

RESEARCH

Open Access



# Adiposity and lipid metabolism indicators mediate the adverse effect of glucose metabolism indicators on oogenesis and embryogenesis in PCOS women undergoing IVF/ICSI cycles

Huahua Jiang<sup>1,2,3,4†</sup>, Manfei Si<sup>1,2,3,4†</sup>, Tian Tian<sup>1,2,3,4</sup>, Huifeng Shi<sup>2,5,6</sup>, Ning Huang<sup>1,2,3,4</sup>, Hongbin Chi<sup>1,2,3,4</sup>, Rui Yang<sup>1,2,3,4</sup>, Xiaoyu Long<sup>1,2,3,4\*</sup> and Jie Qiao<sup>1,2,3,4,7,8\*</sup> 

## Abstract

**Background** Polycystic ovary syndrome (PCOS) women have high incidences of dyslipidemia, obesity, impaired glucose tolerance (IGT), diabetes, and insulin resistance (IR) and are fragile to female infertility. Obesity and dyslipidemia may be the intermediate biological mechanism for the associations between glucose metabolism dysfunction and abnormal oogenesis and embryogenesis.

**Methods** This retrospective cohort study was performed at a university-affiliated reproductive center. A total of 917 PCOS women aged between 20 and 45 undergoing their first IVF/ICSI embryo transfer cycles from January 2018 to December 2020 were involved. Associations between glucose metabolism indicators, adiposity and lipid metabolism indicators, and IVF/ICSI outcomes were explored using multivariable generalized linear models. Mediation analyses were further performed to examine the potential mediation role of adiposity and lipid metabolism indicators.

**Results** Significant dose-dependent relationships were found between glucose metabolism indicators and IVF/ICSI early reproductive outcomes and between glucose metabolism indicators and adiposity and lipid metabolism indicators (all  $P < 0.05$ ). Also, we found significant dose-dependent relationships between adiposity and lipid metabolism indicators and IVF/ICSI early reproductive outcomes (all  $P < 0.05$ ). The mediation analysis indicated that elevated FPG, 2hPG, FPI, 2hPI, HbA1c, and HOMA2-IR were significantly associated with decreased retrieved oocyte count, MII oocyte count, normally fertilized zygote count, normally cleaved embryo count, high-quality embryo count, or blastocyst formation count after controlling for adiposity and lipid metabolism indicators. Serum TG mediated 6.0–31.0% of the associations; serum TC mediated 6.1–10.8% of the associations; serum HDL-C mediated 9.4–43.6% of the associations; serum LDL-C mediated 4.2–18.2% of the associations; and BMI mediated 26.7–97.7% of the associations.

<sup>†</sup>Huahua Jiang and Manfei Si contribute equally to this work.

\*Correspondence:

Xiaoyu Long  
long\_x\_y@163.com  
Jie Qiao  
jie.qiao@263.net

Full list of author information is available at the end of the article



© The Author(s) 2023. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

**Conclusions** Adiposity and lipid metabolism indicators (i.e., serum TG, serum TC, serum HDL-C, serum LDL-C, and BMI) are significant mediators of the effect of glucose metabolism indicators on IVF/ICSI early reproductive outcomes in PCOS women, indicating the importance of preconception glucose and lipid management and the dynamic equilibrium of glucose and lipid metabolism in PCOS women.

**Keywords** PCOS, Mediating effect, Adiposity and lipid metabolism, Glucose metabolism indicators, IVF/ICSI outcomes

## Background

Polycystic ovary syndrome (PCOS), characterized by oligo-anovulation, hyperandrogenism, and polycystic ovarian morphology, is the most common endocrine disease affecting 4 to 21% of reproductive-aged women [1–3]. PCOS has diverse lifelong clinical manifestations of reproductive, metabolic, and psychological disorders, such as anovulatory infertility, dyslipidemia, obesity, impaired glucose tolerance (IGT), Type 2 diabetes mellitus (T2DM), insulin resistance (IR), cardiovascular disease, low self-esteem, psychosexual dysfunction, depression, and anxiety [1, 2, 4, 5]. PCOS women undergoing adverse assisted reproductive technology (ART) are at high risk of adverse IVF outcomes [6]. The pathogenesis of PCOS is complex, including genetic and epigenetic susceptibility, hypothalamic and ovarian dysfunction, hyperandrogenism, hyperinsulinemia, and adiposity-related mechanisms [2, 7].

As common manifestations of PCOS, maternal lipid and glucose metabolism dysregulations have been suggested to contribute to abnormal embryonic development [8–12]. In vitro fertilization (IVF)-based studies indicated that poor embryogenesis and pregnancy outcomes in diabetic and obese mice result from the maternal effects on the oocytes rather than the uterine environment, suggesting an oocyte origin of adverse reproductive outcomes [11, 13]. IVF/intracytoplasmic sperm injection (ICSI) provides an excellent means to study the effects of maternal factors on oogenesis and embryogenesis. The recent clinical studies have shown that dyslipidemia, obesity, glucose homeostasis dysregulation, IR, and metabolic syndrome (MetS) contribute to ART outcomes in PCOS women. A multicenter randomized controlled trial of 1508 PCOS women observed that MetS affects female fecundity in forms of longer infertility duration, poorer ovarian response, fewer retrieved oocytes, fewer available embryos, and lower chance of cumulative live birth [14]. Several studies have also reported the impact of sole obesity and IR on pregnancy outcomes in PCOS women undergoing IVF/ICSI [15–17]. However, these results remain controversial [18–20], and it is not easy to explore causal paths due to the close interactions among metabolite profiles.

The causal roles of body mass index (BMI) and IR in PCOS etiology have been proven via Mendelian randomization (MR) analyses [21–23]. However, the causal relationship between glucose and lipid metabolism in PCOS women remains uncertain. A small MR analysis in the Netherlands indicated that dyslipidemia (i.e., high plasma triglycerides (TGs) levels) seem to be responsible for hyperglycemia and T2DM risk, and large (L) and extra-large (XL) high-density lipoprotein (HDL) lipid components might protect against increasing fasting glucose level [24]. However, a recent MR study based on the genome-wide association summary statistics of Europeans reported bidirectional causality among serum glycemic and lipid traits [25]. Therefore, we assume the potential mediation role of adiposity and lipid metabolism indicators on the associations between glucose metabolism indicators and IVF/ICSI outcomes in PCOS women.

When compared with MR analysis, mediation analysis is another valuable tool for exploring causality by testing whether a variable is a mediator and what proportion of the role it plays [26]. In this study, we aimed to determine whether adiposity and lipid metabolism indicators mediate the effect of glucose metabolism indicators on early reproductive and pregnancy outcomes in PCOS women undergoing IVF/ICSI cycles.

