

MOLECULAR EVOLUTION OF A SUPERFAMILY OF CHEMORECEPTORS IN *MUSCA DOMESTICA* AND *CERATITIS CAPITATA*

BY

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THESIS

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ABSTRACT

The house fly (*Musca domestica*) and the Mediterranean fruit fly (*Ceratitis capitata*) are two important pest species. While both are generalists, the first is a major vector of a variety of human and livestock disease, while the second is a broadly polyphagous herbivore and important crop pest. Each must be able to navigate a complex set of chemical cues to gain access to resources. While it is relatively-well studied in *Drosophila* species, less is known about chemoreception in other dipterans. Generalist species of *Drosophila* retain higher diversity of chemoreceptor genes as compared to the rapid loss characteristic of specialists, but the same may not be true of other dipterans. Here the odorant receptor subfamily in *M. domestica* and the gustatory receptor subfamily in *C. capitata* were annotated, and phylogenetic analyses were performed.

Compared with *Drosophila melanogaster*, the *M. domestica* odor receptor repertoire is considerably larger with 84 models in the gene set, plus OrCo (59 in *D. melanogaster*), and contains expansion via complicated relationships of duplications in some lineages and losses in others. *C. capitata* shows a similar pattern of expansion within its gustatory receptor repertoire, with 81 protein models, which is intermediate between *D. melanogaster* (68 protein models) and *M. domestica* (103 protein models). The many species-specific expansions are the likely result of major changes in the chemosensory ability and ecology of each species. Similarity of ecologies between species appears to be a major driver for gene family evolution. Further exploration of these losses and expansions might ultimately lead to innovations in pest management and provide a base for which to further understand evolution of specialist versus generalist dipterans.

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INTRODUCTION

Among insects, chemoreception plays a strong role in locating and assessing mates, food, and oviposition sites. A large superfamily of seven-transmembrane receptors mediate response to a variety of volatile and soluble chemicals, to include odorants, tastants, and pheromones (Clyne et al., 1999; Hallem & Carlson, 2004; Leal 2013; Suh et al., 2014). This superfamily contains the distantly-related odor receptor (OR) and the gustatory receptor (GR) families (Robertson et al., 2003). In addition, the ionotropic receptor (IR) family, comprised of three-transmembrane domains, is also used in gustation and some aspects of olfaction (Benton, 2009). A fourth, odorant binding protein (OBP), family serves to bind and solubilize odorants then transport them through the sensillar lymph to the membrane-bound ORs (Leal, 2013). In adult flies, the antennae and maxillary palps are covered in sensilla (sensory hairs) and contain the olfactory receptor neuron (ORN) dendrites. These sensilla are well conserved in structure across insect species, consisting of a cuticular wall with multiple pores to allow odors through (Rospars, 1988). Each ORN expresses one to three ligand-binding ORs in with one being the most common for odor-specificity (Vosshall, 2000). In addition, *Orco*, which is necessary for olfaction (Vosshall, et al. 1999), is coexpressed with ORs in a heteromeric complex (Neuhaus et al. 2005). These complexes are thought to form ligand-gated cation channels (Leal, 2013). In *Drosophila*, a combinatorial code appears to contribute to olfactory sensitivity, as multiple ORs often detect the same compounds (Leal, 2013). Gustatory sensilla are distributed over the adult fly rather than being in only a few locations as are ORs (Stocker, 1994). As with ORs and ORNs, GRs are expressed in small subsets of gustatory receptor neurons (GRNs) (Smith, 2001), with some GRNs containing many coexpressed GRs (Thorne et al., 2004).

Conservation of chemoreceptor genes varies in flies and in other insects, with both *Orco* (well-conserved across most insects) (Hallem et al., 2006) and *Gr21* (well-conserved among Diptera and confers a CO₂ response in coexpression with *Gr63*) (Hill et al., 2002, Kwon et al. 2007) being examples of highly conserved genes. However, gene losses and duplications can be indicative of host changes, life history changes, and speciation (Wu and Ting, 2004). Rapid loss of chemoreceptor genes of specialists has been documented in *Drosophila* species (Wu & Ting, 2004; McBride & Arguello, 2007), whereas generalist species retain a higher diversity (Clark et al., 2007). This trend likely relates to the ecology of the species, where specialists may encounter fewer odorants, tastants, and toxic compounds than generalists.

The ecology of the model organism *Drosophila melanogaster*, as well as conspecifics, have been well studied and have genomes, including annotated chemoreceptors, available (Adams et al., 2000; Guo and Kim, 2007) for use in comparisons. Comparisons among species in the genus can provide some insight. However, few full chemoreceptor annotation studies have been completed outside of blood-feeders and *D. melanogaster* (NCBI, 2016). The genome annotations of the tsetse fly (*Glossina morsitans*) (Obiero et al., 2014) shows a continuation of the pattern of rapid gene loss in specialist feeders, but outside of *Drosophila*, little work has been done on patterns of chemoreceptor gene loss and duplication in generalist fly species. As genes which no longer serve a function (such as extraneous duplications) have no selective pressure against mutations and are likely to become pseudogenes (Li et al., 1981), and given the short generation time of flies, retained duplications are likely to be evolutionary important. To gain a more comprehensive understanding of patterns of dipteran gene loss and gain, further analysis at a deeper phylogenetic level is necessary. This study considers two generalist species within the suborder Brachycera (Wiegmann et al., 2011).

House flies, *Musca domestica*; (Diptera: Muscidae), are extreme generalists which are able to feed on a wide variety of decomposing matter and the bacteria it contains (Keiding, 1986; Watson & Martin, 1993). Larvae are unable to leave the substrate in which they develop. As decomposing matter, to include animal feces and carcasses, contains many toxins, pathogens and competitors (Zvereva, 1986a; Zvereva, 1986b) a variety of chemoreceptors likely play an important role in allowing the female to choose the correct oviposition sites. House flies are important vectors of human and animal diseases (Greenberg, 1973; Keiding, 1986, Scott & Lettig, 1962). Dangerous bacterial (including anthrax, typhoid fever, tuberculosis, cholera, and leprosy), fungal, and other pathogens are may be carried and transmitted by house flies. Due to this and their close association with humans and livestock, a comprehensive study of *M. domestica* chemoreception is of particular interest for their control.

The Mediterranean fruit fly, *Ceratitis capitata*; (Diptera: Tephritidae), hereafter referred to as the “medfly”, is a generalist plant feeder that originated in sub-Saharan Africa (Thomas et al., 2001). Its ability to shift host preferences based on availability has enabled the medfly to spread around the world (Malacrida et al., 1992). The medfly attacks over 260 documented fruits, flowers, vegetables, and nuts (Liquido et al., 1990) and is one of the most important pests of fruit crops worldwide. Combined annual costs of damage, control, and prevention in the US alone are in the millions of dollars (Szyniszewska & Tatem, 2014). Mating occurs primarily on the host plant (Kaspi & Yuval, 1999), and both males and females must distinguish between a wide range of chemical cues in order to locate appropriate host plants. As with *M. domestica*, fitness of medfly larvae is affected by competition (Dukas et al., 2001), and suitability of the potential oviposition location in regards to fruit ripeness and species must be determined by the female (Joachim-Bravo & Fernandes, 2001). Though many of the important pest species in

Tephritidae are specialists, chemoreceptor sequences of generalists such as the medfly can serve as models to search for and characterize other tephritid species chemoreceptor genes.

Annotations of the chemoreceptors in these two widespread and highly adaptive species may also offer insights into how flies adapt to new habitats and novel food sources. Such shifts can also result in sympatric speciation (Berlocher & Feder, 2002), allowing insight into processes of natural selection and evolution. Novel food sources such as non-indigenous hosts may serve as drivers for this selection, such as in the case of the *Rhagoletis pomonella* (apple maggot fly) complex. A relatively short (~150 years) shift from hawthorn (*Crataegus* spp.) to apple (*Malus pumila*) with associated loss of gene flow (a rate of 4-6% per generation) between host races underlines how quickly distinct populations can form (Dambroski et al., 2005).

As well as shedding light on speciation, understanding host shifts is especially critical to understand in a worldwide economy in which new and potentially invasive pests are easily transported from their country of origin across the world. Ecological traps—in which an organism is preferentially attracted to a low-quality habitat over a high-quality habitat, have long been studied (Dwernychuk & Boag, 1972), but the importance has only more recently been considered. While polarized light pollution has been considered one of the more well-documented sources of ecological traps for aquatic insects (Horváth et al., 2009), chemical cues cannot be overlooked. The mosquito *Culex pipiens* has shown preference for oviposition in aquatic habitats with non-indigenous blackberry (*Rubus allegheniensis*) detritus despite a low emergence rate in the same habitats versus those of more suitable habitats (Gardener et al., 2015). When presented with plant infusions (lacking in visual cues), the trend was retained (Gardner et al., 2014), suggesting a possibility for new methods of control.

Specific blends of attractive volatiles are already in use for selective trapping of the apple maggot fly (Zhang et al., 1999; Nojima et al., 2003). Conversely, non-host fruit volatiles can act as antagonists, repelling insects (Linn et al. 2005). Even the genotype of the host plant can affect host-finding and oviposition behaviors (Rull & Prokopy, 2004). Furthermore, duplications of chemoreceptors parallel those of resistance, such as the case of cytochrome P450s, which have been implicated in pesticide resistance a variety of insects (Wondji et al., 2009; Bass & Field, 2011). This underscores the usefulness of chemoreceptor annotations, as behavioral assays without ligand identification may not be as fine-tuned as would be ideal for such control methods as the production of better attractants and repellants and in developing insect-resistant crop varieties. The ability to stay one step ahead of resistance by implementing a variety of control methods, to include those based on chemoreception, may prove invaluable in integrative pest management systems. These methods are likely to become increasingly important in the light of pesticide resistance. Cross-resistance of one pesticide to another is documented in mosquitoes (Chandre et al., 1999) and beetles (Mota-Sanchez & Hollingworth, 2006) and is likely a common cause of resistance in insects, making development of new pesticides all the more difficult.

A better understanding of gene families thought to be involved may ultimately improve prevention and control methods for not only medflies and houseflies, but for many current pests, as well as invasive and potentially invasive insect species. With comparisons to the well-studied *D. melanogaster* genome, it is possible to make inferences about the evolution of chemoreceptor families in these two generalist species. While *Drosophila* is more closely related to *Musca*, it is expected that the more similar ecologies of *Drosophila* and *Ceratitis* will have resulted in similar expansions and deletions as compared to the filth fly, *Musca*, though they may be independent gene duplications due to the greater evolutionary distance between the species. Conversely,

genes involved in the detection of odorants and tastants associated with decay are likely to have been retained between *Drosophila* and *Musca*, as—though they feed on different substrates—both feed on decaying food sources.

MATERIALS AND METHODS

Musca domestica methods

Gene families were manually annotated and analyzed with the aid of corrected distance phylogenetic trees. BLAST searches were performed on the available Official Gene Set of proteins in REFSEQ at NCBI. All *D. melanogaster* and available *M. domestica* proteins were used as queries for TBLASTN searches. TextWrangler was used to manually assemble gene models. OBPs, gustatory receptors, and ionotropic receptors were not annotated by the author and the methods will not be discussed within but are instead available in the house fly genome paper (Scott et al., 2014). Gene nomenclature was done in accordance with Drosophila convention, with gene names italicized and proteins not.

