

## REVIEW

# Decline of ergothioneine in frailty and cognition impairment

 Hiroshi Kondoh<sup>1</sup> , Takayuki Teruya<sup>2</sup>, Masahiro Kameda<sup>1</sup> and Mitsuhiro Yanagida<sup>2</sup>
<sup>1</sup> Geriatric unit, Graduate School of Medicine, Kyoto University, Japan

<sup>2</sup> G0 Cell Unit, Okinawa Institute of Science and Technology Graduate University (OIST), Japan

## Correspondence

H. Kondoh, Geriatric unit, Graduate School of Medicine, Kyoto University, Sakyo-ku, Kyoto, 606-8507, Japan

Tel: +81 75 751 3465

E-mail: hkondoh@kuhp.kyoto-u.ac.jp and

M. Yanagida, G0 Cell Unit, Okinawa Institute of Science and Technology Graduate University (OIST), Onna-son, Okinawa, 904-0495, Japan

Tel: +81 98 966 8658

E-mail: myanagid@gmail.com

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Ergothioneine is a well-known antioxidant that is abundant in both human red blood cells and in fission yeast responding to nutritional stress. In frail elderly people, whose ageing organs undergo functional decline, there is a correlation between ergothioneine levels and cognitive, but not skeletal muscle decline. In patients suffering from dementia, including Alzheimer's disease with hippocampal atrophy, deteriorating cognitive ability is correlated with declining ergothioneine levels. S-methyl-ergothioneine, trimethyl-histidine and three other trimethyl-ammonium compounds also decrease sharply in dementia, whereas compounds such as indoxyl-sulfate and quinolinic acid increase, possibly exacerbating the disease. Using these opposing dementia markers, not only diagnosis, but also therapeutic interventions to mitigate cognitive decline may now become possible.

**Keywords:** antioxidant; dementia; ergothioneine; frailty; metabolomics; sarcopenia; starvation; whole blood

Due to ageing of the global population, ageing-related diseases are increasing, including diabetes, atherosclerosis, hypertension, osteoporosis, dementia, cancer and others [1]. Because 'ageing' is a complex multi-faceted process, it is characterised by great individual variability [2,3]. In countries with a high proportion of elderly people, a greater variety of ageing and ageing-related diseases is observed. While the number of healthy elderly people is increasing, the number of bedridden patients and people who need long-term nursing care is increasing as well [1,4]. Patients with ageing-related diseases, such as dementia, frailty and sarcopenia, also require intensive nursing care and family support.

Dementia is a form of cognitive impairment that develops into an irreversible cognitive decline [5,6]. In

its initial stage, it is easily confused with forgetfulness due to ageing. But later, in addition to core symptoms of memory impairment or disorientation, patients are frequently afflicted with behavioural and psychological symptoms of dementia, such as delusions, hallucinations, wandering, aggressiveness, sleep disorders, resistance to nursing care and depression, resulting in severely impaired daily life activities [7]. On the other hand, frailty is defined as a condition in which vulnerability to stress increases due to a decreased physiological reserve capacity [8]. Frailty comprises hypomobility, impaired cognition and social disturbance, while sarcopenic patients show reduced volume and strength of skeletal muscles due to muscle ageing [9,10]. Both frailty and sarcopenia result in life dysfunction and

## Abbreviations

CR, calorie restriction; CV, coefficient of variation; EFS, Edmonton Frailty Scale; ERG, ergothioneine; IF, intermittent fasting; LC-MS, liquid chromatography-mass spectrometry; MMSE, mini mental state examination; MoCA-J, the Japanese version of the Montreal Cognitive Assessment; OA, ophthalmic acid; OCTN, organic cation transporter; PPP, pentose phosphate pathway; RBCs, red blood cells; ROS, reactive oxygen species; Se, selenium; S-methyl-ERG, S-methyl-ergothioneine; WBCs, white blood cells.

nursing care, condemning affected individuals to a bedridden state and death [11,12].

Ergothioneine (ERG), a water-soluble amino acid derivative, was discovered in 1909, when it was isolated from *Claviceps purpurea*, the fungal parasite that causes ergot in rye [13,14]. ERG is found in many organisms, although animals and plants cannot synthesise it themselves [15,16]; indeed, ERG is mainly produced by fungi, yeasts, bacteria and a few other organisms [17–19]. ERG has excellent heat and pH stability and its antioxidative capacity against hydroxyl radicals and peroxynitrite *in vitro* is about three times and about 30 times higher than that of glutathione, another antioxidant respectively [20]. Because vertebrates cannot synthesise ERG, they extract it from food and store it in the brain, liver, kidneys, red blood cells (RBCs), skin, etc. [21–23].

We have established a metabolomic approach using whole blood, which enables us to comprehensively analyse metabolite profiles in cellular and non-cellular components of human blood [24,25]. This whole blood metabolome identified several markers for ageing, fasting, frailty, sarcopenia and dementia. Among them, we noticed a decline in ERG levels in the blood of patients suffering from frailty and dementia, but not from sarcopenia [26–28]. As glucose starvation leads to ERG upregulation in both in human blood and yeast, the antioxidant properties of ERG may protect individuals from ageing-relevant dysfunctions [29,30]. Here we review recent findings on ERG in relation to frailty and dementia, including Alzheimer's disease.

## Whole blood, urinary and salivary metabolomes

Metabolic activity *in vivo* is affected by a subject's genetic background, environmental factors, lifestyle, habits, health conditions, etc. During these activities, cells and tissues generate metabolites, a portion of which are excreted into or generated in body fluids (blood, urine, saliva, cerebrospinal fluid, sweat, tears, faeces, etc.) [31]. These metabolites closely reflect organismal health, physiological homeostatic responses, disease states, etc. [32]. Thus, metabolites are good disease indices.

