

1 **Cancer Risk Assessment: Should New Science Be Applied?**

2 Workgroup Summary

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1 Running Title: New Science need in Cancer Risk Assessment?

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3 Keywords:

4  
5 cancer risk assessment  
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7 adaptive response  
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9 susceptibility  
10 research needs

11  
12 Abbreviations:

13  
14 high-LET – high linear energy transfer  
15 low-LET – low linear energy transfer  
16 CDKN1A – cyclin dependent kinase 1A, also known as CIP1/WAF1  
17 TGF- $\beta$  – transforming growth factor beta  
18 MVK – Moolgavkar, Venson & Knudsen  
19 NIEHS – National Institute for Environmental Health Sciences  
20 NIH – National Institutes of Health  
21 DOE – U.S. Department of Energy

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1 Abstract

2

3 In July of 2002 a workshop was convened that explored some of the intercellular phenomena that

4 appear to condition responses to carcinogen exposure. Effects that result from communication

5 between cells that appear to either increase the sphere of damage or to modify the sensitivity of

6 cells to further damage were of particular interest. Much of the discussion focused on the effects

7 of ionizing radiation that were transmitted from cells directly hit to cells not receiving direct

8 exposure to radiation (bystander cells). In cell culture, increased rates of mutation, chromosomal

9 aberration, apoptosis, genomic instability, and decreased clonogenic survival have all been

10 observed in cells that have experienced no direct radiation. In addition, there is evidence that

11 low doses of radiation or certain chemicals give rise to adaptive responses in which the treated

12 cells develop resistance to the effects of high doses given in subsequent exposures. Data were

13 presented at the workshop indicating that low dose exposure of animals to radiation and some

14 chemicals frequently reduces the spontaneous rate of mutation *in vitro* and tumor responses *in*

15 *vivo*. Finally, it was concluded that considerable improvement in understanding of how genetic

16 variation may modify the impact of these phenomena is necessary before the risk implications

17 can be fully appreciated. The workshop participants discussed the substantive challenge that

18 these data present with respect to simple linear methodologies that are currently used in cancer

19 risk assessment and attempted to identify broad strategies by which these phenomena may start

20 to be used to refine cancer risk assessment methods in the future.

## I. Introduction

### A. The workshop

A workshop was conducted on July 17-19, 2002 to discuss the implications of recent findings in radiation biology for assessment of cancer risks from low doses of chemical and physical agents. The objective of the workshop was to bring together basic scientists who are responsible for observations made in radiobiology, their counterparts from the field of toxicology, scientists whose main interests are risk assessment, and scientists working on incorporation of concepts of mode of action into cancer risk assessments made by various federal agencies (Wiltse and Dellarco, 1996). The regulatory agencies represented at the workshop were Occupational Safety and Health Administration, National Institute of Occupational Safety and Health, Agency for Toxic Substances and Disease Registry, U.S. Environmental Protection Agency, Food and Drug Administration, the Nuclear Regulatory Commission, the Department of Energy and the National Radiological Protection Board (UK).

This workgroup summary has been prepared by reference to the presentations that were made and materials developed from summaries and notes that were made at the workshop. The summary has been circulated to the participants and the resulting comments have been utilized in developing the final draft of the manuscript. Every attempt was made to capture any important differences of opinion that were articulated at the workshop and the subsequent comment period.

### B. Evolution of cancer risk assessment

1 Risk assessment of potentially harmful physical and chemical agents developed gradually  
2 over the past half century (McClellan, 1999; 2002). The major impetus for this science has been  
3 the utilization of its research products in the development of regulatory policy related to  
4 allowable human exposures. However, the approach taken has depended in large part on  
5 whether the data available come from observational or experimental data.

6 In the case of radiation, risk estimates for cancer have been based on data from human  
7 experience, in particular the atomic bomb survivors. Data from animal experiments, and to a  
8 lesser degree cellular studies, have been used to assess the influence of dose rate, dose  
9 distribution, and the type of radiation.

10 In the period of 1945-50, the concept of “tolerance doses”, based upon the assumption that  
11 the effects of radiation had threshold doses, was gradually replaced by the “maximum  
12 permissible limit”, as the possibility that some responses might not have thresholds was  
13 recognized.

14 Dose-response assessments for non-cancer endpoints for chemicals have continued to evolve  
15 based upon the threshold concept. A reference dose is developed that is thought to be below the  
16 threshold. It is derived from a no-observed-adverse-effect-level or a lowest-observed-adverse-  
17 effect-level (or from a benchmark dose in more recent years) that is adjusted by uncertainty  
18 factors (related to the interspecies and low-dose extrapolation of data) and modifying factors  
19 based upon the completeness of the data that are available (NRC, 1983).

20 In the early 1970s a variety of developments occurred that brought greater focus to the  
21 assessment of cancer risks. The enabling legislation that formed the U.S. Environmental  
22 Protection Agency required a closer look at the regulation of chemicals in the environment.  
23 Cancer risk policy first made use of the one-hit model. This was highly criticized because it

frequently did not fit the data available and the basic assumption did not seem to apply generally. This gave way to the formalization of the linearized multistage model (Crump et al., 1976). This model had the advantage of being both biologically plausible and relatively simple in its application and has underpinned much of cancer policy in the past two decades.

Unlike radiation, chemicals that cause cancer have been largely identified by animal studies. For this reason there has been greater reliance on experimental data in risk assessment. These studies have provided evidence that many different types of mechanisms contribute to the development of cancer. The draft guidelines for cancer risk assessment (U.S. EPA, 1996) open up risk assessment to modes or mechanisms of action that would allow either the low dose or across species extrapolation of data to depart from defaults. When these data have been treated formally in models, such as that developed by Moolgavkar, Venzon & Knudson, rates of cell division and cell death, as well as changes in mutation rate, have been utilized to estimate risk at low doses (Moolgavkar and Venzon, 1979). It is recognized that even these more elaborate models remain as oversimplifications.

The main focus of the present workshop was on observations related to phenomena that suggest that a number of indirect effects (e.g. bystander effects) may be determinants of the dose-response curve in low-dose ranges. These phenomena may well introduce non-linear behavior into the dose-response curves for carcinogenic effects. Although the clearest examples are with ionizing radiation, cross-tolerance with chemicals and radiation demonstrate at least one parallel phenomenon exists with chemical carcinogens (Padovani et al., 1995; Tedeschi et al., 1995). The workshop represented one of the first attempts to assemble a group of experts to identify the implications these new findings have for cancer risk assessment in general.

