



# Germline mutation analyses of malignant ground glass opacity nodules in non-smoking lung adenocarcinoma patients

Wenjun Mao<sup>1,\*</sup>, Ruo Chen<sup>1,\*</sup>, Rongguo Lu<sup>1</sup>, Shengfei Wang<sup>1</sup>, Huizhu Song<sup>2</sup>, Dan You<sup>2</sup>, Feng Liu<sup>1</sup>, Yijun He<sup>1</sup> and Mingfeng Zheng<sup>1</sup>

<sup>1</sup>Department of Cardiothoracic Surgery, The Affiliated Wuxi People's Hospital of Nanjing Medical University, Wuxi, Jiangsu, China

<sup>2</sup>Department of Pharmacy, The Affiliated Wuxi People's Hospital of Nanjing Medical University, Wuxi, Jiangsu, China

\*These authors contributed equally to this work.

## ABSTRACT

**Background.** Germline mutations play an important role in the pathogenesis of lung cancer. Nonetheless, research on malignant ground glass opacity (GGO) nodules is limited.

**Methods.** A total of 13 participants with malignant GGO nodules were recruited in this study. Peripheral blood was used for exome sequencing, and germline mutations were analyzed using InterVar. The whole exome sequencing dataset was analyzed using a filtering strategy. KOBAS 3.0 was used to analyze KEGG pathway to further identify possible deleterious mutations.

**Results.** There were seven potentially deleterious germline mutations. NM\_001184790:exon8: c.C1070T in *PARD3*, NM\_001170721:exon4:c.C392T in *BCAR1* and NM\_001127221:exon46: c.G6587A in *CACNA1A* were present in three cases each; rs756875895 frameshift in *MAX*, NM\_005732: exon13:c.2165\_2166insT in *RAD50* and NM\_001142316:exon2:c.G203C in *LMO2*, were present in two cases each; one variant was present in *NOTCH3*.

**Conclusions.** Our results expand the germline mutation spectrum in malignant GGO nodules. Importantly, these findings will potentially help screen the high-risk population, guide their health management, and contribute to their clinical treatment and determination of prognosis.

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Corresponding author

Mingfeng Zheng,  
zhengmfmedical@126.com

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Additional Information and  
Declarations can be found on  
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## INTRODUCTION

Though therapeutic advances have been made using targeted therapy and immunotherapy, lung cancer continues to be the most common cause of cancer-related deaths worldwide (Siegel, Miller & Jemal, 2015). The majority of lung cancers are caused by somatic mutations that accumulate with age and germline mutations could explain a predisposition to cancer development.

Lung cancer is a complex disease that is mainly attributed to smoking (Hung et al., 2008). However, over 10% of lung cancer patients are non-smokers (Subramanian & Govindan,

2007). The development of lung cancer in never-smokers is associated with several potential risk factors, including environmental pollution and genetic predisposition (*Malhotra et al., 2016*). Germline mutations in lung cancer have been studied to some extent (*Ikeda et al., 2014; Liu et al., 2016; Shukuya et al., 2018; Zhang et al., 2017*), including some in familial settings (*Kanwal et al., 2018; Tomoshige et al., 2015*). There are also studies on non-smoking lung cancer cohorts (*Donner et al., 2018; Renieri et al., 2014*). Nonetheless, studies on germline mutations in lung cancer patients fall far short when compared to those on somatic mutations. There is a need to study germline mutations in lung cancer since they are related to the pharmacodynamics, prognosis, and interactions with somatic mutations (*Bartsch et al., 2007; Erdem et al., 2012; Wang et al., 2018; Winther-Larsen et al., 2015*).

GGOs observed on computed tomography are described as hazy areas but preserved broncho-vascular markings (*Austin et al., 1996; Lee et al., 2014*). Advances in high resolution computed tomography and its application in lung cancer screening have led to an increased detection rate of GGOs, with an estimated prevalence of 0.2–0.5% (*Henschke et al., 2006*). While many GGOs are benign and disappear with time, some are persistent and turn malignant. These tumours are frequently found in non-smokers and women lung cancer patients (*Blons et al., 2006; Raz et al., 2006*).

Here, we recruited a total of 13 non-smoking patients with malignant GGO nodules to study their germline mutations using whole exome sequencing (WES). The results provide a better understanding of molecular mechanisms underlying the development of GGOs and their predisposition to turn cancerous.

## MATERIALS & METHODS

### Study subjects

Candidates that were radiologically found to have small GGO nodules in physical checkup or who came to outpatients department for the reason of cough and checked by computed tomography to have small GGO nodules were closely followed up from 6 months to 3 years. When the GGO nodules increased in size or the nodule density increased or the solid components of pulmonary nodules increased, 13 patients were recruited and underwent surgery and thereafter they were histologically confirmed to have malignant GGOs in the Department of Cardiothoracic Surgery at Wuxi People's Hospital affiliated to Nanjing Medical University, China, from April 1st, 2019 to August 30th, 2019. No other treatments were adopted. Written informed consent was obtained from all participants. Blood samples were collected before surgery and their clinical information was recorded. The research project was approved by the institutional review board of Wuxi People's Hospital affiliated to Nanjing Medical University (no: HS2019014).