Improvements in analytical instruments and high-volume data processing techniques have accumulated evidence that information on health, ageing and disease status can be interpreted using quantitative data on metabolites in various body fluids. Metabolomics is an efficient tool to evaluate metabolite profiles, to discover and decipher highly integrated, but complex biological processes [32–34]. As blood is constantly circulating throughout the body, it reflects the metabolic status of

the individual. Human blood comprises cellular and non-cellular components, RBCs, white blood cells (WBCs), platelets and plasma. Many studies on human blood have been conducted on plasma or serum, the non-cellular component [35,36], partly due to the difficulty in handling unstable cellular metabolites [37]. However, as about half of the metabolites in whole blood are derived from RBCs, we established a metabolomic approach to analyse whole blood, plasma and RBCs. The whole blood metabolome includes about 130 metabolites that are implicated in energy production, DNA, RNA, protein and lipid metabolism, mitochondrial respiration, redox homeostasis and so on.

The whole blood metabolome provides valuable information about individuals and diseases. Based on the coefficient of variation (CV: SD divided by the mean), metabolites detected by whole-blood metabolomics are classified into two subgroups: less or highly variable (CV < 0.4 or CV > 0.4) [25]. The former comprises many vitally essential metabolites (ATP, glutathione, phospho-sugars, etc.), while the latter include dietary compounds such as caffeine, carnosine, ERG, 4-aminobenzoate and others. ERG is a sulfur-containing N $\alpha$ ,N $\alpha$ ,N $\alpha$ -trimethyl-L-histidine-derived metabolite that is synthesised by various species of bacteria and fungi [17,18,38]. ERG is generated from trimethyl-histidine, known as hercynine, followed by its conversion into S-methyl-ergothioneine (S-methyl-ERG). ERG functions as an antioxidant by scavenging free radicals and chelating transition metals [39]. Selenoneine, the selenium (Se)-bound form of ERG, is enriched in tuna and other fish, exhibiting higher antioxidative capacity [40], whose status in whole blood metabolome is much unknown. Several antioxidants, for example, ERG and acetyl carnosine, are enriched in the RBC fraction [25,26,30].

Alternatively, other biofluids, such as urine and saliva, can easily be acquired noninvasively and are suitable for daily observation. Production and excretion of urine are fine-tuned *in vivo*. As urine contains not only waste products or toxins, but also various nutrients, their filtration and reabsorption are tightly regulated in kidney. Accordingly, urine has been broadly utilised for diagnosis of renal dysfunction in kidney diseases [41]. Saliva is secreted mainly from salivary glands. It also contains metabolites such as sugars, amino acids, antioxidants and high-energy compounds [42,43]. Daily salivary secretion accounts for 0.75–1.5 L, similar to the quantity of urine, suggesting that both urine and saliva can provide useful information on physiological and pathophysiological conditions [44]. In addition to basic metabolites synthesised in the body, various food-derived metabolites are also detected in urine [45] and saliva [46] as well as in blood [25]. Three ERG-related compounds (ERG,

S-methyl-ERG and hercynine) are also detected in samples of human blood, urine or saliva. ERG is abundant in RBCs but is hardly detected in urine and saliva [25,45,46]. It would be worthy to evaluate the levels of salivary and urinary ERG-related compounds in age-related disease, dementia and frailty, in the near future. The abundance of S-methyl-ERG, like ERG, is much lower in urine and saliva than in blood. However, trimethyl-histidine is more abundant in urine than in blood or saliva (T. Teruya and M. Yanagida, unpublished data).

### Starvation-induced upregulation of ergothioneine in yeast cells and human blood

Oxidative stress, damage to large molecules caused by free radicals and reactive oxygen species (ROS), is well known as a major cause of cellular dysfunction and death, and has been implicated in ageing and ageing-relevant diseases [47–49]. Ectopic expression of ‘radical scavengers’ in experimental models exerts beneficial effects during disease states. In addition, genetic manipulation to extend the life span is accompanied with increased antioxidants, for example, glutathione [50,51]. When calorie intake is reduced by 20%–30%, life span is extended by 20% or more in many model organisms, accompanied by reduced oxidative stress [52]. This is partly due to activation of FOXO transcription factor during calorie limitation, which upregulates a group of radical scavenger genes [53]. Alternatively, small-molecule antioxidants containing sulfur or selenium can ameliorate oxidative damage [54]. However, for the purpose of treatment, calorie restriction (CR) or intermittent fasting (IF) is applicable only to human obesity and diabetes. Fasting studies in model organisms would give us alternative hints.