1 C. Technological advances leading to identification of bystander effects.

2  
3 Bystander effects are defined broadly as responses that occur in cells not directly impacted  
4 by the chemical or physical agent in question. The term arose in radiation biology from the  
5 observation that evidence of genetic damage occurred in many more cells at low doses of high-  
6 LET radiation (or fluences) than could have been “hit” by a particle of ionizing radiation. It is  
7 important to note that to many radiation biologists, the unmodified use of the term bystander  
8 directly refers to the genetic damage in non-hit cells. However, more recent work has  
9 demonstrated that a variety of other effects can be triggered in cells not directly hit by ionizing  
10 radiation. The more recent development of microbeams has allowed the targeting of individual  
11 cells with defined numbers and types of particles. Investigators developed single particle  
12 irradiation systems at Columbia, Gray Laboratory, and Texas A&M that allowed targeting of  
13 single cells *in vitro* to more finely study the effects of very low doses on clastogenic and  
14 mutagenic responses (Hall, 2002; Nelson et al., 1996; Zhou et al., 2000). This technology has  
15 now been expanded to the study of bystander effects on cell transformation (Sawant et al. 2001a)  
16 and has allowed earlier indirect observations to be confirmed.

17 Although presaged by earlier observations, the first formal recognition that bystander effects  
18 occurred for radiation came with the publication by Nagasawa and Little (1992) where sister  
19 chromatid exchanges were noted in far greater numbers of cells than could have been hit by  
20 alpha particles at the low fluences examined. Subsequent work from this same laboratory  
21 demonstrated that gene activation in non-hit cells was communicated from those hit by alpha  
22 particles via gap junctions (Azzam et al., 1998). Mothersill and Seymour (1997) found that  
23 media taken from cells that had been irradiated decreased the clonogenic survival of unirradiated



cells, indicating that all bystander effects are not mediated through gap junctions. In his presentation, Eric Hall pointed out that such communication can be one-way, two-way, or non-existent, depending upon the type of cells involved.

## II. Examples of phenomena induced in irradiated and/or bystander cells

### A. Adaptive responses

A number of responses are induced by radiation that could be termed adaptive responses. The term is often narrowly used in radiation biology and applied to experiments where treatment with a low priming dose of radiation reduces the response of cells to a higher dose delivered a few hours later. Apparently parallel phenomena, which could be viewed as protective adaptations, have been identified without the priming dose in dose-response curves that deviate from simple linear or log-linear functions and have been described at both the cellular and whole animal levels of biological organization. The linkage between these low dose protective (or even beneficial) effects and the classical adaptive response is tenuous at this time. Since there seems to be an intuitive relationship, they will be discussed under the broad heading of adaptive response, but the discussion will focus on modification of adverse outcomes at low doses.

#### 1. Classical adaptive response

Adaptive responses to radiation can be observed as a decreased induction of chromatid aberrations when a small priming dose was followed by high doses of X-rays: Early work used a

1 prior incubation of cells with low levels of tritiated thymidine prior to a large X-ray dose  
2 (Olivieri et al., 1984). Subsequent work demonstrated that low priming doses of X-rays also  
3 resulted in fewer chromatid breaks being observed following the higher challenge dose (Wolff,  
4 1992).

5 Adaptive responses to subsequent  $\alpha$  irradiation that were induced by low doses of  $\gamma$ -  
6 irradiation (2 cGy) have been elegantly demonstrated when only 10% of the cells present were  
7 irradiated using a single particle microbeam (Sawant et al., 2001b). This study was of particular  
8 interest in that the adaptive response was found to reduce the bystander effect (measured as  
9 clonogenic survival) by 50%.

10 Experiments utilizing this same exposure paradigm have extended these observations to  
11 include a wide variety of endpoints. In the case of cell killing, relatively large priming doses  
12 confer protection to subsequent doses of similar magnitude (Raaphorst and Boyden, 1999).  
13 However, the induction of the adaptive response varies widely depending upon cell type and  
14 genetic susceptibility among individuals.

15 Adaptive responses have been demonstrated in mice by administering a low total dose of 10  
16 cGy  $\gamma$ -irradiation delivered at two different dose-rates 24 hours before administering a 1 Gy dose  
17 (Mitchel et al., 1999). Mice receiving the pretreatment were found to have a significantly  
18 prolonged latent period for development of myeloid leukemia. Similar adaptive responses were  
19 induced by 60 min of whole body hyperthermia and injection of 1500 U of interleukin-1,  
20 intraperitoneally. These studies provide an experimental case that demonstrates adaptive  
21 responses can provide protection against radiation-induced disease *in vivo*.

## 22 23 2. Complexities of dose-response curves

1  
2 A variety of experiments have shown that many of the classical responses to irradiation are  
3 more complex than originally thought. The experimental paradigms used for these experiments  
4 vary from that used to demonstrate the adaptive response, but they have been inferred to be  
5 related to adaptive responses (Joiner et al., 1996).

6 Studies of the dose-response relationships for cell killing have revealed responses that are  
7 very non-linear in the low dose range (Joiner et al., 1996). At radiation doses between 0 and 0.3  
8 Gy, cell killing is significantly greater than would be predicted by extrapolating the dose-  
9 response observed at doses  $> 1$  Gy. While the hypersensitive set of cells may come from  
10 populations in different parts of the cell cycle in part, the low-dose hypersensitivity is displayed in  
11 all phases of the cell cycle (Short et al., 1999). The interpretation offered of these results is that  
12 there is an induction of DNA repair mechanisms that protect the cells from killing in the high  
13 dose range.

14 Cells that display bystander effects (measured as reduced clonogenic survival) do not display  
15 the low dose hypersensitivity (Mothersill et al., 2002). Consequently, they are viewed as  
16 separable phenomena.

17 Low doses of  $\gamma$ -irradiation (0.1 to 10 cGy) have been shown to reduce the spontaneous  
18 transformation of cells *in vitro* (Redpath et al. 2001). Data on the reduction of background levels  
19 of damage by low dose radiation were reviewed and related to some cancer responses to  
20 radiation that have been observed in humans and experimental animals (Redpath, 2002). Thus,  
21 the experience with low-LET radiation appears to be somewhat different from the increased rates  
22 of cell transformation induced in cells that are neighbors of cells hit by high-LET  $\alpha$ -particles  
23 (Sawant et al., 2001a).

