



Melatonin in Edible and Non-Edible Plants

Yenilebilen ve Yenilemeyen Bitkilerde Melatonin

Ufuk KOCA ÇALIŞKAN*, Ceylan AKA, Emrah BOR

Gazi University, Faculty of Pharmacy, Department of Pharmacognosy, Ankara, Turkey

ABSTRACT

The concept of melatonin has become more important recently both in plants and in human who utilize plants for nutritional and health purposes. Melatonin, synthesized from L-tryptophan by enzymes, protects plants against difficult conditions. People have consumed these plants for their antioxidant, immunomodulator, antiinflammatory and anticancer effects. In parts of edible and non-edible plants, levels of melatonin are determined by cyclodextrin-modified micellar electrokinetic chromatography, enzyme-linked immuno sorbent assay, radioimmunoassay, high-performance liquid chromatography, liquid chromatography with electrochemical detection, liquid chromatography with fluorimetric detection, liquid chromatography-mass spectrometry, and liquid chromatography-ultraviolet spectrophotometry. In this review, biosynthesis of melatonin in both animal and plants, function of melatonin in plant kingdom, especially in medicinal/edible and nonedible plants, and detection of phytomelatonin content in those plants are presented.

Key words: Melatonin, phytomelatonin, activity of melatonin

ÖZ

Bitkilerde melatonin kavramı, son yıllarda, hem bitkiler hem de beslenme ve sağlığı koruma amacıyla bitkileri kullanan insanlar için oldukça önemli olmuştur. Enzimler aracılığıyla L-triptofandan sentezlenen melatonin bitkiyi zorlu koşullara karşı korumaktadır. İnsanlar antioksidan, immünomodülatör, antienflamatuvlar ve antikanser etkilerinden dolayı bu bitkileri kullanmaktadır. Yenilebilen ve yenilemeyen bitki kısımlarında siklodeskstrinle modifiye edilmiş miseller elektrokinetik kromatografi, enzim bağlı immünsorban deneyi, radyoimmün test, yüksek performanslı sıvı kromatografisi, elektrokimyasal algılamalı sıvı kromatografisi, florometrik algılamalı sıvı kromatografisi, sıvı kromatografisi-kütle spektrometrisi ve sıvı kromatografisi-ultraviyole spektrofotometri yöntemleri ile tespit edilmiştir. Bu derlemede, melatoninun hem hayvanlarda hem de bitkilerde biyosentezi, özellikle tıbbi/yenilebilen ve yenilemeyen bitkilerde melatoninun fonksiyonu ve bu bitkilerde fitomelatonin içeriği sunulmuştur.

Anahtar kelimeler: Melatonin, fitomelatonin, melatonin etkisi

INTRODUCTION

Melatonin (*N*-acetyl-5-methoxytryptamine) means melanophore-contracting hormone (Greek: μαύρος=black; τάσης=tension) firstly was isolated from bovine's pineal gland in 1958.^{1,2} It is a neurohormone secreted by the pineal gland and a derivative of serotonin.³ Serotonin is a monoamine neurotransmitter and one of the precursors (Figure 1), whereas L-tryptophan, like serotonin is the common precursor of melatonin biosynthesis.^{4,5} Both have many influences on health of animal and human being, such as serotonin is used against depression⁶ and also affects behaviours and inward.⁷ Secretion of melatonin increases in the dark on the contrary of light, seasonal and physiological alteration effect levels of melatonin^{8,9} for that reason that has been studied for its hormon like effects and its biological activities for decades.