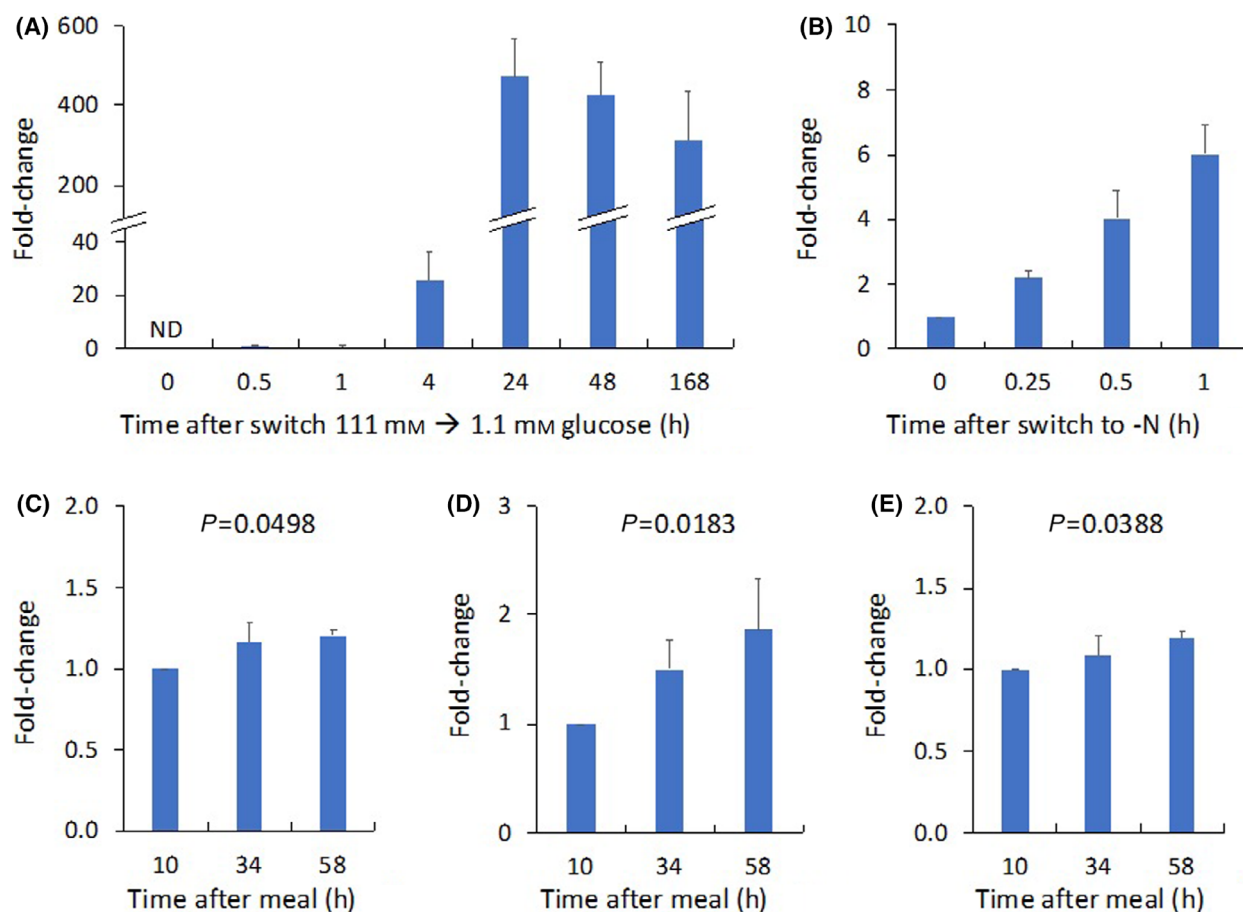
The fission yeast, *Schizosaccharomyces pombe* (*S. pombe*), is an excellent model eukaryote in research for various biological processes, such as proliferation, meiosis, cytoskeletal organisation, metabolism, and chromosome/DNA biology [55–60]. Like human cells, *S. pombe* is equipped with mitochondria, lysosome-like vacuoles, peroxisomes, lipid droplets, endosomes and endoplasmic reticulum, which perform important functions in cellular metabolism [61–63]. While excess calories or glucose accelerates ageing in *S. pombe* [64], responses to limited nutrients are distinct. When nitrogen is absent, cells arrest completely after two rounds of cell division without cell growth [65,66]. The resulting small, round G0 cells remain quiescent and metabolically viable for months, as they possess the ability to reuse intracellular nitrogen. Under glucose

fasting, however, *S. pombe* immediately stops cell division and loses viability within 32 h. These glucose-fasting arrested cells are rod-shaped and arrested at G2 phase of cell cycle with postreplicative DNA [29]. Thus, the study of starvation in fission yeast would give us mechanistic insights in its metabolic mechanism. We performed non-targeted metabolomics in *S. pombe* under starved conditions. We observed that both glucose and nitrogen starvation provoke drastic increases of ERG in *S. pombe* [29,67] (Fig. 1). Ablation of the *egt1*<sup>+</sup> gene, the main ERG biosynthetic enzyme, abolished the boost of ERG in *S. pombe* [68].

Next, we performed non-targeted comprehensive liquid chromatography-mass spectrometry (LC-MS) analysis of whole blood during 58 h fasting by four young, non-obese volunteers [30]. Notably, 44 of 120 metabolites increased 1.5- to 60-fold during this period, suggesting that fasting increases the activity of diverse metabolic pathways. In addition to established fasting markers, such as butyrates, acylcarnitines and branched-chain amino acids, several TCA cycle-related compounds (cis-aconitate, malate, 2-oxoglutarate and succinate) and coenzymes (nicotinamide and pantothenate, a precursor for acetyl-CoA) also increased, reflecting enhanced mitochondrial activity in tissues during fasting. Besides increased metabolites for energy production, previously unappreciated impacts of prolonged fasting were disclosed. Increases of several antioxidants (carnosine, ERG, urate and xanthine), antioxidative marker [ophthalmic acid (OA)] and pentose phosphate pathway (PPP) metabolites were observed. The transporter for ERG, organic cation transporter (OCTN)-1. Interestingly, it was reported that mRNA for OCTN-2, a homologue of OCTN-1, was upregulated in rats under fasting. However, little is known about the activity of human OCTN-1 under fasting [69]. Previously it was observed that ERG levels ( $n = 4$ ) are constant on a daily basis (fig. S1A in Chaleckis *et al.* [25]) (its CV on a daily basis is very low, 0.05). Thus, ERG levels are upregulated not by daily basis, but by longer fasting. As our study was performed in limited numbers ( $n = 4$ ), it would be worthy to perform fasting experiment in larger samples in the future. These findings imply that boost of ERG level may be also adaptive response to fasting stress in both humans and fission yeast, which was not expected previously, implicating that ERG may also exert beneficial effect on ageing-relevant events (Fig. 1).