1 Qualitative differences in responses in different dose ranges are reflected at the molecular  
2 level. *In vitro* studies of changes in gene expression following varying doses of  $\gamma$ -irradiation  
3 reveals 30 genes that were activated at all doses. However, at low doses (0.2 Gy) and low dose  
4 rates, there were 137 genes activated that were not activated at 10 Gy and 16 that were activated  
5 at 10 Gy, but not activated at the lower doses (Amundson et al., 2001).

6 At the workshop, a preliminary analysis on the shape of the dose response in the low dose  
7 region was presented. Radiation studies in experimental animals that have been published in the  
8 open scientific literature were reviewed and subjected to statistical analyses for J-shaped dose  
9 response curves (Krewski, 2002). It was found that the number of studies displaying J-shaped  
10 dose-response curves was significantly greater than could be expected by chance. The J-shape  
11 appeared to occur with both high and low-LET radiation. This finding is remarkable, because  
12 these studies had not been designed to demonstrate such behavior in low doses. Thus, such  
13 effects can occur. Nevertheless, the generality of the low-dose protective effect has yet to be  
14 established.

15 Some treatments of data on cancer mortality in atomic bomb survivors support, at least in  
16 part, the hypothesis that low doses of radiation are protective against certain types of cancer.  
17 Some analyses of these data and those in the literature were discussed (Hoel, 2002). Depending  
18 upon the selection of the dose cutpoints, mortality from leukemia can appear to be less in  
19 survivors receiving low doses of radiation relative to the control population (Chomentowski et al.  
20 2000). However, solid tumor mortality displays no evidence of low dose protection. However,  
21 data involving chronic exposures to radiation (Miller et al., 1989; Hrubec et al., 1989) indicate  
22 that the dose response for some tumors appears linear within statistical confidence intervals, to  
23 the lowest exposures that have been characterized.

## B. Genomic instability

Genomic instability induced by radiation was reviewed (Morgan, 2002). There are differences in the processes responsible for induction of genomic instability that are produced by low and high doses of radiation that may be critical to understanding the implications of these observations for the development of disease.

### 1. Low dose effects

Low doses of radiation to cells in culture increases the numbers of cells that display genomic instability. Genomic instability is observed as increased non-clonal mutations or chromosomal aberrations in either cells that have been subject to direct irradiation or that are bystander cells. The phenomenon has also been described when medium is transferred from irradiated cells to naïve cells.

Genomic instability is induced *in vitro* by very low doses of radiation. In media transfer experiments; evidence of clonogenic cell death in response to  $\gamma$ -irradiation appears to approach a maximum between 1-5 cGy (Seymour and Mothersill, 2000). Genomic instability induced in bystander cells by  $\alpha$ -irradiation generally occurs when cells are cultured under conditions of close cell-to-cell contact and to be largely dependent upon gap junction communication (Azzam et al., 1998; Azzam et al., 2001). Responses are also seen at low doses, but with less evidence of saturation in the dose-response curve. This may be related to heterogeneity of dose/cell that is observed relative to that observed with equivalent energies of  $\gamma$ -irradiation in these dose ranges. Genomic instability has also been observed to be induced in bystander cells by  $\alpha$ -irradiation in

hemopoietic cells, which do not have gap junctions, both in tissue culture experiments (Lorimore et al., 1998) and in mice (Watson et al., 2001). Collectively, these data indicate, by virtue of the very high frequencies of the induced changes, that genomic instability is not directly the result of simple mutation.

## 2. High dose effects

As doses of radiation increase, the influence of genomic instability on cancer outcomes may increase. The factors affecting the dose-dependent increase in genomic instability may not be simple. In experiments presented at the workshop, the differences in the dose-response curves for the induction of chromosomal instability by acute exposures to x-rays, iron ions and  $^{125}\text{I}$ Urd were presented (Morgan, 2002).  $^{125}\text{I}$  seeding was used to construct an exposure system to obtain long-term, low dose rate, irradiation (48 hours) at 50 cGy/day. The medium from these experiments was transferred to non-irradiated cells and was found to contain a death-inducing factor that selectively affected non-irradiated cells. Thus, it appears that the effects of higher doses of radiation delivered at low dose rates can result in the development of selection mechanisms that favor cells with unstable genomes.

## 3. Other causes of genomic instability

A recent paper (Li et al., 2001), not discussed at the workshop, provided evidence that genetic instability can be induced by stresses that do not directly damage DNA. These were observed in a spontaneously arising mammary carcinoma line from Balb/c mice. Heat treatment

(45° C for 30 min), serum starvation, or *in vivo* growth in a syngeneic host all increased the yield of genomically unstable cells. In the same study, genetic instability was also demonstrated with  $\gamma$ -rays and hydrogen peroxide treatments that damaged DNA.

#### C. Other cellular fates.

It has become increasingly clear that the responses of cells from exposure to radiation or carcinogenic chemicals vary widely. Cells that become transformed directly or as a result of induced genomic instability represent only two of several possible cellular fates. The diversity of these fates following exposure to radiation is sometimes difficult to recognize in the primary literature because most studies tend to focus on one aspect of the responses and try to identify the mechanisms that are responsible for the selected endpoint.

Figure 1 displays a number of these alternative fates of cells exposed to cancer-inducing agents and hints at some of the processes that might influence the fate of individual cells in a culture dish or *in vivo*. At the molecular level, damage induced in DNA directly or through the activation of indirect processes stimulates repair of lesions in DNA. Such cells survive and continue to divide, usually after a short delay in the cell cycle to allow for repair. Some cells are killed either directly from DNA damage or secondarily from apoptosis. Other cells become resistant to killing and in some cases this resistance has been associated with the increased ability to repair DNA damage. Some cells, however, apparently become permanently arrested by mechanisms that are separable from the cyclin-dependent kinase activity of CDKN1A (Savell et al., 2001). This permanent arrest appears to be secondary to DNA damage and can involve increased expression of p53 and p21 (te Poele et al., 2002). The important point to be made is

1 that we are just beginning to understand the signaling processes that are triggered by the effects  
2 of chemicals or ionizing radiation. They are likely to occur in both directly damaged and  
3 bystander cells. Undoubtedly there are other factors that condition the relative proportion of  
4 cells having a given fate at different doses of the agent in different cell types and tissues.

#### 5 6 D. Tissue and cell-specific responses.

7  
8 Low dose hypersensitivity, bystander effects, adaptive responses, and genomic instability are  
9 not seen uniformly in cells of differing origins (see for example, Sorensen et al. 2002).