Common problems involved absence of exons in gaps between contigs with scaffolds or off ends of scaffolds (denoted with suffices NTE [N-terminus missing sequence], CTE [C-terminus missing sequence], and INT [internal missing sequence] in the figures, tables, and proteins). Only a few of these gene models were able to be corrected using raw reads (suffix FIX in the figures, tables, and proteins) due to frequent large and complicated introns. Several of the gene models spanned scaffolds, and they had no support other than the agreement of the available exons in both scaffolds, and their appropriate relatedness to similar genes (suffix JOI in the figures, tables, and proteins). These problems are denoted in the Tables section. Genes on short scaffolds that were identical to ones on longer scaffolds were ignored as likely resulting from separate assembly of another haplotype. Extremely short fragments and highly degraded pseudogenes were also excluded. The *OrCo* gene, which is highly conserved across insects, had the last two exons duplicated 4kb downstream as well as a duplication of the first four exons at

the 5' end of another 231kb scaffold (originally modeled as XP_005184813). These were ignored as likely assembly artifacts due to polymorphisms or, if real, as identical fragments.

Pseudogenes were translated as best possible in order to do phylogenetic analysis with intact proteins. Pseudogenes that were at least half the average length of related proteins were included (denoted with suffix PSE in the figures, tables and proteins). CLUSTALX v2.0 (Larkin et al., 2007) on default settings was used to align protein families with relevant *D. melanogaster* families, and these alignments were used to help further correct problematic gene models and of pseudogenes. Pseudogenes with in-frame deletions or insertions, crucial amino acid changes, promoter defects, or other unrecognizable changes were necessarily included as encoding functional proteins and may have artificially inflated the totals.

Poorly-aligned and variable length N-terminal and C-terminal regions were excluded for phylogenetic analysis, as was an internal region of the ORs which does not align with the OrCo proteins. Though potentially misleading, other regions of potentially problematic alignment were retained as they can provide important information for relationships within subfamilies. A combination of model-based corrections of distance between each pair of proteins and distance-based phylogenetic tree building was used for phylogenetic analysis. The BLOSUM62 amino acid exchange matrix within the maximum likelihood phylogenetic program, TREEPUZZLE v5.2 (Schmidt et al., 2002), was used to correct pairwise distances for multiple changes in the past. The resulting corrections were entered into PAUP*v4.0b10 (Swofford, 2003) where a full heuristic distance search was conducted with tree-bisection-and-reconnection branch swapping to search for the shortest tree. Bootstrap analysis with 10,000 replications of neighbor-joining using uncorrected distances was performed to assess the confidence of branches, and they are shown above major branches in the figures. Adobe Illustrator® was used to manually color and

attach labels to lineages and subfamilies. Subfamilies were defined by shared evolutionary ancestry and function as described in *Drosophila*, where the degree of sequence similarity across all ORs is 26 percent or less (though still considered a family due to highly conserved sequences at certain locations throughout the protein) while the degree of subfamily similarity is upwards of 40 percent (Vosshall et al., 1999).

***Ceratitis capitata* methods**

The gustatory gene receptor family was manually annotated and analyzed using maximum likelihood phylogenetic trees. BLASTP searches were performed on both the JAMg Consensus Gene Set v1 (Poelchau et al., 2015) and high- and low-confidence protein sets from NCBI. *D. melanogaster* and *M. domestica* gene relatives were also used as queries for TBLASTN searches. Gene models were assessed for intactness and correctness with regards to length, introns (location, number, and boundaries), and conserved residues. Errors were repaired and named in the WebApollo (Lee et al., 2013) tool for manual gene annotations. Difficult gene models were manually assembled in TextWrangler prior to modification in WebApollo.

The genome assembly was generally straightforward with regards to this gene family. The most common corrections that needed to be made were truncation of exons in gaps between contigs within scaffolds or off of end of scaffolds (denoted with the suffix NTE in all figures, tables, and protein sequences). Raw reads were used to correct these gene models (suffix FIX, as in *M. domestica* results) where possible. Pseudogenes were translated as best possible for the purpose of aligning with intact proteins for phylogenetic analysis (suffix PSE). To qualify for analysis, a pseudogene had to be at least half the average length of related proteins. Several shorter fragments were not included. Protein alignment with *D. melanogaster* proteins and

refinements of gene models and pseudogenes were completed in the same manner as in *M. domestica*.

Alignments were trimmed for phylogenetic analysis using TRIMAI v1.4 (Capella-Gutiérrez et al., 2009), retaining only positions present in more than 80% of the sequences. Phylogenetic analysis was performed using maximum likelihood methods in PHYML v3.0 (Guindon et al., 2010) using default settings, and trees were prepared in FIGTREEv1.4 (Rambaut, 2014) and Adobe Illustrator®.

RESULTS

Musca domestica results

To avoid assumptions of orthology based on numbers, the numbering of MdOr genes starts with *MdOr1* as the ortholog of *DpORN*, a *D. pseudoobscura* gene which was lost from *D. melanogaster*, then roughly follows the *D. melanogaster* cytologically-named genes in order. The MdOr gene set consists of 84 models, plus the *OrCo* gene, compared to 59 models in *D. melanogaster*. While *D. melanogaster* has two sets of proteins encoded through alternative splicing of one gene (66aA/B and 69aA/B), the MdOr set only contains one such alternatively spliced gene (MdOr84A/B), and this model is questionable due to a large gap between them that might contain the C-terminal exons for *MdOr84A*. There are 7 apparent pseudogenes (8%), and 8 genes with missing parts could also be pseudogenes, with the result of 78 total apparently intact OR proteins. While approximately 12 gene fragments were so short and incomplete as to not be included, some may yet represent intact genes.

The automated gene modeling had access to all available insect ORs in GenBank for comparative information. The REFSEQ set used as the official gene set of the genome project was sufficient for building at least partial gene models for all but 2 of the current 78 intact genes. Unlike the case of many insect genome projects, more than half of these (44) models were precisely correct, presumably due to the relatively close relationship between *M. domestica* and *Drosophila*. The remaining 32 models required at least one change, and two new gene models were generated. Pseudogenes and models requiring joining across scaffolds were not included in this count.

As is expected, there was a single conserved ortholog (a) of the DmOr83b (now called Orco), sharing 87% amino acid identity (Vosshall, 2011). These were declared the out-group to root the tree (Figure 4). Fourteen simple 1:1 orthologous relationships appear in the set, such as the relationships of MdOr1JOI and DpOrN near the base of the tree (Figure 1b), sharing 44% amino acid identity, which allowed for confident building of the MdOr1 model across two scaffolds. Despite appearing to be simple orthologs, some cases such as in the pair of MdOr79 and DmOr88a (Figure 1c) (25% amino acid identity) are extremely divergent. While they are the best reciprocal BLAST matches and cluster together confidently in the tree and appear to be orthologous, they do not share microsyntenic neighbors in the two genomes and might instead have lost their true orthologs in each species and be inappropriate comparisons of paralogs.

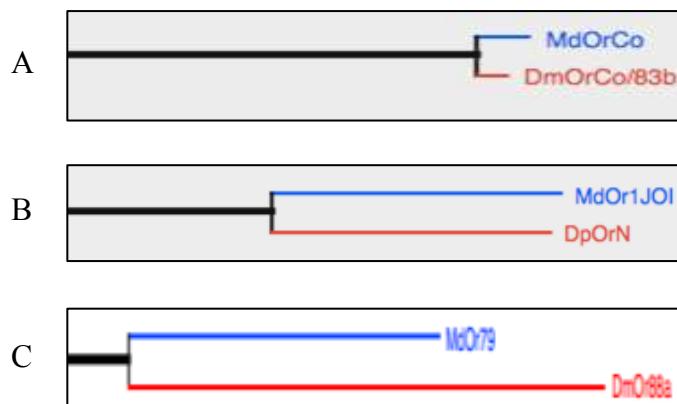


Figure 1. The 1:1 orthologous relationship between DmOrco/83b and MdOrCo (A) as well as DpOrN and MdOr1JOI (B). (C) Shows a more complicated relationship between the seemingly orthologous DmOr88a and MdOr79. The extreme sequence divergence may indicate a paralogous relationship. (Branch lengths across the three diagrams are not on the same scale. See Figure 4 for true lengths.)

A birth-and-death mode of evolution in this large family corresponds to its ecological relevance within a broadly generalist feeder. As well as the simple orthologs, there were a variety of more complicated relationships such as gene duplications in one or both species, large expansions in one species, and apparent gene losses. DmOr1a was duplicated as MdOr2/3 (Figure 2a), while the MdOr80 gene was duplicated as DmOr94a/b in *Drosophila* (Figure 2b).

The set of DmOr85b-d and MdOr71-75 (Figure 2c) is considerably more complicated, and the orthology is less clear. This may be, in part, due to the missing C-termini in two of the *Musca* genes, as well as one being pseudogenized.

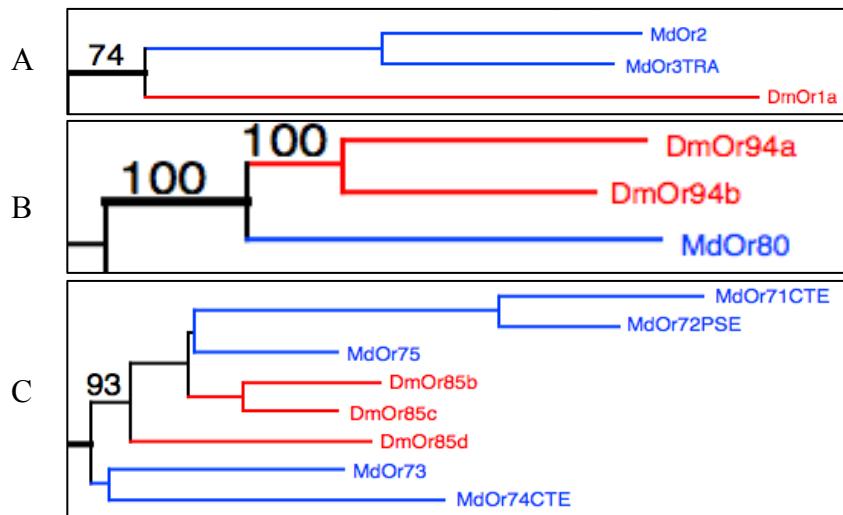


Figure 2. A duplication of DmOr1a in *M. domestica* (A) and the duplication of MdOr80 in *Drosophila* (B). (C) shows a complicated relationship between DmOR85b-d and MdOr71-75. (Branch lengths across the three diagrams are not on the same scale. See Figure 4 for true lengths.)

D. melanogaster contains nine genes related to MdOr22. There is an expansion of MdOr24-33 related to DmOr45a (Figure 3). DmOr67d has expanded to MdOr53-64 in *M. domestica*. Though some of the duplicated genes are in tandem arrays, these expansions are likely old events as each species appears to have undergone considerable gene movement since the separation of these two fly lineages. MdOr24-30 are in four different scaffolds but have the potential to be in a single tandem array save for their apparent move from the other three tandemly arrayed genes (MdOr31-33), which are in microsynteny with DmOr45a.

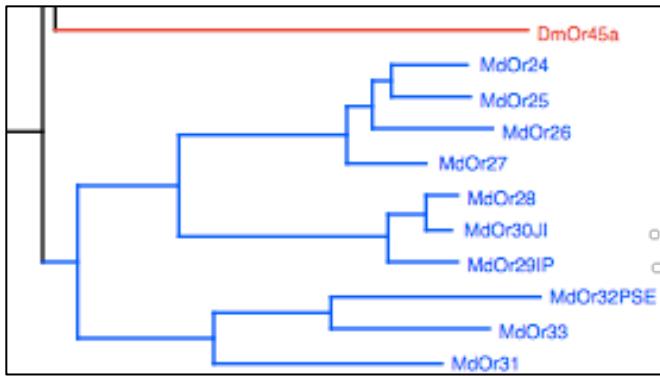


Figure 3. An expansion of a DmOr45a relative to 10 genes in *M. domestica*. (Branch lengths across the three diagrams are not on the same scale. See Figure 4 for true lengths and full relationship.)