### Ergothioneine declines in frailty and dementia, but not in sarcopenia

Various ageing-related diseases share some clinical features, such as frailty, dementia and sarcopenia, with

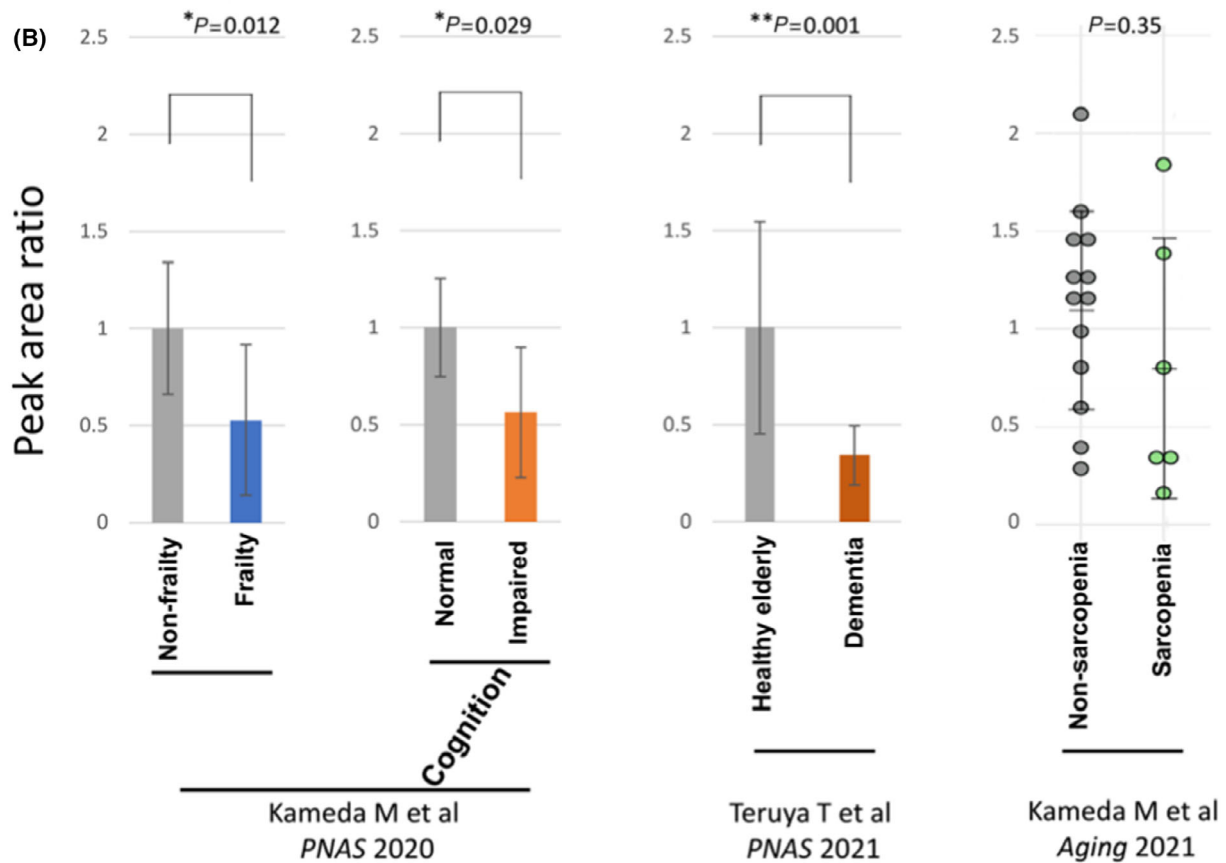
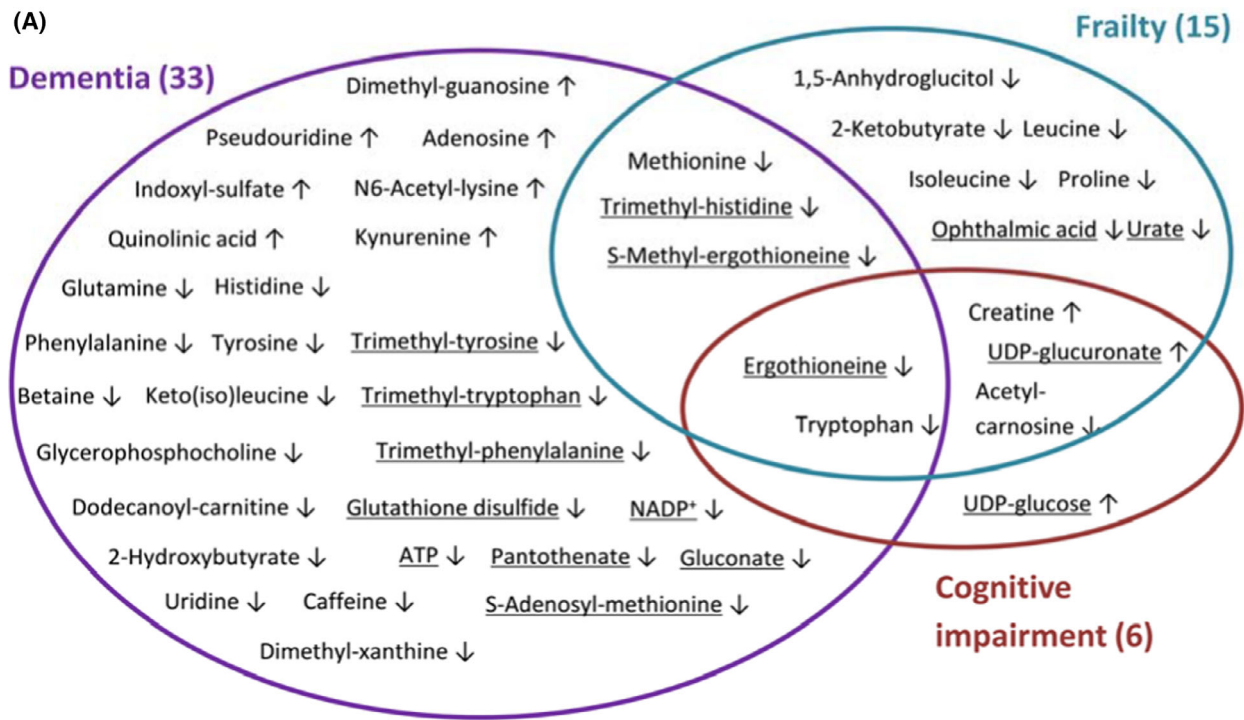


**Fig. 1.** Increased ergothioneine yeast cells upon starvation. Time course of changes in ergothioneine levels, evaluated by LC-MS during starvation in *S. pombe* (A, B) and human blood (C–E). *S. pombe* was grown in medium with low glucose (1.1 mM) (A) or without nitrogen (–N) (B). (C–E) Four human volunteers fasted for 58 h. Ergothioneine was detected in whole blood (C), plasma (D) and RBCs (E) under prolonged fasting. *P*-values were calculated with the Friedman test.

increased needs for life support [70,71]. We performed whole blood metabolomics in relation to these three diseases to clarify the metabolic basis of their pathophysiology [26–28].