Although melatonin was described in organisms such as bacteria, fungi, algae, and vertebrates¹⁰ it was notified in plants

at the end of 1994.^{11,12} Increasing number of studies have proved that there was melatonin in different parts (seed, fruit, leaf, root etc.) of plants and in so much as medicinal herbs.¹³ A major role of melatonin in plants have been discovered that protects plants against damages of changing climate.¹⁴

Biosynthesis of melatonin

Melatonin is synthesised not only in bone marrow cells¹⁵ but also in retina.^{16,17} Thus it is both a hormone and tissue factor.¹⁰ The presence of melatonin was detected in egg, biological fluids like plasma, milk, by developed methods, such as liquid chromatography (LC) with fluorimetric detection, and LC-tandem mass spectrometry (LC-MS/MS).^{18,19} Biosynthesis of melatonin is explained enzymatically from the essential amino acid precursor tryptophan to melatonin. The synthesis includes four different enzymes. The first one is tryptophan hydroxylase (TPH), which forms 5-hydroxytryptophan from tryptophan; the second is aromatic amino acid decarboxylase which forms

*Correspondence: E-mail: ukoca@gazi.edu.tr, Phone: +90 312 202 31 87

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serotonin from 5-hydroxytryptophan; the third is arylalkylamine *N*-acetyl-transferase (AANAT), which forms *N*-acetylserotonin from serotonin; and the last one is *N*-acetylserotonin *O*-methyltransferase (ASMT), which forms the final step to melatonin (Figure 2). AANAT and ASMT is considered that they were speed limiting enzymes.^{4,20}

Biological activity of melatonin

A major role of melatonin is the antioxidant function with free radicals (reactive oxygen species) and reactive nitrogen species scavenging activity²¹⁻²⁵ thus has protective effect against ultraviolet (UV) radiations induced damages.²⁶ Consequently, melatonin can be used for healing of muscle diseases, Parkinson and Alzheimer's due to antioxidant and neuroprotective affects.²⁷⁻³¹ Melatonin is widely used for sleep disorders such as jetlag and insomnia.³² Its administration can relieve daytime and overnight sleep.^{33,34} Clinical and *in vivo* studies showed that melatonin decreased symptoms of depression³⁵⁻³⁷ moreover has immunomodulator function.^{38,39} It regulates immuno fuctions by means of production interleukin (IL)-2, IL-6, IL-12 and interferon gamma.^{40,41,42} An *in vivo* study

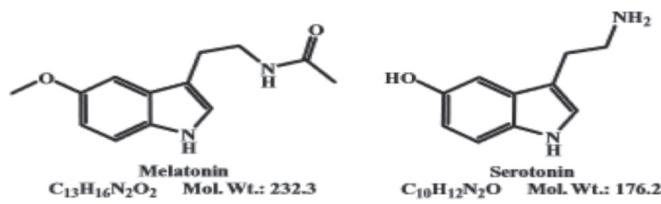


Figure 1. Structure of melatonin and serotonin

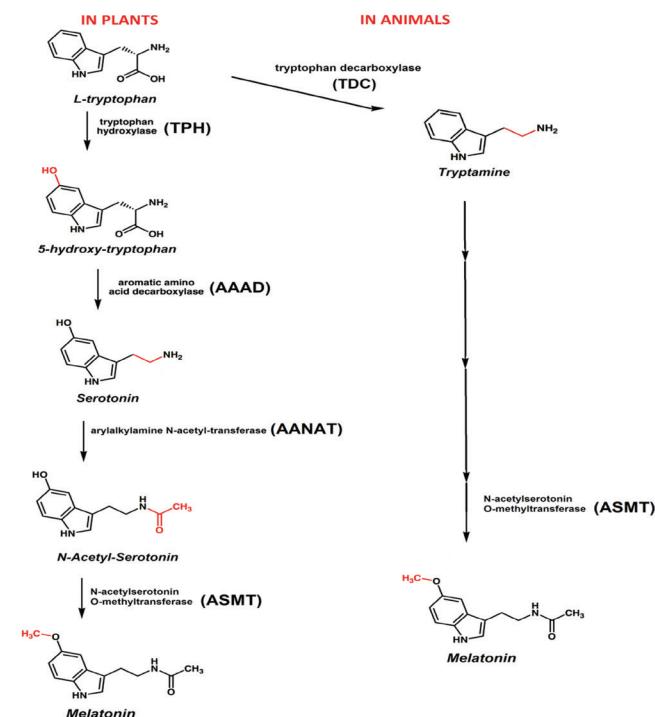


Figure 2. Modified synthesis of melatonin from tryptophan

TPH: Tryptophan hydroxylase, TDC: Tryptophan decarboxylase, AAAD: Aromatic amino acid decarboxylase, AANAT: Arylalkylamine *N*-acetyl- transferase, ASMT: *N*-acetylserotonin *O*-methyltransferase