### DNA extraction, library preparation, capture enrichment, and WES

Genomic DNA was extracted from peripheral blood collected from participants using a DNA blood mini kit (Qiagen, Germantown, MD, USA) following the manufacturer's instructions. DNA concentration and purity were assessed by a Qubit fluorometer (Invitrogen, Carlsbad, CA, USA).

**Table 1** Characteristics of study subjects. Characteristics of study subjects was collected and analyzed.

Characteristics		
Age at Diagnosis	Mean (SD)	61.5 (8.7)
	Range	48–79
Gender	Male (%)	2 (15.4)
	Female (%)	11 (84.6)
Smoking history	Non-smokers (%)	13 (100.0)
	Smokers (%)	0 (0)

WES was conducted on 500 ng of genomic DNA from each participant. Fragment libraries were prepared from sheared samples by sonication, and exons were enriched by hybridisation capture with a SureSelect Human All Exon V6 Kit (Agilent, Santa Clara, CA, USA) according to the manufacturer's protocol. Captured DNA was amplified followed by solid-phase bridge amplification. The paired-end library was sequenced on a NovaSeq 6000 platform (Illumina, San Diego, CA, USA). The data from this study were deposited in NCBI Sequence Read Archive under SRA accession: [PRJNA613408](https://www.ncbi.nlm.nih.gov/sra/PRJNA613408).

### Read alignment, variant calling, variant annotation, and filtering

Trimmomatic-0.36 ([Bolger, Lohse & Usadel, 2014](#)) was used as quality control for raw data and to remove adapters. Clean sequence reads were aligned to the human reference genome (GRCh37/b37 assembly) using Burrows-Wheeler Aligner software (version 0.7.10) ([Li & Durbin, 2009](#)). Picard (version 2.9.2, Broad Institute, Boston, MA, USA) was used to remove duplicates. Variant detection was performed using HaplotypeCaller in the Genome Analysis Toolkit 3.4 (<https://gatk.broadinstitute.org/hc/en-us>) ([DePristo et al., 2011](#)). Variants were annotated using InterVar database. Detailed stepwise filtering strategy for screening potential candidate germline mutations was described in [Supplement 1](#).

### KEGG pathway analysis

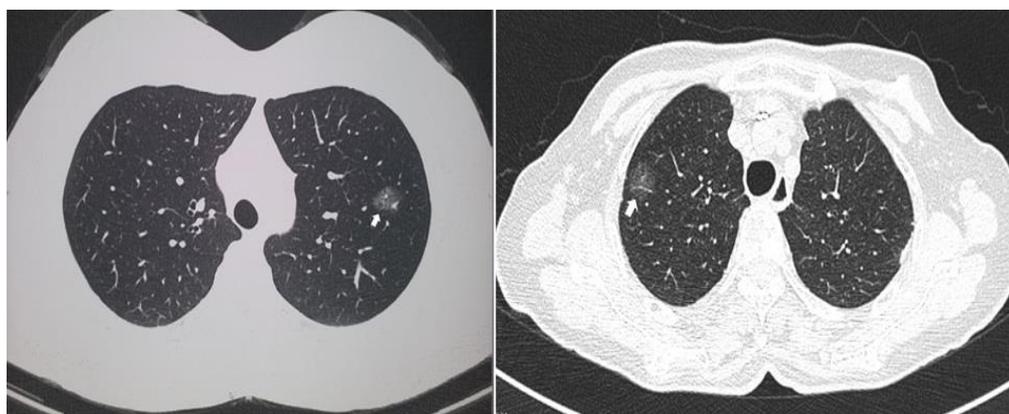
KEGG pathway analysis was conducted *via* KOBAS 3.0 (<http://kobas.cbi.pku.edu.cn/>).

## RESULTS

Clinical information of patients was summarised in [Table 1](#). The mean age at onset of non-small cell lung cancer (NSCLC) in the cases was 61.5 years (range 48–79 years). All cases were non-smokers and 84.6% were females.

Two computed tomographic images are shown as representative of GGO nodules in the study cohort ([Fig. 1](#)). Of the 13 cases, 12 were diagnosed as lung adenocarcinomas while one was diagnosed as an atypical adenomatous hyperplasia. Five cases were adenocarcinoma *in situ*, four were invasive, and three were minimally invasive. Eight of the GGO nodules were located at the right upper lobe, two were at the right lower lobe, and three were at the left upper lobe. Detailed histologic information is presented in [Table 2](#).

We used a stepwise filtering strategy to screen for potential candidate variants ([Fig. 2](#)). Of 83,302 single-nucleotide variants (SNVs) located in exons of the whole exome, our filtering strategy identified 17 potential candidate variants ([Table 3](#)). Of the 17 candidate variants



**Figure 1** Representative of ground glass opacity nodules. Two representative computed tomography images of ground glass opacity nodules. The arrows indicate the nodules.