As there are several highly divergent genes and lineages in one species without any clear orthologous relative in the other it is indicative of the loss of several genes and lineages from each species (at least 8 from *M. domestica*, 12 from *Drosophila*). These losses, plus several expansions in *M. domestica* leads to the relatively larger size of the OR family in *M. domestica*.

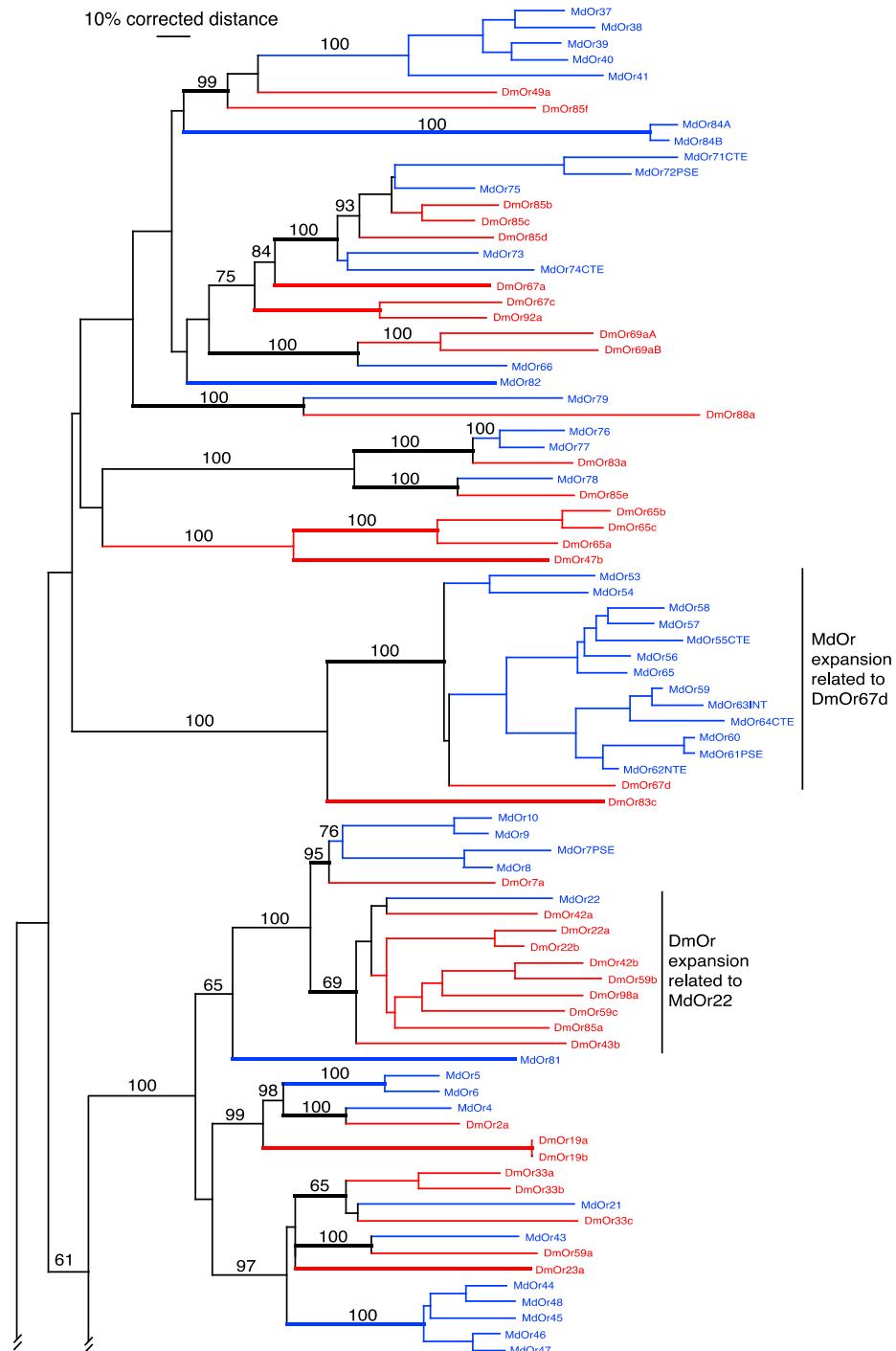


Figure 4. Phylogenetic tree of the *M. domestica* and *D. melanogaster* ORs. This is a corrected distance tree with the OrCo orthologs as the out-group to root the tree. The OrCo orthologs were declared as the out-group to root the tree, based on the basal position of this gene in the OR family in analysis of the entire chemoreceptor superfamily in *D. melanogaster* (Robertson et al. 2003). Comments on major gene lineages are on the right. The *M. domestica* and *D. melanogaster* gene/protein names are highlighted in blue and red, respectively, as are the branches leading to them to emphasize gene lineages. Bootstrap support level in percentage of 10,000 replications of uncorrected distance analysis is shown above major branches. Inferred ancestral and orthologous lineages are highlighted in double thickness. Suffixes after the gene/protein names are: NTE, amino terminus missing; CTE, carboxyl terminus missing; PSE, pseudogene; FIX, fixed assembly.

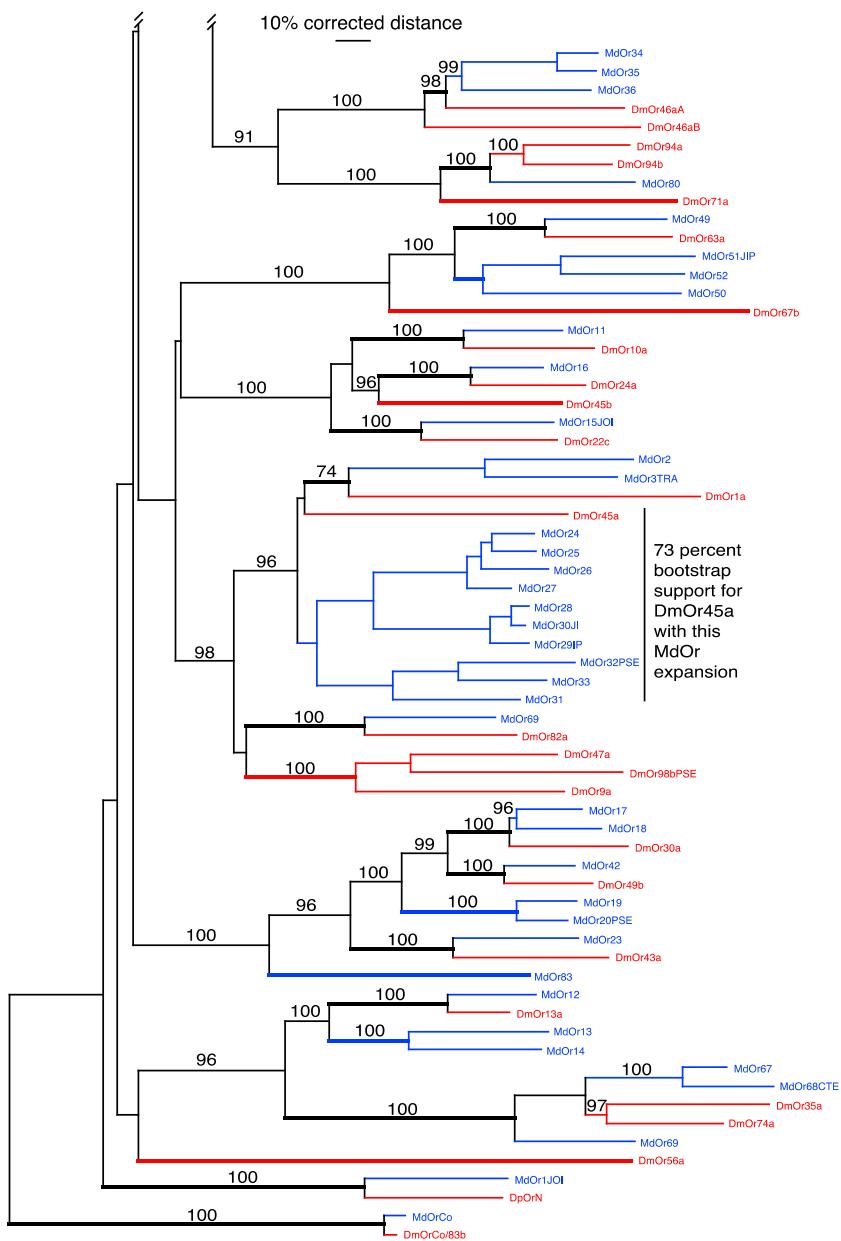


Figure 4. Phylogenetic tree of the *M. domestica* and *D. melanogaster* ORs (cont'd).

Ceratitis capitata results

C. capitata has a GR family that is intermediate in size between these two species with 73 gene models encoding 81 potential proteins through alternative splicing (Table 2). In *D. melanogaster* the GR family consists of 60 genes encoding 68 proteins via alternative splicing of some genes (Robertson et al., 2003). *M. domestica* shows a considerable expansion to 79 genes encoding 103 proteins (with 13 pseudogenes) (41). *C. capitata* has two relatively small species-specific expansions, independent of two relatively small expansions in *D. melanogaster*, compared to three large species-specific subfamily expansions in *M. domestica* (Figure 9).

The CcapGr gene set contains a few fragments, but no long pseudogenes were found. As with both *Drosophila* and *Musca* GRs, some of the genes (five in *C. capitata*) appear to have an unusual form of alternative splicing in which multiple alternative long first exons are spliced into a shared set of C-terminal exons downstream of the last long first exon in these tandem arrays. The resultant proteins differ considerably in most of their sequence, and hence presumably bind different ligands. They are indicated with a lower case letter after the gene number (Table 2).

The carbon dioxide receptors are highly conserved within most of the holometabolous insects, except the Hymenoptera to date, with two proteins represented by DmGr21a/AgGr22 and DmGr63a/AgGr24 constituting the functional receptor (Robertson & Kent, 2009). *Drosophila* species have, however, lost a third member of this subfamily, first recognized as AgGr23, which is present in insects as divergent as *Tribolium*, *Bombyx*, mosquitoes, and *Glossina*, and known in those species as Gr2 (Robertson & Kent, 2009). This gene is an ancient paralog of the DmGr21 or Gr1 lineage. *Musca* does not have this Gr2 lineage, but *Ceratitis* does, so it was apparently lost between the divergence of the tephritid lineage from that leading to the *Musca/Drosophila* split. The importance of this protein is debated, with one study (Lu et al.,

2007) finding that it enhanced perception of carbon dioxide, while another (Erdelyan et al., 2012) found that it did not. Like *Musca*, *Ceratitis* also has a recent duplication of the DmelGr21 or Gr1 lineage and, in an attempt to maintain the naming convention proposed by Robertson and Kent (Robertson & Kent, 2009), these are called CcapGr1.1 and 1.2, while the DmGr63a ortholog is called CcapGr3 (Table 2 and Figure 5). The tsetse fly contains the only other known recent duplication (Gr2 lineage) of a carbon dioxide receptor (Obiero et al., 2014).

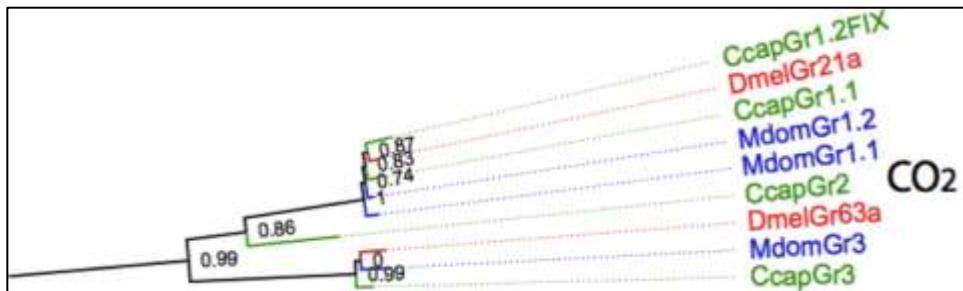


Figure 5. The duplications of DmelGr21a in *C. capitata* and *M. domestica*.