Non-targeted metabolomics of 19 elderly subjects (average  $84.2 \pm 6.9$  years;  $80.5 \pm 4.7$  years for frail subjects vs.  $88.2 \pm 6.8$  years for non-frail subjects) identified 15 frailty markers. Strikingly, these 15 markers include seven metabolites related to antioxidation that decrease in frailty (acetyl-carnosine, ERG, S-methyl-ERG, trimethyl-histidine, OA, 2-ketobutyrate and urate) and three amino acids with radical scavenger properties (methionine, proline and tryptophan; Fig. 2). Recent longitudinal studies have also suggested the involvement of antioxidants in frailty [72]. Three ERG-relevant metabolites (ERG, S-methyl-ERG and trimethyl-histidine) decline and the metabolic or uptake pathway of ERG is much affected in frailty (Fig. 2).

As frail elderly people commonly manifest complex clinical symptoms, including cognitive dysfunction, hypomobility and impaired daily activity, we also evaluated cognitive function using the Japanese version of the Montreal Cognitive Assessment (MoCA-J) and sarcopenia in these same populations [26,27]. Six cognitive markers were identified, five of which are also frailty markers (acetyl-carnosine, ERG, tryptophan, creatine and UDP-glucuronate) (Fig. 2). On the other hand, 22 sarcopenia markers were identified in blood. Most of the metabolites that decreased in blood of sarcopenia are uraemic compounds that increase in kidney dysfunction, including TCA cycle, urea cycle, nitrogen and methylated metabolites. Thus, sarcopenia markers imply a close link between muscle and kidney function. However, little is known on sarcopenia markers in urine. Notably, the 22 sarcopenia markers are distinct from the 15 frailty markers in the blood of same





**Fig. 2.** Ergothioneine in frailty and dementia. (A) Summary of biomarkers identified with whole blood metabolomics in frailty, dementia and cognitive impairment. Arrows indicate increased or decreased metabolites. Note that ergothioneine is a biomarker for all three disease states. (B) The ergothioneine level in whole blood is compared in the indicated disease states. The relative ratios of the peak areas are shown, after normalisation against controls. In the same populations, frailty, cognitive impairment and sarcopenia were diagnosed using criteria in the EFS (Edmonton Frailty Scale), the MoCA-J and the AWGS 2014 (Asian Working Group for Sarcopenia) [26,27]. Dementia was diagnosed according to DSM-IV [28].

subjects, although sarcopenia overlaps clinically with physical frailty. Thus, ERG in blood of sarcopenia is comparable to that in controls. It is possible that ERG transporter OCTN-1 might be affected in these specific diseased states, whose significance in the pathogenesis is still unknown. Further study is required to acquire its mechanistic insights in the future.

Moreover, we quantified metabolites in whole blood of dementia, the majority of which were diagnosed as Alzheimer patients, using non-targeted LC-MS [28]. Thirty-three dementia markers are classified into five groups. Seven metabolites, such as possible neurotoxins increased [73,74], including quinolinic acid, kynurenine and indoxyl-sulfate, while the remaining 26 dementia markers decreased. Six metabolites in this subgroup, enriched in RBCs, contain trimethylated ammonium moieties, ERG derivatives (ERG, S-Methyl-ERG and trimethyl-histidine), trimethyl-tryptophan, trimethyl-tyrosine and trimethyl-phenylalanine (Fig. 2). Pearson's correlation analysis showed that ERG, S-methyl-ERG and trimethyl-histidine were inversely correlated with indoxyl-sulfate, quinolinic acid and kynurenine ( $-0.66 < r < -0.42$ ). Cheah *et al.* [75] showed that in whole blood of subjects with Mild Cognitive Impairment, ERG was reduced by 0.65-fold compared to its levels in healthy individuals. ERG in frailty was 0.53-fold lower than in non-frail subjects, while that in dementia was 0.41-fold lower than in healthy elderly. The reduction of S-methyl-ERG in frailty or dementia was more remarkable 0.48 and 0.20 fold respectively, compared to healthy counterparts, while changes of trimethyl-histidine in frailty and dementia were comparable to those of ERG. A recent large-scale study of metabolomics ( $n = 496$ ) also identified ERG as a dementia marker [76]. We also investigated the correlation between ERG levels in whole blood and the degree of frailty (Edmonton Frailty Scale, EFS score) or cognitive function (mini mental state examination, MMSE score) in dementia patients. The EFS showed a moderate correlation with ERG ( $r = -0.45$ ,  $P = 0.053$ ) and S-methyl-ERG ( $r = -0.51$ ,  $P = 0.026$ ) [26]. On the other hand, the MMSE showed a significant correlation between ERG ( $r = 0.64$ ,  $P = 0.018$ ) and S-methyl-ERG ( $r = 0.62$ ,  $P = 0.024$ ; calculated based on the raw data [28]). The decrease of ERG, S-methyl-ERG and

hercynine in a concerted manner suggests that the intake of ERG might be affected in frailty or dementia. Collectively, ERG, a potent antioxidant, is significantly decreased in dementia, cognitive impairment and frailty, but not in sarcopenia (Fig. 2).