Full-size [DOI: 10.7717/peerj.12048/fig-1](https://doi.org/10.7717/peerj.12048/fig-1)

**Table 2** Clinical information of study subjects. Pathology, tumor size and tumor location of study objects were shown in details.

Patient ID	Pathology	Tumour size (cm)	Tumour location
WL-1	LUAD <sup>a</sup> , invasive	1.2	right upper lobe
WL-2	LUAD <sup>a</sup> , AIS <sup>b</sup>	0.6	left upper lobe
WL-3	Atypical adenomatous hyperplasia	0.5	right upper lobe
WL-4	LUAD <sup>a</sup> , invasive	1.5	left upper lobe
WL-5	LUAD <sup>a</sup> , minimally invasive	1.0	right upper lobe
WL-6	LUAD <sup>a</sup> , minimally invasive	0.6	left upper lobe
WL-7	LUAD <sup>a</sup> , invasive	2.0	right upper lobe
WL-8	LUAD <sup>a</sup> , invasive	2.0	right lower lobe
WL-9	LUAD <sup>a</sup> , AIS <sup>b</sup>	0.8	right upper lobe
WL-10	LUAD <sup>a</sup> , AIS <sup>b</sup>	0.7	right upper lobe
WL-11	LUAD <sup>a</sup> , AIS <sup>b</sup>	0.6	right lower lobe
WL-12	LUAD <sup>a</sup> , AIS <sup>b</sup>	0.8	right upper lobe
WL-13	LUAD <sup>a</sup> , minimally invasive	0.7	right lower lobe

**Notes.**

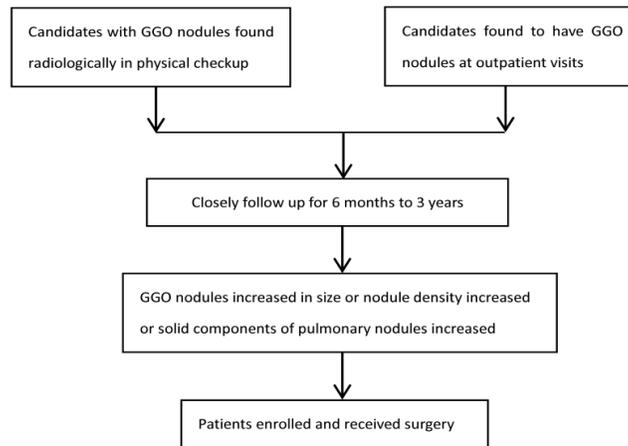
<sup>a</sup>lung adenocarcinoma.

<sup>b</sup>adenocarcinoma in situ.

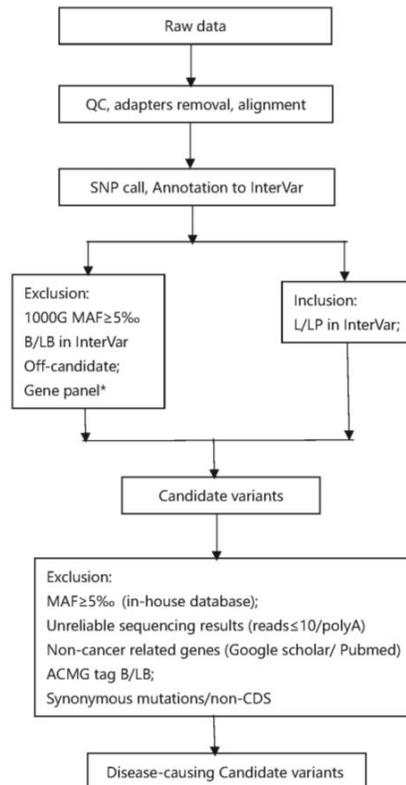
<sup>c</sup>ground glass opacity.

in 17 genes, NM\_000700:exon6:c.A418T in *AXAN1*, NM\_001184790:exon8:c.C1070T in *PARD3*, NM\_001170721:exon4:c.C392T in *BCAR1*, NM\_001127221:exon46:c.G6587A in *CACNA1A*, NM\_001170634:exon5:c.G383A in *FUS*, and NM\_002451:exon6:c.C538T in *MTAP* were present in three cases each. In addition, rs756875895 frameshift in *MAX*, NM\_001199292:exon7:c.C482G in *HSD17B4*, NM\_005732:exon13:c.2165\_2166insT in *RAD50*, NM\_001350128:exon11:c.T1172C in *PPOX*, NM\_001098816:exon28:c.A4751G in *TENM4*, NM\_004004:exon2:c.235delC in *GJB2*, and NM\_001142316:exon2:c.G203C in *LMO2* were present in two cases each. The remaining variants were present in one

A.



B.



**Figure 2** Flowchart of analysis. The stepwise filtering strategy used to screen for potential candidate germline mutations.

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case each. Of the 17 variants, 12 were nonsynonymous mutations, four were frameshift deletions, and one was a stopgain. The distribution of mutations in each patient was shown in Fig. 3.