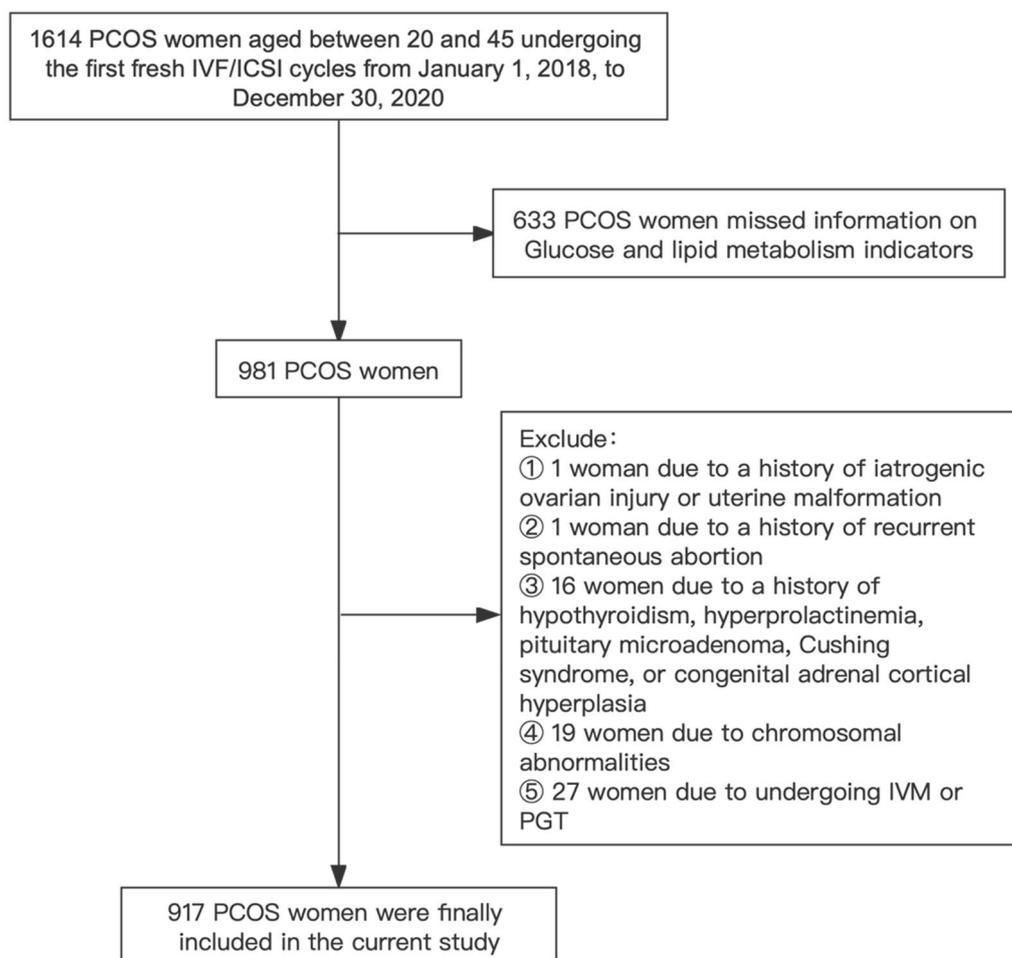
## Materials and methods

### Ethical approval

This study was approved by the Institutional Review Board of Peking University Third Hospital (No. 2021SZ—011). All the patients had previously provided written informed consent.

### Study population

We performed a retrospective cohort study of 917 PCOS patients aged between 20 and 45 undergoing the first fresh IVF/ICSI cycles from January 1, 2018, to December 30, 2020, at the Reproductive Center of Peking University Third Hospital (Fig. 1). Diagnosis of PCOS is based on the 2003 Rotterdam criteria and confirmed with at least two of the following three



**Fig. 1** Flow chart for the study population to investigate associations between sleep characteristics and outcomes of IVF/ICSI

features: hyperandrogenism (clinical or biochemical), menstrual abnormalities (oligo- or amenorrhea), and polycystic ovary morphology ( $\geq 12$  antral follicles (2 to 9 mm in diameter) in unilateral or bilateral ovaries or ovarian volume  $> 10 \text{ cm}^3$  on ultrasonography) [27]. The exclusion criteria were as follows: (1) history of chemotherapy, radiotherapy, ovarian tumors, or ovarian surgery; (2) uterine malformation; (3) history of recurrent spontaneous abortion or autoimmune disease; (4) history of hypothyroidism, hyperprolactinemia, pituitary microadenoma, Cushing syndrome, or congenital adrenal cortical hyperplasia; (5) chromosomal abnormalities; (6) in vitro maturation or preimplantation genetic testing cycles. The information on baseline reproductive and cycle characteristics were collected from the internal electronic medical records. The information on pregnancy outcomes (i.e., biochemical pregnancy, clinical pregnancy, and live birth) was collected by experienced follow-up nurses using telephone interviews.

#### Standardized IVF procedures

We followed the standardized protocols of ovarian stimulation procedures, oocyte retrieval, insemination, embryo transfer, and luteal support as previously described [28, 29]. The initial dose of gonadotropins for ovarian stimulation was individualized based on the BMI and ovarian reserve features. The insemination technique (i.e., conventional insemination, ICSI) was applied based on the sperm quality. The embryos were morphologically graded on Day 3 and Day 5.

#### Definition of study outcomes

Normal fertilization is cognized as zygotes with two pronuclei (2PN). High-quality embryos are defined as 5 to 8-cell embryos with uniform size and less than 30% fragments. Biochemical pregnancy is defined as the positive serum  $\beta$ -hCG level 14 days after ET. Clinical pregnancy is confirmed by ultrasonographic visualization of one or more gestational sacs with fetal heart activity 28 days after ET. Live birth is the delivery of a live-born fetus

beyond 28 weeks of gestation. Deliveries of multiple fetuses were counted as just one live birth.

### Laboratory assessment

Glucose and lipid metabolism indicators were measured within 6 and 3 months before ovarian stimulation, respectively. Fasting plasma glucose (FPG), fasting plasma insulin (FPI), 2-h plasma glucose (2hPG), and 2-h plasma insulin (2hPI) were obtained through a 75 g oral glucose tolerance test (OGTT) and insulin-releasing test after an overnight (at least 8 h) fast. FPG and 2hPG were measured by the hexokinase method (Beckman Access Health Company, Chaska, MN, USA); and FPI and 2hPI were measured by the chemiluminescence method using the Immulite 1000 system (DPC, USA). Glycosylated hemoglobin type A1c (HbA1c) was measured by ion-exchange high-performance liquid chromatography (HLC-723 G8, Tosoh, Japan). The serum total cholesterol (TC), TG, HDL cholesterol (HDL-C), and low-density lipoprotein cholesterol (LDL-C) were determined using a fully automatic biochemical analyzer (7170A, HITACHI, Japan). The intra- and inter-assay variations of all the glucose and lipid metabolism indicators were less than 10%. The technicians in the biochemical laboratory were blinded to other clinical information about the study population.

### Statistical analysis

All the statistics, calculations, and plotting were performed using the R software (version 4.0.3). For patients' baseline reproductive and cycle characteristics, continuous data were presented as median (interquartile range (IQR)), and discrete variables were described as frequencies and percentages ( $n$  (%)). We computed the homeostasis model assessment 2 estimate of insulin resistance (HOMA2-IR) using the HOMA2 Calculator (version 2.2.4) ([www.dtu.ox.ac.uk/homacalculator/index.php](http://www.dtu.ox.ac.uk/homacalculator/index.php), University of Oxford) based on FPG and FPI values [30], and further divided the distribution of HOMA2-IR values into four groups according to quartiles (0.9, 1.3, and 1.9). We analyzed the baseline reproductive and cycle characteristics according to the HOMA2-IR quartile groups by applying Kruskal–Wallis tests for continuous variables and Chi-squared tests (or Fisher's exact test) for categorical variables. Owing to right skewness, FPG, 2hPG, FPI, 2hPI, HbA1c, HOMA2-IR, TC, TG, HDL-C, LDL-C, and BMI were transformed by natural logarithm (ln) when analyses required a normal distribution.

We constructed multivariate generalized linear models (GLMs) to evaluate the associations between glucose metabolism indicators, adiposity and lipid metabolism

indicators, and IVF/ICSI outcomes in PCOS women to assess the feasibility of further mediation analyses. A Poisson distribution and log link function were specified for retrieved oocyte count, MII oocyte count, normally fertilized zygote count, normally cleaved embryo count, high-quality embryo count, and blastocyst formation count, and a binomial distribution and logit link function were specified for biochemical pregnancy, clinical pregnancy, and live birth. According to the statistical consideration and previously reported biological plausibility [31, 32], the following variables were examined as potential covariates and potential confounders: age (continuous), basal follicle-stimulating hormone (FSH) (continuous), antral follicle count (AFC) (continuous), duration of infertility (continuous), infertility type (primary versus secondary infertility), ovarian stimulation regimen (GnRH antagonist protocol, long GnRHa protocol, others), insemination technique (IVF versus ICSI), number of transferred embryos (one vs. two), and timing of embryo transfer (Day 3, Day 5, Day 6). These above-mentioned variables were retained in the final multivariate analyses if they caused a >10% change in the effect estimates based on the "Change-in-Estimate Method" [33].

