

Effects of chlorhexidine, ethylenediaminetetraacetic acid, and sodium hypochlorite on cell viability of human gingival fibroblasts *in vitro*

Abdel-Rahman Youssef^{1,2}, Ehdaa Alturkistani³, Israa Muharrij³, Lina Alsrehi³, Noor Shafei³, Nora Alzahrani³, Mashael Alqahtani¹

¹Department of Basic and Clinical Oral Sciences, Faculty of Dentistry, Umm Al-Qura University, Saudi Arabia, ²Department of Microbiology, Faculty of Medicine, Suez Canal University, Egypt, ³Dental intern, Faculty of Dentistry, Umm Al-Qura University, Saudi Arabia

Abstract

Introduction: The aim of this *in vitro* study was to evaluate the cytotoxic effects of chlorhexidine (CHX), ethylenediaminetetraacetic acid (EDTA), and sodium hypochlorite (NaOCl) irrigants on human gingival fibroblasts.

Materials and Methods: Gingival fibroblasts were cultured in Dulbecco Modified Eagle Medium for 24 h. Then, the cells were exposed for 1 min to different concentrations of CHX, EDTA, and NaOCl and the cell viability was assessed using 3-(4,5-dimethylthiazolyl-2)-2,5-diphenyltetrazolium bromide tetrazolium reduction assay. The percentage of gingival fibroblast viability was compared to control (100% viability).

Results: All concentrations of CHX, EDTA, and NaOCl were cytotoxic to gingival fibroblasts compared to control group and this effect was dose-dependent ($P < 0.0001$). There was an inverse relationship between the concentrations of these irrigants and cell viability. The highest clinically used concentrations of CHX (2%), EDTA (17%) and NaOCl (2.5%) reduced the cell viability to 2.2%, 3.2%, and 1.9%, respectively, compared to the 100% viability of control. However, the lowest concentration of CHX (0.00002%), EDTA (0.00017%), and NaOCl (0.000025%) increased the cell viability to 39.8%, 30.2%, and 44.2%, respectively, in comparison to control. There were no significant differences between the irrigants at clinically used concentrations ($P > 0.05$).

Conclusion: The cytotoxic effect of CHX, EDTA, and NaOCl on gingival fibroblasts was dose-dependent. Further studies are needed to assess and optimize the safety and efficacy of these irrigants *in vivo*.

Keywords: Chlorhexidine, cytotoxicity, ethylenediaminetetraacetic acid, fibroblasts, sodium hypochlorite

Address for correspondence: Dr. Abdel-Rahman Youssef, Department of Basic and Clinical Oral Sciences, Faculty of Dentistry, Umm Al-Qura University, Saudi Arabia.

E-mail: amyoussef@uqu.edu.sa

Submission: 02-10-19 **Revision:** 06-12-19 **Acceptance:** 07-12-19 **Web Publication:** 27-08-20

Access this article online

Quick Response Code:



Website:

www.saudiendodj.com

DOI:

10.4103/sej.sej_149_19

This is an open access journal, and articles are distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 4.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as appropriate credit is given and the new creations are licensed under the identical terms.

For reprints contact: WKHLRPMedknow_reprints@wolterskluwer.com

How to cite this article: Youssef AR, Alturkistani E, Muharrij I, Alsrehi L, Shafei N, Alzahrani N, *et al.* Effects of chlorhexidine, ethylenediaminetetraacetic acid, and sodium hypochlorite on cell viability of human gingival fibroblasts *in vitro*. Saudi Endod J 2020;10:234-9.

