

Accepted Manuscript

Assessment of drinking water quality at the tap using fluorescence spectroscopy

Masoumeh Heibati, Colin A. Stedmon, Karolina Stenroth, Sebastien Rauch, Jonas Toljander, Melle Säve-Söderbergh, Kathleen R. Murphy



PII: S0043-1354(17)30673-5

DOI: [10.1016/j.watres.2017.08.020](https://doi.org/10.1016/j.watres.2017.08.020)

Reference: WR 13143

To appear in: *Water Research*

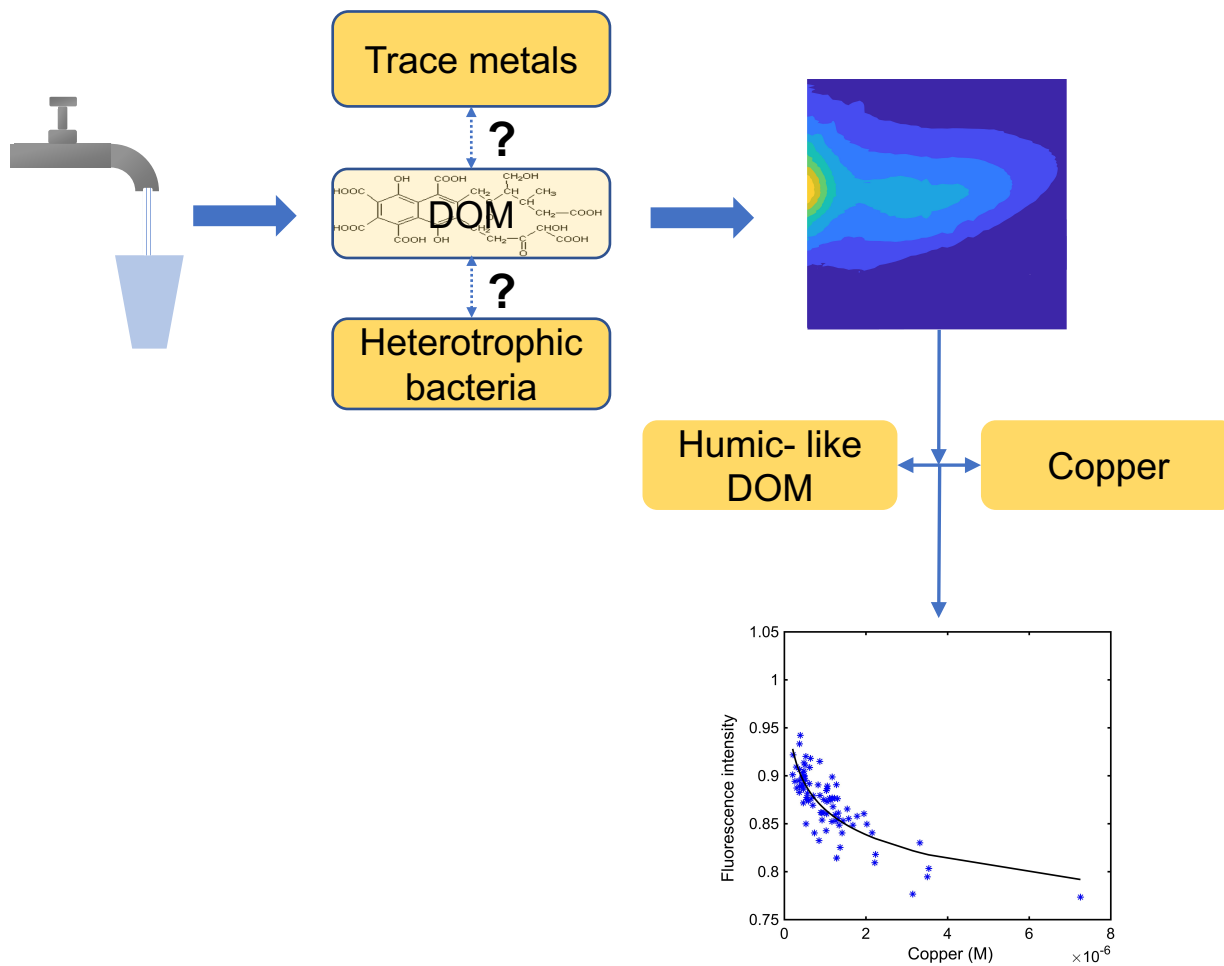
Received Date: 14 February 2017

Revised Date: 23 June 2017

Accepted Date: 8 August 2017

Please cite this article as: Heibati, M., Stedmon, C.A., Stenroth, K., Rauch, S., Toljander, J., Säve-Söderbergh, M., Murphy, K.R., Assessment of drinking water quality at the tap using fluorescence spectroscopy, *Water Research* (2017), doi: 10.1016/j.watres.2017.08.020.

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.



Assessment of drinking water quality at the tap using fluorescence spectroscopy

Masoumeh Heibati ^a, Colin A. Stedmon ^b, Karolina Stenroth
^c, Sebastien Rauch ^a, Jonas Toljander ^d, Melle Säve-Söderbergh
^{d,e}, Kathleen R. Murphy ^a

^a Department of Civil and Environmental Engineering, Water
Environment Technology, Gothenburg, Sweden

^b National Institute for Aquatic Resources, Technical University of
Denmark, Denmark

^c Gästrike Vatten AB, Gävle, Sweden

^d Science Division, National Food Agency, Uppsala, Sweden

^e Institute of Environmental Medicine, Karolinska Institutet, Stockholm,
Sweden

Acronyms and abbreviations:

DOM	Dissolved organic matter
CDOM	Coloured dissolved organic matter
NOM	Natural organic matter
DOC	Dissolved organic carbon
RSD	Relative standard deviation
TT	Triger threshold
HPC	Heterotrophic plate count
LoD	Limit of detection
EEMs	Excitation-emission matrices
PLS	Partial least squares
O-PLS	Orthogonal partial least squares

Abstract: Treated drinking water may become contaminated while travelling in the distribution system on the way to consumers. Elevated dissolved organic matter (DOM) at the tap relative to the water leaving the treatment plant is a potential indicator of contamination, and can be measured sensitively, inexpensively and potentially on-line via fluorescence and absorbance spectroscopy. Detecting elevated DOM requires potential contamination events to be distinguished from natural fluctuations in the system, but how much natural variation to expect in a stable distribution system is unknown. In this study, relationships between DOM optical properties, microbial indicator organisms and trace elements were investigated for households connected to a biologically-stable drinking water distribution system. Across the network, humic-like fluorescence intensities showed limited variation ($RSD = 3.5 - 4.4\%$), with half of measured variation explained by interactions with copper. After accounting for quenching by copper, fluorescence provided a very stable background signal ($RSD < 2.2\%$) against which a $\sim 2\%$ infiltration of soil water would be detectable. Smaller infiltrations would be detectable in the case of contamination by sewage with a strong tryptophan-like fluorescence signal. These findings indicate that DOM fluorescence is a sensitive indicator of water quality changes in drinking water networks, as long as potential interferents are taken into account.

26 Keywords: drinking water distribution, CDOM, natural
27 organic matter (NOM), PARAFAC

28 1. Introduction

29 Between leaving a treatment plant and arriving at the
30 consumer's tap, drinking water enters the distribution network
31 where it resides for periods that typically range from hours to
32 days. During this time, the drinking water may become
33 contaminated via a range of processes. Microbial water quality
34 can deteriorate in networks due to regrowth or entrainment of
35 untreated water through damaged pipes, presenting potentially
36 serious health risks to consumers (WHO 2014). In the United
37 States during 1971-2006, around 10% of disease outbreaks
38 caused by unsafe drinking water have been attributed to
39 deficiencies in the distribution network (Craun et al. 2010).

40 Microorganisms in drinking water distribution systems are
41 either part of the indigenous community or enter the system
42 where the pipe network integrity is compromised. Microbes
43 living in soil pore-waters can be entrained through cracks in
44 pipes and joints during negative pressure events (LeChevallier
45 et al. 2003). Inside the pipes, heterotrophic bacteria utilise
46 available organic substrate in the water as a source of carbon,
47 nutrients and energy. Changing flow conditions in the network
48 can also dislodge biofilms harbouring pathogenic species and

49 create conditions that favour opportunistic species, potentially
50 including pathogens (Manuel et al. 2007).

51 Obtaining rapid and affordable assessments of the microbial
52 quality of drinking water is a famously intractable problem.
53 Microbial indicator species including *Escherichia coli* (*E. coli*),
54 coliforms, *Enterococcus* spp. and total bacterial counts are
55 frequently monitored as proxies for pathogens that are
56 expensive or impractical to measure. Although the presence of
57 *E. coli* and coliforms indicates contamination, their absence
58 does not preclude the presence of other harmful organisms (Wu
59 et al. 2011). Microbial biomass is typically measured via
60 heterotrophic plate counts (HPCs), which quantifies bacteria
61 that grow by consuming organic nutrients, i.e. a small fraction
62 of total microorganisms in drinking water. The actual species
63 quantified by HPC depends on cultivation medium, incubation
64 temperature and incubation time (Allen et al. 2004). HPC levels
65 are not regulated, although abundances above 500 cfu/ml are
66 considered of potential concern, mainly due to interference
67 with the analytical detection of total coliforms. HPC analyses
68 typically take several days to implement assuming a well-
69 equipped laboratory (Allen et al. 2004), hindering a rapid
70 response to adverse measurements. Faster, cultivation-free
71 methods for assessing microbial biomass exist, including
72 adenosine tri-phosphate (ATP) and flow cytometry cell counts
73 (FC), but these methods are still relatively complex to

74 implement and interpret, preventing their widespread use for
75 monitoring distribution networks (Hammes et al. 2008, Van
76 der Wielen and Van der Kooij 2010).