We further performed mediation analyses to evaluate whether adiposity and lipid metabolism indicators are potential mediators for the associations between glucose metabolism indicators (independent variables) and IVF outcomes (dependent variable) in PCOS women using the mediation package in R. Assuming the mediation analysis hold, we estimated the direct effect which refers to the effect of glucose metabolism indicators on IVF/ICSI outcomes in PCOS women when controlling for adiposity and lipid metabolism indicators. We also estimated the indirect effect that represents the mediating effect of adiposity and lipid metabolism indicators. The proportion of the mediation by each adiposity and lipid metabolism indicator was computed as the ratio of the indirect effect to the total effect. All statistical significance levels were set at two-sided  $P < 0.05$ .

## Results

### Participants' characteristics

Table 1 presents the baseline reproductive and cycle characteristics of the 917 PCOS women. They had a median (IQR) age of 30.0 (28.0, 33.0) years and a median (IQR) BMI of 25.6 (22.6, 28.6) kg/m<sup>2</sup>, were predominantly diagnosed with primary infertility (71.3%), and 83.1% underwent GnRH antagonist protocol. When compared with women in the lowest quartile of HOMA2-IR, women in the highest quartile were more likely to have a longer

**Table 1** Baseline reproductive and cycle characteristics of the PCOS women undergoing the first fresh IVF/ICSI cycles

Characteristics	Overall n=917	HOMA2-IR				P
		← More insulin resistant		Less insulin resistant→		
		0–0.9 n=254	>0.9–1.3 n=213	>1.3–1.9 n=234	>1.9 n=216	
Age, years	30.0 (28.0, 33.0)	31.0 (29.0, 33.0)	31.0 (28.0, 33.0)	30.0 (28.0, 33.0)	30.0 (28.0, 33.0)	0.35
BMI, kg/m <sup>2</sup>	25.6 (22.6, 28.6)	22.3 (20.7, 24.5)	24.9 (22.3, 27.3)	27.2 (24.5, 29.5)	28.3 (25.9, 31.1)	<0.001
AFC, n	17 (13, 22)	17 (13, 22)	17 (12, 22)	17 (12, 22)	17 (13, 22)	0.97
Basal FSH, mIU/mL	5.8 (4.7, 7.1)	6.1 (4.7, 7.4)	5.8 (4.7, 7.0)	5.7 (4.7, 6.9)	5.6 (4.6, 7.0)	0.21
FPG, mmol/L	4.9 (4.6, 5.2)	4.7 (4.4, 4.9)	4.9 (4.6, 5.1)	4.9 (4.7, 5.2)	5.3 (4.8, 5.7)	<0.001
2hPG, mmol/L	6.8 (5.8, 8.2)	5.9 (5.2, 6.8)	6.5 (5.7, 7.7)	7.2 (6.2, 8.5)	8.2 (6.9, 10.2)	<0.001
FPI, uIU/mL	12.0 (8.1, 17.0)	6.5 (5.2, 7.6)	10.5 (9.5, 11.4)	14.5 (13.0, 15.9)	22.6 (19.7, 27.0)	<0.001
2hPI, uIU/mL	77.6 (43.5, 131.1)	40.3 (26.9, 59.3)	65.7 (42.8, 94.8)	96.3 (66.0, 134.6)	170.6 (113.6, 241.0)	<0.001
HbA1c, %	5.5 (5.3, 5.7)	5.4 (5.2, 5.6)	5.4 (5.2, 5.7)	5.5 (5.3, 5.7)	5.6 (5.5, 5.9)	<0.001
Serum TC, mmol/L	4.6 (4.1, 5.1)	4.5 (4.0, 5.1)	4.6 (4.0, 5.1)	4.6 (4.1, 5.2)	4.6 (4.0, 5.1)	0.55
Serum TG, mmol/L	1.4 (1.0, 2.1)	1.0 (0.7, 1.5)	1.4 (0.9, 1.9)	1.6 (1.1, 2.2)	1.8 (1.3, 2.7)	<0.001
Serum HDL-C, mmol/L	1.2 (1.0, 1.4)	1.3 (1.1, 1.6)	1.2 (1.1, 1.4)	1.1 (1.0, 1.3)	1.1 (0.9, 1.2)	<0.001
Serum LDL-C, mmol/L	2.9 (2.5, 3.4)	2.8 (2.4, 3.3)	2.9 (2.4, 3.5)	3.0 (2.6, 3.5)	3.0 (2.6, 3.5)	<0.01
Duration of infertility, years	3 (2, 5)	3 (2, 4)	3 (2, 5)	3 (2, 5)	4 (2, 5)	0.02
Infertility type						0.07
Primary	654 (71.3%)	195 (76.8%)	150 (70.4%)	167 (71.4%)	142 (65.7%)	
Secondary	263 (28.7%)	59 (23.2%)	63 (29.6%)	67 (28.6%)	74 (34.3%)	
Ovarian stimulation regimen						0.51
GnRH antagonist	762 (83.1%)	211 (83.1%)	178 (83.6%)	191 (81.6%)	182 (84.3%)	
Long GnRHa	145 (15.8%)	38 (15.0%)	32 (15.0%)	41 (17.5%)	34 (15.7%)	
Others <sup>a</sup>	10 (1.1%)	5 (2.0%)	3 (1.4%)	2 (0.9%)	0 (0.0%)	
Insemination technique						0.02
IVF	674 (73.5%)	168 (66.1%)	161 (75.6%)	177 (75.6%)	168 (77.8%)	
ICSI	243 (26.5%)	86 (33.9%)	52 (24.4%)	57 (24.4%)	48 (22.2%)	
Number of transferred embryos						0.09
1	110 (12.0%)	22 (8.6%)	26 (12.2%)	39 (16.7%)	23 (10.6%)	
2	807 (88.0%)	232 (91.3%)	187 (87.8%)	195 (83.3%)	193 (89.4%)	
Timing of embryo transfer						0.03
Day 3	871 (95.0%)	245 (96.5%)	205 (96.2%)	212 (90.6%)	209 (96.8%)	
Day 5	27 (2.9%)	7 (2.8%)	4 (1.9%)	11 (4.7%)	5 (2.3%)	
Day 6	19 (2.1%)	2 (0.8%)	4 (1.9%)	11 (4.7%)	2 (0.9%)	

Numbers are median ± IQR (range), except for percentages

PCOS, polycystic ovary syndrome; IVF, in vitro fertilization; ICSI, intracytoplasmic sperm injection; HOMA2-IR, homeostasis models assessment 2 estimate of insulin resistance; BMI, body mass index; AFC, antral follicle count; FSH, follicle-stimulating hormone; FPG, fasting plasma glucose; 2hPG, 2-h plasma glucose; FPI, fasting plasma insulin; 2hPI, 2-h plasma insulin; HbA1c, glycosylated hemoglobin type A1C; TC, total cholesterol; TG, triglyceride; HDL-C, high-density liprotein cholesterol; LDL-C, low-density liprotein cholesterol; GnRH, gonadotropin-releasing hormone; GnRHa, gonadotropin-releasing hormone agonist; IQR, interquartile range

duration of infertility ( $P=0.02$ ); higher FPG, 2hPG, FPI, 2hPI, HbA1c, BMI, TG, and LDL-C (All  $P<0.001$ ); and a lower HDL-C ( $P<0.001$ ). Women in the highest quartile of HOMA2-IR were more likely to undergo the conventional IVF procedure (77.8% vs. 66.1%;  $P=0.02$ ) and Day 3 embryo transfer (96.8% vs. 95%;  $P=0.03$ ) when compared with women in the lowest quartile. No other characteristics differed significantly across quartiles of HOMA2-IR.