The medfly has a slightly smaller set of seven sugar receptor subfamily genes (Figure 6) as compared to eight in *D. melanogaster* (Fujii et al., 2015). These genes have been found to represent four major lineages, which duplicated in basal Diptera (Kent & Robertson, 2009). AgGr16 represents one lineage, which was lost from *Drosophila* and *Musca*, and is not found in *Ceratitis*. The other three lineages are each represented by two or three paralogs in *Drosophila*, specifically DmGr61a and 64a, Gr64b/c/d, and 64e/f and 5a, all of which are proposed to have once been in a large tandem array, with the terminal Gr61a and Gr5a genes moving from that array (Kent & Robertson, 2009). These genes were poorly assembled in *Musca* but appeared to have a similar arrangement (Scott et al., 2014). *Ceratitis* has all seven genes in a single perfect tandem array, confirming this model (Table 2). There is a loss of the DmelGr61a/MdomGr4 ortholog in *Ceratitis*, though there is a duplication of its ortholog of DmelGr5a/MdomGr5

(Figure 9). The effect of these losses and duplications of these sugar receptors is unclear, as they are expressed in complicated combinations in subsets of sweet receptor gustatory neurons in *Drosophila* (Fujii et al., 2015), and even a single change could potentially have substantial effects.

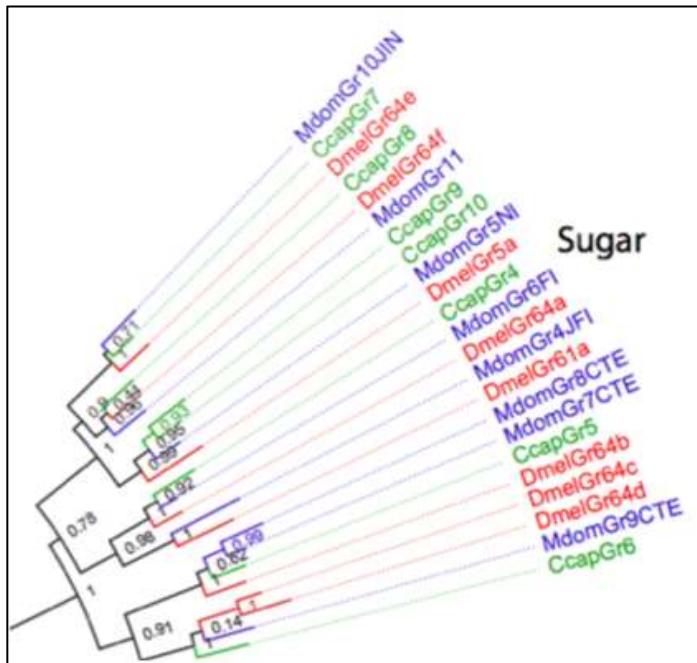


Figure 6. The sugar receptor family contains only 7 genes in *C. capitata* as compared to 8 in *D. melanogaster* and *M. domestica*. (Branch lengths across the three diagrams are not on the same scale. See Figure 8 for true lengths and full relationship.)

The fructose receptor (Miyamoto et al., 2012) and nutrient receptor in the brain (Montell, 2009), DmGr43a, is conserved among Diptera. In *Musca*, there is a species-specific duplication (MdomGr12/13), and in *Ceratitis*, it has further expanded to four genes (Figure 7).

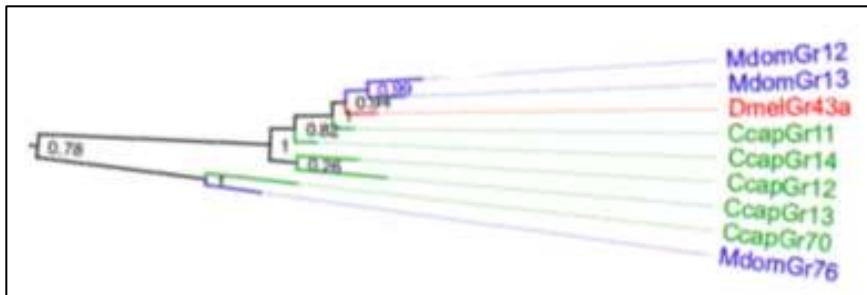


Figure 7. An expansion of the fructose receptor, Gr43a, to 3 genes in *M. domestica* and 5 gene in *C. capitata*. (Branch lengths across the three diagrams are not on the same scale. See Figure 8 for true lengths and full relationship.)

Unlike the multiple novel duplications in *Musca* (MdomGr39a-g) (top of Figure 9), the alternatively-spliced *DmelGr28b* locus ortholog is fairly simple in *Ceratitis* (CcapGr41a-e). The same five alternative-spliced forms are found in both *Drosophila* and *Ceratitis*.

Most of the remaining *Drosophila* GRs are implicated in perception of bitter tastants (Shim, et al., 2015) or have not been functionally characterized. Orthologs of these GRs were named sequentially in *Ceratitis*, with no implications of orthology with the *Musca* genes. There are examples of simple orthologs across all three species (*DmelGr2a/MdomGr16/CcapGr24*, *DmelGr10a/MdomGr42/CcapGr50*, *DmelGr33a/MdomGr38/CcapGr42*, *DmelGr47b/MdomGr65/CcapGr71*, *DmelGr57a/MdomGr66/CcapGr72*, *DmelGr59f/MdomGr71/CcapGr64*, *DmelGr89a/MdomGr73/CcapGr66*, and *DmelGr93a/MdomGr74/CcapGr68*). *DmelGr66a*, a highly-conserved protein, has a simple conserved ortholog in both *Musca* (MdomGr36) and *Ceratitis* (CcapGr38), as well as a duplication of the gene in both flies (MdomGr36 and CcapGr38). Like *Musca* (MdGr14), *Ceratitis* has a single ortholog (CcapGr15) of the *DmelGr32a* gene. It also has four related genes (CcapGr16-19) that are related to *DmelGr68a*, but this lineage was lost from *Musca*.

Complex patterns of gene duplication and loss across the three species, as well as changes in synteny, characterize many of the remaining candidate bitter taste receptors.

CcapGr43-52 are in a mostly tandem array (two genes are in inverted orientation) on a single scaffold (Table 2). Similarly, CcapGr21-34 are two arrays on the same scaffold (Table 2). The expansion of CcapGr55-61 are also in two arrays on the same scaffold as one another (Table 2).

In the tree (Figure 9), most of the bitter taste receptors cluster together on the bottom half and upper left quadrants, except for the Gr66a and 47b/57a lineages, which cluster with the Gr28a/b and Gr43a (fructose) receptors.

DmelGr22a-f was independently amplified to seven genes in a split array in a single scaffold in *Ceratitis* (Table 2, Figure 8a), and DmelGr93b-d is an alternatively-spliced locus in *Ceratitis* (CcapGr67a/b) (Table 2, Figure 8b). Both of these lineages were lost from *Musca*.

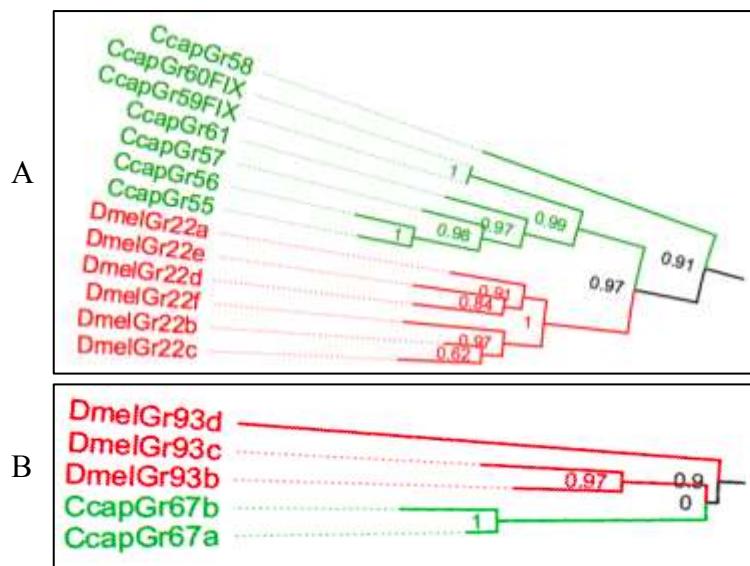


Figure 8. Independent amplification of DmelGr22a-f to seven genes in *C. capitata* (A) and the alternatively-spliced locus CcapGr67a/b as compared to Dmel93b-d (B). Neither lineage is found in *M. domestica*. (Branch lengths across the three diagrams are not on the same scale. See Figure 8 for true lengths.)

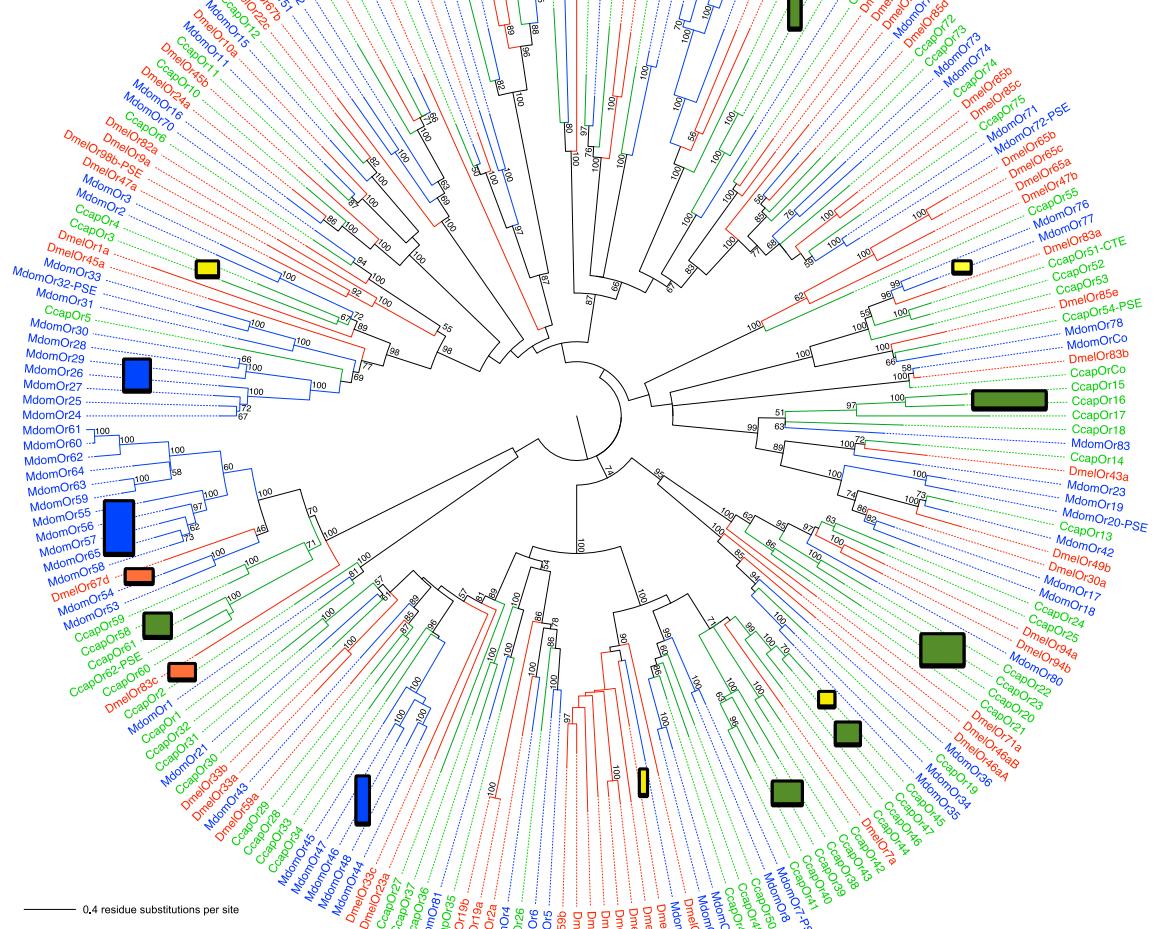


Figure 9. Phylogenetic tree of the *Ceratitis capitata*, *Drosophila melanogaster*, and *Musca domestica* GRs. This maximum likelihood tree was rooted by declaring the carbon dioxide and sugar receptor subfamilies as the outgroup. The *C. capitata*, *M. domestica* and *D. melanogaster* gene/protein names are highlighted in green, blue, and red, respectively, as are the branches leading to them to emphasize gene lineages. Clades discussed in the text are indicated on the outer edge. Suffixes after the gene/protein names are: NTE, amino terminus missing; CTE, carboxyl terminus missing; PSE, pseudogene; FIX, fixed assembly.

