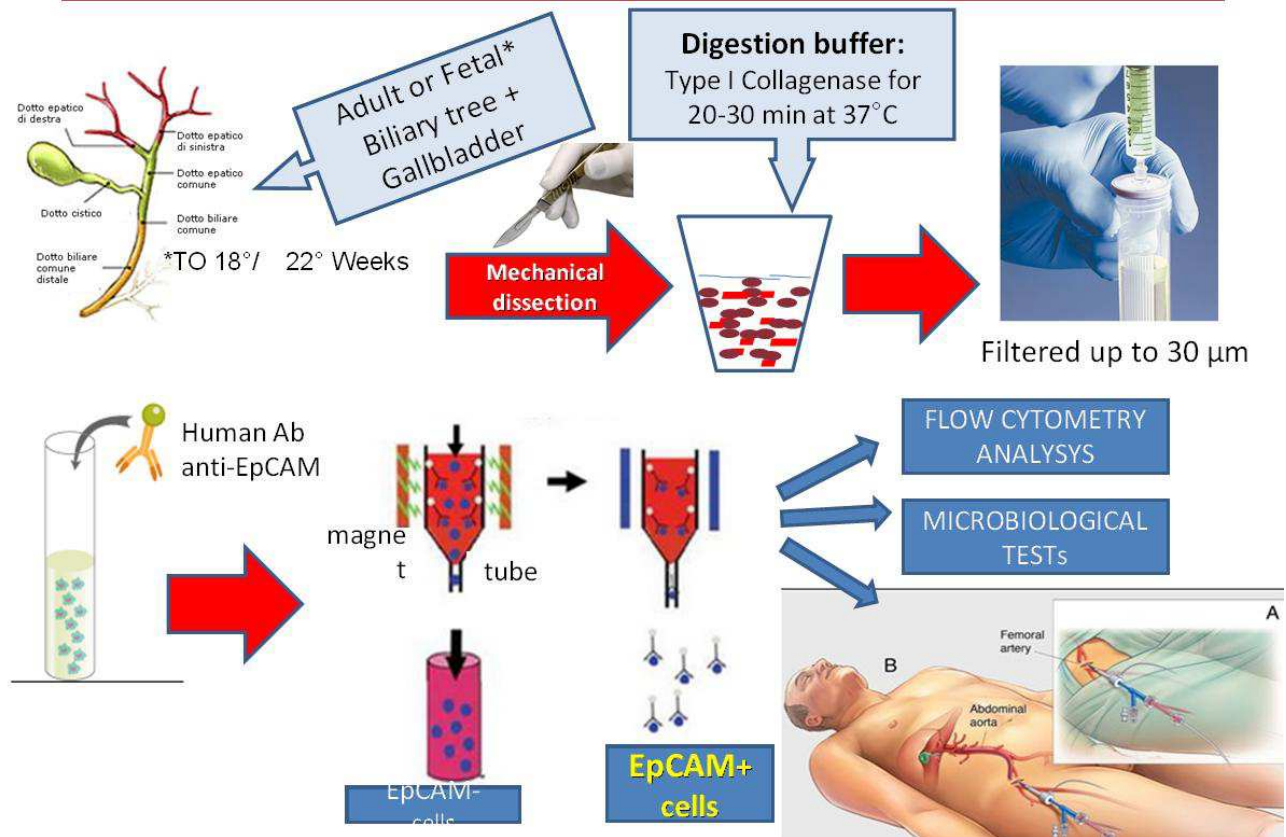


Figure 1

BOX 1. FACTORS POTENTIALLY AFFECTING THE LIVER ENGRAFTMENT OF TRANPLANTED CELLS
<p>Host factors</p> <ul style="list-style-type: none"> • Host liver extracellular matrix: <ul style="list-style-type: none"> - major driver of adhesion molecules expression • Host liver pathologic conditions and related alterations of vasculature perfusion: <ul style="list-style-type: none"> - liver engraftment of hHpSCs or hBTSCs increases in the mouse model of CCl4-induced liver fibrosis with respect to normal liver (23, 16) - engraftment and repopulation of transplanted cells has been demonstrated in genetic mice models of liver repopulation, after acute toxic liver injuries or in mice treated with compounds (retrotransposon and monocrotaline) blocking hepatocyte proliferation. These models provide space for the growth of engrafted cells or a selective advantage for transplanted cells (58)
<p>Factors related to the cell source</p> <ul style="list-style-type: none"> • Cell size • Proliferation: <ul style="list-style-type: none"> - hBTSCs have intrinsic proliferative advantages (15, 16); the percentage of hepatic stem cells expressing proliferation markers was on average 45 times greater than the percentage of mature hepatocytes (31) • Immunogenicity issues: <ul style="list-style-type: none"> - fetal hepatic and biliary stem/progenitor cells express no or minimal class I and II MHC antigens (reviewed in 25) - hBTSCs modulated T-cell apoptosis through the Fas/Fas ligand pathway (25) • Tolerance to toxins and ischemia: <ul style="list-style-type: none"> - high expression of multidrug resistance proteins in stem cells (reviewed in 20 and 21)
<p>Administration Routes</p> <ul style="list-style-type: none"> • Hepatic delivery: <ul style="list-style-type: none"> - via portal vein or hepatic artery or direct injection into the liver • Ectopic implantation: <ul style="list-style-type: none"> - Into the peritoneum or into the spleen
<p>Cell rejection: immunomediated cell rejection</p>