### Glucose metabolism indicators and IVF/ICSI outcomes in PCOS women

The effects of glucose metabolism on IVF/ICSI early reproductive outcomes among PCOS women based on the multivariate GLMs are shown in Table 2. After adjustment for age, AFC, and insemination technique, significant monotonic relationships were found across increasing 2hPG, FPI, 2hPI, HbA1c, and HOMA2-IR with decreasing retrieved oocyte count, MII oocyte

**Table 2** Relative risk (RR) and 95% CI in early reproductive outcomes associated with glucose metabolism indicators among PCOS women undergoing their first IVF/ICSI cycles based on GLIMs (n = 917)

Glucose metabolism indicators <sup>a</sup>	Retrieved oocytes, n		MII oocytes, n		Normally fertilized embryos, n		Normally cleaved embryos, n		High-quality embryos, n		Blastocyst formation, n	
	RR (95% CI)	P	RR (95% CI)	P	RR (95% CI)	P	RR (95% CI)	P	RR (95% CI)	P	RR (95% CI)	P
FPG, mmol/L	0.76 (0.64, 0.89)	<0.001	0.78 (0.66, 0.92)	<0.01	0.86 (0.70, 1.05)	0.14	0.85 (0.69, 1.04)	0.13	0.87 (0.69, 1.09)	0.24	0.52 (0.32, 0.82)	<0.01
2hPG, mmol/L	0.90 (0.84, 0.97)	<0.01	0.91 (0.84, 0.97)	<0.01	0.90 (0.82, 0.98)	0.02	0.90 (0.82, 0.98)	0.02	0.88 (0.80, 0.98)	0.02	0.68 (0.56, 0.83)	<0.001
FPI, uIU/mL	0.90 (0.87, 0.94)	<0.001	0.90 (0.87, 0.94)	<0.001	0.89 (0.85, 0.93)	<0.001	0.89 (0.85, 0.93)	<0.001	0.88 (0.83, 0.92)	<0.001	0.77 (0.69, 0.84)	<0.001
2hPI, uIU/mL	0.97 (0.94, 0.99)	<0.01	0.97 (0.94, 1.00)	0.02	0.94 (0.91, 0.97)	<0.001	0.94 (0.91, 0.97)	<0.001	0.93 (0.90, 0.97)	<0.001	0.87 (0.81, 0.94)	<0.001
HbA1c, %	0.62 (0.47, 0.81)	<0.001	0.64 (0.48, 0.85)	<0.01	0.53 (0.38, 0.76)	<0.001	0.52 (0.37, 0.74)	<0.001	0.49 (0.33, 0.74)	<0.001	0.34 (0.15, 0.72)	<0.01
HOMA2-IR	0.91 (0.87, 0.95)	<0.001	0.91 (0.87, 0.96)	<0.001	0.86 (0.81, 0.91)	<0.001	0.86 (0.81, 0.91)	<0.001	0.86 (0.81, 0.92)	<0.001	0.72 (0.64, 0.82)	<0.001

<sup>a</sup>Adjusted for age (continuous), AFC (continuous), and insemination technique

<sup>a</sup>TTransformed by natural logarithm

count, normally fertilized zygote count, normally cleaved embryo count, high-quality embryo count, and blastocyst formation count (All  $P < 0.05$ ). FPG was negatively associated with retrieved oocyte count, MII oocyte count, and blastocyst formation count (All  $P < 0.01$ ). We also observed that increased 2hPI led to a lower chance of live birth (OR: 1.27; 95% CI 1.06, 1.53;  $P = 0.01$ ) after adjustment for age, AFC, and insemination technique (Table 3).

**Glucose metabolism indicators and adiposity and lipid metabolism indicators in PCOS women**

The associations between glucose metabolism indicators and adiposity and lipid metabolism indicators among PCOS women based on the multivariate GLMs are shown in Table 4. After controlling for age, AFC, and duration of infertility, we found that increasing 2hPG, 2hPI, and HbA1c were significantly associated with increasing BMI, serum TC, serum TG, and serum LDL-C (All  $P < 0.01$ ) and decreasing serum HDL-C (All  $P < 0.001$ ). Besides, increasing FPG, FPI, and

HOMA2-IR were significantly associated with increasing BMI, serum TG, and serum LDL-C (All  $P < 0.05$ ) and decreasing serum HDL-C (All  $P < 0.001$ ).

**Adiposity and lipid metabolism indicators and IVF/ICSI outcomes in PCOS women**

We also explored the effect of adiposity and lipid metabolism indicators on PCOS women's IVF/ICSI early reproductive and pregnancy outcomes. As shown in Additional file 1: Table S1, BMI, serum TG, and serum LDL-C were negatively associated with all six IVF/ICSI early reproductive outcomes (All  $P < 0.05$ ). Serum TC was negatively related to high-quality embryo count ( $P < 0.01$ ). Serum HDL-C was positively associated with all six IVF/ICSI early reproductive outcomes (All  $P < 0.01$ ). However, we found no association between adiposity and lipid metabolism indicators and IVF/ICSI pregnancy outcomes in PCOS women (Additional file 1: Table S2).

**Table 3** Odds ratio (OR) and 95% CI in pregnancy outcomes associated with glucose metabolism indicators among PCOS women undergoing their first IVF/ICSI cycles based on GLMs ( $n = 917$ )

Glucose metabolism indicators <sup>a</sup>	Biochemical pregnancy		Clinical pregnancy		Live birth	
	OR (95% CI)	P	OR (95% CI)	P	OR (95% CI)	P
FPG, mmol/L	0.98 (0.35, 2.73)	0.97	0.91 (0.32, 2.57)	0.85	1.04 (0.40, 3.79)	0.75
2hPG, mmol/L	1.35 (0.86, 2.13)	0.19	1.23 (0.78, 1.95)	0.38	1.57 (0.95, 2.61)	0.08
FPI, uIU/mL	0.98 (0.78, 1.24)	0.87	1.00 (0.79, 1.27)	0.98	1.15 (0.89, 1.49)	0.27
2hPI, uIU/mL	1.13 (0.95, 1.33)	0.17	1.15 (0.97, 1.37)	0.10	1.27 (1.06, 1.53)	0.01
HbA1c, %	6.30 (1.06, 40.46)	0.05	4.90 (0.80, 32.29)	0.09	7.79 (1.08, 62.03)	0.05
HOMA2-IR	1.08 (0.81, 1.46)	0.59	1.02 (0.76, 1.38)	0.87	1.26 (0.91, 1.73)	0.16

Adjusted for age (continuous), AFC (continuous), and insemination technique

<sup>a</sup> Transformed by natural logarithm

**Table 4** Regression coefficients and 95% CI in adiposity and lipid metabolism indicators associated with glucose metabolism indicators among PCOS women undergoing their first IVF/ICSI cycles based on GLMs ( $n = 917$ )

Glucose metabolism indicators	BMI, kg/m <sup>2</sup>		Serum TC, mmol/L		Serum TG, mmol/L		Serum HDL-C, mmol/L		Serum LDL-C, mmol/L	
	β (95% CI)	P	β (95% CI)	P	β (95% CI)	P	β (95% CI)	P	β (95% CI)	P
FPG, mmol/L	1.32 (1.22, 1.43)	<0.001	1.02 (0.93, 1.12)	0.66	1.77 (1.33, 2.36)	<0.001	0.78 (0.70, 0.87)	<0.001	1.16 (1.02, 1.33)	0.03
2hPG, mmol/L	1.20 (1.16, 1.25)	<0.001	1.06 (1.02, 1.11)	<0.01	1.90 (1.68, 2.15)	<0.001	0.83 (0.79, 0.87)	<0.001	1.12 (1.06, 1.19)	<0.001
FPI, uIU/mL	1.16 (1.15, 1.18)	<0.001	1.01 (0.99, 1.03)	0.26	1.51 (1.42, 1.60)	<0.001	0.85 (0.83, 0.87)	<0.001	1.06 (1.02, 1.09)	<0.001
2hPI, uIU/mL	1.09 (1.07, 1.10)	<0.001	1.02 (1.01, 1.04)	<0.01	1.34 (1.28, 1.40)	<0.001	0.91 (0.89, 0.92)	<0.001	1.05 (1.02, 1.07)	<0.001
HbA1c, %	1.78 (1.55, 2.04)	<0.001	1.33 (1.13, 1.56)	<0.001	5.17 (3.18, 8.40)	<0.001	0.65 (0.54, 0.79)	<0.001	1.55 (1.23, 1.95)	<0.001
HOMA2-IR	1.20 (1.18, 1.23)	<0.001	1.02 (1.00, 1.05)	0.09	1.63 (1.51, 1.76)	<0.001	0.83 (0.81, 0.86)	<0.001	1.08 (1.04, 1.12)	<0.001