10 Moreover, the details of the responses to radiation appear to vary between species and strain.

11 These differences make it difficult to generalize the data taken from model systems. If these  
12 phenomena are to be incorporated into risk assessment models, the inability to generalize will  
13 seriously complicate the simple linear extrapolations that have been made up to now. At the  
14 very minimum, the variation in the type and magnitude of responses in different tissues suggest  
15 that low dose extrapolation will have to be tailored to the tumor site.

#### 16 17 E. Dose-dependence of responses

18  
19 An important consideration of the implications of phenomena that occur at low doses of  
20 ionizing radiation may be differences in the effects of high and low-LET radiation. Alpha  
21 particles have a high relative mass and a single track can deposit large amounts of energy  
22 resulting in catastrophic impacts upon the cell that has been hit. This correlates with the ability  
23 of a single hit by a high-LET particle to generate genomic instability in the hit cell and in



1 bystander cells, whereas it takes multiple hits by low-LET radiation to demonstrate such  
2 responses. The traditional view in radiation protection is that quality factors make simple and  
3 accurate adjustments for differences in energy deposition, the spatial differences involved, the  
4 background experience with repairing such damage, the parallel damage induced by metabolism,  
5 and the localized intensity of the damage. That view has been questioned in recent years, but  
6 remains controversial.

7 As suggested in the model of damage done to the genome in bystander cells by  $\alpha$ -particles,  
8 the simplest models that try to take these phenomena into account can result in complex  
9 relationships between dose and response (Brenner and Sachs, 2002). The inability of low-LET  
10 radiation to induce immediate alterations in bystander cells may result in less complex responses  
11 at low doses.

#### 12 13 F. Potential mechanisms underlying bystander effects.

14

15 Understanding of how cells generate, transmit and interpret signals as a result of an event that  
16 occurs in the hit cell is in its infancy. In the context of ionizing radiation, there are systems that  
17 are obvious candidates based upon some of the chemistry that is generated, such as reactive  
18 oxygen and nitrogen species. Information from studies of mechanisms that effect or modify  
19 chemical carcinogenesis adds more complexity to the overall picture. Presentations at the  
20 workshop dealt with examples of mechanisms that were involved in intracellular communication,  
21 intercellular communication, and influences that are exerted at the whole tissue level.

22 The sliding clamp model whereby mismatch repair proteins accumulate at the sites of  
23 mismatches in DNA and appear to serve as a signal for recruiting other repair proteins to these

1 sites was a mechanism that was discussed as an example (Fishel, 2001; 2002). Products of two  
2 of the mismatch repair genes, hMSH2 and hMLH1, have the additional property of promoting  
3 apoptosis, whereas the remaining homologues do not. Cells containing mutations in these genes  
4 are resistant to apoptosis induced by N-methyl-N'-nitro-N-nitrosoguanidine (MNNG). Thus,  
5 mutation in either one of these two genes will protect cells from apoptosis and provide a  
6 selective advantage to an unstable genotype. The question raised was whether loss of signaling  
7 functions of other DNA damage processing proteins could contribute to the selection of  
8 genetically unstable cells that could lead to more rapid development of cancer.

9 The ability of hit cells to contribute to instability in non-hit cells increases the complexity of  
10 carcinogenesis. The role of free radicals that are formed as the result of cells being hit by  
11 ionizing radiation or from exposures to other chemical and physical agents was explored as a set  
12 of mechanisms that might contribute to this phenomena (Spitz, 2002). The mitochondrial  
13 electron transport chain, NADPH oxidase enzymes, xanthine oxidase, and nitric oxide synthases  
14 were identified as the most likely sources of prooxidants that could contribute to bystander  
15 effects and discussed the antioxidant systems that limit the influence of these sources. Recent  
16 studies have demonstrated that ionizing radiation does result in production of reactive  
17 oxygen/nitrogen species. It is postulated that this signal can be transmitted to other mitochondria  
18 by permeability change releasing calcium, providing a means of amplifying the signal (Leach et  
19 al., 2001). More recently, ionizing radiation was shown to increase nitric oxide synthase  
20 activity (Leach et al., 2002). NO is an attractive means of transmitting signals between cells  
21 because its relative stability allows migration over several cell diameters. To date, these studies  
22 have been limited to relatively high doses of radiation (e.g. 2-5 Gy). However, there is ample  
23 evidence that low doses of low-LET radiation (<0.02 Gy) do result in measurable changes in

1 gene expression in cells in culture (Amundson et al., 1999). Although these latter results  
2 certainly have implications for intracellular responses, the extent to which they contribute to  
3 intercellular communication remains to be established.

4 Experiments were described that demonstrated that irradiation of mammary gland stroma  
5 promoted the growth of tumors from transplanted cells that received no direct exposure to  
6 radiation (Barcellos-Hoff, 2002; Barcellos-Hoff and Ravani, 2000). This effect was associated  
7 with activation of latent TGF- $\beta$ , although the precise mechanisms by which tumor cells are  
8 selected have not yet been worked out.

9 These selected topics illustrate that carcinogenic responses in intact organisms involve  
10 complex mechanisms that may well arise indirectly, rather than simply being set in motion by  
11 mutations induced by a carcinogenic agent.

#### 12 13 H. Genetically determined susceptibility

14  
15 The role genetic susceptibility plays in carcinogenic responses was discussed. It was pointed  
16 out that a relatively small fraction of human cancers are accounted for by dominant genes  
17 leaving much of the differences in susceptibility to be accounted for by interactions among  
18 multiple genes. Work that has focused on susceptibility to liver tumors in different strains of  
19 mice and the linkage of susceptibility to specific genes was discussed as an example  
20 (Drinkwater, 2002). Many genes modify susceptibility to cancer and they influence different  
21 stages in the carcinogenic process. In the case of liver cancer in mice, the growth rate of  
22 preneoplastic foci appears to be an important differentiator between susceptible and non-  
23 susceptible strains (Hanigan et al., 1988). However, the relative importance of a particular locus

1 was shown to vary among strains (6-20 fold) and accounts for only a portion of the 100-fold  
2 difference between strains. These data imply that multiple loci influence carcinogenesis and that  
3 they modify different stages. It is probable that similar mechanisms contribute to variation in  
4 susceptibility among people.

5  
6 III. The need for an integrated, quantitative description of phenomena that contribute to or  
7 modulate carcinogenic responses.