We identified four potential deleterious frameshift deletions: rs80338943 in *GJB2*, rs587781454 in *RAD50*, rs756875895 in *MAX*, each occurring in two cases, and a frameshift in *LDLRAP1*, occurring in one case. The first frameshift, rs80338943 in *GJB2*, causing a p.L79fs fusion, was annotated as uncertain significance by InterVar. The second frameshift, rs587781454 in *RAD50* caused a p.K722fs fusion and was annotated as pathogenic from InterVar. The third frameshift, rs756875895 in *MAX*, was annotated as likely pathogenic in InterVar, causing a fusion of p.L52fs. The frameshift in *LDLRAP1* caused a fusion of p.W22fs in one case and was interpreted as pathogenic from InterVar, it might be deleterious in the development of lung cancer. Another potential deleterious variant was a stopgain, rs7755898 in *CYP21A2*, causing a protein change of p.Q289X which was likely pathogenic according to InterVar.

The other interesting candidates were four likely pathogenic SNVs annotated from InterVar: NOTCH 3:p.T357M (present in one case), HSD17B4 p.A161G (present in two cases), PPOX p.L391P (present in two cases), and TENM4 p.Q1584R (present in two cases).

There were five SNVs annotated as uncertain significance by InterVar that were present in three patients: ANXA1:p.I140 F, BCAR1:p.P131L, CACNA1A:p.R2196Q, FUS:p.S128N, and MTAP:p.R180W.

There were two additional candidate variants, LMO2 p.G68A and TTN p.R18629C, that were present in two cases and one case, respectively (Table 3). Their annotations by InterVar were of uncertain significance.

KEGG analysis did not indicate pathways that were related to *AXANI*, *TENM4* and *GJB2*. Pathways of *BCAR1*, *CYP21A2*, *LPLRAP1*, *HSD17B4*, *MTAP*, *PPOX* and *TTN* were not associated with cancer. Pathways derived from *NOTCH3*, *PARD3*, *CACNA1A*, *MAX*, *RAD50*, *FUS* and *LMO2* were cancer-related. The details were shown in a (Table S). Mutations in these genes were considered unlikely to cause cancer, therefore they would not be discussed here.

## DISCUSSION

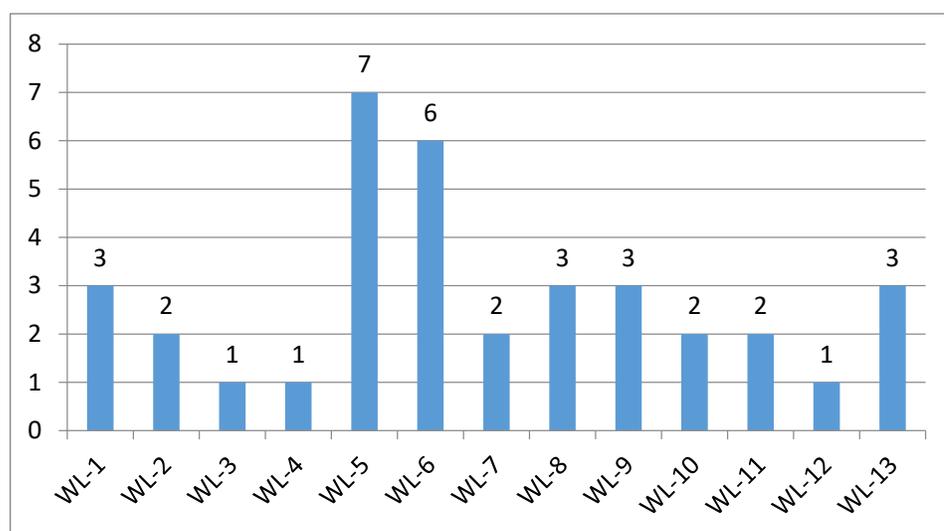
Although there are studies available on genetic mutations of lung cancer, the heritability of lung cancer, especially for GGO nodules, remains understudied compared to sporadic lung cancer. Using WES, our study reports germline mutations in GGO nodules of non-smoker lung cancer patients, largely females.

The discovery of germline mutations is very significant for both basic research and clinical treatment of lung cancer. First, germline mutations may play a role in tumorigenesis. Wang *et al.* (2018) reported that germline mutations interacted with somatic mutations, indicating their role in lung tumorigenesis. Tomoshige *et al.* (2015) also reported that germline mutations could cause familial lung cancer. Second, germline mutations are valuable for prognosis (Erdem *et al.*, 2012). For example, a study by Winther-Larsen *et al.* (2015) found that genetic polymorphism in the epidermal growth

**Table 3** Summary of potentially deleterious germline mutations in lung cancer cases. Annotation of potentially deleterious germline mutations in each gene were described in details.

