



PARP inhibitors for small cell lung cancer and their potential for integration into current treatment approaches

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Abstract: Small cell lung cancer (SCLC) is a very aggressive, highly lethal, neuroendocrine tumor that constitutes 15% of all lung cancer cases. It is characterized by its rapid disease progression and high relapse rate leading to poor survival for diagnosed patients. Recently, poly (ADP-ribose) polymerase inhibitors (PARPi) have emerged as a novel therapeutic strategy for SCLC. Preclinical studies have demonstrated that PARPi possesses cytotoxic activity as a single-agent and in combination with other anti-cancer agents. Predictive biomarkers of response to PARPi, such as *SLFN11*, have also been described in SCLC. This review aims to summarize the recent preclinical investigations and the relevant clinical trials that evaluate PARPi in SCLC. Here, we highlight the potential role of PARPi in a biomarker-selected manner and in combination with chemotherapy, targeted agents, radiotherapy and immunotherapy.

Keywords: Small cell lung cancer (SCLC); poly (ADP-ribose) polymerase inhibitors (PARPi); chemotherapy; radiation sensitizer; immunotherapy

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Introduction

Lung cancer has remained the most common cancer worldwide since 1985, with approximately 1.8 million new cases diagnosed each year (1,2). Among the major lung cancer subtypes, small cell lung cancer (SCLC) represents the sixth leading cause of cancer-related mortality, accounting for 13–15% of all lung cases (3,4). Clinically, SCLC is considered an aggressive and lethal high-grade neuroendocrine malignancy that is pathologically, molecularly, and biologically very distinct from other forms of lung cancer. Almost all SCLC cases have homozygous loss of *RBI*, which encodes the key regulator of the G1-S cell cycle checkpoint, and *TP53*, a gene critical for multiple

DNA damage response (DDR) pathways (5-11). This can, in part, explain the initial responsiveness of SCLC to various DNA damaging agents, such as those that induce covalent DNA adducts and crosslinks [cisplatin, carboplatin, temozolomide (TMZ)], or those that produce single-strand/double-strand DNA breaks (ionizing radiation, etoposide, topotecan, irinotecan).

The standard of care for first-line treatment of SCLC consists of a platinum-based doublet chemotherapy regimen for all patients, concurrent with radiotherapy (RT) for those with limited-stage disease. Despite a high (70–80%) initial response rate to these first-line regimens, the 5-year overall survival (OS) rates for patients diagnosed with SCLC is a dismal 6.5% (12). Until 2019, with the exception

of the FDA approval of atezolizumab with chemotherapy for extensive-stage SCLC (ES-SCLC) patients, the general treatment paradigm has remained unchanged for the previous several decades (13,14). Therefore, novel therapeutic interventions are needed and are an active area of research (15-17).

One such novel therapeutic are inhibitors of poly-(ADP)-ribose polymerase (PARP), which have demonstrated efficacy against SCLC in preclinical and clinical data over the past several years. Poly (ADP-ribose) polymerase inhibitors (PARPi) have the potential to enhance cytotoxic response to chemotherapy, radiotherapy, and immunotherapy in SCLC. This review will highlight the advances made in these areas.

Current application of PARPi in oncology

PARP is a family of proteins that orchestrate various cellular processes and have important roles in DNA repair and genome integrity. PARP1 activates base excision repair (BER) in response to DNA single-strand breaks (SSBs) where PARP1 binds to SSBs and facilitates the recruitment of DNA repair proteins. When PARP1 function is impaired, the BER process is halted and double-strand breaks (DSBs) develop due to a destabilized replication fork (18). As a result, malignancies deficient in the DSB repair pathway of homologous recombination (HR) are vulnerable to PARP inhibition. PARPi were first demonstrated to have efficacy in ovarian cancers with *BRCA1/2* mutations—which are deficient in HR (19). Subsequently, PARPi clinical efficacy has extended to other histologies harboring *BRCA1/2* mutations (19-27) with most PARPi carrying FDA approval for treating *BRCA1/2*-mutated ovarian and breast cancers (Table 1) (30-37).

The various PARPi generally have similar activity in their degree of polymerase inhibition. However, the PARPi differ in their ability to poison and trap PARP to the DNA SSB lesions which consequently devolve into cytotoxic DSBs upon DNA replication (38). Talazoparib has been demonstrated to be the most potent PARP trapper followed by niraparib, then olaparib and rucaparib, with veliparib being the least potent trapper (28,29). These differences between their potency in PARP trapping may inform their observed efficacy and side effect profiles.

Recently, studies have examined the utility of PARPi beyond *BRCA1/2*-mutant tumors (39). Several groups have demonstrated that large chromosomal structural alterations, characteristic of these *BRCA1/2*-mutant cancers, can be

quantitated by three correlated HR deficiency (HRD) metrics: loss of heterozygosity (LOH), large-scale state transition (LST), and telomeric allelic imbalance (N_{TAD}) (40-43). These HRD scores correlate with sensitivity to platinum agents of sporadic triple-negative breast and ovarian cancers (40-42,44,45). This suggests that PARPi may have therapeutic benefits in any malignancy harboring HRD independent of the canonical *BRCA1/2* mutations. As *BRCA* mutations are rare ($\leq 2\%$) in SCLC, this concept of non-*BRCA* dependent PARPi sensitivity lay the foundation for subsequent investigations (10,11).

PARP as a therapeutic target in SCLC

Pre-clinical evidence and studies

Rationale for PARPi in SCLC

In 2012, Byers *et al.* conducted a landmark study where reverse phase protein array (RPPA) for proteomic analysis of 34 SCLC and 74 non-SCLC (NSCLC) cell lines identified potential targets unique to SCLC. The analysis revealed that SCLC cell lines had high PARP1 protein expression relative to NSCLC. In addition, SCLC patient tumors demonstrated high PARP1 protein expression when compared to other neuroendocrine tumors and NSCLC. *In vitro* cell line drug response studies with olaparib and rucaparib confirmed that most SCLC cell lines tested were highly sensitive to treatment with PARPi in contrast to NSCLC cell lines (46). However, PARPi drug sensitivity was not universal for all SCLC cell lines tested. Therefore, interest towards identifying molecular mechanisms of sensitivity as potential predictive biomarkers increased.

Potential predictive biomarkers of PARPi response

Schlafen family member 11 (SLFN11) was recently identified as a putative predictive biomarker for SCLC sensitivity to PARPi. Multiple independent groups demonstrated that high *SLFN11* gene or protein expression levels positively correlate with increased PARPi treatment sensitivity (47-51).