8  
9 Workshop participants expressed varying opinions about how the phenomena discussed  
10 should influence risk assessment. It was apparent from the discussions that there is now  
11 evidence of many effects of radiation that do not arise directly from damage to DNA in the hit  
12 cells, either by a direct hit to DNA or indirectly through the local generation of reactive oxygen.  
13 These “secondary” phenomena are clearly important and may be major determinants of whether  
14 exposure to low doses of radiation will have a cancer outcome. Discussions of topics led to  
15 varying degrees of consensus:

- 16 • **The implications of damage produced by high-LET and low-LET radiation could be**  
17 **quite different.** The amount of energy deposited in an individual cell decreases  
18 uniformly with low-LET radiation to a point where the total energy in any one cell fades  
19 into the background level and may not be able to produce physiological responses such as  
20 the adaptive or bystander effects. Research is needed to determine if the clustered DNA  
21 damage produced at very low doses of low-LET radiation has consequences in terms of  
22 an increased risk of cancer. The relatively low-levels of such damage with low doses of

1 low-LET radiation compared to that seen with high-LET radiation may be more readily  
2 repaired by molecular, cellular, or tissue processes. .

- 3 • **Many, if not all, of the phenomena that were largely identified for ionizing radiation**  
4 **in the presentations were also likely to play a role in cancer induced by chemicals.**

5 There are clear examples where complex and indirectly mediated modes of action play an  
6 important role in the induction of cancer. Data from studies on the mode of action of  
7 arsenic were identified as an example of comparable complexity with a chemical  
8 carcinogen (Luster, 2002; also see Miller et al., 2001).

- 9 • **Low dose extrapolation is even more complex than has been previously appreciated.**

10 Empirical data support a variety of shapes in the dose-response curve, depending largely  
11 upon the radiation quality, the tumor type, and the tissue environment. Some  
12 carcinogenic responses to ionizing radiation appear linear to doses as low as can  
13 reasonably be characterized (e.g. carcinomas) whereas others clearly have non-linear  
14 relationships with low doses (e.g. sarcomas). The main point is that in the former case,  
15 the empirical data are not sufficient to distinguish among linear and a variety of non-  
16 linear alternatives. However, on conceptual grounds and within the realm of  
17 experimental data, rational arguments can be constructed for more complex dose-  
18 response relationships (Brenner and Sachs, 2002; Krewski, 2002).

- 19 • **Responses of tissues and whole organisms will be qualitatively as well as**  
20 **quantitatively different in different dose ranges.** The data that are available clearly  
21 show that different genes are expressed in response to different dose ranges of ionizing  
22 radiation. This is reflected in different cellular behaviors as well. The probability of a

1 predominance of adaptive responses at low doses perhaps reflects the importance of a  
2 living system to be subject to some level of stress.

3 • **Intracellular and intercellular communication play important roles in determining**  
4 **whether there is a carcinogenic outcome to a given dose of an agent.** At this time  
5 there is not an integrated understanding of the signaling pathways involved, the interplay  
6 between them, and what damage may be induced or the nature of protection that may  
7 result.

8 • **Alternative cellular fates and the determinants of these fates are extremely**  
9 **important to new quantitative approaches to risk assessment.** Some risk assessment  
10 methods now explicitly incorporate cellular dynamics (e.g. MVK , see Hazelton et al.,  
11 2001). Clearly, bystander effects, genomic instability, and adaptive responses have  
12 important impacts on cellular dynamics within a tissue, but that it is too early to fully  
13 comprehend how they should be considered in risk assessment. A key consideration is  
14 whether and how these phenomena are involved at different dose ranges.

15  
16 A. Develop a means for organizing information.

17  
18 A critical need identified at the workshop was the need to have a structured means for  
19 identifying the important biological responses that collectively determine whether a cancer will  
20 develop. Hanahan and Weinberg (2000) provided an excellent classification of the traits that  
21 cells must acquire to become cancerous. These traits include self-sufficiency in growth signals,  
22 insensitivity to antigrowth signals, evasion of apoptosis, sustained angiogenesis, limitless  
23 replicative potential, and the ability to invade and metastasize. Carcinogens would necessarily

1 contribute to one or more of these properties. Both qualitative and quantitative data should be  
2 organized into a ‘repository’ that would provide risk assessors a means to organize the  
3 information. In turn the collection of the information into a structured package would enable  
4 researchers to more clearly identify those data gaps that impede the development of more  
5 accurate approaches.

6 Beyond the development of a general understanding of carcinogenesis, account must be  
7 taken of critical differences in the behavior of experimental models and what occurs in intact  
8 humans. Many *in vitro* systems utilize immortalized cells that no longer possess all the controls  
9 on growth present *in vivo*. Interspecies differences have also confounded extrapolation issues in  
10 the past. However, it is important to realize that some of the mechanisms that account for such  
11 differences are coming to be understood. For example, at least part of the basis for  
12 understanding the different numbers of mutations required for transformation of human and  
13 rodent cells have been identified (Hahn and Weinberg, 2002). It appears that adjustments for  
14 differences in target organ specificity of carcinogens may become tractable. Such issues are  
15 extremely important for translating results from experimental systems to assessing risks to  
16 humans.

## 17 18 B. Dose-dependence 19

20 Those phenomena that both contribute to the development of cancer and are modified by  
21 exogenous agents need to have established dose-response relationships to be useful in developing  
22 new approaches to risk assessment. It is frequently not appreciated that the dose-response  
23 relationships contribute qualitative as well as quantitative information. As was emphasized in

1 the conference, behaviors in one dose range frequently do not carry over to another dose range.  
2 Non-linear dose-response ultimately provides information about the underlying biology of the  
3 response, something that must be understood to address low dose issues.

4 Few of the dose-response data obtained in isolated systems are likely to be quantitatively  
5 utilized in risk assessment. The few data that will be transferred to the *in vivo* system (ultimately  
6 in humans) will be used only to the extent they can be shown to be consistent with *in vivo*  
7 observations. Data taken from the radiation biology literature continually reinforce the fact that  
8 phenomena initiated by radiation vary by cell of origin, are modified by the context of the tissue  
9 within which they reside, are affected by differences in physiological state, and are subject to  
10 variations in genetic background.

#### 11 12 C. Integration

13  
14 Concerns of risk necessarily focus on the response of intact humans. Responses of complex  
15 organisms are always conditioned by the physiological state. The physiological state of an  
16 individual will be modified by his/her genetic makeup, lifestyle, and by other factors that occur  
17 in the surrounding environment. Clearly molecular and biochemical effects which are triggered  
18 by an environmental agent can be important in the approach to improvement of risk assessments.