Gene	Position	RS	Ref/Alt	Protein alteration	Genetic model	Type of mutation	InterVar annotation	VAF in patients	VAF in GnomAD_EAS	No. of patients with mutation
ANXA1	Chr 9: 75775752	–	A/T	NM_000700:exon6:c.A418T:p.I140F	–	nonsynonymous SNV	Uncertain significance	0.0833	–	3
NOTCH3	Chr 19: 15281611	–	T/G	NM_001184790:exon8:c.C1070T:p.T357M	AD <sup>a</sup> nonsynonymous SNV	Likely pathogenic	0.0278	–	1	
PARD3	Chr 10: 34671665	<a href="#">rs116642073</a>	G/A	NM_001184790:exon8:c.C1070T:p.T357M	–	nonsynonymous SNV	Uncertain significance	0.0833	0	3
BCAR1	Chr 16: 75269775	<a href="#">rs1047683608</a>	G/A	NM_001170721:exon4:c.C392T:p.P131L	–	nonsynonymous SNV	Uncertain significance	0.0833	0	3
CYP21A2	Chr 6: 32008198	<a href="#">rs7755898</a>	C/T	NM_001128590:exon7:c.C865T:p.Q289X	–	Stopgain	Likely pathogenic	0.0278	0.0001	1
LDLRAP1	Chr 1: 25870253	–	G/-	NM_015627:exon1:c.65delG:p.W22fs	AR <sup>b</sup> frameshift deletion	Pathogenic	0.0278	–	1	
CACNA1A	Chr 19: 13319766	<a href="#">rs373192655</a>	C/T	NM_001127221:exon46:c.G6587A:p.R2196Q	AD	nonsynonymous SNV	Uncertain significance	0.0556	0.0015	3
MAX	Chr 14: 65551007	<a href="#">rs756875895</a>	G/-	NM_001271068:exon3:c.154delC:p.L52fs	AD	Frameshift deletion	Likely pathogenic	0.0278	–	2
HSD17B4	Chr 5: 118814630	<a href="#">rs763363391</a>	C/G	NM_001199292:exon7:c.C482G:p.A161G	AR	nonsynonymous SNV	Likely pathogenic	0.0556	0.0002	2
RAD50	Chr 5: 131931460	<a href="#">rs587781454</a>	-/T	NM_005732:exon13:c.2165_2166insT:p.K722fs	–	Frameshift deletion	Uncertain significance	0.0556	–	2
PPOX	Chr 1: 161140719	–	T/C	NM_001350128:exon11:c.T1172C:p.L391P	AD	nonsynonymous SNV	Likely pathogenic	0.0278	–	2
FUS	Chr 16: 31195580	–	G/A	NM_001170634:exon5:c.G383A:p.S128N	AD	nonsynonymous SNV	Uncertain significance	0.0556	–	3
MTAP	Chr 9: 21854717	<a href="#">rs891972796</a>	C/T	NM_002451:exon6:c.C538T:p.R180W	AD	nonsynonymous SNV	Uncertain significance	0.0556	0	3
TENM4	Chr 11: 78412907	–	T/C	NM_001098816:exon28:c.A4751G:p.Q1584R	AD	nonsynonymous SNV	Likely pathogenic	0.0833	–	2
GJB2	Chr 13: 20763485	<a href="#">rs80338943</a>	G/-	NM_004004:exon2:c.235delC:p.L79fs	AD	Frameshift deletion	Uncertain significance	0.0556	–	2
TTN	Chr 2: 179427779	<a href="#">rs192360370</a>	G/A	NM_003319:exon154:c.C55885T:p.R18629C	AR/AD	nonsynonymous SNV	Uncertain significance	0.0278	0.0038	1
LMO2	Chr 11: 33886202	–	C/G	NM_001142316:exon2:c.G203C:p.G68A	–	nonsynonymous SNV	Uncertain significance	0.0556	–	2

**Notes.**<sup>a</sup>autosomal dominant.<sup>b</sup>autosomal recessive.



**Figure 3 Mutation distribution.** The distribution of germline mutations in each patient.

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factor receptor could predict the outcome in advanced NSCLC patients treated with erlotinib. Third, germline mutations are closely associated with a genetic predisposition to cancer, and screening for germline mutations is beneficial to the susceptible population ([Chen et al., 2015](#)) and for their health management.

In this study, we used a highly selective population, lung adenocarcinoma patients with GGOs, to investigate germline mutations and their possible role in the predisposition to lung cancer. In our cohort, 11 of 13 were females and all were non-smokers. The ethnicity of all patients was Han Chinese. The aforementioned facts were consolidated with the notion that malignant GGO nodules occur frequently in non-smokers and women ([Blons et al., 2006](#); [Raz et al., 2006](#)).

Strong evidence for two deleterious germline mutations ([rs587781454](#) in *RAD50* and [rs756875895](#) in *MAX*) has been shown in lung cancer patients. [rs587781454](#) in *RAD50* was reported as a hereditary predisposition and labelled as pathogenic in ClinVar ([Nykamp et al., 2017](#)). [rs756875895](#) in *MAX* was labelled as likely pathogenic by InterVar. Both variants occurred simultaneously in two females (WL-5 and WL-6). Both had minimally invasive GGO nodules. How these mutations in the same patient affected lung tumorigenesis is worth examining.