77 Dissolved organic matter (DOM) is a heterogeneous mixture
78 of carbon-containing molecules present in all aquatic
79 ecosystems. Globally, DOM plays a key role in carbon and
80 nutrient cycling, and as a substrate for microbial growth, is one
81 of the main risk factors promoting microbiological growth in
82 distribution networks (Camper et al. 2003). DOM optical
83 properties (absorbance and fluorescence) are widely used for
84 studying changes in DOM composition and concentration
85 (Murphy et al. 2013). Although neither spectroscopic technique
86 necessarily directly measures the small bioavailable molecules
87 consumed by heterotrophic bacteria, numerous studies have
88 shown that optical measurements are nevertheless sensitive
89 proxies of the wider DOM pool and track subtle changes in
90 water quality (Stedmon et al. 2011, Stubbins et al. 2014). DOM
91 fluorescence is a sensitive tracer of sewage contamination,
92 correlating with *E. coli* abundances (Baker et al. 2015) and
93 nutrients (Baker and Inverarity 2004) across systems.

94 Absorbance spectroscopy is frequently used to track the
95 abundance of the coloured fraction of dissolved organic matter
96 (CDOM) in drinking water treatment systems (Weishaar et al.
97 2003), including in online applications (Chow et al. 2008).
98 Fluorescence spectroscopy is a much more sensitive

99 technology, and additionally tracks compositional changes in
100 DOM (Stedmon et al. 2011). However, studies of fluorescence
101 in drinking distribution systems are very few. Hambly et al.
102 (2010) surveyed houses serviced by two separate distribution
103 systems (potable and recycled non-potable), and concluded that
104 network cross-connections would be detectable from measuring
105 fluorescence intensities at the tap. However it remain to be seen
106 if organic matter fluorescence in drinking water networks is
107 both stable and predictable enough to offer a sound baseline to
108 identify contamination at point-of-use; and if the signal is
109 correlated to microbial abundances and other chemical
110 constituents in distribution systems.

111 Trace metals leached from pipe materials can potentially
112 interfere with spectroscopic measurements of DOM in drinking
113 water. In the presence of transition metals such as iron, copper
114 and aluminium, metal-DOM complexes can form which absorb
115 more strongly than un-complexed DOM while fluorescing less
116 (Senesi et al. 1991, Yan et al. 2013). Corrosion by cast iron,
117 galvanized iron and steel pipes are the main sources of iron in
118 drinking water (WHO 2014). Copper is seldom used for
119 municipal network pipes but is frequently used in household
120 plumbing and fixtures. The suppression or quenching of DOM
121 fluorescence by various metal ions has been studied in natural
122 aquatic systems (Ryan 1982, Yamashita and Jaffe 2008) and
123 wastewaters (Reynolds and Ahmad 1995). However, it is

124 uncertain whether metals would interfere to any significant
125 extent with DOM spectroscopic measurements in distribution
126 systems where concentrations of DOM and metals are both
127 low.

128 Chlorine is frequently applied at the end of drinking water
129 treatment to limit regrowth and other microbial risks in the
130 distribution network. In chlorinated networks, reactions
131 between organic matter and chlorine break down large DOM
132 molecules, decreasing aromaticity and fluorescence intensities
133 and shifting fluorescence emission spectra (Beggs et al. 2009,
134 Korshin et al. 1999). The effect of chlorine exposure on
135 fluorescence intensities approximately follows an exponential
136 decay curve, with rapid losses occurring at short reaction times
137 (minutes to hours) followed by gradual losses at long exposures
138 (Beggs et al. 2009). Chlorine could therefore be a confounding
139 factor for comparing fluorescence measurements at the tap,
140 particularly when chlorine doses are high and distribution times
141 vary greatly.

142 In this study, relationships between DOM optical properties,
143 microbial indicator organisms and trace element concentrations
144 were investigated in a drinking water distribution network. The
145 purpose was to assess whether DOM optical properties
146 measured at the tap correlate with, and are potential surrogate
147 indicators of, abundances of microbial indicator species. The
148 study area had no reoccurring chemical or biological water

quality issues, allowing determination of baseline conditions in the network and thresholds to be established for recognising significant changes in water quality. Also, since there is much interest in using DOM optical properties for online water quality monitoring, we investigated whether trace elements sourced from within the pipe network interfere with DOM optical measurements at the tap. If significant interferences occur, this may seriously limit the interpretation of online DOM measurements if trace elements are not monitored at the same time.

2. Material and methods

2.1. Sampling and analytical methods

A municipal drinking water distribution network in central eastern Sweden was surveyed. The Gävle distribution system forms a 486 km network of predominantly iron and plastic (polyethylene) pipes. The plant receives groundwater, adjusts the pH with sodium hydroxide, and chlorinates before releasing it into the distribution system. Due to the groundwater source, the outgoing drinking water is moderately hard (calcium and magnesium hardness $> 60 \text{ mgL}^{-1}$). NaClO is dosed at 0.3-0.4 mgL^{-1} producing total chlorine in the outgoing water of 0.1-0.15 mgL^{-1} . Residual chlorine at the plant reacts rapidly with the NOM in the water to produce total chlorine concentrations (total chlorine = residual chlorine + chlorine demand) usually

173 around 0.01-0.06 mgL⁻¹ in the taps of buildings along the
174 network. Thus the levels of free residual chlorine (FRC) in the
175 network are much lower than is typical (>0.2 mgL⁻¹) to ensure
176 a disinfection effect at the point of use (WHO 2014).

177 Drinking water samples were collected in winter (December
178 1-2, 2015) at 87 locations in houses and public buildings
179 connected to the distribution system. Sampling locations were
180 selected so as to encompass the entire range of water residence
181 times experienced by households on the network (0.5-50 h).
182 Water samples were obtained from taps in the kitchen or
183 bathroom, after first flushing for 5 min. Replicate samples (n =
184 2) were collected at a subset of sites (n = 10) to assess
185 experimental and analytical reproducibility; these were both
186 collected and measured completely independently of one
187 another. Samples for microbial analyses and turbidity
188 measurement were collected in sterile plastic (HDPE) bottles,
189 DOM (dissolved organic carbon (DOC), absorbance, and
190 fluorescence) samples in ashed amber glass bottles (DOM), and
191 trace metal samples in acid-washed polyethylene tubes. DOM
192 and metals samples were filtered on-site through pre-flushed,
193 0.45 µm cellulose acetate filters; lab tests indicated no
194 measurable fluorescence after flushing with 120 mL of Milli-Q.
195 Absorbance and fluorescence samples were analysed at
196 Chalmers within 48 h of sampling. Trace metal samples were
197 acidified to 1% v/v with high purity HNO₃ and analysed within

15 days. DOC samples were acidified to pH = 2 with high-purity HCl, stored at 4 °C, and analysed within two months. Microbial samples and turbidity were analysed the following day at a commercial analytical laboratory (Eurofins).

In the laboratory, CDOM fluorescence and absorbance were measured in a 1-cm quartz cuvette using an Aqualog spectrofluorometer (Horiba Scientific). Excitation-emission matrices (EEMs) were obtained with 3 s integration time for excitation wavelengths 220-600 nm at 3-nm intervals and emission wavelengths of 240-800 nm at 2.3-nm intervals. Blank EEMs were acquired daily from ultra-pure water sealed in a quartz fluorometer cell and from MilliQ water. EEMs were spectrally corrected for instrumental biases and concentration effects according to established methods (Murphy et al. 2010). DOC was measured using a Shimadzu TOC-V_{CPH} carbon analyser, using the non-purgeable organic carbon (NPOC) method (EN 1484: 1997).

Concentrations of ten metals (Al, Cd, Cr, Cu, Fe, Mg, Mn, Ni, Pb and Zn) were determined by inductively coupled plasma mass spectrometry (ICP-MS) using a Thermo Scientific iCAP Q spectrometer. The instrument was operated in standard mode for all elements, except for Fe and Ni which were analysed in kinetic energy discrimination (KED) mode with He as collision gas.

Microbial analyses were performed according to standard

223 methods. Culturable heterotrophic bacteria counts were
224 determined using the ISO HPC method 6222-M (ISO 6222:
225 1999) which involves incubation at 22°C for three or seven
226 days. *E. Coli* and coliforms were enumerated by the IDEXX-
227 Colilert method and *Enterococcus* spp. by the IDEXX-
228 Enterolert method. Turbidity was measured using the SS-EN
229 ISO 7027 method. In historical surveys of the distribution
230 system (unpublished data), culturable microorganisms and
231 slow-growing bacteria abundances were generally below 10
232 and 100 cfu/ml, respectively.