## Conclusions and perspectives

In addition to whole blood metabolomics, we analysed both the urine and saliva metabolomics to comprehensively understand ageing-relevant diseases. Our studies of these three types of biofluids showed that ERG levels have no significant age-related differences. Previous studies have suggested that ERG exerts antioxidative and anti-inflammatory effects that are involved in several human diseases, for example, rheumatoid arthritis. We noticed that ERG is increased in human blood and yeast under glucose starvation, implying a protective role for ERG under CR. As fission yeast can produce ERG, it is a useful model organism for investigating the biochemical function of ERG. Although frailty comprises both cognitive impairment and hypomobility, ERG is identified as a biomarker for frailty and dementia, but not for sarcopenia, at least according to our whole blood metabolomics. These findings suggest the significance of ERG in ageing-related diseases: as oxidative damage accelerates organismal ageing and ageing-relevant disorders [77], it is possible that a decline in a radical scavenger such as ERG would promote the progression of frailty, dementia and ageing-related events. Several reports support the notion that ERG supplementation alleviates cognitive impairment and tissue oxidative damage in experimental animal models [78]. Thus, ERG treatment is a potential therapeutic approach for frailty and dementia.

## References

- 1 World Health Organization. World report on ageing and health 2015. WHO Press; 2015: p. 1–246.
- 2 Nagai Y, Metter EJ, Earley CJ, Kemper MK, Becker LC, Lakatta EG, *et al.* Increased carotid artery intimal-medial thickness in asymptomatic older subjects with exercise-induced myocardial ischemia. *Circulation*. 1998;**98**:1504–9.

- 3 Lee C, Dobson AJ, Brown WJ, Bryson L, Byles J, Warner-Smith P, et al. Cohort profile: the Australian longitudinal study on women's health. *Int J Epidemiol*. 2005;**34**:987–91.
- 4 Tantirat P, Suphanchaimat R, Rattanathumsakul T, Noree T. Projection of the number of elderly in different health states in thailand in the next ten years, 2020–2030. *Int J Environ Res Public Health*. 2020;**17**:8703.
- 5 Cummings JL, Mega M, Gray K, Rosenberg-Thompson S, Carusi DA, Gornbein J. The neuropsychiatric inventory: comprehensive assessment of psychopathology in dementia. *Neurology*. 1994;**44**:2308–14.
- 6 Arvanitakis Z, Shah RC, Bennett DA. Diagnosis and management of dementia: review. *JAMA*. 2019;**322**:1589–99.
- 7 Cerejeira J, Lagarto L, Mukaetova-Ladinska EB. Behavioral and psychological symptoms of dementia. *Front Neurol*. 2012;**3**:73.
- 8 Fried LP, Tangen CM, Walston J, Newman AB, Hirsch C, Gottdiener J, et al. Frailty in older adults: evidence for a phenotype. *J Gerontol A Biol Sci Med Sci*. 2001;**56**:M146–56.
- 9 Rosenberg IH. Sarcopenia: origins and clinical relevance. *J Nutr*. 1997;**127**:990S–1.
- 10 Rockwood K, Stadnyk K, MacKnight C, McDowell I, Hebert R, Hogan DB. A brief clinical instrument to classify frailty in elderly people. *Lancet*. 1999;**353**:205–6.
- 11 Dent E, Lien C, Lim WS, Wong WC, Wong CH, Ng TP, et al. The Asia-Pacific Clinical Practice Guidelines for the Management of Frailty. *J Am Med Dir Assoc*. 2017;**18**:564–75.
- 12 Chen LK, Liu LK, Woo J, Assantachai P, Auyeung TW, Bahyah KS, et al. Sarcopenia in Asia: consensus report of the Asian Working Group for Sarcopenia. *J Am Med Dir Assoc*. 2014;**15**:95–101.
- 13 Melville DB. Ergothioneine. *Vitam Horm*. 1959;**17**:155–204.
- 14 Cheah IK, Halliwell B. Ergothioneine; antioxidant potential, physiological function and role in disease. *Biochim Biophys Acta*. 2012;**1822**:784–93.
- 15 Genghof DS. Biosynthesis of ergothioneine and hercynine by fungi and Actinomycetales. *J Bacteriol*. 1970;**103**:475–8.
- 16 Ames BN. Prolonging healthy aging: longevity vitamins and proteins. *Proc Natl Acad Sci USA*. 2018;**115**:10836–44.
- 17 Genghof DS, Inamine E, Kovalenko V, Melville DB. Ergothioneine in microorganisms. *J Biol Chem*. 1956;**223**:9–17.
- 18 Genghof DS, Vandamme O. Biosynthesis of ergothioneine and hercynine by mycobacteria. *J Bacteriol*. 1964;**87**:852–62.
- 19 Borodina I, Kenny LC, McCarthy CM, Paramasivan K, Pretorius E, Roberts TJ, et al. The biology of ergothioneine, an antioxidant nutraceutical. *Nutr Res Rev*. 2020;**33**:190–217.
- 20 Franzoni F, Colognato R, Galetta F, Laurenza I, Barsotti M, Di Stefano R, et al. An in vitro study on the free radical scavenging capacity of ergothioneine: comparison with reduced glutathione, uric acid and trolox. *Biomed Pharmacother*. 2006;**60**:453–7.
- 21 Tokuhito S, Yamada R, Chang X, Suzuki A, Kochi Y, Sawada T, et al. An intronic SNP in a RUNX1 binding site of SLC22A4, encoding an organic cation transporter, is associated with rheumatoid arthritis. *Nat Genet*. 2003;**35**:341–8.
- 22 Tang RMY, Cheah IK, Yew TSK, Halliwell B. Distribution and accumulation of dietary ergothioneine and its metabolites in mouse tissues. *Sci Rep*. 2018;**8**:1601.
- 23 Cheah IK, Halliwell B. Ergothioneine, recent developments. *Redox Biol*. 2021;**42**:101868.
- 24 Chaleckis R, Ebe M, Pluskal T, Murakami I, Kondoh H, Yanagida M. Unexpected similarities between the Schizosaccharomyces and human blood metabolomes, and novel human metabolites. *Mol Biosyst*. 2014;**10**:2538–51.
- 25 Chaleckis R, Murakami I, Takada J, Kondoh H, Yanagida M. Individual variability in human blood metabolites identifies age-related differences. *Proc Natl Acad Sci USA*. 2016;**113**:4252–9.
- 26 Kameda M, Teruya T, Yanagida M, Kondoh H. Frailty markers comprise blood metabolites involved in antioxidation, cognition, and mobility. *Proc Natl Acad Sci USA*. 2020;**117**:9483–9.
- 27 Kameda M, Teruya T, Yanagida M, Kondoh H. Reduced uremic metabolites are prominent feature of sarcopenia, distinct from antioxidative markers for frailty. *Aging*. 2021;**13**:20915–34.
- 28 Teruya T, Chen YJ, Kondoh H, Fukui Y, Yanagida M. Whole-blood metabolomics of dementia patients reveal classes of disease-linked metabolites. *Proc Natl Acad Sci USA*. 2021;**118**:e2022857118.
- 29 Pluskal T, Hayashi T, Saitoh S, Fujisawa A, Yanagida M. Specific biomarkers for stochastic division patterns and starvation-induced quiescence under limited glucose levels in fission yeast. *FEBS J*. 2011;**278**:1299–315.
- 30 Teruya T, Chaleckis R, Takada J, Yanagida M, Kondoh H. Diverse metabolic reactions activated during 58-hr fasting are revealed by non-targeted metabolomic analysis of human blood. *Sci Rep*. 2019;**9**:854.
- 31 Zhang A, Sun H, Wang P, Han Y, Wang X. Recent and potential developments of biofluid analyses in metabolomics. *J Proteomics*. 2012;**75**:1079–88.
- 32 van der Greef J, van Wietmarschen H, van Ommen B, Verheij E. Looking back into the future: 30 years of metabolomics at TNO. *Mass Spectrom Rev*. 2013;**32**:399–415.

