

Accepted Manuscript

Facile syntheses of functionalized toll-like receptor 7 agonists

Babatope Akinbobuyi, Matthew R. Byrd, Charles A. Chang, Myna Nguyen, Zacharie J. Seifert, Anne-Laure Flamar, Gerard Zurawski, Katherine C. Upchurch, SangKon Oh, Stephen H. Dempsey, Thomas J. Enke, John Le, Hunter J. Winstead, José R. Boquín, Robert R. Kane

PII: S0040-4039(14)02040-1
DOI: <http://dx.doi.org/10.1016/j.tetlet.2014.11.126>
Reference: TETL 45510

To appear in: *Tetrahedron Letters*

Received Date: 10 November 2014
Revised Date: 26 November 2014
Accepted Date: 28 November 2014

Please cite this article as: Akinbobuyi, B., Byrd, M.R., Chang, C.A., Nguyen, M., Seifert, Z.J., Flamar, A-L., Zurawski, G., Upchurch, K.C., Oh, S., Dempsey, S.H., Enke, T.J., Le, J., Winstead, H.J., Boquín, J.R., Kane, R.R., Facile syntheses of functionalized toll-like receptor 7 agonists, *Tetrahedron Letters* (2014), doi: <http://dx.doi.org/10.1016/j.tetlet.2014.11.126>

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.



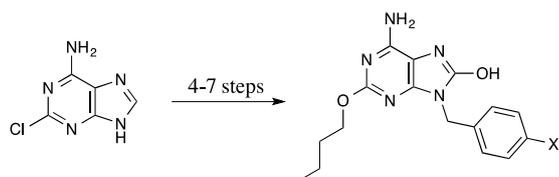
Graphical Abstract

To create your abstract, type over the instructions in the template box below.
 Fonts or abstract dimensions should not be changed or altered.

Facile syntheses of functionalized toll-like receptor 7 agonists

Leave this area blank for abstract info.

Babatope Akinbobuyi, Matthew R. Byrd, Charles A. Chang, Mysel Nguyen, Zacharie J. Seifert, Anne-Laure Flamar, Gerard Zurawski, Katherine C. Upchurch, SangKon Oh, Stephen H. Dempsey, Thomas J. Enke, John Le, Hunter J. Winstead, José R. Boquín, and Robert R. Kane*



9 new TLR-7 agonists; 10-70% overall for 4-7 steps

X = H, I, CO₂H, CN, CONH₂, CCCH₂OH, CCCH₂NH₂, (CH₂)₃NH₂,
 CONHCH₂CH₂NH₂, CONH-PEG₂-NH₂

ACCEPTED MANUSCRIPT



Tetrahedron Letters
journal homepage: www.elsevier.com

Facile syntheses of functionalized toll-like receptor 7 agonists

Babatope Akinbobuyi^a, Matthew R. Byrd^a, Charles A. Chang^b, Maysa Nguyen^a, Zacharie J. Seifert^a, Anne-Laure Flamar^c, Gerard Zurawski^{bc}, Katherine C. Upchurch^b, SangKon Oh^{bc}, Stephen H. Dempsey^d, Thomas J. Enke^d, John Le^d, Hunter J. Winstead^d, José R. Boquín^d, and Robert R. Kane^{abc*}

^a Department of Chemistry and Biochemistry, Baylor University, One Bear Place #97348, Waco, TX 76798, USA

^b Institute of Biomedical Studies, Baylor University, One Bear Place #97224, Waco, TX 76798, USA

^c Baylor Institute for Immunology Research, Baylor Research Institute, 3434 Live Oak Street, Dallas, TX 75204, USA

^d Department of Chemistry, Augustana College, 639 38th Street, Rock Island, IL 61201, USA

ARTICLE INFO

Article history:

Received

Received in revised form

Accepted

Available online

Keywords:

Toll-like receptor

Adenine derivatives

Vaccines

Immune activation

ABSTRACT

Protein conjugates of toll-like receptor 7 agonists have been shown to elicit powerful immune responses. In order to facilitate our studies in this area our group has developed efficient syntheses for a number of functionalized derivatives that retain immune stimulatory activity.