19 The complexity that is introduced when all the factors that can condition the development of  
20 a chronic disease such as cancer are considered will have to be simplified in several ways to be  
21 developed into a useful model or set of models to describe the process. Attempting to develop a  
22 model that includes all variables is likely to be cumbersome. An approach that is likely to be  
23 more profitable is to build dose-response relationships that are reasonable for a primary event



1 and determine how secondary factors are likely to modify the response. This was demonstrated  
2 at the workshop by illustrating how a response depending upon DNA damage might be modified  
3 if the responsible agent also induces DNA repair (Conolly, 2002). An example that differed in  
4 its outcome was recently published by Brenner and Sachs (2002) to describe potential influences  
5 of bystander effects.

#### 6 7 IV. Generalization of principles to cancer risk assessment

8  
9 An important point made in the workshop was that even with a single type of ionizing  
10 radiation the dose response relationships differ in different tumor sites. This probably reflects a  
11 different level of efficiency (e.g. very different levels of inducing apoptotic cell death in different  
12 tissues) and importance for the processes that are activated by radiation in different tissues. In  
13 the genesis of some tumors the effects of radiation could overcome the homeostatic mechanisms  
14 that protect against cancer in that tissue (e.g. mammary gland), whereas tumors do not develop at  
15 other sites until the cell killing effects of radiation begin to play a role (e.g. in the development of  
16 sarcomas).

17 Nevertheless, it is probable that intercellular communication (i.e. bystander effects), genomic  
18 instability, and adaptive responses, as well as direct consequences of DNA damage, play some  
19 role in the control and development of tumors at all sites. What is likely to differ are 1) the  
20 relative importance of these and other variables in the contribution to a carcinogenic response  
21 and 2) the details of the signaling processes that control these cellular responses.

22 Finally, there is the issue of risk within a population vs. the probable risk to any given  
23 individual in the population. The probability that an exogenous agent will produce cancer in a

1 population is a direct function of the distribution of sensitivities within that population. If there  
2 is a broad range of sensitivities in the population and if every individual in a population has some  
3 threshold to a given agent, it becomes difficult to determine whether a population threshold  
4 exists (Lutz, 1999). In a relatively homogeneous population, thresholds or protective effects may  
5 be more easily demonstrable. Similar arguments could be made for adaptive responses. They  
6 may be demonstrable for some tumors in a species, but very difficult for other tumor sites, even  
7 when examining the effects of a single agent.

## 8 9 V. Conclusions and recommendations

10  
11 The workshop made apparent that there are diverse uses made of cancer risk assessments.  
12 Issues varied by federal agency and each agency's mission. In some cases, the issues varied  
13 among multiple missions within a single agency. Fundamentally, each agency has statutory  
14 mandates that determine its risk assessment policies and the mandates do not completely overlap.  
15 It was not possible for the workshop to go more deeply into this very important area. In part  
16 because there was little social-legal expertise among the participants and little time could be  
17 devoted to discussion of these complicated issues. A key point differentiating agencies depends  
18 upon the question of who gains the benefits and who assumes the risk. Obviously, each agency  
19 must continue to develop its own risk management policies in the context of its own mission, the  
20 interest groups it must serve, its responsibilities for protection of the general public health, and  
21 the legal requirements of its enabling legislation.

22 The most wide-ranging question raised in the workshop was whether these scientific  
23 observations had reached a point where they could be used to develop new approaches to risk

1 assessment. The draft cancer risk assessment guidelines do provide for incorporation of data  
2 related to varying modes of action (Wiltse and Dellarco, 1996). There is no reason why the  
3 diverse phenomena discussed in the workshop could not be incorporated in such a construct.  
4 However, the consensus of the group was that to insert these phenomena independently into a  
5 risk assessment paradigm at this time would not be wise. It was pointed out that the observations  
6 should be used to develop a structure that would make apparent the data that are needed to  
7 develop and validate new methodologies.

8 At several stages in the workshop, the availability of new research tools was cited as being  
9 responsible for being able to detect complex phenomena that contribute to carcinogenesis. The  
10 contribution of various genome projects has been not only to provide a basis for understanding  
11 how variations in genetic background affect susceptibility to disease, but also to provide research  
12 tools in the biological sciences not even dreamed of a decade ago. The ability to expose and  
13 observe responses of single cells is a reality today. The ability to detect and quantify changes in  
14 the expression of large numbers of genes simultaneously and the availability of computational  
15 tools to aid in the interpretation of these data has exploded. The time is approaching where much  
16 better experiments can be planned that can help develop an integrated picture of how a wide  
17 variety of biological phenomena can contribute to the development of cancer. The major  
18 difficulty will be to design a set of experiments that can provide the basis for integrating the  
19 information generated into a model(s) that can improve the accuracy of risk assessments. Such a  
20 model could substantially improve the ability to make cost-effective decisions in the risk policy  
21 arena.

22 There can be no doubt of the importance of research spawned through request for proposal  
23 approaches to identify phenomena that are important for risk assessment. The advances that

1 have already been made as a result of programs conducted by several government agencies  
2 illustrate this. However, many of the participants were concerned that the complexity and  
3 magnitude of the data collection and management problem is so great that some more highly  
4 coordinated efforts may also be necessary to make more specific use of the new knowledge that  
5 has been gained through traditional granting mechanisms.

6  
7 The recommendations of the workshop are as follows:

- 8  
9 • Federal and international agencies should continue to fund basic research aimed at  
10 identifying those phenomena that modify carcinogenic responses to various agents in the  
11 environment, whether they be chemical or physical agents. Currently funded programs that  
12 will make contributions to these areas are NIEHS's Toxicogenomics Program and DOE's  
13 Low Dose Radiation Program. It is important to recognize that progress in these two areas is  
14 heavily leveraged off the highly mission-oriented activities of the various genome projects  
15 supported by NIH and DOE.
- 16 • Some effort, perhaps through an interagency initiative, needs to be directed at studies that can  
17 integrate this information by the development of:
  - 18 ○ Consensus teams to identify important variables and data gaps that limit the utility of  
19 current information.
  - 20 ○ A repository of qualitative and quantitative data that documents the nature of the  
21 phenomena thought to affect carcinogenic responses. It would appear that  
22 information should be species (strain) and target organ specific.