Polley *et al.* examined 63 SCLC cell-lines in response to treatment with multiple PARPi (talazoparib, olaparib, niraparib, rucaparib, AZD-2461) in their screen of 103 FDA approved oncology drugs and 423 investigational agents (47). Results indicated that increased gene expression of *SLFN11* correlated with decreased IC_{50} (i.e., inhibitory concentration producing 50% growth inhibition) values to all tested PARPi ($R=-0.42$). Of note, expression levels of neither *PARP1* nor *PARP2* had any predictive value (47).

Table 1 FDA approved PARP inhibitors

Drug	Date of FDA Approval	Population	Indication	Dosing	References	Specificity	Ki	Relative trapping capacity (28,29)
Olaparib	2017	gBRCAm advanced ovarian cancer	Received three or more prior lines of chemotherapy	300 mg BID	(30)	PARP1, PARP2	5 nM, 1 nM	+++
	2018	Recurrent epithelial ovarian, fallopian tube, primary peritoneal cancer	Tumors must have CR/PR to platinum-based chemotherapy	300 mg BID	(31)			
	2018	gBRCAm HER2-negative metastatic breast cancer	Previous treatment with chemotherapy in the neoadjuvant, adjuvant, or metastatic setting	300 mg BID	(32)			
	2019	gBRCAm metastatic pancreatic adenocarcinoma	No disease progression after 16 weeks of a first-line platinum-based chemotherapy regimen	300 mg BID	(30)			
Rucaparib	2018	Recurrent epithelial ovarian, fallopian tube, primary peritoneal cancer	Tumors must have CR/PR to platinum-based chemotherapy	600 mg BID	(33)	PARP1	4 nM	+++
Niraparib	2017	Recurrent epithelial ovarian, fallopian tube, primary peritoneal cancer	Tumors must have CR/PR to platinum-based chemotherapy	300 mg QD	(34)	PARP1, PARP2	3.2 nM, 4 nM	++++
	2019	Advanced ovarian, fallopian tube, primary peritoneal cancer with HRD-positive status	Previous treatment with three or more chemotherapy regimens	300 mg QD	(35)			
Talazoparib	2018	gBRCAm HER2-negative breast cancer, locally advanced or metastatic	Patient selection for therapy is based on FDA-approved companion diagnostic for talazoparib	1 mg QD	(36)	PARP1, PARP2	1.2 nM, 0.9 nM	+++++

Data as of February 2020. PARP, poly (ADP-ribose) polymerase; gBRCAm, germline BRCA-mutated; CR, complete response; PR, partial response; BID, twice daily; QD, once daily; Ki, catalytic inhibitory constant; HRD, homologous recombination deficiency; HER2, human epidermal growth factor receptor 2.

Similarly, a significant correlation between *SLFN11* expression levels and response to talazoparib was identified in a study by Murai *et al.* (48). Importantly, the relationship between *SLFN11* expression and PARPi sensitivity was determined to be causal, where CRISPR-mediated genetic knockout of *SLFN11* in 4 cell lines with high *SLFN11* (prostate DU145, leukemia CCRF-CEM and MOLT4, and Ewing's sarcoma EW8) resulted in resistance to both talazoparib and olaparib relative to their parental cell lines. Further confirmation was achieved when exogenous expression of *SLFN11* was induced in leukemia K562 cells (that have low *SLFN11* endogenous transcript) and resulted in hypersensitivity to both talazoparib and olaparib. Murai *et al.* concluded that *SLFN11* is a dominant determinant of PARPi sensitivity in these cancer cells (48).

Specific to SCLC, Lok *et al.* observed that *SLFN11* gene and protein expression levels correlated to PARPi response in SCLC cell lines and patient-derived xenograft (PDX) mouse models (49). The study showed that high expression of *SLFN11* positively correlated with increased sensitivity to various PARPi (i.e., olaparib, rucaparib, veliparib, and talazoparib) in cell line datasets. Functionally, genetic knockdown and knockout of *SLFN11* in SCLC cell lines conferred resistance to PARPi. Immunohistochemical (IHC) staining of SLFN11 confirmed these findings in multiple PDX models treated with talazoparib (49). These studies suggested that *SLFN11* could be used as a predictive biomarker of response to PARPi monotherapy in SCLC.

Stewart *et al.* reported that there may be two potential biomarkers (SLFN11 and ATM) with predictive capability of PARPi response in SCLC. In this study, 170 proteins quantified by RPPA were investigated as potential predictive biomarkers in response to single-agent treatment with talazoparib. Results revealed that low ATM and high SLFN11 protein expression were significantly associated with treatment response in SCLC PDX models. While high CHK1, IGF1R beta, and IRS1 protein levels correlated with resistance. These findings were validated at the mRNA level which showed the strongest association between talazoparib response and high *SLFN11*, low *ATM*, and low *CHEK1* expression in SCLC PDX models (50).

Multiple laboratories have investigated the mechanisms by which SLFN11 may confer PARPi sensitization (52–54). Mu *et al.* reported the interaction of SLFN11 with replication protein A (RPA) led to the suppression of HR. SLFN11 destabilized RPA1-ssDNA complexes necessary for efficient downstream DNA repair by HR (52). Murai *et al.* observed that SLFN11 was recruited to DNA

damage sites where it binds with RPA and subsequently to the minichromosome maintenance protein complex (MCM) DNA helicase that is essential for DNA replication. Although SLFN11 did not directly engage in replication initiation, SLFN11 unwound heterochromatin, blocked the progression of the replication fork, and ultimately hindered the DNA repair process (53). Li *et al.* reported indirect inhibition of ATM and ATR protein synthesis by SLFN11. The absence of these DNA repair proteins promoted sensitization to the effects of DNA damaging agents (54). Continued investigation into the contribution of these mechanisms to PARPi sensitivity would be of considerable basic and translational interest as these therapeutics are being actively investigated in multiple clinical trials of several histologies, including SCLC.