2009 Elsevier Ltd. All rights reserved.

Pathogens are initially detected by the innate immune system upon the binding of their associated ‘signature’ molecules to germline-encoded pathogen-associated molecular pattern (PAMP) receptors.¹ Toll-like receptor 7 (TLR7) is an endosomal member of the Toll-like receptor family of PAMP recognition proteins.² Virus-associated single stranded RNA is the molecular pattern that activates TLR7, thereby initiating an antiviral immune response involving the maturation of dendritic cells (DC), the secretion of cytokines, and the up-regulation of major histocompatibility complex.^{3,4} The development of immunotherapeutic compounds that act via the activation of PAMP receptors, including TLR7, is an active area of research.

Potent small molecule TLR7 agonists have been discovered, including imidazoquinolines such as compound **1**⁵ and substituted adenine derivatives such as compound **2**⁶⁻⁸ (Figure 1). Unfortunately, the clinical utilization of these compounds is significantly limited by side-effects resulting from the overwhelming generation of cytokines subsequent to systemic administration. Accordingly, the clinical application of compound **1** has been limited to localized administration such as formulation for topical use for the treatment of skin diseases.⁹ A wide variety of derivatives of these compounds have been synthesized in hopes of identifying analogues that elicit especially potent or selective immune responses¹⁰⁻¹⁵ or to provide compounds with improved solubility,¹⁶ bioavailability,¹⁷ or pharmacokinetic properties.^{18,19}

Previous work has demonstrated that TLR7 ligands can be conjugated to a variety of molecules (lipids, peptides, and proteins) while retaining their potent agonist activity *in vitro* and *in vivo*.²⁰⁻²³ Specifically, para substituents on compounds such as **2** are well tolerated, as demonstrated by the conjugation of aldehyde **3** to a to proteins and peptides with retention of TLR7 stimulation.²² Our group is interested in synthesizing anti-DC receptor antibody/TLR7 agonist conjugates in order to facilitate the targeted delivery of these immunostimulatory compounds, thereby potentially avoiding the problems associated with systemic administration. This paper reports the efficient synthesis of functionalized TLR7 agonists **4** that can potentially serve as the starting point for the synthesis of antibody conjugates.

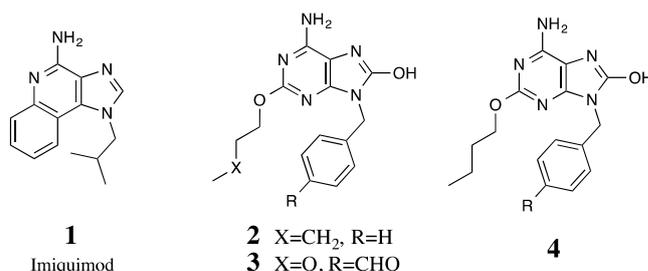
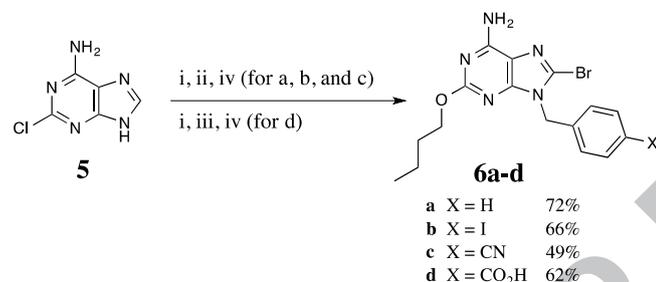


Figure 1. Synthetic TLR-7 agonists

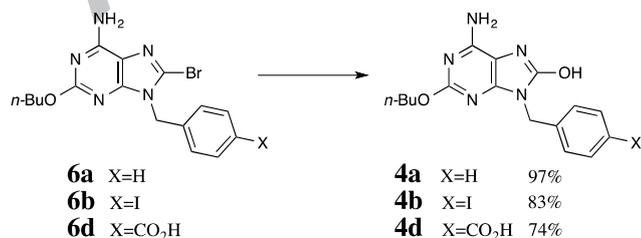
* Corresponding author. Tel.: +1-254-710-4556; fax: +1-254-710-4272; e-mail: Bob_Kane@baylor.edu