<sup>a</sup> Adjusted for age (continuous), AFC (continuous), and duration of infertility

<sup>b</sup> Glucose metabolism, adiposity, and lipid metabolism indicators were transformed by natural logarithm

### Mediation analysis

After adjusting for various confounders and adiposity and lipid metabolism indicators, we found significant adverse effects of FPG on retrieved oocyte count and MII oocyte count (All  $P < 0.05$ ); significant negative effects of 2hPG, 2hPI, and HbA1c on retrieved oocyte count, MII oocyte count, normally fertilized zygote count, normally cleaved embryo count (All  $P < 0.05$ ); and significant adverse effects of FPI and HOMA2-IR on normally cleaved embryo count (All  $P < 0.05$ ) (Table 5). A significant mediating effect by serum TG was observed for the impact of glucose homeostasis and IR on early reproductive outcomes, with 6.0–31.0% proportion mediated. Moreover, we found statistically significant mediation for the effects of glucose homeostasis and IR on early reproductive outcomes by serum TC (6.1–10.8% proportion mediated), serum HDL-C (9.4–43.6% proportion mediated), serum LDL-C (4.2–18.2% proportion mediated), and BMI (26.7–97.7% proportion mediated) (Additional file 1: Tables S3 to S6).

### Discussion

In this study focused on women with PCOS, we observed significant dose-dependent correlations between glucose metabolism indicators and IVF/ICSI early reproductive

outcomes and between glucose metabolism indicators and adiposity and lipid metabolism indicators. Moreover, we observed significant dose-dependent correlations between adiposity and lipid metabolism indicators and IVF/ICSI early reproductive outcomes. Our estimates suggested that serum TG, serum HDL-C, serum LDL-C, and BMI mediate the inverse associations between glucose metabolism indicators and all six IVF/ICSI early reproductive outcomes in PCOS women. We also observed that serum TC mediates the inverse associations between glucose metabolism indicators and high-quality embryo count in PCOS women.

Glucose homeostasis dysregulation, IR, obesity, and dyslipidemia are critical clinical manifestations of PCOS. The metabolisms of glucose and lipid are often intertwined and regulated [34, 35]. Most evidence suggests altered lipid concentrations are secondary to IR, IGT, or diabetes [36]. High-sugar diet-induced diabetes and genetically diabetes (*db/db*) mice can cause obesity and dyslipidemia by either microbiota imbalance or deficiency of leptin receptor [37, 38]. Obesity and dyslipidemia could also contribute to the pathogenesis of IR or glucose homeostasis dysregulation. Evidence from animal studies has proved that obesity and dyslipidemia can cause hyperinsulinemia and IR in high-fat diet

**Table 5** Mediating effects investigating whether serum TG mediated the associations between glucose metabolism indicators and IVF/ICSI early reproductive outcomes in PCOS women

Mediators	Associations	Total effect (95% CI)	Mediating effect (95% CI)	Estimated proportion of mediation
Serum TG	Retrieved oocyte count and FPG	-4.08 (-7.40, -1.39)***	-0.26 (-0.66, -0.02)*	6.0
	MI I oocyte count and FPG	-3.29 (-6.36, -0.86)**	-0.24 (-0.57, -0.02)*	7.1
Serum TG	Retrieved oocyte count and 2hPG	-1.30 (-2.39, -0.39)**	-0.23 (-0.49, -0.01)*	17.8
	MI I oocyte count and 2hPG	-1.13 (-2.13, -0.25)**	-0.22 (-0.45, -0.01)*	19.2
	Normally fertilized zygote count and 2hPG	-0.03 (-1.61, -0.09)*	-0.24 (-0.45, -0.05)*	29.1
	Normally cleaved embryo count and 2hPG	-0.83 (-1.68, -0.12)*	-0.25 (-0.45, -0.08)**	31.0
	Normally cleaved embryo count and FPI	-0.97 (-1.45, -0.55)***	-0.11 (-0.22, -0.01)*	11.1
Serum TG	Retrieved oocyte count and 2hPI	-0.43 (-0.81, -0.10)*	-0.09 (-0.20, -0.01)*	21.5
	MI I oocyte count and 2hPI	-0.35 (-0.70, -0.03)*	-0.09 (-0.18, -0.01)*	25.6
	Normally fertilized zygote count and 2hPI	-0.54 (-0.89, -0.23)***	-0.09 (-0.18, 0.00)*	16.1
	Normally cleaved embryo count and 2hPI	-0.54 (-0.91, -0.24)***	-0.09 (-0.18, -0.02)*	17.7
Serum TG	Retrieved oocyte count and HbA1c	-9.84 (-20.87, -2.74)***	-0.93 (-2.37, -0.10)*	9.5
	MI I oocyte count and HbA1c	-7.95 (-17.33, -1.92)***	-0.86 (-1.93, -0.08)*	10.9
	Normally fertilized zygote count and HbA1c	-9.46 (-20.11, -2.35)***	-1.09 (-2.47, -0.20)*	11.8
	Normally cleaved embryo count and HbA1c	-10.17 (-22.40, -2.92)***	-1.17 (-2.72, -0.26)**	12.1
Serum TG	Normally cleaved embryo count and HOMA2-IR	-0.96 (-1.33, -0.59)***	-0.11 (-0.21, -0.01)*	10.9

Adjusted for age (continuous), AFC (continuous), and insemination technique

Glucose metabolism indicators and serum TG were transformed by natural logarithm

\* $P < 0.05$

\*\* $P < 0.01$

\*\*\* $P < 0.001$

(HFD)-induced obese and genetically obese (*ob/ob*) mice [39]. Lipids can influence glucose metabolism via several pathways, including disturbing insulin secretion from pancreatic  $\beta$ -cells [40], apoptosis [41], stress responses (e.g., oxidative stress, endoplasmic reticulum stress, and autophagy) [42], and mitochondrial dysfunction [43]. As an instrumental variable method investigating the causal effects of traits using genetic predictors, MR has been used to determine the causal paths among glucose, insulin, and lipid metabolism markers in humans [24, 25, 44, 45]. The causal roles of adiposity and lipid metabolism markers for diabetes have been elucidated [24]. Several studies found that elevated BMI, TG, and small HDL particle levels might increase the risk of diabetes, while elevated large HDL particle levels might have a protective effect [24, 46]. Recently, Zhu et al. systematically investigated the associations among serum HDL-C, LDL-C, TG, fasting insulin, and HbA1c in European populations and illustrated the bidirectional causality of lipids and glycaemic traits [25]. They indicated that elevated serum fasting insulin level was a decisive causal factor of low serum HDL-C and high serum TG levels, which in turn would further increase serum fasting insulin [25]. We found significant positive effects of glucose metabolism indicators on serum TC, serum TG, serum LDL-C, and BMI and negative effects of glucose metabolism indicators on serum HDL-C. These findings also prove the close relationships between glucose metabolism indicators and adiposity and lipid metabolism indicators.