Clinical evidence and studies

Single-agent PARPi trials

In a phase I trial (NCT01286987), 113 patients with recurrent breast, ovarian, prostate, and pancreatic cancers containing deficiencies in DNA repair pathways were recruited, in addition to 23 relapsed SCLC patients. This study was designed to assess the safety and efficacy of talazoparib monotherapy. The dose escalation cohort of this study evaluated 39 patients, none of which were SCLC, and determined the maximum tolerated dose (MTD) to be 1.0 mg of talazoparib daily. The subsequent cohort of this study assessed 71 patients, including all 23 SCLC patients, who were administered talazoparib 1.0 mg daily. Talazoparib demonstrated the highest bioavailability and anti-tumor activity in patients presenting with *BRC A* mutations [objective response rate (ORR) >40%] (55). Of the 23 SCLC patients, 2 patients had a partial response (ORR =9%) that lasted between 3–4 months. Both patients previously had an objective response to prior platinum therapy and were platinum-free for up to 6 months. The median progression-free survival (PFS) of these patients was 11.1 weeks [95% confidence interval (CI): 4.3–13.0 weeks]. An additional 4 SCLC patients had stable disease (SD) that lasted at least 16 weeks [clinical benefit rate (CBR), 26% >16 weeks] (55). These data suggest that incorporating a predictive biomarker to select SCLC patients that may benefit from PARPi monotherapy would be prudent.

As such, there is an ongoing biomarker-selected PARPi monotherapy phase II clinical trial (NCT03009682), investigating the efficacy of olaparib in relapsed SCLC patients whose tumors harbor mutations in HR related

genes including *BRCA1/2*, *ATM*, *BLM*, *MRE11*, *RAD50*, *NBN*, *RAD51*, *RAD51* paralogs, *RECQL* family members, and other deleterious HR pathway alterations. Results from this biomarker informed clinical trial are eagerly awaited (summarized in *Table 2*).

Given that the benefit of single-agent PARPi for SCLC therapy may have limited efficacy outside of biomarker-selected patients, combining PARPi with other therapeutics is a rational next step. In subsequent sections, we will review the landscape of PARPi combination studies with chemotherapy, targeted therapeutics, radiotherapy and immunotherapy.

PARPi and drug combinations in SCLC

Chemotherapy remains the standard of care for treating SCLC patients. ES-SCLC patients treated with chemotherapy have a low median survival of about 10 months (63,64) with immunotherapy increasing that to about 12 months (65,66). There is a critical need to improve treatment efficacy and outcomes for SCLC patients that PARPi may be able to contribute toward. Several preclinical studies demonstrated that PARPi chemosensitizes SCLC. These data informed the subsequent development of related clinical trials.

Preclinical evidence and studies

Byers *et al.* first reported that the addition of olaparib to the standard platinum-based chemotherapy regimen, cisplatin and etoposide (CE), potentiated the anti-tumor effects in SCLC (46). Owonikoko *et al.* similarly reported synergism between veliparib combined with a platinum-based agent (cisplatin/carboplatin) and etoposide in SCLC cell lines as well as xenograft mouse models (67). Teicher *et al.* demonstrated enhanced sensitivity to carboplatin/etoposide treatment with talazoparib in some SCLC cell lines (68). Several preclinical reports indicated that PARPi can also sensitize SCLC to other chemotherapeutic agents. For example, Murai *et al.* observed that talazoparib sensitizes cancer cells to the DNA-alkylating agent TMZ, and that sensitization was dependent on *SLFN11* expression (48). Lok *et al.* also found evidence of synergy to the combination of talazoparib and TMZ (49).

Beyond PARPi combinations with chemotherapy, multiple groups have examined combining novel targeted therapies with PARPi (69-71). Lallo *et al.* showed that the combination of PARP inhibitor olaparib and WEE1

inhibitor adavosertib (AZD1775) can significantly improve the efficacy of the single-agent activity of olaparib in SCLC circulating tumor cell patient-derived xenografts (CDX) (69). A study in abstract form by Gay *et al.* also demonstrated synergism between an ATR inhibitor (AZD-6738) and olaparib that increased the cytotoxic effects in SCLC cell lines (70). While Sen *et al.* indicated that a CHK1 inhibitor (LY2606368) synergizes with olaparib to decrease cell viability and cause tumor regression in a triple-knockout *RB^{-/-}/p53^{-/-}/p130^{-/-}* (RPP) genetically engineered mouse model (GEMM) (71). These studies collectively demonstrated the potential of a combinatorial approach as an effective therapeutic strategy for the incorporation of PARPi into the management of SCLC.

Clinical evidence and studies

PARPi combination trials with cisplatin and etoposide

The combination of chemotherapy and PARPi has been studied in clinical trials. Two independent studies evaluated the feasibility of veliparib in combination with CE as a therapeutic strategy in SCLC (59,60). Owonikoko *et al.* completed a phase I/II clinical trial (NCT01642251) to evaluate the safety and efficacy of veliparib combined with CE in SCLC patients. A total of 128 ES-SCLC patients were recruited and treated with four cycles of CE along with veliparib or placebo. The toxicities were balanced between the two treatment groups with the exception of higher grade 3+ lymphopenia (8% *vs.* 0%; $P=0.06$) and neutropenia (49% *vs.* 32%; $P=0.08$) in the veliparib group compared to placebo. The primary endpoint of the study was to examine if the veliparib combination would reduce the PFS hazard ratio by 37.5% as analysed by a one-sided log rank test with an α of 0.10. Results demonstrated a higher median PFS (6.1 *vs.* 5.5 months; one-sided P value =0.06) and median OS (10.3 *vs.* 8.9 months; $P=0.17$) in the veliparib group, which indicated that the addition of veliparib may improve current CE treatment (59). In another phase I dose-escalation study (NCT 02289690), Atrafi *et al.* reported that 16 of the 25 ES-SCLC patients (64%) had confirmed responses to the combination of veliparib and carboplatin/etoposide with this proportion increasing to 83% (5 of 6) of SCLC patients treated at the recommended phase II dose of veliparib. To date, the phase II portion of this study is ongoing (60).