INTRODUCTION

The endodontic root canal treatment is common dental procedure aims to eliminate infection and remove all the inflamed and necrotic tissues in the pulp through the use of intracanal medicaments during the mechanical cleaning and shaping of the involved root canal.^[1,2]

Spreading of the endodontic infection into periapical tissues cause inflammation of the periapical tissues where the periodontal ligament and bone are damaged and replaced by inflamed tissue which is characteristically fibrous and infiltrated by inflammatory cells.^[3]

The ideal irrigation solution should eliminate or reduce the intracanal microbial population and serve as a lubricant during instrumentation.^[4] It should also have low toxic effect to the periodontium to avoid the destructive sequela of irrigant extrusion through apical foramen to the surrounding tissues which prevent regeneration and recovery.^[5]

A widely diverse irrigating solutions are available for endodontic treatment, including sodium hypochlorite (NaOCl), chlorhexidine gluconate (CHX), ethylenediaminetetraacetic acid (EDTA), QMix, and normal saline.^[6]

NaOCl is the most frequently used endodontic irrigant due to its efficiency in dissolving the remnants of necrotic tissue and its antimicrobial activity but is not able to remove the smear layer.^[7] It is usually used at a concentration range of 0.5%–5.25%.^[8] EDTA is a powerful efficient chelating agent used in endodontic treatment,^[9] because of its ability to remove the mineralized portion of smear layer but not soft tissue.^[10] CHX is commonly used as an endodontic irrigant because it has the range of antibacterial and antifungal activity.^[11] Several *in vivo* studies demonstrated the lingering effect of CHX and its anti-microbial activity with residual effects in the root canal system.^[12,13] QMix solution (combination of 17% EDTA and 2% CHX), removes the smear layers and has antimicrobial activity.^[14]

Irrigant extrusion into the periapical area may influence the survival of the cells of pulpal and periapical tissues.^[15] NaOCl can cause injury if it comes in contact with vital tissues.^[16] The fibroblasts, osteoblasts, and endothelial cells are the most affected cells. The periodontal ligament fibroblasts present in the apical constriction are the main cells that first react to irrigant extrusion.^[17] These cells as well as gingival fibroblasts are commonly used in evaluating the toxicity of endodontic materials. Histological studies

have shown that the morphology, and growth rates of gingival fibroblasts and periodontal ligament fibroblasts are similar.^[18,19]

Previous studies evaluating cytotoxicity of intracanal irrigants have showed contradicting results. Vouzara *et al.*^[20] have found 17% EDTA to be less cytotoxic to fibroblasts than 2.5% NaOCl, while Karkehabadi *et al.*^[21] showed that EDTA had the highest toxicity to human periodontal ligament fibroblasts compared to NaOCl and CHX. In addition, Farhad Mollashahi *et al.*^[22] have shown that CHX had the lowest cytotoxicity to stem cells from the human apical papilla compared to EDTA, and NaOCl. However, Vouzara *et al.*^[20] showed CHX to be significantly more cytotoxic to human lung fibroblasts than NaOCl and EDTA. Hence, the aim of this study was to compare the cytotoxicity of CHX, EDTA, and NaOCl irrigants on human gingival fibroblast cells using ten-fold serial dilutions with a starting concentration of 2%, 17%, and 2.5%, respectively.

MATERIALS AND METHODS

Cell culture

Human gingival tissues were collected from healthy adult gingiva at Dental Teaching Hospital after obtaining signed informed consent and approval of ethical committee of Faculty of Dentistry, Umm Al-Qura University, Makkah, Saudi Arabia. The gingival tissue was cut into small pieces and then digested in a solution of 3 mg/mL collagenase Type I and 4 mg/mL dispase (Sigma, USA) for 1 h at 37°C. Single-cell suspensions were obtained by passing cells through a 70 µm cell strainer and cultured in 25 mL flask in Dulbecco Modified Eagle Medium (DMEM, UFC Biotech, KSA) containing 10% fetal bovine serum (FBS; HyClone Thermo Scientific, USA), 100 U/mL penicillin, 100 µg/mL streptomycin (Sigma, USA), and incubated at 37°C in a humidified atmosphere of 5% CO₂.