We found negative associations between adiposity and lipid metabolism indicators and IVF/ICSI early reproductive outcomes in PCOS women. Numerous previously published literature can support our findings. Increasing BMI was found to be associated with poorer ovarian response [47, 48], fewer retrieved oocytes [47–49], fewer mature oocytes [49], lower clinical pregnancy rate [16, 17, 50], lower live birth rate [16, 51, 52], and higher risk of miscarriage [48, 51] among PCOS women undergoing IVF/ICSI cycles. High serum TC level was found to be a risk factor for the live birth of IVF/ICSI cycles in PCOS women [52, 53]. The mechanism of obesity and dyslipidemia for PCOS women may be intricate. By promoting the mobilization of free fatty acids, IR is a recognized cause of dyslipidemia in PCOS [54]. Obesity can alter the adipokine distribution in adipocytes, thus, resulting in abnormal activation of pro-inflammatory response, accumulated toxic reactive oxygen species, and aggravated oxidative stress [55]. Inflammation and oxidative stress have been illustrated to cause abnormal oogenesis and folliculogenesis [56]. The damage of obesity to oocytes and embryos may also be related to the alteration of epigenetic reprogramming [56]. Animal studies have

elucidated that maternal obesity can cause a remarkable reduction of an essential maternal effect protein named *Stella* (aka *Dppa3/Pgc7*) in oocytes and may disrupt the establishment of pronuclear epigenetic asymmetry and impair embryo development via accumulation of maternal 5-hydroxymethylcytosine (*5-hmc*) modifications and DNA lesions [11].

We also revealed inverse associations between PCOS women's glucose metabolism indicators and IVF/ICSI early reproductive outcomes after controlling for various confounders and adiposity lipid metabolism indicators. Evidence from previous clinical and animal studies supports our results. A secondary analysis of a multicenter randomized trial illustrated that MetS in PCOS women had a negative impact on IVF cycle stimulation characteristics and clinical outcomes, including higher and longer doses of gonadotropin, lower peak estradiol levels, fewer retrieved oocytes, fewer available embryos, and lower oocyte utilization rate [14]. In another clinical trial, elevated serum insulin, HOMA-IR, and homocysteine levels were negatively associated with oocyte numbers and maturation in PCOS women [19]. The mechanism by which glucose metabolism dysfunction leads to abnormal oogenesis and embryogenesis remains unclear. T2DM was found to increase oocyte mtDNA mutations [57]. Studies also found that IR contributed to apoptosis, oxidative stress, impaired mitochondrial function, abnormal spindles, and misaligned chromosomes of oocytes; thus, leading to fertilization disorders and arrest of embryo development [58]. Moreover, preconception hyperglycemia can reduce the expression of *Tet3* dioxygenase in the oocytes, which is responsible for 5-methylcytosine (*5-mc*) oxidation and DNA demethylation in the zygote. This epigenetic alteration disturbed the subsequent DNA methylation reprogramming of embryos [12]. Our mediation analysis showed that TG, TC, HDL-C, LDL-C, and BMI mediate the associations between glucose metabolism indicators and IVF/ICSI early reproductive outcomes, suggesting that obesity and dyslipidemia may mediate the adverse effect of glucose homeostasis dysregulation and IR on oocyte and embryo quality. As we have discussed above, glucose homeostasis dysregulation and IR have been associated with dyslipidemia and may be the causality of dyslipidemia [25], possibly by increasing mobilization of free fatty acids [54]. Animal and human studies also supported the adverse effect of obesity and dyslipidemia on IVF/ICSI outcomes via inflammation, oxidative stress, and epigenetic mechanisms [49, 55, 57]. However, we must cautiously interpret the mediation analysis's results because solid assumptions are required to interpret our findings' causality.

To our knowledge, this is the first study that conducted a mediation analysis to assess the potential role of adiposity and lipid metabolism indicators in the effects of glucose metabolism indicators on IVF/ICSI outcomes in PCOS women. Our findings provide insights into the mechanism of abnormal oogenesis and embryogenesis caused by glucose metabolism dysfunction and show the importance of the dynamic equilibrium of glucose and lipid metabolism in oogenesis and embryogenesis in PCOS. However, some limitations of this study warrant consideration. First, given the retrospective nature of this study, we cannot entirely avoid the bias in data collection. However, the bias in data collection can be eliminated to some extent because the data we studied are objective measures with specific definitions. Second, the database did not fully record information on adjunctive therapies, including diet, exercise, or medication (i.e., metformin) before or during IVF treatment. Future well-designed prospective studies should focus on the impact of adjunctive therapies. Third, multiple hypothesis tests were simultaneously performed to assess the associations between glucose metabolism indicators and adiposity and lipid metabolism indicators. Thus, we cannot entirely rule out the possibility that some of our results are found by chance. Fourth, we only included PCOS women from a reproductive center, thus possibly causing selection bias and limiting the extrapolation of our results to the general PCOS population. However, these findings may have significance for those who seek infertility treatment, which accounts for 40 to 72% of PCOS women [2, 59]. Fifth, although we found that abnormal glucose metabolism is associated with PCOS women's adverse pregnancy outcomes in this study, we found no mediating role of obesity and dyslipidemia in this relationship. Further mechanism studies are still needed.

## Conclusions

In conclusion, in the present study on 917 PCOS women undergoing their first IVF/ICSI cycles, we found that glucose metabolism indicators were associated with adiposity and lipid metabolism indicators (i.e., increased TG, TC, LDL-C, and BMI and decreased HDL-C), and adiposity and lipid metabolism indicators were inversely correlated with IVF/ICSI early reproductive outcomes (i.e., reduced retrieved oocyte count, MII oocyte count, normally fertilized zygote count, normally cleaved embryo count, high-quality embryo count, or blastocyst formation count). Mediation analyses suggested that obesity and dyslipidemia may mediate a significant proportion of the adverse effect of glucose homeostasis dysregulation and IR on IVF/ICSI early reproductive

outcomes. Our findings show the importance of pre-conception glucose and lipid management in PCOS women.

## Abbreviations

PCOS	Polycystic ovary syndrome
IVF	In vitro fertilization
ICSI	Intracytoplasmic sperm injection
ART	Assisted reproductive technology
BMI	Body mass index
MR	Mendelian randomization
FPG	Fasting plasma glucose
FPI	Fasting plasma insulin
2hPG	2-Hour plasma glucose
2hPI	2-Hour plasma insulin
HOMA2-IR	Homeostasis model assessment 2 estimate of insulin resistance
GLM	Generalized linear model

## Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s40001-023-01174-8>.

**Additional file 1: Table S1.** Relative risk (RR) and 95% CI in early reproductive outcomes associated with adiposity and lipid metabolism indicators among PCOS women undergoing their first IVF/ICSI cycles based on GLMs ( $n = 917$ ). **Table S2.** Odds ratio (OR) and 95% CI in pregnancy outcomes associated with adiposity and lipid metabolism indicators among PCOS women undergoing their first IVF/ICSI cycles based on GLMs ( $n = 917$ ). **Table S3.** Mediating effects investigating whether serum TC mediated the associations between glucose metabolism indicators and IVF/ICSI early reproductive outcomes in PCOS women. **Table S4.** Mediating effects investigating whether serum HDL-C mediated the associations between glucose metabolism indicators and IVF/ICSI early reproductive outcomes in PCOS women. **Table S5.** Mediating effects investigating whether serum LDL-C mediated the associations between glucose metabolism indicators and IVF/ICSI early reproductive outcomes in PCOS women. **Table S6.** Mediating effects investigating whether BMI mediated the associations between glucose metabolism indicators and IVF/ICSI early reproductive outcomes in PCOS women.

## Acknowledgements

We thank Ms. Lixue Chen at the Reproductive Center of Peking University Third Hospital for providing excellent management of the ART database, which provides us with statistical support. We also thank Dr. Yaxin Guo at the Reproductive Center of Tongji Hospital, Huazhong University of Science and Technology for her moral support and suggestions for manuscript submission.