PARPi combination trials with TMZ

Farago *et al.* reported a phase I/II clinical trial (NCT02446704), where the phase II of this study

Table 2 Clinical trials of PARP inhibitors in SCLC

PARPi	Phase	Treatment	Patient selection criteria	Number of SCLC patients enrolled	Study start year	Trial status	NCT number	Associated references
Monotherapy								
Olaparib	II	N/A	Relapsed SCLC patients harboring HR-pathway gene mutations	28	2016	Recruiting	NCT03009682	N/A
Niraparib	III	N/A	Patients with ES-SCLC	591	2018	Recruiting	NCT03516084	N/A
Talazoparib	I	N/A	Patients with advanced tumors with HR mutations	23	2011	Completed	NCT01286987	(55)
PARPi + drug combination								
Olaparib	I/II	TMZ	Patients with previously treated SCLC	50	2015	Recruiting	NCT02446704	(56)
	I	Adavosertib	Patients with relapsed SCLC	15	2015	Completed	NCT02511795	(57)
	I	CLR101	Patients with relapsed SCLC	123	2016	Recruiting	NCT02769962	N/A
	II	Cediranib	Patients with stage III SCLC	126	2016	Recruiting	NCT02498613	N/A
	II	Cediranib maleate	SCLC patients with stable disease after initial therapy	132	2017	Suspended	NCT02899728	N/A
	II	Ceralasertib	Patients with platinum refractory ES-SCLC	72	2016	Active, not recruiting	NCT02937818	N/A
	II	Ceralasertib	Patients with relapsed SCLC	45	2018	Recruiting	NCT03428607	N/A
Veliparib	II	TMZ	Patients who have returned or have not responded to TMZ	104	2012	Completed	NCT01638546	(58)
	I/II	Cisplatin and Etoposide	Patients with ES- SCLC	128	2012	Completed	NCT01642251	(59)
	I/II	Carboplatin and Etoposide	Patients with ES- SCLC	25	2014	Completed	NCT02289690	(60)
	I/II	Topotecan	Patients sensitive or refractory to chemotherapy	30	2016	Recruiting	NCT03227016	N/A
Niraparib	Ib/II	TMZ	Patients with ES-SCLC with a response to chemotherapy	64	2019	Recruiting	NCT03830918	N/A
Pamiparib	I/II	TMZ	Patients with SCLC	N/A	2017	Recruiting	NCT03150810	N/A
Rucaparib	I/II	PLX038	Patients with SCLC	62	2020	Not yet recruiting	NCT04209595	N/A
Talazoparib	II	TMZ	Patients with previously treated ES-SCLC	28	2018	Recruiting	NCT03672773	N/A
	II	ZN-c3	Patients with SCLC	N/A	2019	Recruiting	NCT04158366	N/A

Table 2 (continued)

Table 2 (continued)

PARPi	Phase	Treatment	Patient selection criteria	Number of SCLC patients enrolled	Study start year	Trial status	NCT number	Associated references
PARPi + RT								
Olaparib	I	RT	Patients with ES-SCLC	24	2018	Recruiting	NCT03532880	N/A
	I	RT + durvalumab	Patients with ES-SCLC	54	2019	Recruiting	NCT03923270	N/A
Talazoparib	I	RT	Patients with ES-SCLC	24	2020	Not yet recruiting	NCT04170946	N/A
PARPi + ICB								
Olaparib	II	PD-L1 ab MEDI4736	Patients with prior platinum-based chemotherapy and 60% have chemo-resistance	20	2015	Recruiting	NCT02484404	(61)
	I/II	PD-L1 ab MEDI4736	Patients with relapsed SCLC	N/R	2016	Recruiting	NCT02734004	(62)
Pamiparib	I	PD1 ab tislelizumab	Patients with ES-SCLC	N/R	2016	Recruiting	NCT02660034	N/A
Rucaparib	II	PD-1 ab nivolumab	Patients with platinum-sensitive SCLC	36	2019	Recruiting	NCT03958045	N/A

Adavosertib (AZD1775), WEE1 inhibitor; ZN-c3, WEE1 inhibitor; Ceralasertib (AZD6738), ATR inhibitor; CLRX, nanoparticle of camptothecin; Cediranib (Pubchem CID: 9933475) and Cediranib maleate (Pubchem CID:11226834), VEGF receptor tyrosine kinase inhibitor; PLX038, a pegylated prodrug of SN38 (an antineoplastic drug); Pamiparib (BGB290), PARP inhibitor; Tislelizumab (BGB-A317), PD1 antibody. Data as of February 2020. SCLC, small cell lung cancer; ES-SCLC, extensive stage small cell lung cancer; PARP, poly (ADP-ribose) polymerase; PARPi, PARP inhibitor; HR, homologous recombination; TMZ, temozolomide; RT, radiotherapy; ICB, immune checkpoint blockade; ab, antibody; PD-L1, programmed-death ligand 1; PD-1, programmed cell death-1; N/A, not applicable; N/R, not reported.

consisted of 50 patients that were administered 200 mg of olaparib orally twice daily combined with 75 mg/m² of TMZ daily on days 1–7 per 21 days. Results from this study showed an ORR of 41.7%, PFS of 4.2 months, and OS of 8.5 months (56).

Another phase II, randomized, double-blinded clinical study conducted by Pietanza *et al.* (NCT01638546) also showed the promise of PARPi in SCLC therapy. In their study, 104 patients with recurrent SCLC were recruited and treated with either TMZ 150–200 mg/m² on days 1–5 per 28 days in combination with veliparib 40 mg twice daily or placebo for days 1–7 per 28 days. Although the results showed no significant differences in 4-month PFS and OS between the two groups, significant differences were observed in the ORR where patients receiving TMZ/veliparib had a higher ORR than TMZ alone (39% *vs.* 14%; *P*=0.016). Interestingly, in an exploratory analysis, significantly prolonged PFS and OS was observed in the TMZ/veliparib treated patients with high protein expression of SLFN11 (58).

PARPi combination trials with other drugs

In addition to combination studies of PARPi with first-line chemotherapeutic agents, clinical trials evaluating the efficacy of other drug combinations with PARPi for SCLC are currently ongoing. In a phase I clinical study (NCT02511795) of 15 relapsed SCLC patients, the relative safety of olaparib in combination with adavosertib is being evaluated (57). Other clinical trials that are ongoing include a phase II study (NCT02498613) of olaparib and cediranib (VEGF inhibitor) in multiple cancer histologies, including SCLC, and a phase II trial (NCT03227016) of veliparib and topotecan in SCLC patients sensitive or refractory to chemotherapy (summarized in *Table 2*).