233 Chlorine was not measured during this survey; however,
234 routine monitoring data are collected approximately monthly
235 and indicate very low residual chlorine in the network. In
236 samples collected immediately before and after this survey (n =
237 13), total chlorine was 0.03 mg/L (median) with a maximum of
238 0.04 mg/L at monitoring sites with distribution times of 8-41
239 hr; these numbers thus represent upper limits for chlorine
240 residuals at the monitoring sites. These low values are
241 consistent with long-term datasets archived with the Swedish
242 Geological Survey (Vattentäktsarkivet 2016): in 2012-2015
243 total chlorine was typically below 0.05 mg/L (90th percentile =
244 0.08 mg/L, n = 400) at monitoring stations along this network.
245 In this study, we use the chlorine reaction time as a proxy for
246 chlorine residuals (Korshin et al. 2002). For all samples in this
247 study, the chlorine reaction time exceeded 57 hours (2.4 days);

248 therefore, it is expected that the Cl residuals at the time of
249 fluorescence analysis were well below the upper limits
250 indicated by the routine monitoring datasets.

251 To simulate the contamination of drinking water pipes by
252 soil, and determine detection thresholds for observing the
253 contamination, a serial dilution was performed of soil water
254 added to drinking water. Soil was obtained from an urban area
255 at a depth of approximate 1 m and its organic carbon content
256 estimated by loss of ignition. The stock solution (2 g soil in 1 L
257 of tap water) was mixed on a magnetic stirrer for 24 h then
258 filtered through cellulose acetate (0.45 μm). The dilution series
259 was prepared by diluting the stock solution using Milli-Q for
260 13 dilution factors between 1 and 1/200. Fluorescence and
261 absorbance were measured the same day and DOC within three
262 days.

263 2.2. Statistical methods

264 2.2.1. Relative standard deviation and detection limits

265 Independently-measured replicate samples were used to
266 assess experimental and analytical error. Relative standard
267 deviations ($\text{RSD} = \text{standard deviation}/\text{mean}$) are independent of
268 scale and were used to compare how precisely different
269 variables could be measured. Analytical detection limits were
270 calculated as three times the standard deviation of triplicate
271 blanks. Trigger thresholds (TT) were also determined, defined
272 as the threshold for recognising a significantly elevated level of

273 a tracer, for example due to its entrainment in the network via a
 274 cracked pipe (equation 1).

$$275 \quad TT = \bar{x} + 3s \quad (1)$$

276 In equation 1, \bar{x} and s are the average and standard deviation
 277 of measurements from samples collected across the whole
 278 network.

279 2.2.2. PARAFAC model

280 The fluorescence measurements generated a three
 281 dimensional dataset of EEMs ($N = 87$, after averaging data
 282 from experimental replicates). Within each EEM, the measured
 283 trilinear data can be modelled as the sum of a limited number
 284 of independently-varying fluorescence signals (Bro 1997).
 285 These independent signals can be quantified using the
 286 PARAFAC algorithm, which identifies the best-fitting
 287 excitation and emission spectra for each independent signal
 288 (termed a ‘component’) and its relative concentration in each
 289 sample. PARAFAC modelling was implemented on the
 290 corrected dataset using the *N-Way* and drEEM toolboxes for
 291 MATLAB according to established methods (Andersson and
 292 Bro 2000, Murphy et al. 2013). Modelling performed with non-
 293 negativity constraints on all modes.

294 PARAFAC models were investigated with two to seven
 295 components, and split-half analysis, jack-knifing, and residual

analysis used to select the most appropriate model. This process identified four independently-varying signals producing a four-component PARAFAC model and their intensities (F1-F4) in each sample (Murphy et al. 2013).

2.2.3. PLS model

Multivariate calibration is often used for process control when it is necessary to predict variables (Y) that are expensive or time-consuming to measure from a set of correlated variables (a matrix of X variables) that are measured more easily. In the context of drinking water monitoring, it would be desirable to predict microbial abundances from one or more easily-obtained chemical measurements. Partial least square (PLS) regression is often used for multivariate calibration since it performs well even when the number of predictor variables is high and some variables correlate with each other. When the PLS model is orthogonalised (O-PLS), all variation correlated to the response variable is compressed in the first latent variable, which greatly simplifies interpretation (Trygg and Wold 2002). In this study, slow-growing bacteria was the only microbial indicator detected at abundances that were high enough to be included in statistical analyses; all other microbial indicator species had high frequencies of non-detection. O-PLS was therefore used to predict slow-growing bacteria (Y) from twelve water quality variables (X, containing F₁, F₂, F₃, F₄, Al,

320 Cu, Pb, Zn, Mn, Fe, DOC, absorbance at 254 nm (A_{254})). O-
321 PLS regression was implemented using the PLS_Toolbox for
322 MATLAB (ver. 8.1, Eigenvector Inc.). Before applying PLS,
323 all predictor variables were transformed using the Cox-Box
324 power transformation to improve adherences to a normal
325 distribution; thereafter, each variable was autoscaled. Replicate
326 measurements were averaged prior to modelling.

327 An iterative process was used to develop the PLS model.
328 Initially, a model was created using all of the chemical data
329 available with the aim to predict slow-growing bacteria
330 abundances across all sites ($N=87$). Subsequently, this model
331 was refined by removing the variables that had least influence
332 on the model (lowest VIP). Still, this model had low predictive
333 power and was not robust during cross-validation. It was then
334 attempted to develop a model only for sites in the southeast
335 parts of the distribution system ($N=37$) since these had
336 generally higher bacterial counts and fewer non-detects. This
337 also produced no robust patterns. Finally, a tentative model was
338 developed for the southeast distribution system ($N=31$ after
339 excluding five sites with low microbial abundances ($< 25\%$
340 percentile, < 7 cfu/ml) and one site with high leverage on the
341 model, and retaining only four parameters as predictor
342 variables (F_4 , Fe, A_{254} , Pb).

343 *Metal complexation model*

344 There are no established models for estimating metal-DOM
 345 complexation parameters from absorbance data. Two widely-
 346 used models for estimating the binding parameters of metal-
 347 ligand complexation from fluorescence data are the Ryan-
 348 Weber model (Ryan 1982) and modified Stern-Volmer model
 349 (Hays et al. 2004). Both assume 1:1 metal to ligand complex
 350 formation. The Ryan-Weber model assumes a linear
 351 relationship between the formed complex and fluorescence
 352 quenching, which may not reflect the full complexity of the
 353 binding mechanism (Hays et al. 2004). In the modified Stern-
 354 Volmer model, a nonlinear relationship is assumed,
 355 parameterized by a quenching constant (K_M) and an initial
 356 fraction (f) of fluorescence contributing to quenching. This
 357 Stern-Volmer model was used to estimate the binding
 358 parameters between PARAFAC components and copper in this
 359 study (equation 2).

$$360 \quad \frac{F_0}{F_0 - F} = \frac{1}{f \cdot K_M \cdot C_M} + \frac{1}{f} \quad (2)$$

361 In equation (2), F and F_0 are fluorescence intensities
 362 corresponding to the measured total copper concentration C_M in
 363 samples containing copper, or in the absence of copper,
 364 respectively. K_M and f are the conditional stability constant and
 365 the fraction of initial fluorescence affected by metal binding.
 366 The K_M and f values were determined in this study from the
 367 relative fluorescence intensity of each component (equation 2)

368 plotted against the inverse concentration of copper.

369 *Effect of chlorine*

370 Chlorine residuals decrease as a function of reaction time,
371 and while rapid changes occur at short reaction times, at longer
372 exposure times (e.g. a day or more), the rate of change can be
373 assumed to be linear (Korshin et al. 2002). We defined the
374 chlorine reaction time for a given sample as the time delay
375 between chlorination at the plant and fluorescence analysis in
376 the laboratory, which is assumed equal to the sum of its
377 distribution time and the delay between sampling and analysis.
378 To investigate whether chlorine exposure could have been a
379 confounding factor in fluorescence measurements, general
380 linear models were used to model fluorescence as a function of
381 chlorine reaction time, both in the presence and the absence of
382 a potential interaction with copper.

383 3. Results

384 3.1. Microbial and chemical water quality

385 3.1.1. Microbial indicators

386 Abundances of microbial indicator species were low or
387 below detection limits across the entire distribution network.
388 *Escherichia*. Abundances of *E. coli*, coliforms and
389 *Enterococcus spp.* were below detection limits (< 1 per 100 ml)
390 at all sites. Slow-growing bacteria abundances varied between
391 0-110 cfu/ml, and culturable microorganisms between 0-30

cfu/ml. Due to fewer non-detects, slow-growing bacteria was used as the primary indicator of microbial abundance in all statistical analyses. Among paired replicate samples, the RSD of slow-growing bacteria abundance averaged 31% (Table 1).

3.1.2. Trace metals

All trace metals were detected at concentrations well below the health limits recommended by the World Health Organization (WHO 2011) (Table 1). No health limits exist for Fe, Al and Zn due to the very low concentrations of these metals in drinking water relative to levels that produce toxicological effects. Coefficients of variation for each metal are presented in Table 1. Variation among replicate measurements of Al, Pb, Cd and Cr was high (RSD > 50%), and concentrations were near the analytical detection limits. For all other trace metals, RSD was below 17%.

Table 1 to be inserted here.