showed that melatonin have potential anticonvulsant activity.⁴³ Melatonin effects vascular system.⁴⁴ Studies showed that melatonin suppress proliferation of cancer cell line and induces apoptosis tumor cell and also it is promising for the treatment of prostat cancer, and breast cancers.^{25,45-53} A study has also emitted that melatonin can be effective on malaria.⁵⁴

Melatonin in plants-phytomelatonin

First evidence of the presence of melatonin in organisms was obtained in *Lingulodinium polyedrum* (syn. *Gonyaulax polyedra*) and *Pyrocystis acuta*, which were unicellular organisms. Scientists detected melatonin metabolite 5-methoxytryptamine and the melatonin analogue *N,N*-dimethyl-5-methoxytryptamine in those living organisms.⁵⁵⁻⁵⁷ By following studies melatonin was determined in the members of alga, bacteria, fungi, plant families. Level of melatonin, although differs from plant to plant, that was observed higher than level of melatonin in animal blood.⁵⁸⁻⁶⁰ Melatonin level varies both from plant to plant and also tissues/organs of same plant, moreover, temperature, pH, effects of present metal ions's, sensitivity of analytics and extraction methods cause these diversities. For example, melatonin of *Datura metel* L. (devil's trumpet) differed from flowers and leaves. In addition, melatonin of *Lycopersicon esculentum* Mill. varied by region.^{11,61-64} Presence of melatonin in different plants were shown in Table 1.

Biosynthesis of phytomelatonin

Plant melatonin biosynthesis pathway firstly was determined owing to *Hypericum perforatum* L. (St John's wort).^{4,59} Synthesis in plants is complicated on the contrary in animals (Figure 2). Initial enzyme is tryptophan decarboxylase (TDC) instead of TPH. TDC forms tryptamine from essential amino acid tryptophan. The last enzyme is ASMT (Figure 2).^{65,66} Plants take melatonin also by their roots apart from biosynthesis.^{67,68} Although its biosynthetic pathway and metabolic mechanisms are unclear, the presence of melatonin in plants is a wide concept.⁶⁹

Functions of phytomelatonin

Melatonin has roles in plants similar to animals, that protects plants against extreme conditions such as temperature change, UV exposure, environmental pollution, toxins, drought oxidative and (a) biotic stress. Exogenous melatonin applied to *Arabidopsis* (thale cress) leaves has demonstrated preservative potency against high salinity, cold and dryness, additionally plant has developed tolerance biotic and abiotic stresses.⁷⁰ Corn embryo proteome was improved due to exogenous melatonin.⁷¹ Moreover, harmful effects of salt diminished by melatonin in faba bean.⁷² Conservation aspects of melatonin were studied in a variety of plants such as wheat, oat, barley, canary grass, tobacco, Chinese liquorice, soybean, cucumber, tomato.^{14,67,73-79} The studies also has shown that melatonin has regulatory role in growth of thale cress, specially growth of flowers and fruits.⁸⁰ Reports, which investigated effect of exogenous melatonin on both tomato's and maize's seeds, have confirmed this case too.^{78,81} Melatonin plays an important role to maintain the vitality of the plants.⁸²