- 33 Suhre K, Shin SY, Petersen AK, Mohny RP, Meredith D, Wägele B, et al. Human metabolic individuality in biomedical and pharmaceutical research. *Nature*. 2011;**477**:54–60.
- 34 Patti GJ, Yanes O, Siuzdak G. Innovation: metabolomics: the apogee of the omics trilogy. *Nat Rev Mol Cell Biol*. 2012;**13**:263–9.
- 35 Lawton KA, Berger A, Mitchell M, Milgram KE, Evans AM, Guo L, et al. Analysis of the adult human plasma metabolome. *Pharmacogenomics*. 2008;**9**:383–97.
- 36 Nishino T, Yachie-Kinoshita A, Hirayama A, Soga T, Suematsu M, Tomita M. In silico modeling and metabolome analysis of long-stored erythrocytes to improve blood storage methods. *J Biotechnol*. 2009;**144**:212–23.
- 37 Gil A, Siegel D, Permentier H, Reijngoud DJ, Dekker F, Bischoff R. Stability of energy metabolites—an often overlooked issue in metabolomics studies: a review. *Electrophoresis*. 2015;**36**:2156–69.
- 38 Dubost N, Ou B, Beelman R. Quantification of polyphenols and ergothioneine in cultivated mushrooms and correlation to total antioxidant capacity. *Food Chem*. 2007;**105**:727–35.
- 39 Motohashi N, Mori I, Sugiura Y, Tanaka H. Metal complexes of ergothioneine. *Chem Pharm Bull*. 1974;**22**:654–7.
- 40 Yamashita Y, Yabu T, Yamashita M. Discovery of the strong antioxidant selenoneine in tuna and selenium redox metabolism. *World J Biol Chem*. 2010;**1**:144–50.
- 41 Tesch GH. Review: serum and urine biomarkers of kidney disease: a pathophysiological perspective. *Nephrology*. 2010;**15**:609–16.
- 42 Owen-Smith B, Quiney J, Read J. Salivary urate in gout, exercise, and diurnal variation. *Lancet*. 1998;**351**:1932.
- 43 Takeda I, Stretch C, Barnaby P, Bhatnager K, Rankin K, Fu H, et al. Understanding the human salivary metabolome. *NMR Biomed*. 2009;**22**:577–84.
- 44 Sanchez-Pablo MA, Gonzalez-Garcia V, del Castillo-Rueda A. Study of total stimulated saliva flow and hyperpigmentation in the oral mucosa of patients diagnosed with hereditary hemochromatosis. Series of 25 cases. *Med Oral Patol Oral Cir Bucal*. 2012;**17**:e45–9.
- 45 Teruya T, Goga H, Yanagida M. Aging markers in human urine: a comprehensive, non-targeted LC-MS study. *FASEB Bioadv*. 2020;**2**:720–33.
- 46 Teruya T, Goga H, Yanagida M. Human age-declined saliva metabolic markers determined by LC-MS. *Sci Rep*. 2021;**11**:18135.
- 47 Harman D. Aging: a theory based on free radical and radiation chemistry. *J Gerontol*. 1956;**11**:298–300.
- 48 Sohal RS, Weindruch R. Oxidative stress, caloric restriction, and aging. *Science*. 1996;**273**:59–63.
- 49 Liguori I, Russo G, Curcio F, Bulli G, Aran L, Della-Morte D, et al. Oxidative stress, aging, and diseases. *Clin Interv Aging*. 2018;**13**:757–72.
- 50 Lopez-Torres M, Perez-Campo R, Rojas C, Cadenas S, Barja G. Maximum life span in vertebrates: relationship with liver antioxidant enzymes, glutathione system, ascorbate, urate, sensitivity to peroxidation, true malondialdehyde, in vivo H<sub>2</sub>O<sub>2</sub>, and basal and maximum aerobic capacity. *Mech Ageing Dev*. 1993;**70**:177–99.
- 51 Gusarov I, Shamovsky I, Pani B, Gautier L, Eremina S, Katkova-Zhukotskaya O, et al. Dietary thiols accelerate aging of *C. elegans*. *Nat Commun*. 2021;**12**:4336.
- 52 Pamplona R, Barja G. Mitochondrial oxidative stress, aging and caloric restriction: the protein and methionine connection. *Biochim Biophys Acta*. 2006;**1757**:496–508.
- 53 Lehtinen MK, Yuan Z, Boag PR, Yang Y, Villén J, Becker EB, et al. A conserved MST-FOXO signaling pathway mediates oxidative-stress responses and extends life span. *Cell*. 2006;**125**:987–1001.
- 54 Battin EE, Brumaghim JL. Antioxidant activity of sulfur and selenium: a review of reactive oxygen species scavenging, glutathione peroxidase, and metal-binding antioxidant mechanisms. *Cell Biochem Biophys*. 2009;**55**:1–23.
- 55 Nurse P. Genetic control of cell size at cell division in yeast. *Nature*. 1975;**256**:547–51.
- 56 Yamamoto M. Regulation of meiosis in fission yeast. *Cell Struct Funct*. 1996;**21**:431–6.
- 57 Kovar DR, Sirotkin V, Lord M. Three's company: the fission yeast actin cytoskeleton. *Trends Cell Biol*. 2011;**21**:177–87.
- 58 Yanagida M. The role of model organisms in the history of mitosis research. *Cold Spring Harb Perspect Biol*. 2014;**6**:a015768.
- 59 Masuda F, Ishii M, Mori A, Uehara L, Yanagida M, Takeda K, et al. Glucose restriction induces transient G2 cell cycle arrest extending cellular chronological lifespan. *Sci Rep*. 2016;**6**:19629.
- 60 Hoffman CS, Wood V, Fantes PA. An ancient yeast for young geneticists: a primer on the *Schizosaccharomyces pombe* model system. *Genetics*. 2015;**201**:403–23.
- 61 Schafer B. Genetic conservation versus variability in mitochondria: the architecture of the mitochondrial genome in the petite-negative yeast *Schizosaccharomyces pombe*. *Curr Genet*. 2003;**43**:311–26.
- 62 Takegawa K, Iwaki T, Fujita Y, Morita T, Hosomi A, Tanaka N. Vesicle-mediated protein transport pathways to the vacuole in *Schizosaccharomyces pombe*. *Cell Struct Funct*. 2003;**28**:399–417.