3.1.3. Turbidity, DOM and DOC

Turbidity was low across the entire distribution network (0 – 0.27 FNU; average 0.14 FNU), and below detection (<0.1 FNU) at almost one third of sites. Spectroscopic measurements

(absorbance and fluorescence) indicated low variability in the concentration and composition of optically-active DOM (Figure 1). Across the network, absorbance varied most at short excitation wavelengths (Figure 1a) and fluorescence at short excitation and emission wavelengths where protein-like fluorescence is observed (Figure 1b). Overall, absorbance was more variable than fluorescence (RSD = 10% for A_{254} , compared to 4% for humic-like peaks).

Figure 1 to be inserted here.

A four-component PARAFAC model explained 99.9% of the total variance in the fluorescence EEM dataset (Figure 2). Based on published interpretations of components with similar spectral properties (Coble 1996), the first three components (F_1 : 314/408 nm F_2 : 359/443 nm and F_3 : 389/508) represent humic-like DOM and component 4 (F_4 : 290/351) represents tryptophan-like DOM. Each fluorescence component was present at intensities exceeding the method detection limits in every sample.

Figure 2 to be inserted here.

Variation in fluorescence intensities could not be explained by differences in chlorine reaction time. In general linear

models of fluorescence intensities regressed against chlorine exposure time and/or copper, chlorine exposure time explained no more than 5% of total variability in any fluorescence component ($R^2 \leq 0.05$), compared to copper, which explained 56 - 63% of total variability for the humic-like components (F1 - F3) but less than 1% of the variability in the protein-like component (Supporting Information, Tables S1-S4).

DOC concentrations varied from 2.5 to 8.8 mg/L across the distribution network, with mean and median concentrations of 4.5 ppm and 3.5 ppm, respectively (Figure 3a). Two distinct DOC distributions could be observed; one with low DOC similar to DOC in the outgoing water from the plant (< 4 mg/L) and a second which was normally distributed with mean of approximately 7 mg/L. No geographical pattern could be detected that explained these two distributions. At the same time, the result could not be explained by contamination or by analytical error as samples were analysed in random order, and replicate samples spanned both distributions and differed by at most 16% (see the Supporting Information). Instead, this result indicates that an additional source of DOC was present either in the distribution network or else in the household pipe network, potentially including plastic piping and rubber seals in tap fittings.

Figure 3 to be inserted here.

3.2. Effect of water residence time on water quality

No correlation was observed between water residence time and any of the individual chemical or microbial parameters measured in the distribution system (Figure 3). Also, no variation in chemical or microbial parameters could be attributed to the time of day when sampling took place. However, qualitative trends were observed for some parameters. When sites were divided in three groups having low ($< 25^{\text{th}}$ percentile, < 7 cfu/ml), medium (25^{th} - 75^{th} percentile, 7-40 cfu/ml) or high ($> 75^{\text{th}}$ percentile, ≥ 40 cfu/ml) slow-growing bacteria abundance, sites with high slow-growing bacteria were often located in the southeast region of the distribution network (Figure 4a). Also, when divided in groups representing low (< 17 h), medium (17-29 h), or high (≥ 29 h) water residence times, sites with long residence times mainly clustered in the same region (Figure 4b). In this southeast region, the average travel time was almost 10 hours longer than at other locations and the average slow-growing bacteria abundance was almost 1.7 times greater than the average for the remaining sites.

Figure 4 to be inserted here.

3.3. Predicting microbial abundance from chemical variables

480 Four chemical variables (protein-like fluorescence F_4 , A_{254} ,
481 Fe and Pb) were most useful for predicting slow-growing
482 bacteria abundances in the southeast network where
483 distribution times were longest. The PLS model of the
484 southeast network explained 33% of the measured variation in
485 slow-growing bacteria abundances and 61% of the measured
486 variation in these four chemical parameters (RMSECV=2.7,
487 RMSEC=2.1, N=31). Only tentative conclusions can be drawn
488 from the model due to its restricted geographical range and
489 relatively low predictive ability ($R^2_{cv} = 35\%$). Along the only
490 axis relevant to predicting microbial abundance, A_{254} and
491 protein-like fluorescence were negatively correlated to slow-
492 growing bacteria. This could occur if these autotrophic bacteria
493 exerted top-down control on the abundance of protein-like
494 fluorophores, or if protein-like fluorescence and bacterial
495 abundance were both influenced by a third parameter but in
496 opposite directions. Bacteria abundances were positively
497 correlated with Fe, which is a potential food source for some
498 types of autotrophic bacteria (Kirchman et al. 2000) but was
499 not a significant ingredient in the HPC growth medium.

500 3.4. Copper and fluorescence/absorbance interaction

501 Copper concentrations were negatively correlated with each
502 of the three humic-like fluorescence components, with Pearson
503 correlation coefficients of 77 - 78% (Figure 5a, Table S5). At

the same time, a positive correlation was observed between absorbance and copper concentrations (Figure 5b). Copper did not correlate with protein-like fluorescence. For each humic-like component, the modified Stern-Volmer model provided a reasonable fit to the fluorescence data, with copper explaining 37 - 49% of the measured variation in fluorescence intensities. This fit is illustrated for component F_2 in Figure 5. A better fit to the dataset was obtained using a linear-fit ($R^2 = 56 - 62\%$) or a power-regression model ($R^2 = 62 - 63\%$). Assuming the Stern-Volmer model, the $\log K_m$ values for the three humic-like components in this study are comparable with values reported in earlier studies (Table 2).

Figure 5 inserted here.

Table 2 inserted here.

The strong correlations between copper and fluorescence enabled the fluorescence data to be corrected for copper quenching by calculating what fluorescence intensities would have been in the absence of copper (i.e. at the intercept $[Cu] = 0$). Fluorescence intensities across the network were significantly less variable after copper correction, as illustrated by reduced coefficients of variation. Thus in the presence of copper, the coefficients of variation were between 3.6 - 4.1% (Table 3). After correcting the fluorescence data using the

528 Stern-Volmer model, RSD decreased to 2.2 - 2.4%, while
529 simple power or linear fitting reduced RSD even further to 1.3 -
530 2.2%.

531 3.5. Trigger threshold for detecting entrained contaminants

532 Trigger thresholds (TT) for observing significant changes in the
533 levels of each chemical and microbial parameter in the
534 distribution network are presented in Table 4. Relative trigger
535 thresholds ($TT_{rel} = \text{threshold}/\text{mean}$) for humic-like fluorescence
536 were low (1.1 - 1.3), reflecting high measurement precision and
537 stable fluorescence intensities across the network, and
538 indicating that a sample with fluorescence intensity only 10%
539 higher than the network average could be identified as being an
540 outlier. Due to the highly variable DOC concentrations in the
541 distribution system, soil water entrainment would have been
542 undetectable on the basis of DOC. Trace metals had higher
543 relative trigger thresholds than fluorescence (1.4 - 6.2) and
544 would need to change by a larger relative amount before they
545 would be distinguishable from natural variation. For slow-
546 growing bacteria with TT around 3.9, a sample would not
547 appear to be an outlier so long as microbial abundances were
548 less than 390% of the network average (i.e. <120 cfu/mL in this
549 study). Trigger thresholds could not be determined for other
550 microbial indicator species, due to too many non-detects.

Table 3 inserted here.

Table 4 inserted here.

551 4. Discussion

552 DOM optical properties are well-established water-quality
553 tracers including for the treatment of drinking water (Murphy et
554 al. 2011, Shutova et al. 2014, Stedmon et al. 2011). However,
555 few DOM data have been reported from point-of-use in
556 distribution networks, and it is unknown how much variability
557 can be expected from spectroscopic measurements in stable
558 systems. The network in this study had no known microbial
559 issues, according to both this study and long-term (bimonthly)
560 monitoring by the municipality. All houses sampled on the
561 network produced samples with non-detectable levels of *E.*
562 *coli*, enterococci and coliforms, together with low abundances
563 of culturable bacteria (3-day and 7-day HPC). Globally,
564 abundances of HPC bacteria vary widely in drinking water
565 distribution systems ($< 0.02\text{-}10^4$ cfu/ml) depending on a range
566 of factors including DOC and source water quality, treatment
567 efficiency, distribution time, disinfection residual, and pipe
568 condition (Allen et al. 2004). Elevated abundances of slow-
569 growing bacteria were observed in this study in the section of
570 the distribution network with longest water residence time;

571 even so, concentrations were always below 110 cfu/ml and
572 never approached levels for concern. Only weak correlations
573 were observed between DOM optical measurements and HPC
574 bacterial abundances, and only at locations where water
575 residence time and microbial abundances were highest,
576 suggesting that most observed variability was due to noise.

577 Due to a general lack of published reports on DOM in
578 distribution networks, few data could be located for comparing
579 to the current dataset. Tryptophan-like fluorescence was
580 previously measured in Australian potable and recycled water
581 networks (Hambly et al. 2010), where it was assessed as a
582 tracer of cross-connections. Intensities in that study were
583 measured in situ and reported in arbitrary units so cannot be
584 directly compared with the current study, however the relative
585 standard deviation of tryptophan-like fluorescence
586 measurements in the Australian study was approximately three
587 times higher than in the current study (RSD = 33% and 11%,
588 respectively). This is not surprising, because in-situ
589 fluorometers are generally much less sensitive than benchtop
590 fluorometers and produce noisier data. Additionally,
591 tryptophan-like fluorescence depends on microbial activity
592 (Moran et al. 2000), which would have been suppressed by the
593 winter temperatures in Sweden in comparison to Australian
594 conditions.