Table 1. Levels of melatonin in plants with the detection methods

Family	Latin name	Part	Quantity		Method	Ref.
			ng/g	pg/g		
Actinidiaceae	<i>Actinidia chinensis</i> Planch.	Fruit	24	RIA	61	
Amaranthaceae	<i>Basella alba</i> L.	Leaf	39	RIA	61	
Amaryllidaceae	<i>Allium cepa</i> L.	Bulb	32	RIA	61	
Amaryllidaceae	<i>Allium fistulosum</i> L.	Bulb	86	RIA	61	
Anacardiaceae	<i>Pistacia lentiscus</i> L.	Leaf	581	ELISA	96	
Anacardiaceae	<i>Pistacia lentiscus</i> L.	Whole fruit	536±129	ELISA	96	
Anacardiaceae	<i>Pistacia palaestina</i> Boiss.	Leaf	498	ELISA	96	
Apiaceae	<i>Angelica keiskei</i> Koidz.	Leaf and stem of leaf	624	RIA	61	
Apiaceae	<i>Apium graveolens</i> L.	Seed	7	HPLC-ECD	97	
Apiaceae	<i>Coriandrum sativum</i> L.	Seed	7	HPLC-ECD	97	
Apiaceae	<i>Daucus carota</i> L.	Root	55	RIA	61	
Apiaceae	<i>Foeniculum vulgare</i> Mill.	Seed	28	HPLC-ECD	97	
Apiaceae	<i>Pimpinella anisum</i> L.	Seed	7	HPLC-ECD	97	
Arecaceae	<i>Phoenix dactylifera</i> L.	Whole fruit	469	ELISA	96	
Asparagaceae	<i>Asparagus officinalis</i> L.	Shoot	10	RIA	61	
Asparagaceae	<i>Ophiopogon japonicus</i> (L.f.) Ker Gawl.	Whole plant	198	HPLC-FD-MS	90	
Asteraceae	<i>Glebionis coronari</i> (L.) Cass. ex Spach	Leaf	417	RIA	61	
Asteraceae	<i>Dendranthema morifolium</i> (Ramat.) Tzvelev	Whole plant	160	HPLC-FD-MS	90	
Asteraceae	<i>Helianthus annuus</i> L.	Seed	29	HPLC-ECD	97	
Asteraceae	<i>Petasites japonicus</i> F. Schmidt	Shoot	50	RIA	61	
Asteraceae	<i>Silybum marianum</i> (L.) Gaertn.	Seed	2	HPLC-ECD	97	
Araceae	<i>Colocasia esculenta</i> (L.) Schott	Tuber	55	RIA	61	
Araceae	<i>Peltandra virginica</i> (L.) Raf. ex Schott	Whole plant	585	HPLC-FD-MS	90	
Brassicaceae	<i>Arabidopsis</i> spp.	Leaf	548±26	SPE, CD-ME-KC	98	
Brassicaceae	<i>Brassica campestris</i> L.	Leaf	657	RIA	61	
Brassicaceae	<i>Brassica hirta</i> Moench	Seed	189	HPLC-ECD	97	
Brassicaceae	<i>Brassica nigra</i> (L.) W. D. J. Koch	Seed	129	HPLC-ECD	97	
Brassicaceae	<i>Brassica oleracea</i> L.	Leaf	107	RIA	61	
Brassicaceae	<i>Raphanus sativus</i> L.	Whole plant	485	HPLC-FD-MS	90	
Brassicaceae	<i>Raphanus sativus</i> L.	Root	113	RIA	61	
Bromeliaceae	<i>Ananas comosus</i> (L.) Merr.	Fruit	36	RIA	61	
Caprifoliaceae	<i>Lonicera etrusca</i> hort. ex Tausch	Leaf	521	ELISA	96	
Caprifoliaceae	<i>Lonicera etrusca</i> hort. ex Tausch	Seed	403	ELISA	96	
Caprifoliaceae	<i>Lonicera japonica</i> Thunb.	Whole plant	140	HPLC-FD-MS	90	
Caprifoliaceae	<i>Viburnum tinus</i> L.	Leaf	613	ELISA	96	
Cucurbitaceae	<i>Cucumis sativus</i> L.	Fruit	25	RIA	61	
Ephedraceae	<i>Ephedra campylopoda</i> C. A. Mey.	Leaf	178	ELISA	96	
Ephedraceae	<i>Ephedra campylopoda</i> C.A.Mey.	Seed	379	ELISA	96	
Fabaceae	<i>Glycyrrhiza uralensis</i> Fisch. ex DC.	Whole plant	112	HPLC-FD-MS	90	
Fabaceae	<i>Lupinus albus</i> L.	Seed (Cotyledone)	1.28±0.06	HPLC-FD	99, 100	
Fabaceae	<i>Medicago sativa</i> L.	Seed	16	HPLC-ECD	97	
Fabaceae	<i>Trigonella foenum-graceum</i> L.	Seed	43	HPLC-ECD	97	
Juglandaceae	<i>Juglans nigra</i> L.	Fruit	3.5±1.0	HPLC-ECD	101	