There was one likely pathogenic variant in *NOTCH 3* (WL-13). The expression of *NOTCH 3* was inversely associated with the sensitivity to platinum-based chemotherapy in patients with NSCLC. The *NOTCH 3* protein, rather than the gene polymorphism, is associated with the chemotherapy response and prognosis of advanced NSCLC patients ([Shi et al., 2014](#)).

Though annotated as uncertain significance by InterVar, three patients carried variants in *BCAR1* (WL-7, WL-10 and WL-13) and *CACNA1A* (WL-5, WL-6 and WL-9). Increased expression of *BCAR1* was associated with poor prognosis and carcinogenesis

in NSCLC (*Deng et al., 2013; Huang et al., 2012*). Overexpression of *CACNA1A* predicted a poor prognosis in NSCLC (*Zhou et al., 2017*). There were one additional candidate variants, LMO2 p.G68A in WL-1 and WL-8. Collectively, these findings suggest that germline mutations may function by regulating gene expression and thereby affect cancer development and/or prognosis.

Our study has limitations. First, the sample size is small. In our study, only non-smoker patients with malignant GGOs were enrolled. Second, gene expression was not investigated. Finally, the identified germline mutations have not been validated. These limitations restrict conclusions about their causative effects on tumorigenesis and their roles as biomarkers for prognosis or for treatment response.

## CONCLUSIONS

In summary, our results demonstrate potentially deleterious germline mutations in GGO nodules in non-smoking lung adenocarcinoma patients. These findings significantly expand the spectrum of genetic variants that may affect the response to therapies and patient survival and possibly increase the risk of being germline mutation carriers. However, due to the small patient samples, our observations encourage further studies. In future, prospective studies, expanding enrolled patients and functional studies should be performed to better understand their causative roles in tumorigenesis and prognosis, and to better manage patients' health.

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### Competing Interests

The authors declare there are no competing interests.

## Author Contributions

- Wenjun Mao, Ruo Chen and Mingfeng Zheng conceived and designed the experiments, authored or reviewed drafts of the paper, and approved the final draft.
- Rongguo Lu, Shengfei Wang and Feng Liu performed the experiments, authored or reviewed drafts of the paper, collected specimens, and approved the final draft.
- Huizhu Song analyzed the data, authored or reviewed drafts of the paper, and approved the final draft.
- Dan You analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the paper, and approved the final draft.
- Yijun He performed the experiments, authored or reviewed drafts of the paper, collected the clinical information, and approved the final draft.

## Human Ethics

The following information was supplied relating to ethical approvals (i.e., approving body and any reference numbers):

The Ethical Committee of Clinical New Technology and Medical Research Wuxi People's Hospital approves this research project (no: HS2019014).

## Data Availability

The following information was supplied regarding data availability:

Data are available in the National Center for Biotechnology Information Sequence Read Archive: [PRJNA613408](https://www.ncbi.nlm.nih.gov/sra/PRJNA613408).

## Supplemental Information

Supplemental information for this article can be found online at <http://dx.doi.org/10.7717/peerj.12048#supplemental-information>.

## REFERENCES

- Austin JH, Muller NL, Friedman PJ, Hansell DM, Naidich DP, Remy-Jardin M, Webb WR, Zerhouni EA. 1996.** Glossary of terms for CT of the lungs: recommendations of the Nomenclature Committee of the Fleischner Society. *Radiology* **200**(2):327–331 DOI [10.1148/radiology.200.2.8685321](https://doi.org/10.1148/radiology.200.2.8685321).
- Bartsch H, Dally H, Popanda O, Risch A, Schmezer P. 2007.** Genetic risk profiles for cancer susceptibility and therapy response. *Recent Results in Cancer Research* **174**:19–36 DOI [10.1007/978-3-540-37696-5\\_2](https://doi.org/10.1007/978-3-540-37696-5_2).
- Blons H, Cote JF, Le Corre D, Riquet M, Fabre-Guilevin E, Laurent-Puig P, Danel C. 2006.** Epidermal growth factor receptor mutation in lung cancer are linked to bronchioloalveolar differentiation. *American Journal of Surgical Pathology* **30**(10):1309–1315 DOI [10.1097/01.pas.0000213285.65907.31](https://doi.org/10.1097/01.pas.0000213285.65907.31).
- Bolger AM, Lohse M, Usadel B. 2014.** Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* **30**(15):2114–2120 DOI [10.1093/bioinformatics/btu170](https://doi.org/10.1093/bioinformatics/btu170).
- Chen HY, Yu SL, Ho BC, Su KY, Hsu YC, Chang CS, Li Y-C, Yang S-Y, Hsu P-Y, Ho Hao, Chang Y-H, Chen C-Y, Yang H-I, Hsu C-P, Yang T-Y, Chen Kun-Chieh,**