DISCUSSION

While functional characterization of genes in both *Musca* and *Ceratitis* is necessary to be sure of their roles, it is possible to extrapolate potential roles based on the known ligands in *Drosophila*, especially in well-characterized subfamilies.

Several large species-specific expansions among the ORs are the likely result of major changes in the chemosensory ability and ecology of each species. As an example, DmOr45a mediates larval repulsion from aversive chemicals (Cobb and Dannet, 1994; Hallem et al., 2004; Kreher et al., 2005), and it is possible that MdOr24-30 proteins serve a similar function. The MdOr53-64 gene expansion may be involved in pheromone sensing in *M. domestica*, as the related DmOr67d receptor is known to sense the male-specific pheromone 11-cis-vaccenyl acetate (Kurtovic et al., 2007).

While the expansion of the DmGr43a/MdomGr12/13 fructose receptors in *Ceratitis* appears on the tree to only include one gene that is truly orthologous (Figure 4), separate phylogenetic analysis of this lineage in diverse insects supports these genes actually being a species-specific expansion. The hessian fly (*Mayetiola destructor*) (Anderson et al., 2014) and the silkworm (*Bombyx mori*) (Wanner & Robertson, 2008) have both duplicated this lineage, and *Tribolium castaneum* has 10 paralogs (Richard et al., 2008), suggests duplications in this lineage are not uncommon. This may be an example of subfunctionalization, though the specific role of these duplications is not clear.

The various splice forms of the DmelGr28b locus are expressed in gustatory cells as well as in the brain and elsewhere (Montell, 2013), and some have also been implicated in perception of light and of warmth (Montell, 2013; Ni et al., 2014). While there was not an expansion in *Ceratitis*, the expansion in *Musca*, as well as the unusual expression in *Drosophila*, highlights

this locus as one of potential interest for understanding ecological and behavioral differences in flies.

The duplication of the DmelGr66a protein, which is required for detection of caffeine and other bitter tastants (Lee et al., 2009), indicates a loss from the *Drosophila* lineage. The lost gene was presumably also involved in detection of bitter tastants in *Drosophila* and potentially is for the two other species, as well.

DmelGr32a is involved in courtship through expression in a small set of gustatory receptor neurons on the male foreleg and mediates rejection of non-conspecific females as targets of courtship (Miyamoto & Amrein, 2008). The related DmelGr68a gene is also a candidate pheromone receptor expressed on the foretarsi of males (Bray & Amrein, 2003), and the expansion of this lineage in *Ceratitis* (CcapGr16-19) indicates a more complex set of cuticular hydrocarbons and other pheromones involved in sex and species recognition in *Ceratitis* (Siciliano et al., 2014).

In *Drosophila*, DmelGr10a/b, DmelGr26a-c, DmelGr47, and Dmel59a-d are on four different chromosome arms. However in both *Musca* and *Ceratitis*, the relatives MdomGr42-64 (Scott et al., 2014) and CcapGr43-52 are in mostly tandem array on a single scaffold. This situation presumably reflects higher genomic flux in the *Drosophila* lineage after the separation from the *Musca* lineage. CcapGr21-34 are on two arrays on the same scaffold, but the related *Musca* expansion (Mdom16-19, 22-29, and 41) are on several scaffolds (Table 1). In *Drosophila*, the relatives (DmelGr22a-f and 85a) are on two different chromosomes.

The addition of *Ceratitis* GRs to the comparison of *Drosophila* and *Musca* corrected some of the several instances of possible gene loss in one or the other species amongst the candidate bitter taste receptors, showing them to be artifacts of the phylogenetic analysis of these

sometimes highly-divergent proteins. DmelGr39b and MdomGr19 now cluster in the tree. The addition of *Ceratitis* also lends support to and improved understanding of some of the other gene losses. Both *Musca* and *Ceratitis* appear to have lost DmelGr9a and 10b, which given the relationship of the three species to one another, indicates independent losses.

The loss of several DmelGr22a-f/CcapGr55-61 and DmelGr93b-d/CcapGr67a-b lineages in *Musca* may indicate they have a common role of fruit tasting, as *Musca* does not require this role. Similarly, *Musca* has lost the DmelOr7a lineage, a receptor for several fruit odors in *Drosophila* (Hallem & Carlson, 2006) which has been expanded to 10 genes in *Ceratitis* (Handler et al, 2016, in press in *Genome Biology*).

In summary, while maintaining a conserved set of receptors for carbon dioxide, sugars, most bitter tastants, and light and warmth, with an expansion of fructose receptors, *Ceratitis* is more similar to *Drosophila* than *Musca* in maintaining and expanding receptors that might mediate perception of fruits, while being differentiated from both in having expansions of receptors implicated in perception of courtship chemicals. This may be due to similar ecologies, despite *Ceratitis* being more basal phylogenetically than either *Musca* or *Drosophila* (Wiegmann et al., 2011).

As with *Drosophila* species, similar ecologies appear to be an important driver for gene family evolution. While *D. melanogaster*, *M. domestica*, and *C. capitata* are all generalists, *Musca* stands out in its role as a filth fly when compared to *Drosophila* and *Ceratitis*. Further exploration of losses and expansions via functional characterization and Ka/Ks ratios may offer important insight into the evolution of chemoreceptors across these species, as well as providing a base for which to further understand evolution of specialist versus generalist dipterans.

TABLES

Table 1. Details of MdomOR family genes and proteins. Columns are: Gene – the gene and protein name we are assigning (suffixes are PSE – pseudogene; FIX – assembly was repaired; JOI – gene model spans scaffolds; NTE – N-terminus unidentified; CTE C-terminus unidentified; INT- internal regions unidentified; multiple suffixes are abbreviated to single letters); Ortholog – the *Drosophila melanogaster* ortholog, if relevant; OGS – the official gene number in the 17508 REFSEQ proteins (prefix is XP_00); Scaffold – the genome assembly scaffold ID (amongst 20,487 scaffolds in assembly v2.0.2); Coordinates – the nucleotide range from the first position of the start codon to the last position of the stop codon in the scaffold; Strand – + is forward and - is reverse; Introns – number of introns in the coding region; AAs – number of encoded amino acids in the protein; Comments – comments on the OGS gene model, repairs to the genome assembly, and pseudogene status (numbers in parentheses are the number of obvious pseudogenizing mutations).

Gene	Ortholog	OGS	Scaffold	Coordinates	Strand	Introns	AAs	Comments
OrCo	OrCo/83b	5175278	1259	14618-39294	+	6	478	Fine as is
Or1JOI	DpOrN	-	1870	<1-704	-	5	410	Join across scaffolds
		5190411	3803	1138->12456	-			
Or2	Or1a	5191432	6947	296-4448	-	4	397	Adjust intron
Or3FIX	Or1a	5192136	926	7639->8205	+	4	394	Fix assembly
		5192135	926	<1-1303	+			
Or4	Or2a	5188267	2	1376434-1380567	-	2	403	Fine as is
Or5	-	5187291	19768	143389-145795	-	2	409	Fine as is
Or6	-	5187290	19768	138355-141817	-	2	392	Fine as is
Or7PSE	Or7a	5190908	469	31864-35096	-	2	301	Pseudogene (2)
Or8	Or7a	5190907	469	18390-27574	-	3	395	Fine as is
Or9	Or7a	5190906	469	1710-3196	-	3	435	Fine as is
Or10	Or7a	5189494	2052	6088-7581	+	3	437	Fine as is
Or11	Or10a	5178484	18678	341323-347596	+	4	404	Fine as is
Or12	Or13a	5191323	622	48571-53025	-	4	429	Fine as is
Or13	-	5189591	2206	16714-25745	-	5	458	Fine as is
Or14	-	5189590	2206	850-7860	-	5	461	Multiple changes
Or15JOI	Or22c	5187358	19788	164641->167811	+	5	398	Join across scaffolds
		5191818	840	<1-15841	+			
Or16	Or24a	5181476	18895	47341-53063	-	5	399	Fine as is
Or17	Or30a	5181900	18939	276808-280725	-	5	373	Fine as is
Or18	Or30a	5187208	19749	35029-43469	+	5	373	Fine as is
Or19	-	5190189	3153	9025-11972	+	4	390	Fine as is
Or20PSE	-		20052	61513-67014	-	4	378	Pseudogene (1)
Or21	Or33a-c	5185739	19448	85849-87055	+	1	372	Fine as is
Or22	multiple	5187966	19928	114212-116248	+	2	433	Fine as is
Or23	Or43a	5175768	172	5489-13041	-	6	375	Remove final exon

Table 1. Details of MdomOR family genes and proteins (cont'd).

Or24	Or45a	5184443	19219	250742-255417	+	4	409	Fine as is
Or25	Or45a	5191600	707	52120-53601	-	4	409	Fine as is
Or26	Or45a	5191599	707	45043-49510	-	4	410	Extend N-terminus
Or27	Or45a	5191598	707	28376-37151	-	4	410	Remove double exon
Or28	Or45a	5191597	707	21161-23884	-	4	401	Fine as is
Or29IP	Or45a	-	707	17156-18697	-	4	379	Pseudogene (1)
Or30JI	Or45a	-	6047	<1-630	-	4	392	Join across scaffolds
		-	11325	2175->2947	-			
Or31	Or45a	5176399	18582	264366-270628	+	4	412	Fine as is
Or32PSE	Or45a	5176400	18582	279858-283416	+	4	401	Pseudogene (1)
Or33	Or45a	-	18582	286433-293007	+	4	401	New gene model
Or34	Or46aA/B	5190201	3215	667-4827	+	2	390	Fine as is
Or35	Or46aA/B	5180017	18775	6136-10301	+	2	387	Change intron
Or36	Or46aA/B	5190949	4876	3163-7162	-	2	388	Extend N-terminus
Or37	Or49a/85f	5180070	18780	379848-399553	-	3	405	Remove an intron
Or38	Or49a/85f	5180069	18780	367541-370610	-	3	416	First half of model
Or39	Or49a/85f	5180069	18780	362526-365181	-	4	417	Second half of model
Or40	Or49a/85f	5180068	18780	355026-357699	-	3	405	Fine as is
Or41	Or49a/85f	5180067	18780	350440-347195	+	3	415	Extend N-terminus
Or42	Or49b	5179498	18733	464-7606	+	5	371	Fine as is
Or43	Or59a	5184787	19276	22286-28146	+	1	398	Fine as is
Or44	-	5182355	18985	218891-220146	+	1	381	Multiple changes
Or45	-	5182356 /7	18985	224017-225238	+	1	385	Join two models
Or46	-	5182358	18985	228629-231719	+	1	381	Extend N-terminus
Or47	-	5182359	18985	235409-238825	+	1	379	Fine as is
Or48	-	5186318	19571	23826-25032	+	1	381	Fine as is
Or49	Or63a	5178182	18661	230762-246942	-	9	415	Fine as is
Or50	-	5182424	1899	14153-21208	-	8	418	Fine as is
Or51JP	-	-	1899	24301->37688	+	9	374	Join across scaffolds
		5189964	282	<1-5485	+			Pseudogene (2)
Or52	-	5189964	282	9213-18804	+	9	415	Second part of model
Or53	Or67d	5191896	86	43081-47468	+	4	392	Fine as is
Or54	Or67d	5191897	86	59515-63895	+	4	391	Fine as is
Or55CTE	Or67d	5190382	371	<35334-39602	-	2	283	C-terminus missing
Or56	Or67d	5190383	371	45516-50769	+	3	389	Remove an exon

Table 1. Details of MdomOR family genes and proteins (cont'd).