## Author contributions

JQ and XL conceived and designed the study. HJ and MS collected the data. HJ analyzed the data and drafted the manuscript. MS participated in drafting the manuscript. TT and HS provided statistical advice. HC, RY, and NH participated in the revision process. All authors contributed to the article and approved the submitted version.

## Funding

This work was supported by the National Key Research and Development Program of China (No. 2021YFC2700605, 2022YFC2702901), the National Natural Science Foundation of China (No. 81701407, 82171626, 82171632, and 82201839), and the Beijing Science and Technology Planning Project (No. Z191100006619085).

## Availability of data and materials

The data underlying this article will be shared on reasonable request to the corresponding author.

## Declarations

### Ethics approval and consent to participate

This study was approved by the Institutional Review Board of Peking University Third Hospital (No. 2021SZ—011). All the patients had previously provided written informed consent.

### Competing interests

All the authors disclosed no conflict of interest.

### Author details

<sup>1</sup>Center for Reproductive Medicine, Department of Obstetrics and Gynecology, Peking University Third Hospital, Beijing 100191, China. <sup>2</sup>National Clinical Research Center for Obstetrics and Gynecology, Peking University Third Hospital, Beijing, China. <sup>3</sup>Key Laboratory of Assisted Reproduction, Peking University, Ministry of Education, Beijing, China. <sup>4</sup>Beijing Key Laboratory of Reproductive Endocrinology and Assisted Reproductive Technology, Beijing, China. <sup>5</sup>Department of Obstetrics and Gynecology, Peking University Third Hospital, Beijing, China. <sup>6</sup>National Centre for Healthcare Quality Management in Obstetrics, Beijing, China. <sup>7</sup>Beijing Advanced Innovation Center for Genomics, Peking University, Beijing, China. <sup>8</sup>Peking-Tsinghua Center for Life Sciences, Peking University, Beijing, China.

Received: 16 March 2023 Accepted: 14 June 2023

Published online: 03 July 2023

## References

- Zhou K, Zhang J, Xu L, Lim CED. Chinese herbal medicine for subfertile women with polycystic ovarian syndrome. *Cochrane Database Syst Rev*. 2021;6:Cd007535.
- Joham AE, Norman RJ, Stener-Victorin E, Legro RS, Franks S, Moran LJ, et al. Polycystic ovary syndrome. *Lancet Diabetes Endocrinol*. 2022;10:668–80.
- Lizneva D, Suturina L, Walker W, Brakta S, Gavrilova-Jordan L, Azziz R. Criteria, prevalence, and phenotypes of polycystic ovary syndrome. *Fertil Steril*. 2016;106:6–15.
- Zhu T, Cui J, Goodarzi MO. Polycystic ovary syndrome and risk of type 2 diabetes, coronary heart disease, and stroke. *Diabetes*. 2021;70:627–37.
- Damone AL, Joham AE, Loxton D, Earnest A, Teede HJ, Moran LJ. Depression, anxiety and perceived stress in women with and without PCOS: a community-based study. *Psychol Med*. 2019;49:1510–20.
- Sha T, Wang X, Cheng W, Yan Y. A meta-analysis of pregnancy-related outcomes and complications in women with polycystic ovary syndrome undergoing IVF. *Reprod Biomed Online*. 2019;39:281–93.
- Diamanti-Kandarakis E, Dunaif A. Insulin resistance and the polycystic ovary syndrome revisited: an update on mechanisms and implications. *Endocr Rev*. 2012;33:981–1030.
- Diamond MP, Pettway ZY, Logan J, Moley K, Vaughn W, DeCherney AH. Dose-response effects of glucose, insulin, and glucagon on mouse pre-embryo development. *Metabolism*. 1991;40:566–70.
- Bermejo-Alvarez P, Rosenfeld CS, Roberts RM. Effect of maternal obesity on estrous cyclicity, embryo development and blastocyst gene expression in a mouse model. *Hum Reprod*. 2012;27:3513–22.
- Finger BJ, Harvey AJ, Green MP, Gardner DK. Combined parental obesity negatively impacts preimplantation mouse embryo development, kinetics, morphology and metabolism. *Hum Reprod*. 2015;30:2084–96.
- Han L, Ren C, Li L, Li X, Ge J, Wang H, et al. Embryonic defects induced by maternal obesity in mice derive from Stella insufficiency in oocytes. *Nat Genet*. 2018;50:432–42.
- Chen B, Du YR, Zhu H, Sun ML, Wang C, Cheng Y, et al. Maternal inheritance of glucose intolerance via oocyte TET3 insufficiency. *Nature*. 2022;605:761–6.
- Wyman A, Pinto AB, Sheridan R, Moley KH. One-cell zygote transfer from diabetic to nondiabetic mouse results in congenital malformations and growth retardation in offspring. *Endocrinology*. 2008;149:466–9.
- He Y, Lu Y, Zhu Q, Wang Y, Lindheim SR, Qi J, et al. Influence of metabolic syndrome on female fertility and in vitro fertilization outcomes in PCOS women. *Am J Obstet Gynecol*. 2019;221:138.e131–138.e112.
- Chang EM, Han JE, Seok HH, Lee DR, Yoon TK, Lee WS. Insulin resistance does not affect early embryo development but lowers implantation rate in in vitro maturation-in vitro fertilization-embryo transfer cycle. *Clin Endocrinol (Oxf)*. 2013;79:93–9.
- Bailey AP, Hawkins LK, Missmer SA, Correia KF, Yanushpolsky EH. Effect of body mass index on in vitro fertilization outcomes in women with polycystic ovary syndrome. *Am J Obstet Gynecol*. 2014;211(163):e161–166.
- Sheng Y, Lu G, Liu J, Liang X, Ma Y, Zhang X, et al. Effect of body mass index on the outcomes of controlled ovarian hyperstimulation in Chinese women with polycystic ovary syndrome: a multicenter, prospective, observational study. *J Assist Reprod Genet*. 2017;34:61–70.
- Fedorcsák P, Dale PO, Storeng R, Tanbo T, Abyholm T. The impact of obesity and insulin resistance on the outcome of IVF or ICSI in women with polycystic ovarian syndrome. *Hum Reprod*. 2001;16:1086–91.
- Nafiye Y, Sevtap K, Muammer D, Emre O, Senol K, Leyla M. The effect of serum and intrafollicular insulin resistance parameters and homocysteine levels of nonobese, nonhyperandrogenemic polycystic ovary syndrome patients on in vitro fertilization outcome. *Fertil Steril*. 2010;93:1864–9.
- Kakiroglu Y, Doger E, Vural F, Kopuk SY, Vural B. Impact of insulin resistance and obesity on intracytoplasmic sperm injection outcomes in young women with polycystic ovary syndrome. *North Clin Istanbul*. 2017;4:218–24.
- Day FR, Hinds DA, Tung JY, Stolk L, Stykarsdottir U, Saxena R, et al. Causal mechanisms and balancing selection inferred from genetic associations with polycystic ovary syndrome. *Nat Commun*. 2015;6:8464.
- Zhao Y, Xu Y, Wang X, Xu L, Chen J, Gao C, et al. Body Mass Index and Polycystic Ovary Syndrome: a 2-sample bidirectional Mendelian randomization study. *J Clin Endocrinol Metab*. 2020;105:1778–84.
- Wang C, Wu W, Yang H, Ye Z, Zhao Y, Liu J, et al. Mendelian randomization analyses for PCOS: evidence, opportunities, and challenges. *Trends Genet*. 2022;38:468–82.
- Liu J, van Klinken JB, Semiz S, van Dijk KW, Verhoeven A, Hankemeier T, et al. A Mendelian randomization study of metabolite profiles, fasting glucose, and type 2 diabetes. *Diabetes*. 2017;66:2915–26.
- Zhu Z, Wang K, Hao X, Chen L, Liu Z, Wang C. Causal graph among serum lipids and glycemic traits: a Mendelian randomization study. *Diabetes*. 2022;71:1818–26.
- Lee H, Cashin AG, Lamb SE, Hopewell S, Vansteelandt S, VanderWeele TJ, et al. A guideline for reporting mediation analyses of randomized trials and observational studies: the AGReMA statement. *JAMA*. 2021;326:1045–56.
- Revised 2003 consensus on diagnostic criteria and long-term health risks related to polycystic ovary syndrome (PCOS). *Hum Reprod*. 2004;19:41–47.
- Junsheng L, Quan B, Ying L, Xiaoying Z, Ming L, Ping Z, et al. The impact of surgical sperm retrieval on the in vitro fertilization outcomes of infertile patients with temporary ejaculation failure on their oocyte retrieval day. *Fertil Steril*. 2013;99:362–6.
- Wang H, Gao H, Chi H, Zeng L, Xiao W, Wang Y, et al. Effect of levothyroxine on miscarriage among women with normal thyroid function and thyroid autoimmunity undergoing in vitro fertilization and embryo transfer: a randomized clinical trial. *JAMA*. 2017;318:2190–8.
- Wallace TM, Levy JC, Matthews DR. Use and abuse of HOMA modeling. *Diabetes Care*. 2004;27:1487–95.
- Broekmans FJ, Kwee J, Hendriks DJ, Mol BW, Lambalk CB. A systematic review of tests predicting ovarian reserve and IVF outcome. *Hum Reprod Update*. 2006;12:685–718.
- Yan J, Wu K, Tang R, Ding L, Chen ZJ. Effect of maternal age on the outcomes of in vitro fertilization and embryo transfer (IVF-ET). *Sci China Life Sci*. 2012;55:694–8.
- Greenland S. Modeling and variable selection in epidemiologic analysis. *Am J Public Health*. 1989;79:340–9.
- Chen L, Chen XW, Huang X, Song BL, Wang Y, Wang Y. Regulation of glucose and lipid metabolism in health and disease. *Sci China Life Sci*. 2019;62:1420–58.
- Chaurasia B, Summers SA. Ceramides in metabolism: key lipotoxic players. *Annu Rev Physiol*. 2021;83:303–30.