PARPi and RT in SCLC

RT is an effective treatment strategy for various cancers and has been proven to improve local tumor control and survival rates for limited-stage SCLC patients when combined with chemotherapy (72,73), while reports of the effectiveness of thoracic RT for ES-SCLC have been mixed (74–76). Results from a phase III randomized clinical study by Slotman *et al.* in 495 ES-SCLC patients reported that consolidative radiation therapy (cRT; 30 Gy/10 fractions over 2 weeks) to the thorax when added to prophylactic cranial irradiation (PCI) significantly improved the secondary endpoints of

PFS at 6 months (24% *vs.* 7%, *P*=0.001) and OS at 2 years (13% *vs.* 3%, *P*=0.004) but not its primary endpoint of OS at 1 year (33% *vs.* 28%, *P*=0.066) (75). In another clinical study (RTOG0937), 86 ES-SCLC patients were randomized to PCI alone or to cRT (45 Gy/15 fractions over 3 weeks) to intrathoracic disease and up to 4 extracranial metastases in addition to PCI. This more aggressive RT strategy did not improve OS at 1 year (60.1% in PCI alone *vs.* 50.8% in PCI + cRT) (76). With these mixed results for the role of RT in ES-SCLC, novel strategies to understand and improve the application of RT for SCLC patients are warranted.

Pre-clinical evidence and studies

PARP1 plays a crucial role in several DDR pathways, and inhibiting the function of PARP1 perturbs the cell's ability to respond to RT-induced DNA damage. Indeed, Owonikoko *et al.* showed that SCLC cell lines were significantly sensitized to RT when combined with veliparib (67). PARP trapping also contributed to radiosensitization as documented by Laird *et al.* who observed that increased radiosensitization of SCLC cell lines and PDXs was most prominent with the more potent PARP trapper, talazoparib, as compared to veliparib, a lower potency PARP trapper (77). In addition to understanding the mechanisms by which radiosensitivity is achieved, pharmacodynamic imaging biomarkers may also help advance the application of PARPi in these settings (78,79).

Clinical evidence and studies

PARPi combination trials with radiotherapy

The preclinical data led to the development of a phase I clinical trial (NCT03532880) to assess the safety of olaparib with cRT in SCLC patients. In this trial, ES-SCLC patients that have undergone 4–6 cycles of a platinum-based agent and etoposide are being recruited. Patients will receive varying doses (ranging from 0 to 300 mg) of olaparib orally, twice daily, and cRT (30 Gy/10 fractions) similar to the Slotman *et al.* study (75). In addition, a similarly designed phase I study (NCT04170946) will examine this approach with talazoparib combined with the same cRT dose in ES-SCLC patients (summarized in *Table 2*). Of note, there are studies of olaparib in combination with RT in other histologies that are out of the scope of this review, but are listed here (NCT01562210, NCT02227082,

NCT02229656).

PARPi and immune oncology in SCLC

Increasing evidence demonstrates the cGAS-STING pathway as a relevant mechanistic link between DNA damage and innate immune activation (80,81). The activation of the cGAS-STING pathway leads to the recruitment and phosphorylation of TANK binding kinase 1 (TBK1) and interferon regulatory factor 3 (IRF-3), which ultimately activates the production of type I interferons (IFNs) (82). The secretion of IFNs, along with several other chemokines, promotes the recruitment of cytotoxic CD8+ T lymphocytes to tumor sites to effectively kill cancer cells (83).

Harding *et al.* demonstrated that DNA damage mediates innate immune activation in a cell cycle and cGAS-STING dependent manner. The inhibition of cell cycle progression or the impairment of the cGAS-STING pathway resulted in the loss of innate immune activity (80). Similarly, results from Mackenzie *et al.* highlighted the role that genomic instability plays in driving the formation of micronuclei following DNA damage where the cGAS molecule enters and triggers the subsequent downstream STING phosphorylation during the telophase of the cell cycle (81).

Preclinical evidence and studies

These findings prompted the investigation of novel approaches to harness DNA damage-induced innate immune activation for cancer therapy. Olaparib was subsequently reported by Sen *et al.* to activate the cGAS-STING pathway in SCLC, which enhanced the phosphorylation of TBK1 and IRF3 and ultimately stimulated the secretion of chemokines CCL5 and CXCL10. Olaparib also significantly increased the protein and surface expression levels of programmed-death ligand 1 (PD-L1) in SCLC models. The combination of olaparib and an anti-PD-L1 antibody was further evaluated in RPP immune-competent GEMMs. An evident increase in CD8+ cytotoxic T cell infiltration was accompanied by a decrease in tumor volume in these models (84).

Clinical evidence and studies

PARPi combination trials with immunotherapy

Despite these promising preclinical findings, a recent clinical trial (NCT02484404) by Thomas *et al.* found

that the combination of olaparib and a PD-L1 inhibitor (durvalumab) had modest efficacy. In this trial, 20 relapsed ES-SCLC patients were treated with 300 mg of olaparib twice a day and 1,500 mg of durvalumab every four weeks. Only 2 (10.5%) of the evaluable 19 patients had a complete or partial response to this treatment; the median PFS was 1.8 months. Grade 3 or 4 adverse effects were reported in 45% patients, with most being hematologic (61). In another phase II study (NCT02734004) that included 38 relapsed SCLC patients, Krebs *et al.* reported in abstract form that the combination of olaparib and durvalumab was well tolerated and 2 patients had confirmed partial or complete responses, however the primary endpoint of disease control rate (complete response, partial response and stable disease) at 12 weeks of 29% was in the futility region for this Bayesian designed study (62). An additional phase I trial (NCT02660034) that also includes relapsed SCLC is currently ongoing to assess the safety of PARP inhibitor pamiparib (BGB290) combined with anti-PD1 antibody tislelizumab (BGB-A317) (summarized in *Table 2*). Additional research is needed to optimize these novel immunotherapy-based combinations.

Conclusions

The prognosis for patients diagnosed with SCLC remains poor due to the aggressive nature and the frequent acquired resistance of this disease. The lack of effective and durable therapies contributes to its grim prognosis. Continual advances in our understanding of SCLC biology has shed light on targetable vulnerabilities, including PARP. Preclinical and growing clinical evidence suggest that PARPi can enhance treatment response in SCLC by acting as sensitizers of chemotherapy, targeted therapies, radiotherapy, and immunotherapy. Additionally, identifying biomarkers of SCLC treatment response to PARPi may optimize patient selection strategies. PARPi could be part of a future wave of novel therapeutics with the potential to impact SCLC patient outcomes.