- Quantitative *in vivo* studies that are inclusive of parameters that can impact the carcinogenic response at low doses. These studies must be designed to identify non-linear dose-responses, if they are present, in the development of pathology as well as the behavior of molecular and cellular responses.
- Develop and refine computational approaches for carcinogenesis that not only integrate the different molecular and cellular processes that contribute to carcinogenesis in a quantitative way, but to efficiently identify deficiencies in the available data to increase the utility of such models in risk assessment.

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- 5

References:

Amundson SA, Do, KT, Fornace AJ Jr. 1999. Induction of stress genes by low doses of gamma rays. *Rad Res* 152:225-231.

Amundson SA, Bittner M, Meltzer P, Trent J, Fornace AJ Jr. 2001. Induction of gene expression as a monitor of exposure to ionizing radiation. *Rad Res* 156:657-661.

Azzam EI, de Toledo SM, Gooding T, Little JB. 1998. Intercellular communication is involved in the bystander regulation of gene expression in human cells exposed to very low fluences of alpha particles. *Radiat Res* 15:497-504.

Azzam EI, de Toledo SM, Little JB. 2001. Direct evidence for the participation of gap junction-mediated intercellular communication in the transmission of damage signals from  $\alpha$ -particle irradiated to nonirradiated cells. *Proc National Acad Sci (USA)* 98:473-478.

Brenner DJ, Sachs RK. 2002. Do low dose-rate bystander effects influence radon risks? *Int J Radiat Biol* 78:593-604.

Barcellos-Hoff MH. 2002. Presentation at the workshop.

Barcellos-Hoff MH, Ravani SA. 2000. Irradiated mammary gland stoma promotes the expression of tumorigenic potential by unirradiated epithelial cells. *Cancer Res*. 60:1254-1260

Chomentowski M, Kellerer AM and Pierce DA. 2000. Radiation dose dependencies in the atom bomb survivor cancer mortality data: A model free visualization. Rad. Res. 153:289-294.

Conolly RB. 2002. Presentation at the workshop.

Crump KS, Hoel DG, Langley CH, Peto R. 1976. Fundamental carcinogenic processes and their implications for low dose risk assessment. Cancer Res 36:2973-2979.

Drinkwater NR. 2002. Presentation at the workshop.

Fishel R. 2001. The selection for mismatch repair defects in hereditary nonpolyposis colorectal cancer: Revising the mutator hypothesis. Cancer Res 61:7369-7474.

Fishel R. 2002. Presentation at the workshop.

Hall EJ. 2002. Presentation at the workshop.

Hahn WC, Weinberg RA. 2002. Modeling the molecular circuitry of cancer. Nature Rev 2:331-341.

Hanhahn D, Weinberg RA. 2000. The hallmarks of cancer. Cell 100:57-70.



1 Hanigan MH, Kemp CJ, Ginsler JJ, Drinkwater NR. 1988. Rapid growth of preneoplastic  
2 lesions in hepatocarcinogen-sensitive C3H/HeJ male mice relative to C57BL/6J male mice.  
3 Carcinogenesis 9:885-890.

4  
5 Hazelton WD, Luebeck EG, Heidenreich WF, Moolgavkar SH. 2001. Analysis of a historical  
6 cohort of Chinese tin miners with arsenic, radon, cigarette smoke, and pipe smoke exposures  
7 using the biologically based two-stage clonal expansion model. Radiat Res 156:78-94.

8  
9 Henshaw DL, Eatough JP, Richardson RB. 1990. Is radon a causative factor in inducing  
10 myeloid leukemia and other cancers in adults and children? In: 29<sup>th</sup> Hanford Symposium on  
11 Health and the Environment, Indoor Radon and Lung Cancer: Reality or Myth, Ed: FT Cross  
12 Battelle Press, Columbus, Richland pp. 935-959.

13  
14 Hoel DG. 2002. Presentation at the workshop.

15  
16 Hrubec Z, Boice JD Jr, Monson RR, Rosenstein M. 1989. Breast cancer after multiple chest  
17 fluoroscopies: Second follow-up of Massachusetts's women with tuberculosis. Cancer Res  
18 49:229-234.

19  
20 Joiner MC, Johns H. 1988. Renal damage in the mouse: the response to very small doses per  
21 fraction. Radiat Res 114:385-398.

1 Joiner MC, Lambin P, Malaise EP, Robson T, Arrand JE, Skov KA, Marples B. 1996.  
2 Hypersensitivity to very-low single radiation doses: its relationship to the adaptive response and  
3 induced radioresistance. *Mutat Res* 358:171-183.  
4  
5 Krewski, D 2002. Presentation at the workshop.  
6  
7 Leach JK, Van Tuyle G, Lin P-S, Schmidt-Ullrich R, Mikkelsen RB. 2001. Ionizing radiation-  
8 induced, mitochondria-dependent generation of reaction oxygen/nitrogen. *Cancer Res* 61:3894-  
9 3901.  
10  
11 Leach JK, Black SM, Schmidt-Ullrich RK, Mikkelsen RB 2002. Activation of constitutive nitric  
12 oxide synthase activity is an early signaling event induced by ionizing radiation. *J Biol Chem*  
13 277:15400-15406.  
14  
15 Li C-Y, Little JB, Hu K, Zhang W, Zhang L, Dewhirst MW, Huang Q. 2001. Persistent genetic  
16 instability in cancer cells induced by non-DNA-damaging stress exposures. *Cancer Res* 61:428-  
17 432.  
18  
19 Lorimore SA, Kadhim MA, Pocock DA, Papworth D, Stevens L, Goodhead DT, Wright EG.  
20 1998. Chromosomal instability in the descendents of unirradiated surviving cells after  $\alpha$ -particle  
21 irradiation. *Proc Nat Acad Sci* 95:5730-5733.  
22  
23 Luster MI. 2002. Presentation at the workshop.

1  
2 Lutz WK. 1999. Dose-response relationships in chemical carcinogenesis reflect differences in  
3 individual susceptibility. Consequences for cancer risk assessment, extrapolation, and  
4 prevention. *Human & Exp. Toxicol.* 18:707-712.

5  
6 McClellan RO. 1999. Human health risk assessment. A historical overview and alternative  
7 paths forward. *Inhalation Toxicol* 11:477-418.

8  
9 Miller AB, Howe GR, Sherman GJ, Lindsay JP, Yaffe MJ, Dinner PJ, Risch HA, Preston DL.  
10 1989. Mortality from breast cancer after irradiation during fluoroscopic examinations in patients  
11 being treated for tuberculosis. *New Engl J Med* 321:1285-1289.