Gingival fibroblast treatment

Gingival fibroblasts at passage 3 were seeded at 10,000 cells/well in 96 well-plate and incubated in DMEM with fetal bovine serum and penicillin/streptomycin and incubated at 37°C in a humidified atmosphere of 5% CO₂ for 24 h. When the cells reached 70% confluency, they were exposed for 1 min to 10-fold serial dilutions of chlorhexidine, EDTA, and NaOCl starting with concentration of 2%, 17%, and 2.5%, respectively. The control cells were left untreated. After 1 min, the cells were washed 3X in PBS and reapplication of complete DMEM and the cell viability were assessed using 3-(4,5-dimethylthiazolyl-2)-2,5-diphenyltetrazolium bromide (MTT) assay.

3-(4,5-Dimethylthiazolyl-2)-2,5-diphenyltetrazolium bromide cell viability assay

The MTT assay was used to evaluate the cytotoxic effect of CHX, EDTA and NaOCl on gingival fibroblasts. The cell viability was assessed with a MTT (Sigma). The MTT assay method developed by Carmichael *et al.*^[23] was followed. Briefly, the medium was removed and replaced with 100 µl/well complete DMEM containing 0.5 mg/ml of MTT and incubated for 3 h at 37°C. At the end of the incubation period, the medium was removed and DMSO: Isopropanol (1:1) solvent solution was added to dissolve formazan crystals. The optical density was read at 570 nm by a spectrophotometric Microplate Reader (SpectroStar Nano, BMG Lab). The optical density values obtained for each irrigant were used to calculate cell viability relative to untreated control, which is set at 100%.

Statistical analysis

The data were collected, tabulated, and analyzed using one-way ANOVA using Graphpad prism 7. The cell viability experiments were performed in triplicate and the results are expressed as mean \pm standard error of the mean and differences were significant if a $P < 0.05$.

RESULTS

All tested concentrations of CHX [Figure 1], EDTA [Figure 2], and NaOCl [Figure 3] were cytotoxic to the fibroblasts and this effect was dose-dependent ($P < 0.0001$). There was an inverse relationship between the concentrations of CHX, EDTA, and NaOCl with the cell viability.

The percentage of fibroblast cell viability of CHX, EDTA, and NaOCl were compared to control that represent 100%. The cell viability was 2.2% for CHX at the highest concentration (2%) and 39.8% at the lowest concentration (0.00002%), whereas EDTA at highest

concentration (17%) reported 3.2% viability and at the lowest concentration (0.00017%) it was 30.2%. Similarly, the viability of the cells exposed to high concentration of NaOCl (2.5%) was reduced to 1.9% but the lowest concentration (0.000025%) increased the cell viability to 44.2%. When we compared the cytotoxic effect of the three irrigants at clinically used concentrations we found that all irrigants were cytotoxic to the fibroblasts ($P < 0.0001$) and there were no significant differences among them ($P > 0.05$).

DISCUSSION

Cell viability assays are used to determine the cytotoxic effect of test material. The frequently used assays include the tetrazolium or resazurin reduction and protease activity assays that measure general metabolism or an enzymatic activity of the viable cells. Another commonly used assay is the luminogenic ATP assay which is the fastest and most sensitive, whereas tetrazolium or resazurin reduction assays are cheap with satisfactory performance. The most commonly used compounds in Tetrazolium reduction assays include viable MTT, MTS, XTT, and WST-1.^[24] In this study, MTT assay was used to assess the cytotoxic effects of three endodontic irrigation solutions, CHX, EDTA, and NaOCl on the human gingival fibroblasts. It was found that all concentrations were cytotoxic to gingival fibroblasts compared to control group and this effect was dose-dependent. The results of the current study confirmed previous findings that NaOCl, EDTA, and CHX are cytotoxic to periodontal ligament fibroblasts^[21] and fibroblast cell line in dose-dependent manner.^[20,25] It has been found that higher concentrations of NaOCl (1%, 3%, and 5%) were cytotoxic to periodontal ligament fibroblasts^[26] and NaOCl concentrations $>0.05\%$ caused complete death of human dermal fibroblasts.^[27]