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References

1. Cruz CSD, Tanoue LT, Matthay RA. Lung cancer: epidemiology, etiology, and prevention. *Clin Chest Med* 2011;32:605-44.
2. Bray F, Ferlay J, Soerjomataram I, et al. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* 2018;68:394-424.
3. Govindan R, Page N, Morgensztern D, et al. Changing epidemiology of small-cell lung cancer in the United States over the last 30 years: analysis of the surveillance, epidemiologic, and end results database. *J Clin Oncol* 2006;24:4539-44.
4. GBD. Global, regional, and national age-sex-specific mortality for 282 causes of death in 195 countries and territories, 1980-2017: a systematic analysis for the Global Burden of Disease Study 2017. *Lancet* 2018;392:1736-88.
5. Helin K, Holm K, Niebuhr A, et al. Loss of the retinoblastoma protein-related p130 protein in small cell lung carcinoma. *Proc Natl Acad Sci* 1997;94:6933-8.
6. Kaye FJ. RB and cyclin dependent kinase pathways: defining a distinction between RB and p16 loss in lung cancer. *Oncogene* 2002;21:6908-14.
7. Ahrendt SA, Halachmi S, Chow JT, et al. Rapid p53 sequence analysis in primary lung cancer using an oligonucleotide probe array. *Proc Natl Acad Sci* 1999;96:7382-7.
8. Takahashi T, Takahashi T, Suzuki H, et al. The p53 gene is very frequently mutated in small-cell lung cancer with a distinct nucleotide substitution pattern. *Oncogene* 1991;6:1775-8.
9. D'amico D, Carbone D, Mitsudomi T, et al. High frequency of somatically acquired p53 mutations in small-cell lung cancer cell lines and tumors. *Oncogene* 1992;7:339-46.
10. George J, Lim JS, Jang SJ, et al. Comprehensive genomic profiles of small cell lung cancer. *Nature* 2015;524:47-53.
11. Rudin CM, Durinck S, Stawiski EW, et al. Comprehensive genomic analysis identifies SOX2 as a frequently amplified gene in small-cell lung cancer. *Nat Genet* 2012;44:1111-6.
12. Howlader N, Noone A, Krapcho M, et al. SEER Cancer Statistics Review, 1975-2016. National Cancer Institute. 2019. Available online: https://seer.cancer.gov/csr/1975_2016/
13. FDA approves atezolizumab for extensive-stage small cell lung cancer. 2019. Available online: <https://www.fda.gov/drugs/drug-approvals-and-databases/fda-approves-atezolizumab-extensive-stage-small-cell-lung-cancer>
14. NCCN. Small cell lung cancer (version 2.2019). Available online: https://www.nccn.org/professionals/physician_gls/pdf/sclc.pdf. Accessed April 12, 2019.
15. Sabari JK, Lok BH, Laird JH, et al. Unravelling the biology of SCLC: implications for therapy. *Nat Rev Clin Oncol* 2017;14:549.
16. Gazdar AF, Bunn PA, Minna JD. Small-cell lung cancer: what we know, what we need to know and the path forward. *Nat Rev Cancer* 2017;17:725.
17. Poirier JT, George J, Owonikoko TK, et al. New approaches to small cell lung cancer therapy: from the

- laboratory to the clinic. *J Thorac Oncol* 2020;S1556-0864:30056-3.
18. Satoh MS, Lindahl T. Role of poly(ADP-ribose) formation in DNA repair. *Nature* 1992;356:356-8.
 19. Fong PC, Boss DS, Yap TA, et al. Inhibition of poly(ADP-ribose) polymerase in tumors from BRCA mutation carriers. *N Engl J Med* 2009;361:123-34.
 20. Fong PC, Yap TA, Boss DS, et al. Poly(ADP)-ribose polymerase inhibition: frequent durable responses in BRCA carrier ovarian cancer correlating with platinum-free interval. *J Clin Oncol* 2010;28:2512-9.
 21. Litton JK, Rugo HS, Ettl J, et al. Talazoparib in patients with advanced breast cancer and a germline BRCA mutation. *N Engl J Med* 2018;379:753-63.
 22. Golan T, Hammel P, Reni M, et al. Maintenance olaparib for germline BRCA-mutated metastatic pancreatic cancer. *N Engl J Med* 2019;381:317-27.
 23. Ledermann J, Harter P, Gourley C, et al. Olaparib maintenance therapy in platinum-sensitive relapsed ovarian cancer. *N Engl J Med* 2012;366:1382-92.
 24. Moore K, Colombo N, Scambia G, et al. Maintenance olaparib in patients with newly diagnosed advanced ovarian cancer. *N Engl J Med* 2018;379:2495-505.
 25. González-Martín A, Pothuri B, Vergote I, et al. Niraparib in patients with newly diagnosed advanced ovarian cancer. *N Engl J Med* 2019;381:2391-402.
 26. Bryant HE, Schultz N, Thomas HD, et al. Specific killing of BRCA2-deficient tumours with inhibitors of poly(ADP-ribose) polymerase. *Nature* 2005;434:913-7.
 27. Farmer H, McCabe N, Lord CJ, et al. Targeting the DNA repair defect in BRCA mutant cells as a therapeutic strategy. *Nature* 2005;434:917-21.
 28. Murai J, Huang SY, Das BB, et al. Trapping of PARP1 and PARP2 by Clinical PARP Inhibitors. *Cancer Res* 2012;72:5588-99.
 29. Murai J, Huang SY, Renaud A, et al. Stereospecific PARP trapping by BMN 673 and comparison with olaparib and rucaparib. *Mol Cancer Ther* 2014;13:433-43.
 30. FDA. FDA approves olaparib for gBRCAm metastatic pancreatic adenocarcinoma. 2019. Available online: <https://www.fda.gov/drugs/resources-information-approved-drugs/fda-approves-olaparib-gbrcam-metastatic-pancreatic-adenocarcinoma>.
 31. FDA. FDA approved olaparib (LYNPARZA, AstraZeneca Pharmaceuticals LP) for the maintenance treatment of adult patients with deleterious or suspected deleterious germline or somatic BRCA-mutated (gBRCAm or sBRCAm) advanced epithelial ovarian, fallopian tube or primary peritoneal cancer who are in complete or partial response to first-line platinum-based. 2018. Available online: <https://www.fda.gov/drugs/fda-approved-olaparib-lynpa-za-astrazeneca-pharmaceuticals-lp-maintenance-treatment-adult-patients>
 32. FDA. FDA approves olaparib for germline BRCA-mutated metastatic breast cancer. 2018. Available online: <https://www.fda.gov/drugs/resources-information-approved-drugs/fda-approves-olaparib-germline-brca-mutated-metastatic-breast-cancer>
 33. FDA. FDA approves rucaparib for maintenance treatment of recurrent ovarian, fallopian tube, or primary peritoneal cancer. 2018. Available online: <https://www.fda.gov/drugs/resources-information-approved-drugs/fda-approves-rucaparib-maintenance-treatment-recurrent-ovarian-fallopian-tube-or-primary-peritoneal>
 34. FDA. FDA approves maintenance treatment for recurrent epithelial ovarian, fallopian tube or primary peritoneal cancers. 2017. Available online: <https://www.fda.gov/news-events/press-announcements/fda-approves-maintenance-treatment-recurrent-epithelial-ovarian-fallopian-tube-or-primary-peritoneal>
 35. FDA. FDA approves niraparib for HRD-positive advanced ovarian cancer 2019. Available online: <https://www.fda.gov/drugs/resources-information-approved-drugs/fda-approves-niraparib-hrd-positive-advanced-ovarian-cancer>
 36. FDA. FDA approves talazoparib for gBRCAm HER2-negative locally advanced or metastatic breast cancer. 2018. Available online: <https://www.fda.gov/drugs/drug-approvals-and-databases/fda-approves-talazoparib-gbrcam-her2-negative-locally-advanced-or-metastatic-breast-cancer>
 37. FDA. FDA approves olaparib tablets for maintenance treatment in ovarian cancer. 2017. Available online: <https://www.fda.gov/drugs/resources-information-approved-drugs/fda-approves-olaparib-tablets-maintenance-treatment-ovarian-cancer>
 38. Pommier Y, O'Connor MJ, de Bono J. Laying a trap to kill cancer cells: PARP inhibitors and their mechanisms of action. *Sci Transl Med* 2016;8:362ps17.
 39. Castroviejo-Bermejo M, Cruz C, Llop-Guevara A, et al. A RAD51 assay feasible in routine tumor samples calls PARP inhibitor response beyond BRCA mutation. *EMBO Mol Med* 2018. doi: 10.15252/emmm.201809172.
 40. Abkevich V, Timms KM, Hennessy BT, et al. Patterns of genomic loss of heterozygosity predict homologous recombination repair defects in epithelial ovarian cancer. *Br J Cancer* 2012;107:1776-82.