595 For any water quality tracer, the more predictable its
596 concentration within the distribution network, the easier it
597 would be to detect contaminated water entrained through
598 damaged pipes. In the current study, fluorescence was the most
599 sensitive tracer among the suite of parameters measured due to
600 high measurement precision and low variability across the
601 network. The minimum amount of contaminated water
602 detectable in practice depends upon the characteristics of the
603 contaminant and the drinking water: as the difference between
604 the two end-members increases, smaller entrainments can be
605 detected. In the network, the average F_1 fluorescence intensity
606 was 0.4 RU. If mixed with our soil water sample ($F_1 = 1.4$ RU),
607 then the contaminated water would need to represent at least
608 4% of the total sample volume before it could be detected on
609 the basis of fluorescence. After taking copper concentrations
610 into account, a 2% infiltration of soil water would be
611 detectable. Note that if the fluorescence signal of the entrained
612 soil water decreases significantly due to interactions with
613 copper, chlorine or other interferents, this would reduce overall
614 sensitivity for detecting an infiltration event by fluorescence
615 spectroscopy.

616 In comparison to humic-like tracers, tryptophan-like
617 fluorescence exhibited higher measurement variation (RSD =
618 11%) even though its fluorescence was not quenched by
619 copper. This variability could not be attributed to any other

parameters monitored in this study, and probably reflects the higher lability as well as greater risk for contamination of this peak by trace amounts of organic matter. To provide comparable sensitivity to a humic-like tracer, tryptophan-like fluorescence would need to be at least ten times higher in the contaminated end-member than in the drinking water end-member. This would not be unusual if the contaminant were sewage, where tryptophan-like fluorescence intensities frequently exceed drinking water levels by several orders of magnitude (Baker et al. 2015, Sorensen et al. 2015). *E. coli* concentrations and tryptophan fluorescence in environmental samples have been shown to correlate approximately linearly over a seven-log range (Baker et al. 2015). If so, tryptophan-like fluorescence could be a sensitive tracer of entrained sewage due to its low detection threshold coupled with high measurement precision.

Copper reduced the measured intensities of humic-like fluorescence in this study, as has been observed in other aquatic systems (Xu et al. 2013, Yamashita and Jaffe 2008). The main source of copper is likely to have been the corrosion of interior copper plumbing in the buildings (WHO 2011). Copper also represented around 0.7% of the pipe materials in the municipal distribution system. Humic-like fluorescence varied inversely with copper across sampling sites, with copper explaining 63% of the variation in fluorescence measurements under a linear

645 regression model, compared with 43% for the modified Stern-
646 Volmer model. The modest fit of the Stern-Volmer model may
647 be due to relatively low copper concentrations in this study
648 ($\text{Cu/DOC} < 1/50$) compared to the ranges typically studied
649 ($\text{Cu/DOC} < 1/25$) (Reynolds and Ahmad 1995). The initial
650 fraction of fluorescence contributing to quenching was also
651 smaller than previously reported, possibly due to competition
652 with calcium and magnesium ions for copper-binding sites
653 (Ryan 1982). The suppression of DOM fluorescence by copper
654 should thus be expected to vary between distribution systems,
655 between sections of a network, and between nearby buildings
656 on the network.

657 Water suppliers in some cities internationally have already
658 made significant investments in online spectrophotometers for
659 monitoring distribution systems, mainly using absorbance
660 spectroscopy (Anon. 2013). In this study, absorbance was a less
661 sensitive water quality tracer than was humic-like fluorescence;
662 a 20% increase in A_{254} relative to the system average would be
663 needed to trigger an outlier compared to a 10% increase for
664 fluorescence, although absorbance exhibits a smaller natural
665 range. For online instrumentation, however, the optimal choice
666 of online technology depends greatly on instrument cost and
667 reliability. Also, although chlorine reaction time was not a
668 confounding factor in this study due to a low chlorine dose and
669 long exposure times, differential chlorine exposure could

670 introduce artefacts that particularly affect online fluorescence
671 monitoring, especially if chlorine doses are high and
672 distribution times vary from hours to days.

673 For all parameters, since measurement variation increases
674 over spatial and temporal scales, the detection threshold
675 calculated in this study would almost certainly improve if
676 comparing measurements at the tap with measurements at
677 various local hubs located in network pipes, instead of with
678 measurements from all households on the network. This would
679 limit spatial and temporal variation, the effects of different
680 household plumbing, and different degrees of chlorine
681 exposure. Also, by comparing network hubs with one another,
682 problems originating in the main pipe network could be
683 isolated more easily.

684 5. Conclusions

685 • Organic matter fluorescence measurements in a
686 functional and stable drinking water distribution system
687 were well above detection limit and exhibited high
688 measurement precision and low fluctuations across the
689 network. Four independently varying fluorescence
690 components were detected.

691 • Potential contamination in the distribution
692 system that results in visible wavelength fluorescence

693 exceeding the network average by 10% would be easily
694 detectable.

695 • In-situ fluorometers should be capable of
696 sensitively monitoring water quality changes in
697 distribution systems between source and consumers,
698 although issues related to reliability, sensitivity and
699 calibration present technical hurdles worthy of further
700 development and investigation.

701 • Trace metals can interfere with spectroscopic
702 measurements in the distribution system and increase
703 detection thresholds for observing significant changes in
704 organic matter quality. It is therefore important to
705 consider trace metals when investigating DOM
706 fluorescence as a potential tracer of contamination in
707 unfamiliar networks.

708

709 Acknowledgment

710 We thank Heather Reader for her assistance with DOC
711 analyses and Julia Tirén Ström and John Östblom sampling
712 assistance. Funding for this study was provided by: Swedish
713 Civil Contingencies Agency, the Swedish Research Council for
714 Environment, Agricultural Sciences and Spatial Planning
715 (FORMAS grant 2013–1214), Danish Council for Independent
716 Research-Natural Sciences (DFF—1323-00336) and
717 Nordic5Tech collaborative funding (DTU).

References

- 718 Allen, M.J., Edberg, S.C. and Reasoner, D.J. (2004)
 719 Heterotrophic plate count bacteria - what is their significance in
 720 drinking water? *International Journal of Food Microbiology*
 721 92(3), 265-274.
- 722 Andersson, C.A. and Bro, R. (2000) The N-way toolbox for
 723 MATLAB. *Chemometrics and Intelligent Laboratory Systems*
 724 52(1), 1-4.
- 725 Anon. (2013), City of Zurich monitors the quality of its
 726 drinking water with s::can, Vienna, Austria, (Access date):
 727 2017-06-10, [http://www.s-](http://www.s-can.at/medialibrary/references/Reference_Zurich_web.pdf)
 728 [can.at/medialibrary/references/Reference_Zurich_web.pdf](http://www.s-can.at/medialibrary/references/Reference_Zurich_web.pdf).
- 729 Baker, A., Cumberland, S.A., Bradley, C., Buckley, C. and
 730 Bridgeman, J. (2015) To what extent can portable fluorescence

- 731 spectroscopy be used in the real-time assessment of microbial
732 water quality? *Science of the Total Environment* 532, 14-19.
- 733 Baker, A. and Inverarity, R. (2004) Protein-like fluorescence
734 intensity as a possible tool for determining river water quality.
735 *Hydrological Processes* 18(15), 2927-2945.
- 736 Beggs, K.M.H., Summers, R.S. and McKnight, D.M. (2009)
737 Characterizing chlorine oxidation of dissolved organic matter
738 and disinfection by-product formation with fluorescence
739 spectroscopy and parallel factor analysis. *Journal of*
740 *Geophysical Research: Biogeosciences* 114(G4), n/a-n/a.
- 741 Bro, R. (1997) PARAFAC. Tutorial and applications.
742 *Chemometrics and Intelligent Laboratory Systems* 38(2), 149-
743 171.
- 744 Camper, A.K., Brastrup, K., Sandvig, A., Clement, J., Spencer,
745 C. and Capuzzi, A.J. (2003) Effect of distribution system
746 materials on bacterial regrowth. *Journal (American Water*
747 *Works Association)* 95(7), 107-121.
- 748 Chow, C., Fabris, R. and Dixon, M. (2008) Case studies using
749 S::CAN on-line monitoring system, Adelaide.
- 750 Coble, P.G. (1996) Characterization of marine and terrestrial
751 DOM in seawater using excitation emission matrix
752 spectroscopy. *Marine Chemistry* 51(4), 325-346.
- 753 Craun, G.F., Brunkard, J.M., Yoder, J.S., Roberts, V.A.,
754 Carpenter, J., Wade, T., Calderon, R.L., Roberts, J.M., Beach,
755 M.J. and Roy, S.L. (2010) Causes of outbreaks associated with
756 drinking water in the United States from 1971 to 2006. *Clinical*
757 *Microbiology Reviews* 23(3), 507-528.
- 758 EN 1484: (1997) Water Quality: Guidelines for the
759 determination of Total Organic Carbon (TOC) and Dissolved
760 Organic Carbon (DOC), BSI.
- 761 Hambly, A.C., Henderson, R.K., Storey, M.V., Baker, A.,
762 Stuetz, R.M. and Khan, S.J. (2010) Fluorescence monitoring at
763 a recycled water treatment plant and associated dual
764 distribution system - Implications for cross-connection
765 detection. *Water Research* 44(18), 5323-5333.
- 766 Hammes, F., Berney, M., Wang, Y., Vital, M., Koester, O. and
767 Egli, T. (2008) Flow-cytometric total bacterial cell counts as a
768 descriptive microbiological parameter for drinking water
769 treatment processes. *Water Research* 42(1-2), 269-277.
- 770 Hays, M.D., Ryan, D.K. and Pennell, S. (2004) A modified
771 multisite stern-volmer equation for the determination of