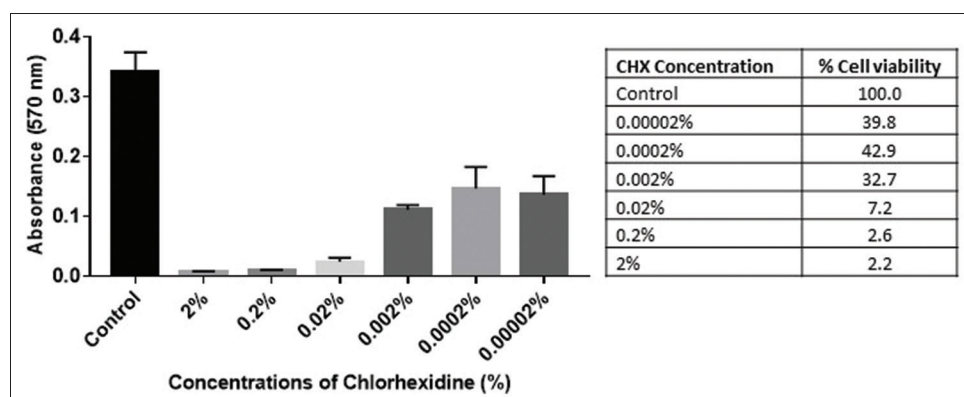


Figure 1: Cell viability of gingival fibroblasts after treatment with CHX. The cells were incubated with different concentrations of CHX for 1 min and the cell viability was measured by MTT assay. All concentrations of CHX were cytotoxic to the fibroblasts and this effect was dose-dependent ($P < 0.0001$). CHX: Chlorhexidine, MTT: 3-(4,5-Dimethylthiazolyl-2)-2,5-diphenyltetrazolium bromide

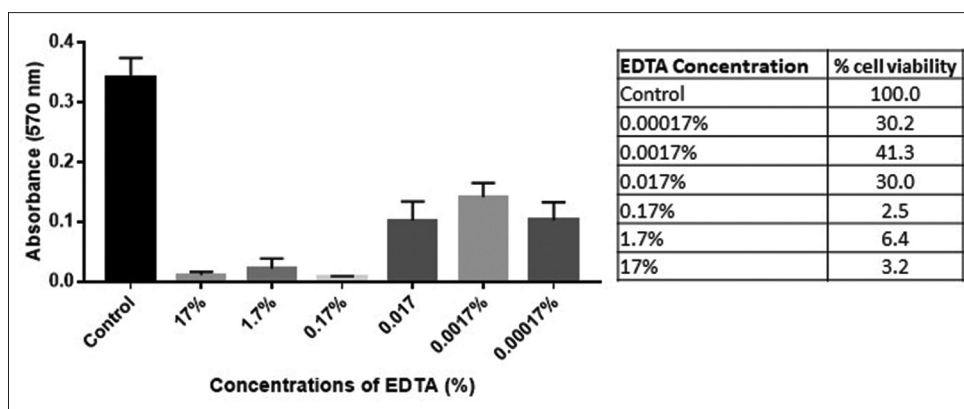


Figure 2: Cell viability of gingival fibroblasts after treatment with EDTA. The cells were incubated with different concentrations of EDTA for 1 min and the cell viability was measured by MTT assay. All concentrations of EDTA were cytotoxic to the fibroblasts and this effect was dose-dependent ($P < 0.0001$). EDTA: ethylenediaminetetraacetic acid, MTT: 3-(4,5-Dimethylthiazolyl-2)-2,5-diphenyltetrazolium bromide