Table 1. Continue

Lamiaceae	<i>Salvia miltiorrhiza</i> Bunge	Whole plant	187	HPLC-FD-MS	90
Lauraceae	<i>Laurus nobilis</i> L.	Leaf	8331	ELISA	96
Lauraceae	<i>Laurus nobilis</i> L.	Whole fruit	3710	ELISA	96
Lauraceae	<i>Laurus nobilis</i> L.	Seed	6060	ELISA	96
Lauraceae	<i>Laurus nobilis</i> L.	Pulp	1820	ELISA	96
Liliaceae	<i>Asparagus aphyllus</i> L.	Leaf	142	ELISA	96
Liliaceae	<i>Ruscus aculeatus</i> L.	Leaf	954	ELISA	96
Liliaceae	<i>Smilax aspera</i> L.	Leaf	443	ELISA	96
Linaceae	<i>Linum usitatissimum</i> L.	Seed	12	HPLC-ECD	97
Meliaceae	<i>Melia azedarach</i> L.	Leaf	1579	ELISA	96
Meliaceae	<i>Melia azedarach</i> L.	Whole fruit	585	ELISA	96
Moraceae	<i>Morus alba</i> L.	Leaf	1510	HPLC-FD-MS	90
Moraceae	<i>Morus</i> spp.	Leaf	990	ELISA	96
Moraceae	<i>Ficus carica</i> L.	Leaf	12.915	ELISA	96
Moraceae	<i>Ficus carica</i> L.	Whole fruit	3963	ELISA	96
Myrtaceae	<i>Feijoa sellowiana</i> (O. Berg) O. Berg	Leaf	1529	ELISA	96
Myrtaceae	<i>Myrtus communis</i> L.	Leaf	291	ELISA	96
Myrtaceae	<i>Myrtus</i> spp.	Leaf	490	ELISA	96
Oleaceae	<i>Olea europaea</i> L.	Leaf	4306	ELISA	96
Oleaceae	<i>Olea europaea</i> L.	Pulp	532	ELISA	96
Oleaceae	<i>Phillyrea latifolia</i> L.	Leaf	6337	ELISA	96
Oleaceae	<i>Phillyrea latifolia</i> L.	Seed	439	ELISA	96
Oleaceae	<i>Phillyrea latifolia</i> L.	Pulp	589	ELISA	96
Papaveraceae	<i>Papaver somniferum</i> L.	Seed	6	HPLC-ECD	97
Poaceae	<i>Avena sativa</i> L.	Seed	1796	RIA	61
Poaceae	<i>Avena sativa</i> L.	Seed	90.6±7.7	HPLC-ECD	102
Poaceae	<i>Hordeum vulgare</i> L.	Seed	378	RIA	61
Poaceae	<i>Hordeum vulgare</i> L.	Seed	82.3±6.0	HPLC-ECD	102
Poaceae	<i>Hordeum vulgare</i> L.	Seed	0.09±0.01	HPLC-FD	99
Poaceae	<i>Hordeum vulgare</i> L.	Seed	0.58±0.05	HPLC-FD	99
Poaceae	<i>Oryza sativa</i> L. subsp. <i>japonica</i> Shig. Kato	Seed	1006	RIA	61
Poaceae	<i>Phalaris canariensis</i> L.	Seed	26.7±2.2	HPLC-ECD	102
Poaceae	<i>Triticum</i> spp.	Seed	124.7±14.9	HPLC-ECD	102
Poaceae	<i>Triticum</i> spp.	Seed	2	HPLC-UV	102
Poaceae	<i>Triticum</i> spp.	Seed	4	HPLC-UV	102
Poaceae	<i>Zea mays</i> L.	Seed	1366	RIA	61
Poaceae	<i>Zea mays</i> L.	Seed	0.011*10-9-2.034*10-9	HPLC	103
Resedaceae	<i>Ochradenus baccatus</i> Delile	Leaf	474	ELISA	96
Resedaceae	<i>Ochradenus baccatus</i> Delile	Whole fruit	488	ELISA	96
Rhamnaceae	<i>Rhamnus alaternus</i> L.	Leaf	306±75	ELISA	96
Rhamnaceae	<i>Rhamnus palaestina</i> Boiss.	Whole fruit	907	ELISA	96
Rhamnaceae	<i>Rhamnus palaestina</i> Boiss.	Seed	547	ELISA	96