- 63 Jourdain I, Sontam D, Johnson C, Dillies C, Hyams JS. Dynamin-dependent biogenesis, cell cycle regulation and mitochondrial association of peroxisomes in fission yeast. *Traffic*. 2008;**9**:353–65.
- 64 Dutcher SK, Roux AE, Leroux A, Alaamery MA, Hoffman CS, Chartrand P, et al. Pro-aging effects of glucose signaling through a G protein-coupled glucose receptor in fission yeast. *PLoS Genet*. 2009;**5**:e1000408.
- 65 Costello G, Rodgers L, Beach D. Fission yeast enters the stationary phase G0 state from either mitotic G1 or G2. *Curr Genet*. 1986;**11**:119–25.
- 66 Su SS, Tanaka Y, Samejima I, Tanaka K, Yanagida M. A nitrogen starvation-induced dormant G0 state in fission yeast: the establishment from uncommitted G1 state and its delay for return to proliferation. *J Cell Sci*. 1996;**109**:1347–57.
- 67 Sajiki K, Pluskal T, Shimanuki M, Yanagida M. Metabolomic analysis of fission yeast at the onset of nitrogen starvation. *Metabolites*. 2013;**3**:1118–29.
- 68 Pluskal T, Ueno M, Yanagida M. Genetic and metabolomic dissection of the ergothioneine and selenoneine biosynthetic pathway in the fission yeast, *S. pombe*, and construction of an overproduction system. *PLoS One*. 2014;**9**:e97774.
- 69 Luci S, Hirche F, Eder K. Fasting and caloric restriction increases mRNA concentrations of novel organic cation transporter-2 and carnitine concentrations in rat tissues. *Ann Nutr Metab*. 2008;**52**:58–67.
- 70 Dent E, Kowal P, Hoogendijk EO. Frailty measurement in research and clinical practice: a review. *Eur J Intern Med*. 2016;**31**:3–10.
- 71 Tamura Y, Ishikawa J, Fujiwara Y, Tanaka M, Kanazawa N, Chiba Y, et al. Prevalence of frailty, cognitive impairment, and sarcopenia in outpatients with cardiometabolic disease in a frailty clinic. *BMC Geriatr*. 2018;**18**:264.
- 72 Marron MM, Wendell SG, Boudreau RM, Clish CB, Santanasto AJ, Tseng GC, et al. Metabolites associated with walking ability among the oldest old from the CHS all stars study. *J Gerontol A Biol Sci Med Sci*. 2020;**75**:2371–8.
- 73 Stone TW, Mackay GM, Rorrest CM, Clark CJ, Darlington LG. Tryptophan metabolites and brain disorders. *Clin Chem Lab Med*. 2003;**41**:852–9.
- 74 Adesso S, Magnus T, Cuzzocrea S, Campolo M, Rissiek B, Paciello O, et al. Indoxyl sulfate affects glial function increasing oxidative stress and neuroinflammation in chronic kidney disease: interaction between astrocytes and microglia. *Front Pharmacol*. 2017;**8**:370.
- 75 Cheah IK, Feng L, Tang RMY, Lim KHC, Halliwell B. Ergothioneine levels in an elderly population decrease with age and incidence of cognitive decline; a risk factor for neurodegeneration? *Biochem Biophys Res Commun*. 2016;**478**:162–7.
- 76 Wu LY, Cheah IK, Chong JR, Chai YL, Tan JY, Hilal S, et al. Low plasma ergothioneine levels are associated with neurodegeneration and cerebrovascular disease in dementia. *Free Radic Biol Med*. 2021;**177**:201–11.
- 77 Harman D. Free radical theory of aging: an update: increasing the functional life span. *Ann N Y Acad Sci*. 2006;**1067**:10–21.
- 78 Halliwell B, Cheah IK, Tang RMY. Ergothioneine - a diet-derived antioxidant with therapeutic potential. *FEBS Lett*. 2018;**592**:3357–66.