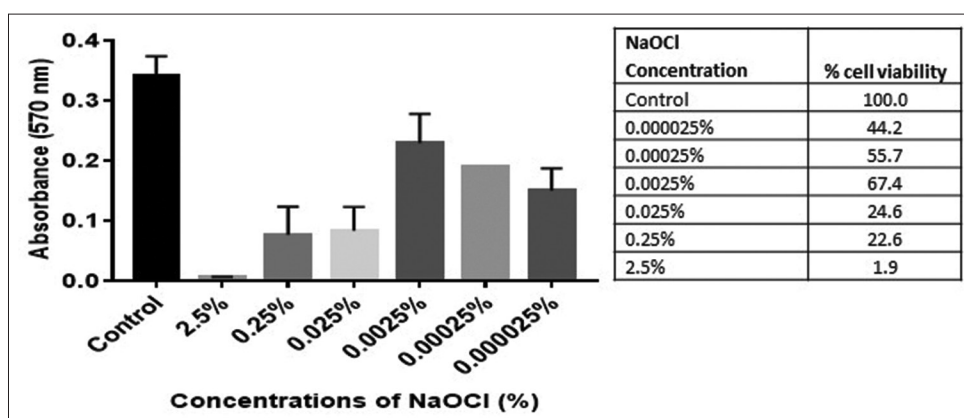


Figure 3: Cell viability of gingival fibroblast after treatment with NaOCl. The cells were incubated with different concentrations of NaOCl for 1 min and the cell viability was measured by MTT assay. All concentrations of NaOCl were cytotoxic to the fibroblast and this effect was dose-dependent ($P < 0.0001$). NaOCl: Sodium hypochlorite, MTT: 3-(4,5-Dimethylthiazolyl-2)-2,5-diphenyltetrazolium bromide

The results of the present study showed that the used concentrations of CHX (2%), EDTA (17%), and NaOCl (2.5%) reduced the cell viability dramatically and there were no significant differences between these irrigants. These findings are in accordance with other *in vitro* studies that have shown that the used concentration of CHX (2%), EDTA (17%), and NaOCl (2.5%) CHX (2%) are cytotoxic to fibroblasts.^[20,25,28] On the contrary, Karkehabadi *et al.*^[21] showed that EDTA had the highest toxicity to human periodontal ligament fibroblasts compared to NaOCl and CHX, while Vouzara *et al.*^[20] found CHX to be significantly more cytotoxic to human lung fibroblasts than NaOCl and EDTA.

The cytotoxicity of CHX is associated with the inhibition of protein synthesis^[29] or the induction of H_2O_2 and superoxide radicals,^[30] whereas the cytotoxic effect EDTA might be related to its ability to penetrate cell membrane and intracellular chelation of ions.^[31] On the other hand, NaOCl cytotoxicity can be attributed to its high pH which interferes with the integrity of cytoplasmic

membrane^[32] or inhibition of DNA synthesis leading to growth arrest.^[27]

Cytotoxicity of irrigation solutions may differ due to variation in the concentration of the solution, exposure length, composition of the exposure medium, and cell type and the evaluation methods used.^[33] Teixeira *et al.*^[25] have shown that NaOCl was able to maintain human fibroblasts cell viability only with the 0.01% dilution while 0.05% and 0.1% dilutions were cytotoxic. A recent study has demonstrated that CHX at low concentration (0.002%) did not affect the proliferation of human gingival fibroblasts but at higher concentration ($\geq 0.04\%$) suppressed cell proliferation.^[34] In addition, Babich *et al.*^[35] demonstrated that cytotoxicity values were 0.106, 0.011, and 0.0045 mmol/L after 1-, 24-, and 72-h exposures of CHX to gingival epithelial cell, respectively. Furthermore, the cytotoxicity was reduced when fetal bovine serum in the exposure medium was increased from 2% to 8% and the medium amended with albumin, lecithin, and heat-killed *Escherichia coli*.^[35]

In general, *in vitro* cytotoxicity tests are only indicative of what may happen in clinical situations when dealing with root canal treatment and they cannot be directly implemented in clinical practice.^[20]

This study has some potential limitations. One of these limitations is that only MTT assay was used to examine cell viability and not using apoptosis and necrotic cell death pathways that remain to be studied. Furthermore, the experiment was conducted on cultured cells, and the outcome represents the response of these cells without considering the host defense mechanism.

CONCLUSION

The cytotoxic effect of CHX, EDTA and NaOCl on gingival fibroblasts was dose-dependent. Further studies are needed to assess and optimize the safety and efficacy of these irrigants *in vivo*.

Financial support and sponsorship

Nil.

Conflicts of interest

There are no conflicts of interest.