Or57	Or67d	5190384	371	55524-57553	+	3	401	Fine as is
Or58	Or67d	5190385	371	60142-64096	+	3	392	Change N-terminus
Or59	Or67d	5184154	19194	200473-205423	+	3	401	Fine as is
Or60	Or67d	5187726	1986	5170-6520	-	3	393	Fine as is
Or61PSE	Or67d	5187725	1986	551-1914	-	3	393	Pseudogene (1)
Or62NTE	Or67d	5187292	19769	<155517-156592	+	3	315	N-terminus missing
Or63INT	Or67d	-	19329	84501-94471	+	3	340	Internal exon missing
Or64CTE	Or67d	5185047	19329	100221->101311	+	3	320	C-terminus missing
Or65	Or67d	5176290	18578	784217-787484	+	3	392	Add N-terminal exon
Or66	Or69aA/B	5180133	18785	70492-77903	+	4	414	Fine as is
Or67	Or35a/74a	5188535	20038	58824-61223	-	4	405	Fine as is
Or68CTE	Or35a/74a	5188534	20038	<52090-53182	-	3	325	C-terminus missing
Or69	Or35a/74a	5185292	19367	744621-746455	+	4	404	Fine as is
Or70	Or82a	5189776	254	14537-21410	-	5	382	Extend N-terminus
Or71CTE	Or85b-d	5186501	19612	<131090-132093	-	1	314	First part of model
Or72PSE	Or85b-d	5186501	19612	117647-123777	-	3	407	Pseudogene (1)
Or73	Or85b-d	-	19612	109012-113608	-	3	430	New gene model
Or74CTE	Or85b-d	5186500	19612	27601-33931	+	3	431	Last exon missing
Or75	Or85b-d	-	19158	246021-256220	-	3	422	New gene model
Or76	Or83a	5189143	20298	44324-57416	-	5	453	Fine as is
Or77	Or83a	5189142	20298	30730-38016	-	5	480	Fine as is
Or78	Or85e	5182622	19003	148795-151719	+	4	462	Fine as is
Or79	Or88a	5179127	18710	611653-616793	+	3	409	Fine as is
Or80	Or94a/b	5185000	19322	151735-155579	+	3	394	Fine as is
Or81	-	5180861	18837	332652-347139	-	2	393	Fine as is
Or82	-	5181716	18920	328080-337530	-	4	399	Fine as is
Or83	-	5186825	19662	78318-83939	+	6	381	Fine as is
Or84A	-	-	18719	42819-57451	+	3	354	Alternatively spliced?
Or84B	-	5179301	18719	54021-57451	+	3	354	Fine as is

Table 2. Details of CcapGR family genes and proteins. Columns are: Gene – the gene and protein name we are assigning (suffixes are PSE – pseudogene; FIX – assembly was repaired); Ortholog – the *Drosophila melanogaster* ortholog, if relevant; Scaffold – the genome assembly scaffold ID; Coordinates – the nucleotide range from the first position of the start codon to the last position of the stop codon in the scaffold; Strand – + is forward and - is reverse; Introns – number of introns in the coding region; AAs – number of encoded amino acids in the protein.

Gene	Scaffold	Coordinates	Strand	Introns	AAs
Gr1.1	NW_004522754.1	2538981-2541774	-	3	453
Gr1.2FIX	NW_004524179.1	616410-619268	-	3	457
Gr2	NW_004522955.1	907124-961389	+	6	418
Gr3	NW_004523769.1	2394633-2398158	+	6	489
Gr4	NW_004524024.1	276530-286395	+	7	422
Gr5	NW_004524024.1	289459-300345	+	8	416
Gr6	NW_004524024.1	308241-319444	+	8	420
Gr7	NW_004524024.1	322859-334335	+	8	433
Gr8	NW_004524024.1	345036-355741	+	8	506
Gr9	NW_004524024.1	359168-371927	+	7	464
Gr10	NW_004524024.1	376169-383933	+	6	414
Gr11	NW_004523199.1	807240-816067	-	11	421
Gr12	NW_004523199.1	824877-833605	+	11	395
Gr13	NW_004523199.1	837796-847745	-	11	402
Gr14	NW_004523199.1	851100-864679	-	11	417
Gr15	NW_004523845.1	205555-207051	-	3	437
Gr16	NW_004524079.1	1250919-1252448	+	3	443
Gr17a	NW_004524079.1	1254576-1258977	+	3	379
Gr17b	NW_004524079.1	1256140-1258977	+	3	429
Gr18	NW_004524079.1	1261932-1263539	+	3	446
Gr19	NW_004524079.1	1266078-1270355	+	3	443
Gr20	NW_004523642.1	688336-694829	+	3	409
Gr21	NW_004524356.1	2552687-2555918	+	2	424
Gr22	NW_004524356.1	2560080-2561917	+	2	375
Gr23	NW_004524356.1	2562016-2565166	-	2	438
Gr24	NW_004524356.1	2566717-2570944	-	4	417
Gr25	NW_004524356.1	2576218-2577563	-	2	403
Gr26	NW_004524356.1	2579304-2580819	+	2	396
Gr27	NW_004524356.1	2582966-2584281	-	2	399
Gr28	NW_004524356.1	2586065-2587412	+	2	404
Gr29	NW_004524356.1	2589835-2591633	+	2	403
Gr30	NW_004524356.1	2593164-2594467	-	2	397

Table 2. Details of CcapGR family genes and proteins (cont'd).

Gr31	NW_004524356.1	2597365-2598731	-	2	411
Gr32	NW_004524356.1	2601901-2603262	+	2	403
Gr33	NW_004524356.1	4729555-4732392	+	5	396
Gr34a	NW_004524356.1	4719310-4726698	-	4	390
Gr34b	NW_004524356.1	4719310-4723149	-	4	390
Gr35	NW_004524112.1	2125168-2127267	+	3	405
Gr36	NW_004524112.1	2129067-2132367	+	3	405
Gr37	NW_004524112.1	2134777-2136158	+	3	379
Gr38	NW_004523802.1	5867855-5871692	+	5	525
Gr39	NW_004523913.1	6812795-6816553	+	5	496
Gr40	NW_004523676.1	890106-897989	-	4	483
Gr41a	NW_004524023.1	9289156-9311740	-	3	441
Gr41b	NW_004524023.1	9289156-9306998	-	3	467
Gr41c	NW_004524023.1	9289156-9298844	-	3	472
Gr41d	NW_004524023.1	9289156-9294457	-	3	439
Gr41e	NW_004524023.1	9289156-9292399	-	3	448
Gr42	NW_004524023.1	9286477-9288048	-	3	451
Gr43	NW_004524212.1	2960679-2961953	+	1	402
Gr44	NW_004524212.1	2957329-2958615	-	0	428
Gr45	NW_004524212.1	2955262-2956508	-	1	394
Gr46	NW_004524212.1	2953066-2954355	-	1	392
Gr47	NW_004524212.1	2949693-2951060	-	1	425
Gr48	NW_004524212.1	2948143-2949535	+	1	436
Gr49	NW_004524212.1	2933559-2934826	-	1	402
Gr50	NW_004524212.1	2882872-2884177	-	1	408
Gr51	NW_004524212.1	2880687-2881968	-	1	404
Gr52	NW_004524212.1	2878439-2879951	-	1	408
Gr53	NW_004523199.1	4152001-4153685	-	1	469
Gr54	NW_004523199.1	4167641-4166385	-	1	400
Gr55	NW_004523680.1	206055-209471	-	1	394
Gr56	NW_004523680.1	203007-204368	-	1	430
Gr57	NW_004523680.1	199138-200373	-	1	388
Gr58	NW_004523680.1	748826-753133	+	1	434
Gr59FIX	NW_004523112.1	540-1307	+	2	256
	NW_004523680.1	771367-771936	+	NA	165
Gr60FIX	NW_004523680.1	776656-777901	+	2	347

Table 2. Details of CcapGR family genes and proteins (cont'd).

Gr61	NW_004523680.1	786169-787409	+	1	389
Gr62	NW_004523814.1	1342584-1345881	-	2	437
Gr63	NW_004523814.1	1350456-1352809	-	2	433
Gr64	NW_004523814.1	1356884-1360185	+	3	420
Gr65a	NW_004522866.1	5997294-6010015	+	1	486
Gr65b	NW_004522866.1	6003391-6010015	+	1	476
Gr66	NW_004523714.1	2082558-2083767	+	1	383
Gr67a	NW_004523968.1	1996565-2005973	+	1	410
Gr67b	NW_004523968.1	2001212-2005973	+	1	407
Gr68	NW_004523968.1	2043588-2046271	-	1	408
Gr69	NW_004524379.1	2209083-2210393	-	2	397
Gr70	NW_004522889.1	271092-281773	-	3	454
Gr71	NW_004523692.1	354117-355701	-	2	412
Gr72	NW_004523836.1	3169564-3171000	-	2	409

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APPENDIX

87 MdOr proteins in FASTA format

>MdOrCo

MQANLQPTKYTGLVADLMPNIKLMKYSGLFMHAFTGGSALLKNVYSSIHLVLIVLQFI
FILVNMALNADEVNELSGNTITALFFTHCITKFVYIAVNQKNFYRTLNIWNQPNSHPLF
AESDARYHSIALAKMRKLFLVMLTTVASAVA WITITFFGESVKFATDKETNSTITVPIP
RLPIKSFYPWDASSGMFYMISFGYQAYYLLFSMVHSNLCDVLFCSWLIFACEQLQHLK
GIMKPLMELSASLDTYRPNSAALFRSLANSKSELIQNEEKEPVNDLDMSGIYSTKADW
GAQFRAPSTLQTFTNGINGGNPNGLKKQEMMVRSIAKYWVERHKHVVRLVAAGDTY
GAALLLHMLTSTIKLTLLAYQATKITGVNVY AFTVIGY LGY ALAQVFHFCIFGNRLIEE
SSSVMEAAYSCHWYDGSEEAKTFVQIVCQQCQKAMSI SGAKFTVSDLFASVLGAV
VTYFMVLVQLK

>MdOr1JOI

MKDKFKTFMRDFFPSNVEKG EIGSVKLNIWLAQITGVPIIGLKDESSLIKNLILLYGIFTT
TVVTIFIYTGFEMYDLYMNWHDLDSL TQNTCLSLTHVSGAIKTVNIIFHLPRLEGVIRKL
KHVTKTYIKSEKQLVVFYDGEVENKLVL SIYIGIVGFTGF MGMIMLYMPEAVAGKIFPY
RVL PDWMPQQQLLYMGLS VIFAIQIIAVDYL NVTIINQIRQLN ILNLA FDDLIVETQ
ANSRET KSLVLYKDDPVKR MDSIVEHH CLLGELRQETEDIFS QPILWQFMTS VIIFAMT
GFQATVRSSGSSAAVLIYACGCICELFVYCWFGNEVSEQSKTLGTSGFHSSWYHFD
RRYGKSLI FLTNAQRPFVFTAGGF MGLSLPSFTGILSKSY SYI ALLRQIY GK