- 772 conditional stability constants and ligand concentrations of soil
773 fulvic acid with metal ions. *Analytical Chemistry* 76(3), 848-
774 854.
- 775 ISO 6222: (1999) Water quality - Enumeration of culturable
776 micro-organisms -- Colony count by inoculation in a nutrient
777 agar culture medium.
- 778 Kirchman, D.L., Meon, B., Cottrell, M.T., Hutchins, D.A.,
779 Weeks, D. and Bruland, K.W. (2000) Carbon versus iron
780 limitation of bacterial growth in the California upwelling
781 regime. *Limnology and Oceanography* 45(8), 1681-1688.
- 782 Korshin, G.V., Kumke, M.U., Li, C.-W. and Frimmel, F.H.
783 (1999) Influence of chlorination on chromophores and
784 fluorophores in humic substances. *Environmental Science &*
785 *Technology* 33(8), 1207-1212.
- 786 Korshin, G.V., Wu, W.W., Benjamin, M.M. and Hemingway,
787 O. (2002) Correlations between differential absorbance and the
788 formation of individual DBPs. *Water Research* 36(13), 3273-
789 3282.
- 790 LeChevallier, M.W., Gullick, R.W., Karim, M.R., Friedman,
791 M. and Funk, J.E. (2003) The potential for health risks from
792 intrusion of contaminants into the distribution system from
793 pressure transients. *Journal of Water and Health* 1(1), 3-14.
- 794 Manuel, C.M., Nunes, O.C. and Melo, L.F. (2007) Dynamics of
795 drinking water biofilm in flow/non-flow conditions. *Water*
796 *Research* 41(3), 551-562.
- 797 Moran, M.A., Sheldon, W.M. and Zepp, R.G. (2000) Carbon
798 loss and optical property changes during long-term
799 photochemical and biological degradation of estuarine
800 dissolved organic matter. *Limnology and Oceanography* 45(6),
801 1254-1264.
- 802 Murphy, K.R., Butler, K.D., Spencer, R.G.M., Stedmon, C.A.,
803 Boehme, J.R. and Aiken, G.R. (2010) Measurement of
804 dissolved organic matter fluorescence in aquatic environments:
805 an interlaboratory comparison. *Environmental Science and*
806 *Technology* 44(24), 9405-9412.
- 807 Murphy, K.R., Hambly, A., Singh, S., Henderson, R.K., Baker,
808 A., Stuetz, R. and Khan, S.J. (2011) Organic matter
809 fluorescence in municipal water recycling schemes: toward a
810 unified: PARAFAC model. *Environmental Science and*
811 *Technology* 45(7), 2909-2916.

- 812 Murphy, K.R., Stedmon, C.A., Graeber, D. and Bro, R. (2013)
 813 Fluorescence spectroscopy and multi-way techniques.
 814 PARAFAC. *Analytical Methods* 5(23), 6557-6566.
- 815 Reynolds, D.M. and Ahmad, S.R. (1995) The effect of metal-
 816 ions on the fluorescence of sewage waste-water. *Water*
 817 *Research* 29(9), 2214-2216.
- 818 Ryan, D.K.W., J. H. (1982) Copper(II) complexing capacities
 819 of natural waters by fluorescence quenching. *Environmental*
 820 *Science and Technology* 16, 866-872.
- 821 Senesi, N., Miano, T.M., Provenzano, M.R. and Brunetti, G.
 822 (1991) Characterization, differentiation, and classification of
 823 humic substances by fluorescence spectroscopy. *Soil Science*
 824 152(4), 259-271.
- 825 Shutova, Y., Baker, A., Bridgeman, J. and Henderson, R.K.
 826 (2014) Spectroscopic characterisation of dissolved organic
 827 matter changes in drinking water treatment: From PARAFAC
 828 analysis to online monitoring wavelengths. *Water Research* 54,
 829 159-169.
- 830 Sorensen, J.P.R., Lapworth, D.J., Marchant, B.P., Nkhuwa,
 831 D.C.W., Pedley, S., Stuart, M.E., Bell, R.A., Chirwa, M.,
 832 Kabika, J., Liemisa, M. and Chibesa, M. (2015) In-situ
 833 tryptophan-like fluorescence: A real-time indicator of faecal
 834 contamination in drinking water supplies. *Water Research* 81,
 835 38-46.
- 836 Stedmon, C.A., Sereďyńska-Sobecka, B., Boe-Hansen, R., Le
 837 Tallec, N., Waul, C.K. and Arvin, E. (2011) A potential
 838 approach for monitoring drinking water quality from
 839 groundwater systems using organic matter fluorescence as an
 840 early warning for contamination events. *Water Research*
 841 45(18), 6030-6038.
- 842 Stubbins, A., Lapierre, J.F., Berggren, M., Prairie, Y.T.,
 843 Dittmar, T. and del Giorgio, P.A. (2014) What's in an EEM?
 844 Molecular signatures associated with dissolved organic
 845 fluorescence in boreal Canada. *Environmental Science and*
 846 *Technology* 48(18), 10598-10606.
- 847 Trygg, J. and Wold, S. (2002) Orthogonal projections to latent
 848 structures (O-PLS). *Journal of Chemometrics* 16(3), 119-128.
- 849 Van der Wielen, P. and Van der Kooij, D. (2010) Effect of
 850 water composition, distance and season on the adenosine
 851 triphosphate concentration in unchlorinated drinking water in
 852 the Netherlands. *Water Research* 44(17), 4860-4867.

- 853 Vattentäcksarkivet (2016), (Access date): 2017-06-10,
854 <https://www.sgu.se/grundvatten/vattentaktsarkivet/>.
- 855 Weishaar, J.L., Aiken, G.R., Bergamaschi, B.A., Fram, M.S.,
856 Fujii, R. and Mopper, K. (2003) Evaluation of specific
857 ultraviolet absorbance as an indicator of the chemical
858 composition and reactivity of dissolved organic carbon.
859 *Environmental Science and Technology* 37(20), 4702-4708.
- 860 WHO (2011) Guidelines for drinking-water quality, World
861 Health Organization, Geneva.
- 862 WHO (2014) Water safety in distribution systems, World
863 Health Organization, Geneva.
- 864 Wu, J., Long, S.C., Das, D. and Dorner, S.M. (2011) Are
865 microbial indicators and pathogens correlated? A statistical
866 analysis of 40 years of research. *Journal of Water and Health*
867 9(2), 265-278.
- 868 Xu, H., Yan, Z., Cai, H., Yu, G., Yang, L. and Jiang, H. (2013)
869 Heterogeneity in metal binding by individual fluorescent
870 components in a eutrophic algae-rich lake. *Ecotoxicology and*
871 *Environmental Safety* 98, 266-272.
- 872 Yamashita, Y. and Jaffe, R. (2008) Characterizing the
873 interactions between trace metals and dissolved organic matter
874 using excitation-emission matrix and parallel factor analysis.
875 *Environmental Science and Technology* 42(19), 7374-7379.
- 876 Yan, M., Dryer, D., Korshin, G.V. and Benedetti, M.F. (2013)
877 In situ study of binding of copper by fulvic acid: Comparison
878 of differential absorbance data and model predictions. *Water*
879 *Research* 47(2), 588-596.

Table 1. Water quality parameters in the drinking water distribution network. • = no data, - = no limit.

Parameter	Min ¹	Max ¹	Median ¹	RSD across sites ¹ (%)	RSD between replicates (%)		Outgoing ²	LoD ³	Health limit ⁴
					Median	Max			
Fe (µg/l)	1	20	4	75	7	17	< 20	0.74	-
Al (µg/l)	0.50	8	2	47	14	79	< 10	1.72	-
Cu (µg/l)	10	500	50	87	3	14	< 20	0.07	2000
Pb (µg/l)	<0.01	0.50	0.07	100	34	110	•	0.01	10
Zn (µg/l)	2	100	5	170	1	11	•	0.04	-
Mn (µg/l)	0.05	2	0.40	100	7	10	< 10	0.05	400
Ni (µg/l)	0.25	1.75	1.70	14	2	5	•	0.01	70
Cd (µg/l)	0.02	0.17	0.05	47	15	71	•	0.01	3
Cr (µg/l)	0.08	1	0.24	54	15	75	•	0.04	50
Mg (mg/l)	5.4	6.8	6.0	5	2	3	4	0.0005	-
F 1 (RU)	0.34	0.42	0.40	4	1	1.9	•	3 e-4	-
F 2 (RU)	0.25	0.31	0.29	4	1	1.4	•	<1 e-6	-
F 3 (RU)	0.19	0.23	0.22	4	0.7	1.1	•	4 e-4	-
F 4 (RU)	0.13	0.23	0.14	11	4	14	•	1 e-6	-
DOC (mg/l)	2.5	8.9	3.5	41	8	13	2.5	0.16	-
A ₂₅₄ (cm ⁻¹)	0.04	0.08	0.05	10	2	19	•	0.001	-
Cl (mg/l)	•	•	0.03	•	•	•	0.1		
Slow-growing bacteria (cfu/ml)	< 1	110	17	100	25	94	•	1	-

¹ Data are from samples collected in houses along the network.