Table 1. Continue

Rhamnaceae	<i>Rhamnus palaestina</i> Boiss.	Pulp	409	ELISA	96
Rhamnaceae	<i>Ziziphus jujuba</i> Lam.	Whole plant	146	HPLC-FD-MS	90
Rhamnaceae	<i>Ziziphus jujuba</i> Mill. var. <i>spinosa</i> (Bunge) Hu ex H. F. Chou	Whole plant	256	HPLC-FD-MS	90
Rhamnaceae	<i>Ziziphus spina-christi</i> (L.) Willd.	Leaf	1324	ELISA	96
Rosaceae	<i>Crataegus aronia</i> (Willd.) Bosc	Leaf	341	ELISA	96
Rosaceae	<i>Crataegus azarolus</i> L.	Leaf	435	ELISA	96
Rosaceae	<i>Fragaria magna</i> Thuill.	Fruit	12	RIA	61
Rosaceae	<i>Malus domestica</i> Borkh.	Fruit	48	RIA	61
Rosaceae	<i>Prunus amygdalus</i> Stokes	Seed	39	HPLC-ECD	97
Rosaceae	<i>Prunus avium</i> L.	Fruit (harvested around middle May-'Burlat')	0.224±0.012	HPLC-MS	104
Rosaceae	<i>Prunus avium</i> L.	Fruit (harvested 6 days after 'Burlat')	0.027±0.024	HPLC-MS	104
Rosaceae	<i>Prunus avium</i> L.	Fruit (harvested 31 days after 'Burlat')	0.006±0.007	HPLC-MS	104
Rosaceae	<i>Prunus avium</i> L.	Fruit (harvested 33 days after 'Burlat')	0.06±0.02	HPLC-MS	104
Rosaceae	<i>Prunus avium</i> L.	Fruit (harvested 37 days after 'Burlat')	0.115±0.033	HPLC-MS	104
Rosaceae	<i>Prunus avium</i> L. http://www.ipni.org/ipni/idPlantNameSearch.do;jsessionid=3F8C9196D5F394AC6484CACBAF1C2FEE?id=160672-3&back_page=%2Fipni%2FeditSimplePlantNameSearch.do%3Bjsessionid%3D3F8C9196D-5F394AC6484CACBAF1C2FEE%3Ffind_wholeName%3DPrunus%2Bavium%26output_format%3Dnormal	Fruit (harvested 44 days after 'Burlat')	0.048±0.022	HPLC-MS	104
Rosaceae	<i>Prunus cerasus</i> L.	Fruit	1.07±0.35-2.18±0.26	HPLC-ECD	105
Rosaceae	<i>Prunus cerasus</i> L.	Fruit	5.57±0.38-19.59±2.76	HPLC-ECD	105
Rosaceae	<i>Prunus cerasus</i> L.	Fruit (Montmorency frozen)	12.3±2	HPLC-MS	106
Rosaceae	<i>Prunus cerasus</i> L.	Fruit (Balaton frozen)	2.9±0.6	HPLC-MS	106
Rosaceae	<i>Prunus cerasus</i> L.	Fruit (Balaton individually quick frozen powder)	1.7±0.5	HPLC-MS	106
Rosaceae	<i>Prunus cerasus</i> L.	Fruit (Montmorency individually quick frozen powder)	7.5±0.9	HPLC-MS	106
Rosaceae	<i>Rubus idaeus</i> L.	Whole plant	387	HPLC-FD-MS	90
Rosaceae	<i>Rubus sanctus</i> Schreb.	Leaf	805	ELISA	96
Rubiaceae	<i>Rubia tenuifolia</i> d'Urv.	Leaf	905	ELISA	96
Rubiaceae	<i>Rubia tenuifolia</i> d'Urv.	Whole fruit	339	ELISA	96
Rubiaceae	<i>Rubia tenuifolia</i> d'Urv.	Seed	539	ELISA	96
Santalaceae	<i>Osyris alba</i> L.	Leaf	844	ELISA	96
Schisandraceae	<i>Schisandra chinensis</i> (Turcz.) K. Koch	Whole plant	86	HPLC-FD-MS	90
Scrophulariaceae	<i>Scrophularia nodosa</i> L.	Whole plant	342	HPLC-FD-MS	90
Solanaceae	<i>Lycium barbarum</i> L.	Seed	103	HPLC-ECD	97