>MdOr2

MYNNVDGKTRQDLEFLDVQYRALIRVGL DIGAIRGKD FLNDRGKF LIYGIITTYLQYGL
ILFAVHIFGVQIDKASAALSMFNQGSLLMLKV SILIFKSNRLLKLIWDMNLLATMANEP
ERETWLSENRF SKVIGNIYSTACIASVILSISIPII FMSYEHF KGLEVSLKLPFDGEFP YEHL
GIPIFILNYILSVIYVYTLLCWTIGIDLFGWLIHAVSGHF RILRLK VEMA AKKIDE HGNH
LDFVQD IGAIVRYHIKT LGFDALNEIFGQI FWAEVA FSCLQMCFLIFTLNNGSDKRMIP
FNAMVFTAISIQMMIYCFGGEKIKSENEMFCFDIYSKFPWEK MYPSEKRMMLPLQRS
QQDAALRGLFFELDRNLLVYIYRTAFSYNTLLGAMKE

>MdOr3FIX

MSDTERQNLDYLPVQFGAFMVLGL DIGVTRRS ALLKSGWTFLFNILCTVMEYGFANF
VINSITDIDAITSSLSMFNQGM LTFKVLVMVFKGDEMLKLIWDMNRLARGANAKEW
EIWISENRMGKWI ALGYYCCYIAATIMA VMPWLFMLYEYVQGRGVH LRLPFQLQFF
FVSGNGFHISI YYIGTLLV VRAWF NMSVGIDTLFGWYI FAVSGHFRILRH KIKETALKI
DAYDNHRDFVSDVAAFVSYHNRTLKF TENLNRLYGEILWSEISM SCLQLCFLLYSLTN
DENFANIPFHFFASAAITMQLMIYCFGGEKLNENDMLCHDIY MAMPWEK MYPSEKK
LMLLPLLRTQREISLKGLYFVINVNLLVIFKTA FFSFITLLGAMKEI

>MdOr4

MTNALTDNNKNIYSKLD TNVAF EYHW KVWRWTG IKPPQDMNPQLYRLYAI VLNFLA
TVLFPLSLIANVFFTQNLQQLCENLTITISDCQSNLKFINVFLVRHQLDRIKSILRRLDR
VQDDKEFAVLKSAIATARSSFLIFFRLYSFGTT SVVKVALAESRSLLFP AWFGV NWDG

NLSTYVVVIVYQFFGLAVQALQNVANDSYPPAYLVILSAHMRALEIRVKAVGQFRQEG
MQQPLTLSAEEQAKCLKEFNECIKDYLNILKLHSIIQRIISKACLAQFACSLVQCTVGL
HFMYVVDAANYEAQLMSIIFVAVTLEAFVICYFGHMMMSLQSSNLTYAFYSCGWLAQ
SPEFKRNLIITLMRTQRTSTIRAGSYIPVDLPTFVVLMKYAYSVFTLLIRFK

>MdOr5

MALQPMASSSSASNKIHTWQAFRNHWILWKFCGLHPPKRNSRWFNPYLIYAIVLNVT
TTLMFPITLIVDLILSQNLTELCENLVYTITDVICSLKFNFITVRHKLLEVWRWILERLDVR
ATTPEQRQELRHGIQTSHKWFMAFFRFYTCAVITSQLVVYLSKERVLMPYPSWFPWDW
KASKRNFIAHCYQVYTVSVQTVQNLGSDTYPQAYIVVILAHIRALGLRIKALGEALS
ATAAGDVSSPSSSKKLSDDELYRELVNCVKDHQIVHELYLTIQECISKTCLAQFVATG
LAQCTIGVYIYVGSDFSRLLNSFMFFGAITIEILLCYFGDLYCRANDFLIDAIYDCNWID
KDERFKKALLLLQRSQQADCLKAGNLIPVRLPTFVKIMKTAYSAFTVLNEVN

>MdOr6

MSVLFSHPNTWEAFKYHWLLWKWCGLQPPSRDSKWFRPYLAYAIIFNLTTILFPLSL
VLDLTLSQLTEIFQNLVVTVTVFSSLKFVNVLIRRKLLEVRFLLERLDVRANTEEQ
QQELKNGIAMAHKCFMIFRLYVCAITSQLVVYFSSERVLMPYPSWLPWDWRESKRY
FLFAICFQIYAVSAQLSQNLGNDTYPQAYIVVILAHIRALALRIKHLGVVSTVPAGEGKL
SQEDFYRELRQCVDHEHVHELYLTIQECLSTTCLAQFIATGLAQCIIGVYIYVGDDFS
RLLNSLVFFGAVTIEILVLCYFGDLYCQANEFLIDAIYATNWMDRDGRFKKALLVLQ
RAQVTNCLKAGNLTPVMLPTFVTIMKTAYSVFTVLNKVN

>MdOr7PSE

AVKISKKVATKQALTNYICFRVVGIGHVTKSNPPLYIVYAIVIHSLLTVFTPISFTSYFR
KTDQDFNVGVFLTSIQAVINVYGCAIKILLIYYKTKLEAAEKLMDKMDQHCRAEDEI
QUELFNIIDLGRKIVLGYITAYWTYTTMTYISALVSGVPSYSINLFFLDWKRSKREFYLAS
FLEYVLVTWTCLQQVANDSYGTIYVCILRGHVRVLLRIRKMGRKVDQTADQNLEEL
KSCIKDHDLLLELYNIISPVISRTIFLQFSITAVILGITLIXIAKFSFSLYTLIKQMGIKERLG
L

>MdOr8

MSKQTVKVIKKVATKQALTLYGCFRVMGIHFTKSHTHLYLIYVIVIHSLLTVFTPISFT
TSYFRKTDENFNMGVFLTSVQAIVNVYGCIVKIFFLVYYKKLEAAEKLMDQMDQHC
QADDEIQEIYNIRNLGRRIIIGYGIAYWYIYTMTYISALASGVPSYSNLFLIDWRRSKLE
FYVASFIEYFLTSWTCFQQVANDSYGTIYVCILRGHVRVLLRIRKMGRKVNRTADQNL
EELKTCIKDHKEIELYNAISPVISRTIFLQFSITAAILGITLVNIAIFASSITAMAASAFYIV
AVSVEIFPLCYYANCLLYDSDTLLATEIFHSAWIGQDRRYRKMLIFFIQRSTQKSMELWAG
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>MdOr9

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CAIKIVLMYFLLPQLLQCDPVLERLDKRCTSPEEKDLVRRFISHGNRFVILFGMAYWSY
ASSTCISAVLFHRLPYNLYNPLL DATAKS FV LGF VEMMP MYLACS QQV V DDS Y A
VIYTQILRTHLKALVFR LQH L N DD HRN EN G VISPEAEERN IEN L KLC IID HKN II E LY TRV

APVISITLFVQFTITASLLGVTLINILIFATNTASIVASCFYVLAVVVEIFPLCYYAQCLMN
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>MdOr10

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VVGCSIKIILMYFLLPQLLKCEPIFERLDGRCTSREEKDLIRQFVHDGNRLVVLFTVAYW
SYSSSTCISAFLFGRLPYNIYNPFIDANASRGYFILAVFMEMVPMDIACFQQVVDDSYA
VIYTQILRTHLQALLIRLQHLNDDAADLDDEAQERNVEKLKCIIDHKSIIELYNRVAP
VISITIFVQFTITASLLGSTLINILIFATNTASIVASCFYVLAVVVEVFPLCYYAQCLMDEN
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>MdOr11

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DMEHKAVYNRIQSSVMAARVNFAFLAGFITCTAYNLKPLILVWIFWSKGKDLMWL
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VKLLFRKFEKGKLTLPTEAAAYVEQLILIKRHNVIEMTDFRKRYSIITLAHFVSASMVI
GASIFEMLTYTGFGRFIYLGYTVAALSQLAVCYGGTLVAENSIYLATVVFKCNWYIC
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>MdOr12

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QFVEDIWINKKQHPHVAAVCSRNMRTFRIMTVLLSCLISMVCVPLLVLFVDVGLDAD
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>MdOr13

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FPTVFPYDTQTHWAYIFTYIFLSYAGYIAVSLFYAMDAILAYFISFVAGQFEILHADIARL
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LFANYGTSTFLICFVGQFMIAGLKSFGDFMRFFMAVTGQLFIVCKLGNLLITQST
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>MdOr14

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PECHAEFLANYRGESTGSKKNDFIFLHNLYVKRLHELATAHDELIRFSMDLEKLFSQL
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>MdOr15JOI

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FPYFPGMFLILTASGVMTVFSFSIVDGYYVCTTVFICSIFKIIQQDIGSIFDELKDCEHATD
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>MdOr16

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LAMPIYPITFLLVHWHGYITVLSFVAGDGLFLGFCFYFSTLLKALQQDLTEVLGVIDET
KKYRKLTESEKVMSLSKIIRRNEIADLTMKLSSIMVEITLCHFITSSVIIGTSVIDLLLFA
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>MdOr17

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RFISKVNLLMGFTSCVGFNMYPFATSKVLPFGMYVPGVDKYESPYYQICFLFQIITPA
GCCMYIPFTNLIVSFILFGILMCKVLQHKLRLNLKDVSEKARTVIWCIKYQLQLINFVD
TINDLTTFTFLFEFMAFGAMLCAMLFLIIIVETVAQMCIICIYIFMIFAQSIVMYYFANEL
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>MdOr18

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NISRVNLVMGTCVISFLIYPIFATTKVLPGIYVPGVDKNISPYYEICFIVQTVMAPIGCC
MFIPFTNMIVAIMLFAILMCRMQRKLRHLCHVTSEEARATIWCICKYQTELIRYVNTIN
DLITYTNLLEFLAFGAMLCAMMFTLVTETVSQMCLICVYILMIFAQSTILYYYANKVF
DESLNVGTAAYESEWFVDVDVDTQRTLRLLILRAQKPCAILVGRVYPMNLELLQSLNT
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>MdOr19

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EYDFEIRQIIRKYSMDMVLKVSINKINTMGILTGLGFSMFPIMAEEREFIFGMYVPYLNEYQ
TPWYEILLA VQSVLNLSGMCTFIPFAGMFVSFLVFAMAISKVLQYKLSKLSTEISSKLAE
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>MdOr20PSE

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MMERDAEICRIMEDHTAMVLKISKINLIMGMLTTXEFIGYVPPYLEYQSPWYEILLTG
QSFLNLSAMCIFIATAMFISYFMFAIAISKVLQYKLSRTCTEVSSKIVEEKIVECIKLHLR
LISFIEQINELCGFIALMDFLFVIVLCIMLLSFVLVKTVTQKCIITVYISMVFTQAFLYY
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>MdOr21

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WAVLMYQFMGVTTQIVQNLVNDAPAGVLLCLISGHVRLGMRVSRIGHTDSKKTENEN
LADLGKLFKLVEDTQSYYQLILYISGGLNICVAVVYLIFFVESLTAYLYSAFILAITIEIY
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>MdOr22

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SSTFLVAVINSKPPYQVFNPFFLWKENVWKFTMQAGFEYMMIAFHCFQQALLDSPV
FITIIRTHLHILTRRISRLGSISTMTSDERYEALVQCVDHKNIMGLYSIFCPVISGTMFVQ
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>MdOr23

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SVLYMPFVSIFVTSSFGKTALQILQHRLKDIFEIYDDDETREALKCAHYYNRLTRFI
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>MdOr24

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KIIAHQAAGNHNEKEFLKSVVECNYHRRIIQMSERFSEVYQGLVFIKFLVSCMQLACL
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>MdOr25

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NIIAHKAMGNHNEEKFLKSIIDCVKYHRRIIQMSERFSEVYKLVFFKFLVSCMQLACLS
FIIPLGGEIADQLFNLSFLMAVTTQLMLYCHGGQKIQDMMSISVNWAIYESFHWHDLSIK
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>MdOr26