Abbreviations: hHpSCs, human hepatic stem/progenitor cells; hBTSCs human biliary stem/progenitor cells; CCL4, carbon tetrachloride; MHC, major histocompatibility complex

<p>BOX 2. STRATEGIES ADOPTED TO INCREASE LIVER ENGRAFTMENT AND REPOPULATION</p> <p>Host pre-conditioning (clinical and pre-clinical evidences obtained in hepatocyte transplantation):</p> <ul style="list-style-type: none"> • Preparative hepatic irradiation in human and animal models (56) • Hepatic resection in human and animal models (59) • Portal vein embolization in animal models (60, 61) <p>Use of matrix components and grafting strategies (pre-clinical studies):</p> <ul style="list-style-type: none"> • Preparative injection of fibrin in animal models of hepatocyte transplantation improves the engraftment (32) • Bio-matrix scaffold components increase proliferation and differentiation of human hepatic stem/progenitor cells <i>in vitro</i> (62) • Hyaluronic acid-coating increase the engraftment of human biliary tree stem cells in mice w/o damage when injected by vascular route into portal vein (47) • Grafting strategies have been devised for transplantation of human hepatic stem cells (hHpSCs) embedded into a mix of soluble signals and extracellular matrix biomaterials (hyaluronans, type III collagen, laminin): grafted cells remained localized into the livers, resulting in a larger bolus of engrafted cells in the host livers, with respect to transplantation by direct injection or via a vascular route, under quiescent conditions and with potential for more rapid expansion under conditions of liver injury (46) • Combination strategies: <ul style="list-style-type: none"> - advanced grafting devices directly transplanted; - liver scaffolds transplantation (63) <p>Cotransplantation (pre-clinical stages):</p> <ul style="list-style-type: none"> • Some evidence exist that cotransplantation of stromal cells facilitates the process of hepatocyte engraftment and proliferation most likely by the release of cytokines and growth factors in mice (64) • Pietrosi and co-workers treated 9 patients affected by end-stage liver disease with intrasplenic infusion of a total cell population obtained from the fetal liver, containing cells expressing typical MSC and HSC markers and found that the procedure was safe and well tolerated with positive effects on clinical scores and on encephalopathy (28) 	<p>Cell sources</p> <ul style="list-style-type: none"> - Stem cell therapies of liver diseases with hepatic and biliary stem/progenitor cells are based on favorable biologic properties of stem cells would improve cell engraftment and repopulation, and attenuate immunoreaction (see ref.# 27-29). However, no clinical trial comparing different cell types in humans is available. - In a competitive liver repopulation mouse model (heterozygous alb-uPA^{tg1/2} mice), engraftment and repopulation capacities of mature hepatocytes were always superior to early and late hepatoblasts as well as embryonic stem (ES)- and iPS cell-derived cells. Any of the tested postnatal non-hepatocyte cells, such as hematopoietic stem cells or mesenchymal stromal cells, did not engraft long term and did not repopulate the liver (58) <p>Administration Route:</p> <ul style="list-style-type: none"> - No trial exists in animal models comparing administration via portal vein versus hepatic artery. The former is mainly used for hepatocyte transplantation, while the latter for hepatic or biliary tree/stem cell transplantation - Direct injection versus vascular route: direct injected hHpSCs embedded in bio-matrices remained localized into the livers, resulting in a larger bolus of engrafted cells in the host with respect to transplantation via vascular route (46) <p>Cell rejection:</p> <ul style="list-style-type: none"> - Pharmacology regimens limit immunerejection of human hepatocytes. - Hepatocytes microencapsulated in alginate limit immunoreaction thanks to the limited access of transplanted cells to host immune cells (65) - The use of fetal or adult hepatic and biliary stem/progenitor cells alleviated the immunogenicity
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issues since these cells express no or minimal class I and II MHC antigens, and since hBTSCs modulated T-cell apoptosis through the Fas/Fas ligand pathway (reviewed in 25). No immunosuppression have been required in clinical trials (27, 29)

Abbreviations: hHpSCs, human hepatic stem/progenitor cells; hBTSCs human biliary stem/progenitor cells; CCL4, carbon tetrachloride; MHC, major histocompatibility complex; MSC, mesenchymal stem cell; HSC, hematopoietic stem cells, ESCs, embryonic stem cells; iPSCs, induced pluripotent stem cells

Bulletpoints

- Liver cell therapy aimed to sustain function, regeneration, and to reduce the scarring
- Hepatocyte transplantation determined clinical benefits in inborn errors of metabolism
- Stemcells from fetal or adult liver is a suitable alternative
- Multipotent stemcells reside in PBGs and crypts of gallbladder in humans
- hBTSCs have been used in cirrhotic patients with significant benefits