36. Mooradian AD. Dyslipidemia in type 2 diabetes mellitus. *Nat Clin Pract Endocrinol Metab.* 2009;5:150–9.
37. Suriano F, Vieira-Silva S, Falony G, Roumain M, Paquot A, Pelicaen R, et al. Novel insights into the genetically obese (ob/ob) and diabetic (db/db) mice: two sides of the same coin. *Microbiome.* 2021;9:147.
38. Kawano Y, Edwards M, Huang Y, Bilate AM, Araujo LP, Tanoue T, et al. Microbiota imbalance induced by dietary sugar disrupts immune-mediated protection from metabolic syndrome. *Cell.* 2022;185:3501–3519. e3520.
39. Jimenez V, Jambina C, Casana E, Sacristan V, Muñoz S, Darriba S, et al. FGF21 gene therapy as treatment for obesity and insulin resistance. *EMBO Mol Med.* 2018;10:e8791.
40. Hao M, Head WS, Gunawardana SC, Hasty AH, Piston DW. Direct effect of cholesterol on insulin secretion: a novel mechanism for pancreatic beta-cell dysfunction. *Diabetes.* 2007;56:2328–38.
41. Rütti S, Ehses JA, Sibilier RA, Prazak R, Rohrer L, Georgopoulos S, et al. Low- and high-density lipoproteins modulate function, apoptosis, and proliferation of primary human and murine pancreatic beta-cells. *Endocrinology.* 2009;150:4521–30.
42. Janikiewicz J, Hanzelka K, Kozinski K, Kolczynska K, Dobrzyn A. Islet  $\beta$ -cell failure in type 2 diabetes—within the network of toxic lipids. *Biochem Biophys Res Commun.* 2015;460:491–6.
43. Lowell BB, Shulman GI. Mitochondrial dysfunction and type 2 diabetes. *Science.* 2005;307:384–7.
44. White J, Swerdlow DI, Preiss D, Fairhurst-Hunter Z, Keating BJ, Asselbergs FW, et al. Association of lipid fractions with risks for coronary artery disease and diabetes. *JAMA Cardiol.* 2016;1:692–9.
45. Fall T, Xie W, Poon W, Yaghootkar H, Mägi R, Knowles JW, et al. Using genetic variants to assess the relationship between circulating lipids and type 2 diabetes. *Diabetes.* 2015;64:2676–84.
46. Yuan S, Larsson SC. An atlas on risk factors for type 2 diabetes: a wide-angled Mendelian randomisation study. *Diabetologia.* 2020;63:2359–71.
47. McCormick B, Thomas M, Maxwell R, Williams D, Aubuchon M. Effects of polycystic ovarian syndrome on in vitro fertilization-embryo transfer outcomes are influenced by body mass index. *Fertil Steril.* 2008;90:2304–9.
48. Yang W, Yang R, Lin M, Yang Y, Song X, Zhang J, et al. Body mass index and basal androstenedione are independent risk factors for miscarriage in polycystic ovary syndrome. *Reprod Biol Endocrinol.* 2018;16:119.
49. Beydoun HA, Stadtmauer L, Beydoun MA, Russell H, Zhao Y, Oehninger S. Polycystic ovary syndrome, body mass index and outcomes of assisted reproductive technologies. *Reprod Biomed Online.* 2009;18:856–63.
50. Jungheim ES, Lanzendorf SE, Odem RR, Moley KH, Chang AS, Ratts VS. Morbid obesity is associated with lower clinical pregnancy rates after in vitro fertilization in women with polycystic ovary syndrome. *Fertil Steril.* 2009;92:256–61.
51. Qiu M, Tao Y, Kuang Y, Wang Y. Effect of body mass index on pregnancy outcomes with the freeze-all strategy in women with polycystic ovarian syndrome. *Fertil Steril.* 2019;112:1172–9.
52. Gao L, Li M, Wang Y, Zeng Z, Xie Y, Liu G, et al. Overweight and high serum total cholesterol were risk factors for the outcome of IVF/ICSI cycles in PCOS patients and a PCOS-specific predictive model of live birth rate was established. *J Endocrinol Invest.* 2020;43:1221–8.
53. Jiang X, Lu X, Cai M, Liu Y, Guo Y. Impact of dyslipidemia on the cumulative pregnancy outcomes after first ovarian stimulation. *Front Endocrinol (Lausanne).* 2022;13:915424.
54. Ezeh U, Arzumanyan Z, Lizneva D, Mathur R, Chen YH, Boston RC, et al. Alterations in plasma non-esterified fatty acid (NEFA) kinetics and relationship with insulin resistance in polycystic ovary syndrome. *Hum Reprod.* 2019;34:335–44.
55. Dlodla PV, Nkambule BB, Jack B, Mkandla Z, Mutize T, Silvestri S, et al. Inflammation and oxidative stress in an obese state and the protective effects of gallic acid. *Nutrients.* 2018;11:23.
56. Gonnella F, Konstantinidou F, Di Berardino C, Capacchietti G, Peserico A, Russo V, et al. A systematic review of the effects of high-fat diet exposure on oocyte and follicular quality: a molecular point of view. *Int J Mol Sci.* 2022;23:8890.
57. Li L, Wu CS, Hou GM, Dong MZ, Wang ZB, Hou Y, et al. Type 2 diabetes increases oocyte mtDNA mutations which are eliminated in the offspring by bottleneck effect. *Reprod Biol Endocrinol.* 2018;16:110.
58. Ou XH, Li S, Wang ZB, Li M, Quan S, Xing F, et al. Maternal insulin resistance causes oxidative stress and mitochondrial dysfunction in mouse oocytes. *Hum Reprod.* 2012;27:2130–45.
59. Teede H, Deeks A, Moran L. Polycystic ovary syndrome: a complex condition with psychological, reproductive and metabolic manifestations that impacts on health across the lifespan. *BMC Med.* 2010;8:41.

## Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more [biomedcentral.com/submissions](https://biomedcentral.com/submissions)