REFERENCES

- Garberoglio R, Becce C. Smear layer removal by root canal irrigants. A comparative scanning electron microscopic study. *Oral Surg Oral Med Oral Pathol* 1994;78:359-67.
- Tomson PL, Simon SR. Contemporary cleaning and shaping of the root canal system. *Prim Dent J* 2016;5:46-53.
- Liao J, Al Shahrani M, Al-Habib M, Tanaka T, Huang GT. Cells isolated from inflamed periapical tissue express mesenchymal stem cell markers and are highly osteogenic. *J Endod* 2011;37:1217-24.
- Chiniforush N, Pourhajbagher M, Shahabi S, Bahador A. Clinical approach of high technology techniques for control and elimination of endodontic microbiota. *J Lasers Med Sci* 2015;6:139-50.
- Navarro-Escobar E, González-Rodríguez MP, Ferrer-Luque CM. Cytotoxic effects of two acid solutions and 2.5% sodium hypochlorite used in endodontic therapy. *Med Oral Patol Oral Cir Bucal* 2010;15:e90-4.
- Singla MG, Garg A, Gupta S. MTAD in endodontics: An update review. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2011;112:e70-6.
- Dutner J, Mines P, Anderson A. Irrigation trends among American association of endodontists members: A web-based survey. *J Endod* 2012;38:37-40.
- Chaugule VB, Panse AM, Gawali PN. Adverse reaction of sodium hypochlorite during endodontic treatment of primary teeth. *Int J Clin Pediatr Dent* 2015;8:153-6.
- Serper A, Calt S. The demineralizing effects of EDTA at different concentrations and pH. *J Endod* 2002;28:501-2.
- Raisingani D, Meshram GK. Cleanliness in the root canal system: An scanning electron microscopic evaluation of manual and automated instrumentation using 4% sodium hypochlorite and EDTA (glyde file prep)-An *in vitro* study. *Int J Clin Pediatr Dent* 2010;3:173-82.
- Gomes BP, Vianna ME, Zaia AA, Almeida JF, Souza-Filho FJ, Ferraz CC. Chlorhexidine in endodontics. *Braz Dent J* 2013;24:89-102.
- White RR, Hays GL, Janer LR. Residual antimicrobial activity after canal irrigation with chlorhexidine. *J Endod* 1997;23:229-31.
- Mohammadi Z. Chlorhexidine gluconate, its properties and applications in endodontics. *Iran Endod J* 2008;2:113-25.
- Jose J, Krishnamma S, Peedikayil F, Aman S, Tomy N, Mariodan JP. Comparative evaluation of antimicrobial activity of QMiX, 2.5% sodium hypochlorite, 2% Chlorhexidine, guava leaf extract and *Aloe vera* Extract Against *Enterococcus faecalis* and *Candida albicans* – An *in-vitro* study. *J Clin Diagn Res* 2016;10:ZC20-3.
- Trevino EG, Patwardhan AN, Henry MA, Perry G, Dybdal-Hargreaves N, Hargreaves KM, et al. Effect of irrigants on the survival of human stem cells of the apical papilla in a platelet-rich plasma scaffold in human root tips. *J Endod* 2011;37:1109-15.
- Gatot A, Arbelle J, Leiberman A, Yanai-Inbar I. Effects of sodium hypochlorite on soft tissues after its inadvertent injection beyond the root apex. *J Endod* 1991;17:573-4.
- Wahjuningrum DA, Elizabeth ME, Puteri FH, Mardiyah AA, Subiyanto A. Cytotoxicity assay of sodium hypochlorite and QMix on cultured human periodontal ligament fibroblast cells. *J Int Oral Health* 2019;11:204-7.
- Hou LT, Yaeger JA. Cloning and characterization of human gingival and periodontal ligament fibroblasts. *J Periodontol* 1993;64:1209-18.
- Somerman MJ, Archer SY, Imm GR, Foster RA. A comparative study of human periodontal ligament cells and gingival fibroblasts *in vitro*. *J Dent Res* 1988;67:66-70.