Table 1. Continue

Solanaceae	<i>Lycium barbarum L.</i>	Whole plant	530	HPLC-FD-MS	90
Solanaceae	<i>Lycopersicon esculentum Mill.</i>	Fruit	32	RIA	61
Solanaceae	<i>Solanum elaeagnifolium Cav.</i>	Whole fruit	7895	ELISA	96
Solanaceae	<i>Solanum elaeagnifolium Cav.</i>	Seed	5604	ELISA	96
Solanaceae	<i>Solanum elaeagnifolium Cav.</i>	Pulp	7392	ELISA	96
Solanaceae	<i>Solanum nigrum L.</i>	Whole fruit	323±46	ELISA	96
Styracaceae	<i>Styrax officinalis L.</i>	Leaf	4069	ELISA	96
Theaceae	<i>Camellia sinensis (L.) Kuntze</i>	Leaf	386±21	CD-MEKC	98
Tiliaceae	<i>Tilia cordata L.</i>	Leaf	410±16	CD-MEKC	98
Verbenaceae	<i>Lantana camara L.</i>	Leaf	389	ELISA	96
Xanthorrhoeaceae	<i>Aloe vera (L.) Burm. f.</i>	Whole plant	516	HPLC-FD-MS	90
Zingiberaceae	<i>Elettaria cardamomum Maton</i>	Seed	15	HPLC-ECD	97
Zingiberaceae	<i>Zingiber officinale Roscoe</i>	Rhizome	584	RIA	61

RIA: Radioimmunoassay, ELISA: Enzyme linked immunosorbent assay, HPLC: High performance liquid chromatography, ECD: Electrochemical detection, FD: Fluorescence detector, MS: Mass spectrometry, SPE: Solid phase extraction, CD: Cyclodextrin, MEKC: Micellar electrokinetic chromatography, UV: Ultraviolet