The synthesis of the known⁶⁻⁸ non-functionalized TLR agonist **4a** was initially explored. Benzoylation of commercially available chloroadenine **5** (which can also be conveniently synthesized on a large scale by amination of 2,6-dichloropurine¹⁸) proceeded smoothly, affording the desired product in good yield by simple precipitation (Scheme 1). This route avoids the formation of isomers (requiring chromatographic separation) observed upon benzylation of 2,6-dichloropurine. Introduction of the alkoxy substituent (again, with a simple isolation by precipitation) followed by bromination provides **6a** in good yield. In our hands the bromination proceeded very slowly in CH₂Cl₂ or CHCl₃ requiring a very large excess of bromine and multiple Na₂S₂O₃ washes to isolate the pure product. However, in acetic acid with sodium acetate the bromination proceeds very efficiently to afford pure product that can be isolated by filtration from the reaction mixture. Finally, while **6a** could be hydrolyzed using a two-step procedure (methanolysis followed by acidic cleavage of the methyl ether), we found that simple treatment with a solution of NaOH in water/methanol solvent gave direct access to the desired product in one step (Scheme 2). Overall, our optimized procedure was scalable and afforded the desired TLR7 agonist **4a** in four steps with 70% overall yield without chromatographic purification.



Scheme 1. Reagents and conditions: i. ArCH₂Br, K₂CO₃, DMSO, rt, ii. *n*-BuONa/*n*-BuOH, reflux, iii. *n*-BuONa/*n*-BuOH, reflux, then add H₂O, reflux, iv. Br₂, CH₂Cl₂, rt 12 h *or* Br₂, AcOH, AcONa, rt, 1 h.

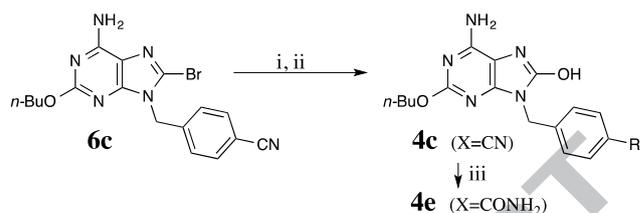
A similar reaction sequence also readily afforded iodide **4b** (55% for 4 steps). However when preparing **4c** we observed that yields were adversely impacted by the concomitant formation of nitrile hydrolysis/alcoholysis side-products during the reactions with sodium butoxide and methanolic sodium hydroxide. Nitrile **4c** was therefore most reproducibly isolated via a two-step methanolysis/hydrolysis protocol (which also provided amide **4e**; Scheme 3). In order to avoid the formation of similar mixtures during the preparation of **4d**, water was added directly to the sodium butoxide reaction mixture after chloride displacement was complete. Refluxing the butoxide/water mixture completed the nitrile hydrolysis to afford, after bromination under standard conditions, carboxylic acid **6d** in good yield. Hydrolysis of **6d** then afforded the useful TLR7 agonist **4d**.



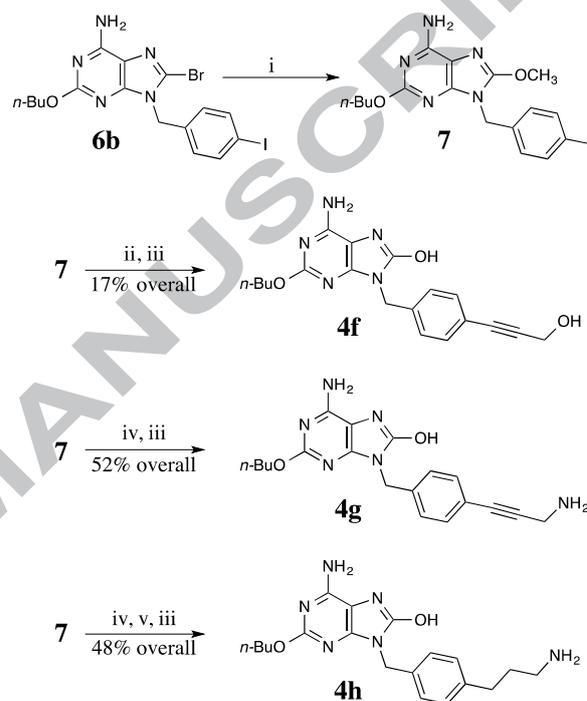
Scheme 2. Reagents and conditions: NaOH, CH₃OH, reflux.