41. Birkbak NJ, Wang ZC, Kim JY, et al. Telomeric allelic imbalance indicates defective DNA repair and sensitivity to DNA-damaging agents. *Cancer Discov* 2012;2:366-75.
42. Popova T, Manie E, Rieunier G, et al. Ploidy and large-scale genomic instability consistently identify basal-like breast carcinomas with BRCA1/2 inactivation. *Cancer Res* 2012;72:5454-62.
43. Watkins JA, Irshad S, Grigoriadis A, et al. Genomic scars as biomarkers of homologous recombination deficiency and drug response in breast and ovarian cancers. *Breast Cancer Res* 2014;16:211.
44. Telli ML, Jensen KC, Vinayak S, et al. Phase II Study of Gemcitabine, Carboplatin, and Iniparib As Neoadjuvant Therapy for Triple-Negative and BRCA1/2 Mutation-Associated Breast Cancer With Assessment of a Tumor-Based Measure of Genomic Instability: PrECOG 0105. *J Clin Oncol* 2015;33:1895-901.
45. Isakoff SJ, Mayer EL, He L, et al. TBCRC009: A Multicenter Phase II Clinical Trial of Platinum Monotherapy With Biomarker Assessment in Metastatic Triple-Negative Breast Cancer. *J Clin Oncol* 2015;33:1902-9.
46. Byers LA, Wang J, Nilsson MB, et al. Proteomic profiling identifies dysregulated pathways in small cell lung cancer and novel therapeutic targets including PARP1. *Cancer Discov* 2012;2:798-811.
47. Polley E, Kunkel M, Evans D, et al. Small Cell Lung Cancer Screen of Oncology Drugs, Investigational Agents, and Gene and microRNA Expression. *J Natl Cancer Inst* 2016;108.
48. Murai J, Feng Y, Yu GK, et al. Resistance to PARP inhibitors by SLFN11 inactivation can be overcome by ATR inhibition. *Oncotarget* 2016;7:76534-50.
49. Lok BH, Gardner EE, Schneeberger VE, et al. PARP inhibitor activity correlates with SLFN11 expression and demonstrates synergy with temozolomide in small cell lung cancer. *Clin Cancer Res* 2017;23:523-35.
50. Stewart CA, Tong P, Cardnell RJ, et al. Dynamic variations in epithelial-to-mesenchymal transition (EMT), ATM, and SLFN11 govern response to PARP inhibitors and cisplatin in small cell lung cancer. *Oncotarget* 2017;8:28575.
51. Thomas A, Murai J, Pommier Y. The evolving landscape of predictive biomarkers of response to PARP inhibitors. *J Clin Invest* 2018;128:1727-30.
52. Mu Y, Lou J, Srivastava M, et al. SLFN11 inhibits checkpoint maintenance and homologous recombination repair. *EMBO Rep* 2016;17:94-109.
53. Murai J, Tang S-W, Leo E, et al. SLFN11 blocks stressed replication forks independently of ATR. *Mol Cell* 2018;69:371-84. e6.
54. Li M, Kao E, Malone D, et al. DNA damage-induced cell death relies on SLFN11-dependent cleavage of distinct type II tRNAs. *Nat Struct Mol Biol* 2018;25:1047-58.
55. de Bono J, Ramanathan RK, Mina L, et al. Phase I, dose-escalation, two-part trial of the PARP inhibitor talazoparib in patients with advanced germline BRCA1/2 mutations and selected sporadic cancers. *Cancer Discov* 2017;7:620-9.
56. Farago AF, Yeap BY, Stanzione M, et al. Combination Olaparib and Temozolomide in Relapsed Small-Cell Lung Cancer. *Cancer Discov* 2019;9:1372-87.
57. Hamilton E, Falchook GS, Wang JS, et al. Abstract CT025: Phase Ib study of adavosertib in combination with olaparib in patients with refractory solid tumors: Dose escalation. *Cancer Research* 2019;79:CT025.
58. Pietanza MC, Waqar SN, Krug LM, et al. Randomized, Double-Blind, Phase II Study of Temozolomide in Combination With Either Veliparib or Placebo in Patients With Relapsed-Sensitive or Refractory Small-Cell Lung Cancer. *J Clin Oncol* 2018;36:2386-94.
59. Owonikoko TK, Dahlberg SE, Sica GL, et al. Randomized Phase II Trial of Cisplatin and Etoposide in Combination With Veliparib or Placebo for Extensive-Stage Small-Cell Lung Cancer: ECOG-ACRIN 2511 Study. *J Clin Oncol* 2019;37:222-9.
60. Atrafi F, Groen HJM, Byers LA, et al. A Phase I Dose-Escalation Study of Veliparib Combined with Carboplatin and Etoposide in Patients with Extensive-Stage Small Cell Lung Cancer and Other Solid Tumors. *Clin Cancer Res* 2019;25:496-505.
61. Thomas A, Vilimas R, Trindade C, et al. Durvalumab in Combination with Olaparib in Patients with Relapsed SCLC: Results from a Phase II Study. *J Thorac Oncol* 2019;14:1447-57.
62. Krebs M, Ross K, Kim S, et al. P1.15-004 An Open-Label, Multitumor Phase II Basket Study of Olaparib and Durvalumab (MEDIOLA): Results in Patients with Relapsed SCLC. *J Thorac Oncol* 2017;12:S2044-5.
63. Lara Jr PN, Natale R, Crowley J, et al. Phase III trial of irinotecan/cisplatin compared with etoposide/cisplatin in extensive-stage small-cell lung cancer: clinical and pharmacogenomic results from SWOG S0124. *J Clin Oncol* 2009;27:2530.
64. Rossi A, Di Maio M, Chiodini P, et al. Carboplatin- or cisplatin-based chemotherapy in first-line treatment of small-cell lung cancer: the COCIS meta-analysis of