- Vouzara T, Koulaouzidou E, Ziouti F, Economides N. Combined and independent cytotoxicity of sodium hypochlorite, ethylenediaminetetraacetic acid and chlorhexidine. *Int Endod J* 2016;49:764-73.
- Karkehabadi H, Yousefifakhr H, Zadsirjan S. Cytotoxicity of endodontic irrigants on human periodontal ligament cells. *Iran Endod J* 2018;13:390-4.
- Farhad Mollashahi N, Saberi E, Karkehabadi H. Evaluation of cytotoxic effects of various endodontic irrigation solutions on the survival of stem cell of human apical papilla. *Iran Endod J* 2016;11:293-7.
- Carmichael J, DeGraff WG, Gazdar AF, Minna JD, Mitchell JB. Evaluation of a tetrazolium-based semiautomated colorimetric assay: Assessment of chemosensitivity testing. *Cancer Res* 1987;47:936-42.
- Riss TL, Moravec RA, Niles AL, Duellman S, Benink H, Worzella TJ, et al. Cell viability assays. In: Sitampalam GS, Coussens NP, Brimacombe K. editors. *Assay Guidance Manual* [Internet]. Bethesda (MD): Eli Lilly & Company and the National Center for Advancing Translational Sciences; 2004. Available from: <https://www.ncbi.nlm.nih.gov/pubmed/23805433?report=docsum>. [Last updated on 2016 Jul 01].
- Teixeira PA, Coelho MS, Kato AS, Fontana CE, Bueno CE, Pedro-Rocha DG. Cytotoxicity assessment of 1% peracetic acid, 2.5% sodium hypochlorite and 17% EDTA on FG11 and FG15 human fibroblasts. *Acta Odontol Latinoam* 2018;31:11-5.
- Singh A, Kakkar P, Pant AB. Comparative evaluation of cytotoxic effects of MTAD and sodium hypochlorite using lactate dehydrogenase and trypan blue assays: An *in vitro* study. *Saudi Endod J* 2018;8:189-95.
- Hidalgo E, Bartolome R, Dominguez C. Cytotoxicity mechanisms of sodium hypochlorite in cultured human dermal fibroblasts and its bactericidal effectiveness. *Chem Biol Interact* 2002;139:265-82.
- Liu JX, Werner J, Kirsch T, Zuckerman JD, Virk MS. Cytotoxicity evaluation of chlorhexidine gluconate on human fibroblasts, myoblasts, and osteoblasts. *J Bone Jt Infect* 2018;3:165-72.
- Chang YC, Huang FM, Tai KW, Chou MY. The effect of sodium hypochlorite and chlorhexidine on cultured human periodontal ligament cells. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2001;92:446-50.
- Yeung SY, Huang CS, Chan CP, Lin CP, Lin HN, Lee PH, et al. Antioxidant and pro-oxidant properties of chlorhexidine and its interaction with calcium hydroxide solutions. *Int Endod J* 2007;40:837-44.
- Hülsmann M, Heckendorff M, Lennon A. Chelating agents in root canal treatment: Mode of action and indications for their use. *Int*

- Endod J 2003;36:810-30.
32. Estrela C, Estrela CR, Barbin EL, Spanó JC, Marchesan MA, Pécora JD. Mechanism of action of sodium hypochlorite. Braz Dent J 2002;13:113-7.
33. Mohammadi Z, Abbott PV. The properties and applications of chlorhexidine in endodontics. Int Endod J 2009;42:288-302.
34. Wyganowska-Swiatkowska M, Kotwicka M, Urbaniak P, Nowak A, Skrzypczak-Jankun E, Jankun J. Clinical implications of the growth-suppressive effects of chlorhexidine at low and high concentrations on human gingival fibroblasts and changes in morphology. Int J Mol Med 2016;37:1594-600.
35. Babich H, Wurzbürger BJ, Rubin YL, Sinensky MC, Blau L. An *in vitro* study on the cytotoxicity of chlorhexidine digluconate to human gingival cells. Cell Biol Toxicol 1995;11:79-88.