² Data reported by the WTP in the finished water leaving the plant.

³ Analytical limits of detection (LoD) were determined from procedural blanks.

⁴ Health limits are from WHO guidelines (WHO 2011).

Table 2. Conditional stability constants ($\log K_M$) and initial fraction, f , calculated using the modified Stern-Volmer model for humic-like fluorescence components in this study, compared to similar components in published studies.

Complexation parameters in this study			(Kirchman et al. 2000, Yamashita and Jaffe 2008)			(Xu et al. 2013)		
PARAFAC component	$\log K_M$	f	PARAFAC component	$\log K_M$	f	PARAFAC component	$\log K_M$	f
F ₁	6.24	0.12	Component 1	4.91	0.54	Humic-like fluorescence	5.10	0.80
F ₂	6.36	0.14	Component 6	5.45	0.30			
F ₃	6.25	0.14	Component 2	4.81	0.61			

Table 3. Variation (RSD x 100%) in fluorescence intensities across the distribution network. Uncorrected data are compared with data corrected for quenching by copper using a modified Stern-Volmer model, a linear model, and a power-fit model.

Componen	Uncorrect	Stern-	Linear	Power
F ₁ :	3.6	2.2	1.9	1.9
F ₂ :	3.8	2.2	2.1	2.0
F ₃ :	4.1	2.4	1.3	2.2
F ₄ :	11	-	-	-

Table 4. Trigger thresholds for detecting outliers in the drinking water distribution network.

Parameter	Network mean	Network std. dev.	TT	TT/mean	Min.	Max.	Median
Fe ($\mu\text{g/l}$)	5.4	4.1	17.7	3.3	1	20	4
Al ($\mu\text{g/l}$)	2.8	1.3	6.7	2.4	0.50	8	2
Cu ($\mu\text{g/l}$)	69.1	61.4	253	3.7	10	500	50
Pb ($\mu\text{g/l}$)	0.08	0.09	0.35	4.4	<0.01	0.50	0.07
Zn ($\mu\text{g/l}$)	8.4	14.6	52.2	6.2	2	100	5
Mn ($\mu\text{g/l}$)	0.38	0.41	1.64	4.3	0.05	2	0.40
Ni ($\mu\text{g/l}$)	1.17	0.17	1.68	1.4	0.25	1.75	1.70
Cd ($\mu\text{g/l}$)	0.06	0.03	0.15	2.5	0.02	0.17	0.05
Cr ($\mu\text{g/l}$)	0.28	0.15	0.73	2.6	0.08	1	0.24
Mg (mg/l)	6.1	0.33	7.09	1.16	5.4	6.8	6.0
F_1 (RU)							
- Uncorrected	0.40	0.014	0.44	1.1	0.34	0.42	0.4
- Corrected for [Cu]	0.44	0.009	0.46	1.06	0.42	0.46	0.44
F_2 (RU)							
- Uncorrected	0.29	0.010	0.32	1.1	0.25	0.31	0.29
- Corrected for [Cu]	0.33	0.007	0.35	1.06	0.31	0.34	0.33
F_3 (RU)							
- Uncorrected	0.22	0.009	0.24	1.1	0.19	0.23	0.22
- Corrected for [Cu]	0.25	0.005	0.27	1.07	0.24	0.26	0.25
F_4 (RU)	0.14	0.02	0.19	1.3	0.13	0.23	0.14
DOC (mg/l)	4.6	1.9	10.34	2.24	2.5	8.9	3.5
A_{254} (cm^{-1})	0.05	0.005	0.06	1.20	0.04	0.08	0.05
Slow - growing bacteria (cfu/ml)	30.9	29.8	120.3	3.9	<1	110	17

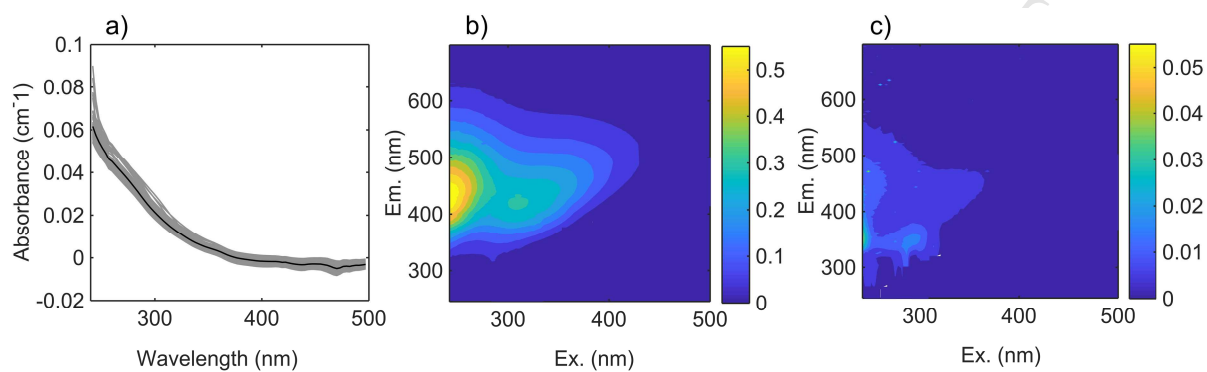


Figure 1. Variation in optical properties across the distribution network. (a) Absorbance spectra (grey lines) compared to the average spectrum (black line); (b) Average fluorescence; (c) standard deviation of fluorescence; observe the change in scale

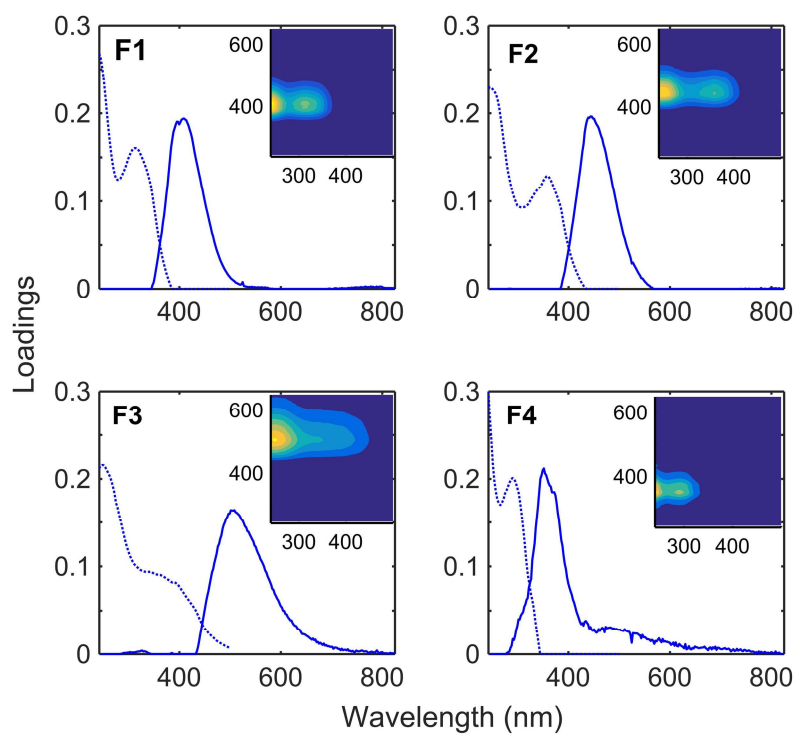


Figure 2. Spectral properties of four independently-varying fluorescent components (F1 - F4) identified in the drinking water network. Inserts show excitation wavelengths on horizontal axis and emission wavelengths on vertical axis.