Phytomelatonin in diets

The most popular drinks, which are tea, coffee, beer and wine contain melatonin. Not only melatonin but also its isomers (tryptophan-ethylester) were determined in wine and bread.⁸³⁻⁸⁵ A study reported that regular coffee consumption remarkably decreases the prevalence of human prostate cancer.⁸⁶⁻⁸⁸ Scientists introduced that melatonin in wine besides the other secondary metabolites, had protective effect against heart injury.⁸⁹ Melatonin was determined high amount in Chinese medicinal herbs. Some of them were *Viola philippica* Cav., *Uncaria rhynchophylla* Miq., *Morus alba* L. and *Phellodendron amurense* Rupr.⁹⁰ In Mediterranean diet, melatonin was found in some foods. It's thought that melatonin can have positive effects on health via synergic effects with other compounds.⁹¹ Dietary supplement/melatonin supplement preparations have been consumed for different purposes by people mostly in Europe and the United States than the other countries.⁹²

Determination of phytomelatonin levels in plants

Melatonin has been detected in fruits, leaves, roots, and seeds of a considerable variety of plant species. Various methods, such as cyclodextrin-modified micellar electrokinetic chromatography, enzyme-linked immuno sorbent assay, radioimmunoassay (RIA), high-performance LC (HPLC), HPLC-electrochemical detection, HPLC-fluorescence detector, HPLC-MS and HPLC-UV spectrophotometry (UV) can be applied in order to determine melatonin levels in plants.

The first step in determining the levels of melatonin in plants is to find the right extraction method, which have been tried by different authors. The first identification method of melatonin in plants was described by Van Tassel et al.⁹³ in a congress communication in 1993. The authors had detected melatonin in tomato fruits (*Solanum lycopersicum* L.) by using RIA and gas chromatography attached with MS, but the results were not published extensively until 1995.⁹⁴

Nowadays, most of the researchers have been utilizing liquid nitrogen treated-plant tissue, which were extracted with organic solvents such as methanol, chloroform, or ethyl acetate. Analysis of these extracts by LC and identification by MS are the most used and recommended techniques for the detection and quantification of melatonin in plants. Due to the developed technology of LC coupled to time-of-flight/MS has also been applied for the melatonin detection in recent years.⁹⁵

Biotechnology

A biotechnologic study showed that transgenic plant rich on account of melatonin had more antioxidative activity and higher yield than regular plants.¹⁰⁷⁻¹⁰⁹ When activity of ASMT enzyme-catalyzed from *N*-acetylserotonin to melatonin and isolated firstly from rice in plants- was increased by overexpression, the level of melatonin has also increased.^{110,111} A study demonstrated that since 6-hydroxymelatonin was not determined in rice, melatonin 2-hydroxylase has been dominant enzyme in melatonin production.¹¹²

CONCLUSION

Melatonin has been studied to treat some symptoms and diseases in human over the years. Melatonin supplements have proven significant results for treating insomnia and other circadian rhythms caused sleep disorders, moreover, jet lag and shift work, headache, various cancers, gallbladder stones, tinnitus, rheumatoid arthritis, Alzheimer's disease, and psychiatric disorders have also tried to be eased with melatonin. Besides, it is known that melatonin is a powerful antioxidant and it improves the immune system. According to recent research, melatonin has also a great anti-aging effect.

Melatonin is a hormone that naturally produced by pineal glad in human brain especially at night-time, however, smoking, using

alcohol, excessive coffee consumption, some medications and disorders can suppress the production of the melatonin. Therefore melatonin should be taken externally such as synthetic melatonin supplements, or from natural resources which produce or contain melatonin. Furthermore, taking nutrients, which contain tryptophan, can increase the secretion of melatonin in the body. For instance, eating strawberries, apples, cherry/juice, rice, pistachios, almonds, spinach, cabbage, onions, tomatoes, cucumber, linseed and sunflower seeds, thistle, fenugreek and mustard; drinking teas such as fennel and anise tea.

In this study, our aim was to bring attention to melatonin in plants, which has important roles in plants as well as in animals. Many scientists have laboured to identify and quantify the levels of melatonin in plants. Although there are numbers of studies completed in plants still more studies have been needed to analyse the levels and their absorption and efficiency of melatonin directly from plants, teas and pharmaceutical preparations.

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