- Hsu K-H, Tseng J-S, Hsia Jiun-Yi, Chuang C-Y, Yuan S, Lee M-H, Liu C-H, Wu G-I, Hsiung CA, Chen Y-M, Wang C-L, Huang M-S, Yu C-J, Chen K-Y, Tsai Y-H, Su W-C, Chen H-W, Chen JJW, Chen C-J, Chang G-C, Yang P-C, Li KC. 2015. R331W missense mutation of oncogene YAP1 is a germline risk allele for lung adenocarcinoma with medical actionability. *Journal of Clinical Oncology* 33(20):2303–2310 DOI 10.1200/JCO.2014.59.3590.
- Deng B, Sun Z, Jason W, Yang P. 2013. Increased BCAR1 predicts poor outcomes of non-small cell lung cancer in multiple-center patients. *Annals of Surgical Oncology* 20(Suppl 3):S701–S708 DOI 10.1245/s10434-013-3184-2.
- DePristo MA, Banks E, Poplin R, Garimella KV, Maguire JR, Hartl C, Philippakis AA, Del Angel G, Rivas MA, Hanna M, McKenna A, Fennell TJ, Kernytisky Andrew M, Sivachenko AY, Cibulskis K, Gabriel SB, Altshuler D, Daly MJ. 2011. A framework for variation discovery and genotyping using next-generation DNA sequencing data. *Nature Genetics* 43(5):491–498 DOI 10.1038/ng.806.
- Donner I, Katainen R, Sipila LJ, Aavikko M, Pukkala E, Aaltonen LA. 2018. Germline mutations in young non-smoking women with lung adenocarcinoma. *Lung Cancer* 122:76–82 DOI 10.1016/j.lungcan.2018.05.027.
- Erdem L, Giovannetti E, Leon LG, Honeywell R, Peters GJ. 2012. Polymorphisms to predict outcome to the tyrosine kinase inhibitors gefitinib, erlotinib, so-rafenib and sunitinib. *Current Topics in Medicinal Chemistry* 12(15):1649–1659 DOI 10.2174/156802612803531333.
- Henschke CI, Shaham D, Yankelevitz DF, Kramer A, Kostis WJ, Reeves AP, Vazquez M, Koizumi J, Miettinen OS. 2006. CT screening for lung cancer: significance of diagnoses in its baseline cycle. *Clinical Imaging* 30(1):11–15 DOI 10.1016/j.clinimag.2005.07.003.
- Huang W, Deng B, Wang RW, Tan QY, He Y, Jiang YG, Zhou JH. 2012. BCAR1 protein plays important roles in carcinogenesis and predicts poor prognosis in non-small-cell lung cancer. *PLOS ONE* 7(4):e36124 DOI 10.1371/journal.pone.0036124.
- Hung RJ, McKay JD, Gaborieau V, Boffetta P, Hashibe M, Zaridze D, Mukeria A, Szeszenia-Dabrowska N, Lissowska J, Rudnai P, Fabianova E, Mates D, Bencko V, Foretova L, Janout V, Chen C, Goodman G, Field JK, Liloglou T, Xinarianos G, Cassidy A, McLaughlin J, Liu G, Narod S, Krokan HE, Skorpen F, Elvestad MB, Hveem K, Vatten L, Linseisen J, Clavel-Chapelon F, Vineis P, Bueno-de-Mesquita HB, Lund E, Martinez C, Bingham S, Rasmuson T, Hainaut P, R Elio, Ahrens W, Benhamou S, L P, Trichopoulos D, Holcátová I, Merletti F, Kjaerheim K, Agudo A, Macfarlane G, Talamini R, Simonato L, Lowry R, Conway DI, Znaor A, Healy C, Zelenika D, Boland A, Delepine M, Foglio M, Lechner D, Matsuda F, Blanche H, Gut I, Heath S, Lathrop M, Brennan P. 2008. A susceptibility locus for lung cancer maps to nicotinic acetylcholine receptor subunit genes on 15q25. *Nature* 452(7187):633–637 DOI 10.1038/nature06885.
- Ikeda K, Shiraishi K, Eguchi A, Osumi H, Matsuishi K, Matsubara E, Fujino K, Shibata H, Yoshimoto K, Mori T, Omori H, Suzuki M. 2014. Association of a genetic variant