Iodide **7**, which was prepared in good yield by the reaction of **6b** with methoxide, was prepared in order to investigate functionalization using Sonogashira conditions.²⁴ We were pleased to find that **7** reacted cleanly with functionalized alkynes (followed by deprotection of BOC-protected amines) to provide

arenes **4f**, **4g**, and **4h** (Scheme 4). Notably, compounds **4g** and **4h** contain primary amines that can be used to conjugate to proteins or other compounds.

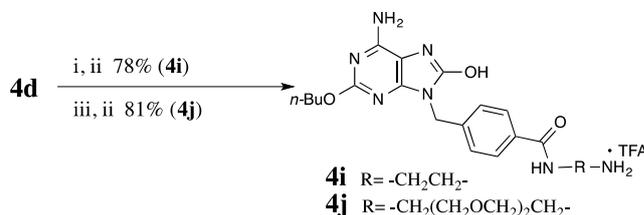


Scheme 3. Reagents and conditions: i. CH₃ONa, CH₃OH, reflux, 61%, ii. HCl (conc), 43%, iii. HCl (conc), 15%.



Scheme 4. Reagents and conditions: i. CH₃ONa, CH₃OH, reflux, 86%, ii. Pd(PPh₃)₂Cl₂, CuI, Et₃N, then propargyl alcohol, reflux, iii. NaI, TMSCl, CH₃CN, reflux, iv. Pd(PPh₃)₂Cl₂, CuI, Et₃N, then BOC-propargyl amine, reflux, v. Pd/C, H₂, MeOH, rt

Finally, carboxylic acid **4d** has been linked, using COMU as a coupling agent, to two different mono-BOC-protected diamines which were cleanly deprotected using TFA to afford two additional amino-substituted TLR7 agonists **4i** and **4j** (Scheme 5).



Scheme 5. Reagents and conditions: i. BOC-ethylenediamine, DIPEA, COMU, DMSO, DCM, 0°-rt, 2 h, ii. TFA, DCM, rt, 1 h, iii. BOC-NH-CH₂CH₂OCH₂CH₂OCH₂CH₂-NH₂, DIPEA, COMU, DMSO, DCM, 0°-rt, 2 h.

Preliminary biological evaluations (TLR7 reporter cell line assays and cytokine release from peripheral blood mononuclear cells) have demonstrated that each of these new compounds (**4b-j**) are active TLR7 agonists (data not shown). Several of these new compounds are well functionalized for protein conjugation, and we are presently attaching them to targeting proteins and characterizing their biological properties.

Acknowledgments

This work was supported by funding from Baylor University (URC), the Baylor Health Care System Foundation, and the NIH (U19 AI057234).