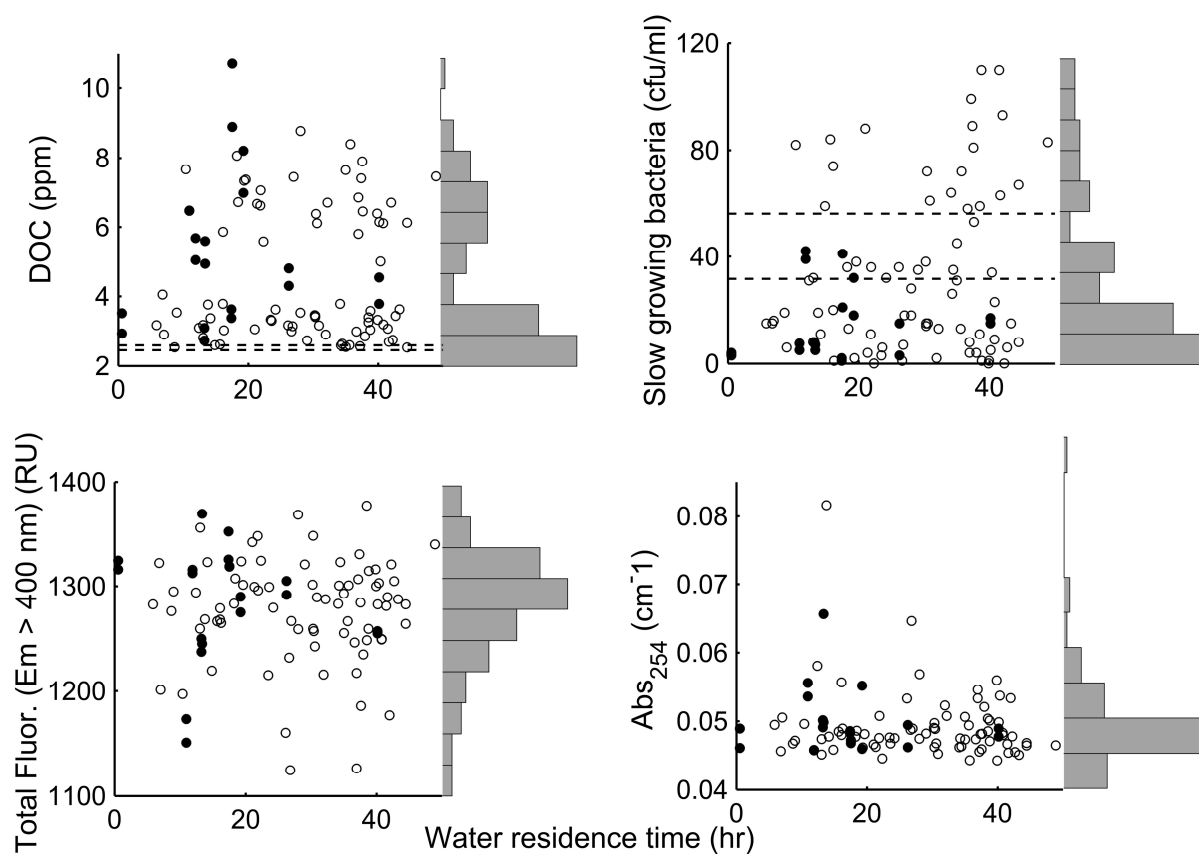


Figure 3. The distribution of DOC, slow-growing bacteria, total fluorescence (Em>400 nm) and A254 versus water residence time. Replicated samples are shown as filled circles. Histograms of the data are shown to the right of each plot

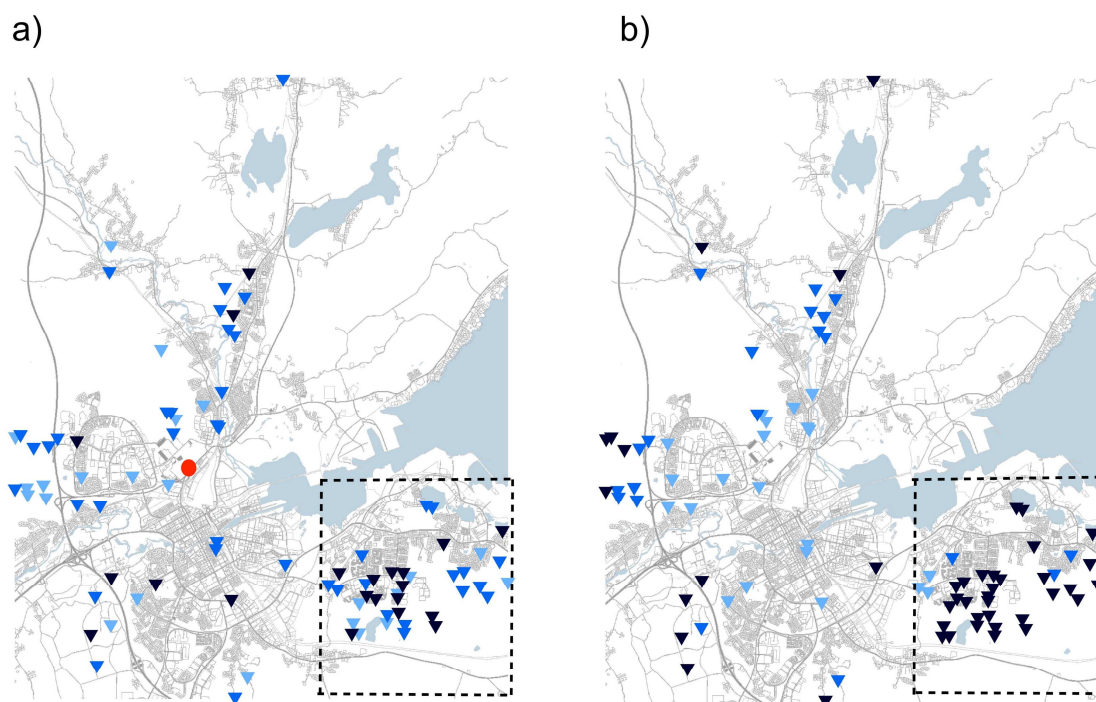


Figure 4. Spatial distributions of (a) slow-growing (7-day) culturable bacteria, and (b) water residence time, at houses in the Gävle distribution network. Sites are classified according to category ranging from high (darkest triangle) to low (lightest triangle). The water treatment plant (WTP) is shown as a red circle. Sites within the southeast network are shown enclosed in a dashed square.

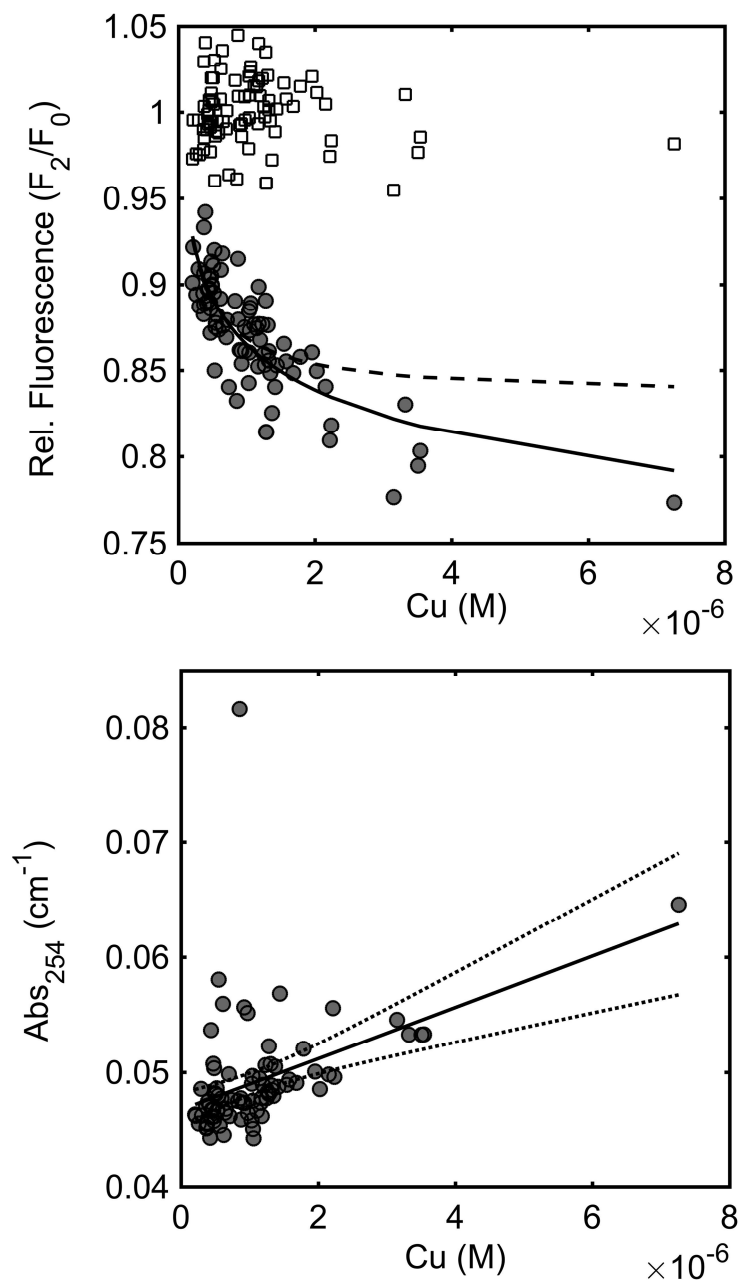


Figure 5. Correlations between DOM optical measurements and copper in the distribution network. (a) Relative fluorescence intensity (fluorescence / 0.33) of component F2 versus copper concentration. The solid line fits a power-regression model ($R^2 = 0.62$), the dashed line a modified Stern-Volmer model ($R^2 = 0.43$). Closed circles show uncorrected data and open squares show corrected data assuming zero copper present under the power model. (b) A_{254} versus copper concentration, $R^2 = 0.2$.

Highlights:

- A biologically-stable drinking water network exhibited stable chemical properties.
- Copper leached from pipes quenched visible wavelength fluorescence.
- Fluorescence would be sensitive tracer of entrained soil water or sewage.