- individual patient data. *J Clin Oncol* 2012;30:1692-8.
65. Paz-Ares L, Dvorkin M, Chen Y, et al. Durvalumab plus platinum-etoposide versus platinum-etoposide in first-line treatment of extensive-stage small-cell lung cancer (CASPIAN): a randomised, controlled, open-label, phase 3 trial. *Lancet* 2019;394:1929-39.
 66. Horn L, Mansfield AS, Szczesna A, et al. First-Line Atezolizumab plus Chemotherapy in Extensive-Stage Small-Cell Lung Cancer. *N Engl J Med* 2018;379:2220-9.
 67. Owonikoko TK, Zhang G, Deng X, et al. Poly (ADP) ribose polymerase enzyme inhibitor, veliparib, potentiates chemotherapy and radiation in vitro and in vivo in small cell lung cancer. *Cancer Med* 2014;3:1579-94.
 68. Teicher BA, Silvers T, Selby M, et al. Small cell lung carcinoma cell line screen of etoposide/carboplatin plus a third agent. *Cancer Med* 2017;6:1952-64.
 69. Lallo A, Frese KK, Morrow CJ, et al. The Combination of the PARP Inhibitor Olaparib and the WEE1 Inhibitor AZD1775 as a New Therapeutic Option for Small Cell Lung Cancer. *Clin Cancer Res* 2018;24:5153-64.
 70. Gay CM, Tong P, Li L, et al. Abstract 2822: ATR inhibitors are active as single agents and in combination with PARP1 and ATM inhibitors in molecularly distinct subsets of small cell lung cancer models. *Cancer Res* 2018;78:2822.
 71. Sen T, Tong P, Stewart CA, et al. CHK1 Inhibition in Small-Cell Lung Cancer Produces Single-Agent Activity in Biomarker-Defined Disease Subsets and Combination Activity with Cisplatin or Olaparib. *Cancer Res* 2017;77:3870-84.
 72. Turrisi AT, 3rd, Kim K, Blum R, et al. Twice-daily compared with once-daily thoracic radiotherapy in limited small-cell lung cancer treated concurrently with cisplatin and etoposide. *N Engl J Med* 1999;340:265-71.
 73. Faivre-Finn C, Snee M, Ashcroft L, et al. Concurrent once-daily versus twice-daily chemoradiotherapy in patients with limited-stage small-cell lung cancer (CONVERT): an open-label, phase 3, randomised, superiority trial. *Lancet Oncol* 2017;18:1116-25.
 74. Jeremic B, Shibamoto Y, Nikolic N, et al. Role of radiation therapy in the combined-modality treatment of patients with extensive disease small-cell lung cancer: A randomized study. *J Clin Oncol* 1999;17:2092-9.
 75. Slotman BJ, van Tinteren H, Praag JO, et al. Use of thoracic radiotherapy for extensive stage small-cell lung cancer: a phase 3 randomised controlled trial. *Lancet* 2015;385:36-42.
 76. Gore EM, Hu C, Sun AY, et al. Randomized Phase II Study Comparing Prophylactic Cranial Irradiation Alone to Prophylactic Cranial Irradiation and Consolidative Extracranial Irradiation for Extensive-Disease Small Cell Lung Cancer (ED SCLC): NRG Oncology RTOG 0937. *J Thorac Oncol* 2017;12:1561-70.
 77. Laird JH, Lok BH, Ma J, et al. Talazoparib Is a Potent Radiosensitizer in Small Cell Lung Cancer Cell Lines and Xenografts. *Clin Cancer Res* 2018;24:5143-52.
 78. Carney B, Kossatz S, Lok BH. Target engagement imaging of PARP inhibitors in small-cell lung cancer. *Nat Commun* 2018;9:176.
 79. Laird J, Lok BH, Carney B, et al. Positron-Emission Tomographic Imaging of a Fluorine 18-Radiolabeled Poly(ADP-Ribose) Polymerase 1 Inhibitor Monitors the Therapeutic Efficacy of Talazoparib in SCLC Patient-Derived Xenografts. *J Thorac Oncol* 2019;14:1743-52.
 80. Harding SM, Benci JL, Irianto J, et al. Mitotic progression following DNA damage enables pattern recognition within micronuclei. *Nature* 2017;548:466-70.
 81. Mackenzie KJ, Carroll P, Martin CA, et al. cGAS surveillance of micronuclei links genome instability to innate immunity. *Nature* 2017;548:461-5.
 82. Tanaka Y, Chen ZJ. STING specifies IRF3 phosphorylation by TBK1 in the cytosolic DNA signaling pathway. *Sci Signal* 2012;5:ra20.
 83. Furtos MB, Kacha AK, Kline J, et al. Host type I IFN signals are required for antitumor CD8+ T cell responses through CD8{alpha}+ dendritic cells. *J Exp Med* 2011;208:2005-16.
 84. Sen T, Rodriguez BL, Chen L, et al. Targeting DNA Damage Response Promotes Antitumor Immunity through STING-Mediated T-cell Activation in Small Cell Lung Cancer. *Cancer Discov* 2019;9:646-61.

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