References

1. Medzhitov, R.; Janeway, C.A. Jr. *Cell* **1997**, *91*, 295–298.
2. Takeda, K.; Akira, S. *Int. Immunol.* **2005**, *17*, 1–14.
3. Aderem, A.; Ulevitch, R.J. *Nature* **2000**, *406*, 782–787.
4. Jarrossay, D.; Napolitani, G.; Colonna, M.; Sallusto, F.; Lanzavecchia, A. *Eur. J. Immunol.* **2001**, *31*, 3388.
5. Hemmi, H.; Kaisho, T.; Takeuchi, O.; Sato, S.; Sanjo, H.; Hoshino, K.; Horiuchi, T.; Tomizawa, H.; Takeda, K.; Akira, S. *Nat. Immunol.* **2002**, *3*, 196–200.
6. Naito, T.; Nakagawa, S.; Okita, T.-A.; Yamashita, H.; Yamasaki, T.; Kamei, H.; Tomatsu, K.; Imanishi, H.; Kawaguchi, H. *Chem. Pharm. Bull.* **1982**, *30(6)*, 2011–2019.
7. Hirota, K.; Kazaoka, K.; Niimoto, I.; Kumihara, H.; Sajiki, H.; Isobe, Y.; Takaku, H.; Tobe, M.; Ogita, H.; Ogino, T.; Ichii, S.; Kurimoto, A.; Kawakami, H. *J. Med. Chem.* **2002**, *45*, 5419–5422.
8. Hirota, K.; Kazaoka, K.; Niimoto, I.; Sajiki, H. *Org. Biomol. Chem.* **2003**, *1(8)*, 1354–1365.
9. Chang, Y. C.; Madkan, V.; Cook-Norris, R.; Sra, K.; Tyring, S. *South. Med. J.* **2005**, *98*, 914–920.
10. Shukla, N.M.; Mutz, C.A.; Malladi, S.S.; Warshakoon, H.J.; Balakrishna, R.; David S.A. *J. Med. Chem.* **2012**, *55*, 1106–1116.
11. Shukla, N.M.; Salunke, D.B.; Balakrishna, R.; Mutz, C.A.; Malladi, S.S.; David, S.A. *PLoS ONE* **2012**, *7(8)*, e43612.
12. Weterings, J.J.; Khan, S.; van der Heden, G.J.; Melief, C.J.M.; Overkleef, H.S.; van der Burg, S.H.; Ossendorp, F.; van der Marel, G.A.; Filippov, D.V. *Bioorg. Med. Chem. Lett.* **2009**, *19*, 2249–2251.
13. Christopherson, M.S.; Broom, A.D. *Nucleic Acids Res.* **1991**, *19*, 5719–5724.
14. Hirota, K.; Kazaoka, K.; Sajiki, H. *Bioorg. Med. Chem.* **2003**, *11*, 2715–2722.
15. Jin, G.; Wu, C.C.N.; Tawatao, R.I.; Chan, M.; Carson, D.A.; Cottam, H.B. *Bioorg. Med. Chem. Lett.* **2006**, *16*, 4559–4563.
16. Nakamura, T.; Wada, H.; Kurebayashi, H.; McNally, T.; Bonnert, R.; Isobe, Y. *Bioorg. Med. Chem. Lett.* **2013**, *23*, 669–672.
17. Kurimoto, A.; Ogino, T.; Ichii, S.; Isobe, Y.; Tobe, M.; Ogita, H.; Takaku, H.; Sajiki, H.; Hirota, K.; Kawakami, H. *Bioorg. Med. Chem. Lett.* **2004**, *12*, 1091–1099.
18. Kurimoto, A.; Ogino, T.; Ichii, S.; Isobe, Y.; Tobe, M.; Ogita, H.; Takaku, H.; Sajiki, H.; Hirota, K.; Kawakami, H. *Bioorg. Med. Chem.* **2003**, *11*, 5501–5508.
19. Kurimoto, A.; Hashimoto, K.; Nakamura, T.; Norimura, K.; Ogita, H.; Takaku, H.; Bonnert, R.; McNally, T.; Wada, H.; Isobe, Y. *J. Med. Chem.* **2010**, *53*, 2964–2972.
20. Weterings, J.J.; Khan, S.; van der Heden, G.J.; Drijfhout, J.W.; Melief, C.J.M.; Overkleef, H.S.; van der Burg, S.H.; Ossendorp, F.; van der Marel, G.A.; Filippov, D.V. *Bioorg. Med. Chem. Lett.* **2006**, *16*, 3258–3261.
21. Shukla, N.M.; Lewis, T.C.; Day, T.P.; Mutz, C.A.; Ukani, R.; Hamilton, C.D.; Balakrishna, R.; David, S.A. *Bioorg. Med. Chem. Lett.* **2011**, *21*, 3232–3236.
22. Wu, C.C.N.; Hayashi, T.; Takabayashi, K.; Sabet, M.; Smee, D.F.; Guiney, D.D.; Cottam, H.B.; Carson, D.A. *Proc. Nat. Acad. Sci. USA* **2007**, *104(10)*, 3990–3995.
23. Chan, M.; Hayashi, T.; Kuy, C.S.; Gray, C.S.; Wu, C.C.N.; Corr, M.; Wrasidlo, W.; Cottam, H.B.; Carson, D.A. *Bioconjugate Chem.* **2009**, *20*, 1194–1200.
24. Chinchilla, R.; Carmen Najera, C. *Chem. Rev.* **2007**, *107*, 874–922.

Supplementary Material

Complete experimental and analytical data for all new compounds.