- of CYP19A1 with multicentric development of lung adenocarcinomas. *Annals of Surgical Oncology* 21(3):939–945 DOI 10.1245/s10434-013-3362-2.
- Kanwal M, Ding XJ, Ma ZH, Li LW, Wang P, Chen Y, Huang Y-C, Cao Y. 2018.** Characterization of germline mutations in familial lung cancer from the Chinese population. *Gene* 641:94–104 DOI 10.1016/j.gene.2017.10.020.
- Lee HY, Choi YL, Lee KS, Han J, Zo JI, Shim YM, Moon JW. 2014.** Pure ground-glass opacity neoplastic lung nodules: histopathology, imaging, and management. *American Journal of Roentgenology* 202(3):W224–233 DOI 10.2214/AJR.13.11819.
- Li H, Durbin R. 2009.** Fast and accurate short read alignment with Burrows-Wheeler transform. *Bioinformatics* 25(14):1754–1760 DOI 10.1093/bioinformatics/btp324.
- Liu Y, Kheradmand F, Davis CF, Scheurer ME, Wheeler D, Tsavachidis S, Armstrong G, Simpson C, Mandal D, Kupert E, Anderson M, You M, Xiong D, Pikielny C, Schwartz AG, Bailey-Wilson J, Gaba C, De Andrade M, Yang P, Pinney SM, Amos CI, Spitz MR. 2016.** Focused analysis of exome sequencing data for rare germline mutations in familial and sporadic lung cancer. *Journal of Thoracic Oncology* 11(1):52–61 DOI 10.1016/j.jtho.2015.09.015.
- Malhotra J, Malvezzi M, Negri E, La Vecchia C, Boffetta P. 2016.** Risk factors for lung cancer worldwide. *European Respiratory Journal* 48(3):889–902 DOI 10.1183/13993003.00359-2016.
- Nykamp K, Anderson M, Powers M, Garcia J, Herrera B, Ho YY, Kobayashi Y, Patil N, Thusberg J, Westbrook M, Topper S. 2017.** Sherlock: a comprehensive refinement of the ACMG-AMP variant classification criteria. *Genetics in Medicine* 19(10):1105–1117 DOI 10.1038/gim.2017.37.
- Raz DJ, He B, Rosell R, Jablons DM. 2006.** Current concepts in bronchioloalveolar carcinoma biology. *Clinical Cancer Research* 12(12):3698–3704 DOI 10.1158/1078-0432.CCR-06-0457.
- Renieri A, Mencarelli MA, Cetta F, Baldassarri M, Mari F, Furini S, Piu P, Ariani F, Dragani TA, Frullanti E. 2014.** Oligogenic germline mutations identified in early non-smokers lung adenocarcinoma patients. *Lung Cancer* 85(2):168–174 DOI 10.1016/j.lungcan.2014.05.020.
- Shi C, Qian J, Ma M, Zhang Y, Han B. 2014.** Notch 3 protein, not its gene polymorphism, is associated with the chemotherapy response and prognosis of advanced NSCLC patients. *Cellular Physiology and Biochemistry* 34(3):743–752 DOI 10.1159/000363039.
- Shukuya T, Patel S, Shane-Carson K, He K, Bertino EM, Shilo K, Otterson GA, Carbone DP. 2018.** Lung cancer patients with germline mutations detected by next-generation sequencing and/or liquid biopsy. *Journal of Thoracic Oncology* 13(2):e17–e19 DOI 10.1016/j.jtho.2017.09.1962.
- Siegel RL, Miller KD, Jemal A. 2015.** Cancer statistics, 2015. *CA: A Cancer Journal for Clinicians* 65(1):5–29 DOI 10.3322/caac.21254.
- Subramanian J, Govindan R. 2007.** Lung cancer in never smokers: a review. *Journal of Clinical Oncology* 25(5):561–570 DOI 10.1200/JCO.2006.06.8015.

- Tomoshige K, Matsumoto K, Tsuchiya T, Oikawa M, Miyazaki T, Yamasaki N, Mishima H, Kinoshita A, Kubo T, Fukushima K, Yoshiura K-i, Nagayasu T. 2015.** Germline mutations causing familial lung cancer. *Journal of Human Genetics* **60(10)**:597–603 DOI [10.1038/jhg.2015.75](https://doi.org/10.1038/jhg.2015.75).
- Wang Y, Wang C, Zhang J, Zhu M, Zhang X, Li Z, Dai J, Ma H, Hu Z, Jin G, Shen H. 2018.** Interaction analysis between germline susceptibility loci and somatic alterations in lung cancer. *International Journal of Cancer* **143(4)**:878–885 DOI [10.1002/ijc.31351](https://doi.org/10.1002/ijc.31351).
- Winther-Larsen A, Nissen PH, Jakobsen KR, Demuth C, Sorensen BS, Meldgaard P. 2015.** Genetic polymorphism in the epidermal growth factor receptor gene predicts outcome in advanced non-small cell lung cancer patients treated with erlotinib. *Lung Cancer* **90(2)**:314–320 DOI [10.1016/j.lungcan.2015.09.003](https://doi.org/10.1016/j.lungcan.2015.09.003).
- Zhang Y, Zhang L, Li R, Chang DW, Ye Y, Minna JD, Roth JA, Han B, Wu X. 2017.** Genetic variations in cancer-related significantly mutated genes and lung cancer susceptibility. *Annals of Oncology* **28(7)**:1625–1630 DOI [10.1093/annonc/mdx161](https://doi.org/10.1093/annonc/mdx161).
- Zhou X, Wang W, Zhang S, Wang X, Tang Z, Gu J, Li J, Huang J. 2017.** CACNA1B (Cav2.2) overexpression and its association with clinicopathologic characteristics and unfavorable prognosis in non-small cell lung cancer. *Disease Markers* **2017**:6136401 DOI [10.1155/2017/6136401](https://doi.org/10.1155/2017/6136401).