12  
13 Miller WH Jr, Schipper HM, Lee JS, Singer J, Waxman S. 2002. Mechanisms of action of  
14 arsenic trioxide. *Cancer Res* 62:3893-3903.

15  
16 Mitchell REJ, Jackson JS, McCann RA, Boreham DR 1999. The adaptive response modifies  
17 latency for radiation-induced myeloid leukemia in CBA/H mice. *Rad Res* 152:273-279.

18  
19 Moolgavkar SH, Venzon DJ. 1979. Two-event models for carcinogenesis: Incidence curves for  
20 childhood and adult tumors. *Math Biosci* 47:55-77.

21  
22 Morgan WF. 2002. Presentation at the workshop.

1 Mothersill C, Seymour CB. 1997. Medium from irradiated human epithelial cells, but not  
2 human fibroblasts reduces the clonogenic survival of unirradiated cells. *Int J Radiat Biol*  
3 71:421-427.

4

5 Mothersill C, Seymour CB, Joiner MC. 2002. Relationship between radiation-induced low-dose  
6 hypersensitivity and the bystander effect. *Radiat Res* 17:526-532.

7

8 Nagasawa H, Little JB. 1992. Induction of sister chromatid exchanges by extremely low doses  
9 of alpha particles. *Cancer Res* 52:6394-6396.

10

11 Nelson JM, Brooks AL, Metting NF, Khan MA, Buschbom A, Duncan A, Miick R, Braby LA.  
12 1996. Clastogenic effects of defined numbers of 3.2 MeV alpha particles on individual CHO-K1  
13 cells. *Radiat Res* 145:568-574.

14

15 NRC. 1983. Risk Assessment in the Federal Government. Managing the Process. Washington  
16 DC. National Academy Press.

17

18 Olivieri G, Bodycote J, Wolff S. 1984. Adaptive response of human lymphocytes to low  
19 concentrations of radioactive thymidine. *Science* 223:594-597.

20

21 Padovani L, Apolloni M, Anzidei P, Tedeschi B, Caporossi D, Vernole P, Mauro F. 1995. Do  
22 human lymphocytes exposed to the fallout of the Chernobyl accident exhibit an adaptive  
23 response? 1. Challenge with ionizing radiation. *Mutat Res* 332:33-38.

1  
2 Raaphorst GP, Boyden S. 1999. Adaptive response and its variation in human normal and  
3 tumour cells. *Int J Radiat Biol* 75:865-873.  
4  
5 Redpath JL. 2002. Presentation at the workshop.  
6  
7 Redpath JL, Liang D, Taylor TH, Christie C, Elmore E. 2001. The shape of the dose-response  
8 curve for radiation-induced neoplastic transformation in vitro. Evidence for an adaptive  
9 response against neoplastic transformation at low doses of low-LET radiation. *Radiat Res*  
10 156:700-707.  
11  
12 Savell J, Rao S, Pledger WJ, Wharton W. 2001. Permanent growth arrest in irradiated human  
13 fibroblasts. *Radiat Res* 155:554-563.  
14  
15 Sawant SG, Randers-Pehrson G, Geard CR, Brenner DJ, Hall EJ. 2001a. The bystander effect in  
16 radiation oncogenesis: I. Transformation in C3H 10T1/2 cells in vitro can be initiated in the  
17 unirradiated neighbors of irradiated cells. *Radiat Res* 155:397-401.  
18  
19 Sawant SG, Randers-Pehrson G., Metting NF, Hall EJ. 2001b. Adaptive response and the  
20 bystander effect induced by radiation in C3H 10T1/2 cells in culture. *Radiat Res* 156:177-188.  
21  
22 Seymour CB, Mothersill C. 1997. Relative contribution of bystander and targeted cell killing to  
23 the low dose region of the radiation dose-response curve. *Radiat Res* 153:508-511.

1

2 Short S, Mayes C, Woodcock M, Johns H, Joiner, MC. 1999. Low dose hypersensitivity in the

3 T98G glioblastoma cell line. *Int J Radiat Biol* 7:847-855.

4

5 Simmons PT, Portier CJ. 2002. Toxicogenomics: the new frontier in risk analysis.

6 *Carcinogenesis* 23:903-905.

7

8 Sorensen KJ, Attix CM, Christian AT, Wyrobeck AJ, Tucker JD. 2001. Adaptive response

9 induction and variation in human lymphoblastoid cell lines. *Mutat Res* 519:15-24.

10

11 Spitz DD. 2002. Presentation at the workshop.

12

13 Tedeshi B, Caporossi D, Vernole P, Padovani L, Appoloni M, Anzidei P, Mauro F. 1995. Do

14 human lymphocytes exposed to fallout of the Chernobyl accident exhibit an adaptive response?

15 2. Challenge with bleomycin. *Mutat Res* 332:39-44.

16

17 te Poele RH, Okorokov AL, Jardine L, Cummings J, Joel SP. 2002. DNA damage is able to

18 induce senescence in tumor cells in vitro and in vivo. *Cancer Res* 62:1876-1883.

19

20 Watson GE, Lorimore SA, Macdonald DA, Wright EG. 2001. *In vivo* chromosomal instability

21 and transmissible aberrations in haemopoietic stem cells induced by high- and low-LET

22 radiations. *Int J Radiat Biol* 77:409-417.

23

1 Wiltse J, Dellarco VL. 1996. U.S. Environmental Protection Agency guidelines for carcinogen  
2 risk assessment: Past and future. *Mutat Res* 365:3-15.

3  
4 Wolff S. 1992. Is radiation all bad? The search for adaptation. *Radiat Res* 131:117-123.

5  
6 Zhou H, Randers-Pehrson G, Waldren CA, Vannis D, Hall EJ, Hei TK. 2000. Induction of a  
7 bystander mutagenic effect of alpha particles in mammalian cells. *Proc. Natl. Acad Sci. (USA)*  
8 97:2099-2104.

9

1 Figure 1. A schematic rendering of key pathways involved in chemical and radiation-induced  
2 carcinogenesis. Classical approaches to risk assessment have focused on the generation of stable  
3 mutations or aberrations by direct action of a carcinogen. New research has identified diverse  
4 responses that occur as a result of low-level carcinogen exposure. These processes have the  
5 potential of magnifying or limiting the probability that these mutations will be expressed as a  
6 carcinogenic response.