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MMVTHKSSEIQNDEIFMKSIVECIQYHLRILEISKRFSEAYQHLVLIKFLISCLQLACLSFI
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>MdOr27

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ERTMMLRAIGEHNEEKFITAIIECVKYHRRIIQMAERFNDVYKGLVFIKFLISCLQLACL
SFQIPSGGEIADLLFSLFLISVTTQLMLYCHGGQKIQDMSTSVSLAIYEHFQWHDLSVK
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>MdOr28

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VRGGDFAFGQVFHMLFLMSISIQMMLYCYGGQRIKDESASISVAIYEYFHWDLLCPKSR
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>MdOr29IP

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LEAKQEELEIIADENRXDTVKSFSFYMTVLTGILALTAPYYVDPKGSYLGYFTVHIWT

CIAIYAVLNTTLAADSLFSWIFHNISAHFAILRERLICVAFSETEGKQSYANLKKSLAEY
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>MdOr30JI

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KHSENGVFFTANSLFWVYKTAGSFVTLMMSVSDTSK

>MdOr31

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GYSLVYIWDFIYFIACSAVSTESMFSWLVCNIIAQFRILMHRLEVASRQVMSTRPMTA
SHHVDDDDDNPLMGELDPQAGMVDAIIACVKFHRRRTLRTQELNSLYGAIIFVKFIVSG
TQICCLAFHLVRGNNSLFNVAYLCMFLSAAALQLILYCYNGQRLKDESLLVTTKIYSIF
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>MdOr32PSE

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NSRDVLHCTVYGSASFSTGFFGILHRLLRPSSSTSNCWYHILPXYLWDYSHLP
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PEVSVSREQERLIFDCVRFHNRNLVQELNLVYGGIIFVKFVVSSVQICCSAFFLNSFG
ASQSMAKLMYQFLLSAVALQLMLYCYNGQRLTDVSFQVATKVYSTFPWSKMPASTK
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>MdOr33

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RKDIRICTLYCIVIYGTFFGMLPFARTILGYLRNGYLVYVTPVASPSLWNYDTVHGY
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EEQHRLINDCIRFHNRTLVLKELNRVYGAITFKFVVSSIQICCSVFFVSSDSKESAFN
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QRSCEMRGVFFRIDLSFVWVFKTAGSLIAVLQTIIDEAQ

>MdOr34

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VKIFRQTHRLCGTVRNFYLVISLCALNVVMLTQYIFDNSELPLSLYNPINIDTKLRYRLM
YLYQYVAVSICCYMNIAFDSISASFMIHIKGQLDILCDRLEHLGMDQESRDEDITRQLK

NCVKYYGDIIVRIAENLISFPISIQIACSVLVLVANFYAMSLSDPGDYANFIKFLIYQL
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NPTYTFNLAAFTSIVNSSYSYFALLKRINS

>MdOr35

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QLFRESQRLLARTVRNFYTTISLCALNALLFTQYIIDTQLPMSIYNPINTDTKLRFVLVYI
YQYLAWSVCCYTNIAFDSISASFMIHAKGQLDILCDRLKHLGMDSETSDEEITAQLKNC
VKYYGDIIVKIAEDLISFPISVQIACSVLVLVANFYAMSLSDFANFIKFLIYQLCMS
QIYILLYFPSEVTSKSEEVPYHLYCSKWANWSASNRKLTLMMTRFDIPIRIKSINPTYTF
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>MdOr36

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TEWQIYCQAIDLSRRVRNYYASLSVSALSALFLSQFLGDEQELPASIYYPFQLNTNWKY
GLMYVYQCVSLAILCFVNNGFDLTASFFINIKGQLDVLGMRLQTIGVGVRDQRRILKK
LKDCIRNYQRILRMTHLMEELVRIPMSVQIGGSVFVLIANFYSMSMLSDNADMGIFAKL
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SIKSINRCYSFNLAFTAIVNSSYSYFALLKRINS

>MdOr37

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YEPMEKRQQKDLKVMAELVRKHQVLLEADDLKEIFSLLVLVMLFSTVATLFCAAVY
VLTQGINKNVLGYMAFLPTSLGQYFMVCYYGQLIINKSLQIGEAAYSQTWYNGCQSY
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>MdOr38

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RFCEIFPISMEDRREYRVNDYYWPRWITCMVVQCGAIAVIIFSPAATLKDYFLAILKF
GFSDAKFSYHILYEEHTYIVDHQRPTGYIFIYSVLAGTQYAVIFNICPDIWLVAYAIQL
CMHFDYISRLENYPEKERSHKDLEVVAKLVKKHQILLDLRKTSILVLIIMLFS
TVVTLFGAAVYVLTQGINSVNLGYLAFLPTTLGQYFMVCYYGQLIINKSLRIGDAAYS
QTWYNGCQSYKKSILAILGRSQSQCEINAGGFQTTNLKAFEGVIRMTFQLFAVWRTLM
EPK

>MdOr39

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LLQRFRDIFPISREDRLAYRVNDYYWPKWITTLYMQLFALSIILFLPFVEAVYEYFGAL
LTVGYANAKFGYYRMYPETTYGINHYNPLGYIIVYTMDIMNGHYCTVWMMGPDVWL

VAFSIQLCMHFDYVSRTLENYKPSKERAQDLRVLAELVRKHQTVLELADDVQENFS
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FAIWSTMSSK

>MdOr40

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DRLAYRVDEYYWPKWITLIMQLCALTIILFVPAESIFEYVGALISLGYNAGFGYY
RMYEETSYGFHHNFLGYVVSYSLDVMNALYSAIWMICTDIWLVAFALQLCMHFDYI
SRTLENYEPHKERSQDDEKVLAGLVRKHQTILEADELKINFSLAIIVMLFSTISMLFGA
AELVLTQGITTHVIGLAYVPTSVGQFFMVCYYGQLIINKSLRVSEAAYSQTWYNGSQ
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>MdOr41

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LREIYPKAGRERILYRVNAFFWPWKWMLTVIYFYFGAVAFIVLSPLESVIVFVIGVGRLG
WNEAQFGYIKLYDIPYSFDHRSPFAYVLTYSIELFHAQFVIICNCVCGDIWLLCYAMQLC
MHLDYLIKILEHYEPRVEHHLRDTQFIAGFSQKHQILLNIADDVNTVFGVQLLLILISTA
ATICCAGIYTLTQGVGKELLEYVAFLPCVVGQYYLICFYGQRLVSSSENVGAAYNHA
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DKKN

>MdOr42

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VMARGNLFLGLLTCIGFGLYPLLAERVLPFGSIIPGIDEYQSPFYECWYVFQMLITPGV
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TVNRLTTFMFLLEFVAFGTLLCALLFLLIFVDSAAQAIIVCAYTMIFCQILALYWYANE
LKEQNLSIAAAAYETEWFTYEIPVQKLILLMIMRAQKPCTIKVGNIPMTLELFQALLN
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>MdOr43

MAPSMEINSNEFFKINRTCWKLLGLGMLMVEGHKTNGQRKMSTNLYMVWAIVINLM
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HHEEVHRFRQDIRSRIMNIQRLYFVVYCGVGISVLAFLSKEQRLFYSGWFPFDWRSS
LGNYAAAISYQCIPIFFQMMQTFCNDSFSPIALCVSAHIELLYMRVVRIGQDKNGKMR
ETTTLQEDEEELNRCVLDQMNLYELYNTMQNIISWAMFIQFFVSVVNNCVAIVALLFF
VTDVFERIYYVIYILAMGIQLFPTCYGSDFVLLFEKLHYAVFSCNWIGQSFSKRHM
IFTERSLRETVALAGGIFPIHLDFFGTCKATYSLFAVVMTMK

>MdOr44

MTEEPNTKALFKTHFIAWRILGMSPPDNYRPLYWIYSILLNIFVTIGYPLHLIFGLFTSTT
MYEIIQNVAINFTCSVACAMKTIAIWWRFNKVDVMFEIIQRQDQRFTSHEEIALRKEVY
PPVRRILLFSILCTFIGISGESAVLVTGLLTWNLMLMYKAYFPFDVFASTKNYMAAHLYQ

FIGISYLILQNVVNDTGFASHLCLRSQVRMLNIRVTKIGHDPKKSREENNQELLECIKV
HKDLLEYRRQLEIISIYMFFQILIAALNMCVVLVFIILFVRDIFTLAYYVSYLTSMIFEIL
PSCYYGTLLEDEFEDLAYALFSCNWPKQTLEFKKNLRIVAEQAKRRIYVTAWLFRINN
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>MdOr45

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MKNVVYPQVRFVIRLFYVICGFLSLFGELESVAGLLGNWRIQYKAYFPFDPYANTKN
YVIAHVYQLGVNFTLVQNIVNDTFASSHLALLRGQVDMALARVAKIGHDPQKTQRE
NNQQLLECIRDHEDLLEYRQILEEIISVYMFQILLCGLNMCVILVYMVIFVRNDVITLS
YYSTHLIGVMCEILPSCYYGTLLEDAFQDIAHALFSCNWMDQDLEFKKNLRIFIENSSR
RIYVTAWLFRINNNNAFIVACKNTYTLFALVMNLK

>MdOr46

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VYPPIRSLINLFYILCSMVALSAESSLIFNGLRGSWALMYQAYFPFDPGSSGNVVAHI
YQFIGIITYVTQNLVNDTFAGAHLSSLGGQVRLGMRVAEIGHDPKSLAENNKA
CIHDHLDLLEYRRKVEDVISLYMFQILFSSMNMCVVLVFMLLFVKDTFTMSYLFYF
VGMIFEVLPSYCFTILEDEFQELSYTLFKCNWADQNVVFKKNLRIFVEQASRRIHVTA
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>MdOr47

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MREKVHPIKWLILFYILCSMVAISAEVSLVVNGLGSWLLMYQAYFPFDPGSSMNY
AVAHIYQLIGLVYTQNLVNDTFAGANLSLLGGQVHLLGMRVANIGHDPNKSMEEN
NKELLD CIHDHLDLLEYRRKVEDVISLYMFQILFSSMNMCVVLVFMLLFVKDPFTMI
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RIEVTAWLFRINNNFLTAVKGSYSIFSL

>MdOr48

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VVHPPVRIILAFTILCSVIAASGESSIONLGNWTLMHKGYFPFDISNNTRNYAIAH
LYQIIGLSYMLQNVVNDTFAASHMCLLRGQVQMLNVRIAKIGHDPKKSREQNNQEFL
ECIKIHDKDLLEYRRQLEIISVYMFQILVAAFNMCII VFIILFVKDVFTLIYYILYFS
FEILPSCYYGTLLEDEFQDFAYALFSCNWPDQDVGFKKNLRIVAEFASRRIYVTAWLFR
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>MdOr49

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FPMSERLQAKIDEILDASWKNINGQLIFYICCCAAISNYFFMALFQNIYHTWKETPNYE
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KVLNMLIENSTSADVPKERRVEYLRYCIYQYQRISDYTDELKIYKHISLTQFLLSLLV
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>MdOr50

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LGIFDLAIDCKQYLLQEINTILDSSWRHIKYQVNFFTSCMMACGFYMFSCIAANYYYT
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VHCASLFEILHMLLEHVDDIPQSERVDYLLCCARLHVRIYNYYAKINGMYKNPSLAQC
VLSMLVLCVVMFMASIGLEEDITLFVKMLCFLCAAGLQIAIYCNGQKIITQSEKSPDA
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NLNGID

>MdOr51JIP

MYLALKETKANQILKYWKWIAFTSGCNIVYKTKFMKLFKLILNMSLAISAAIGCYGQA
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NLINPSKILKDISAIMDQSWSMSIKFQLNFFIGNVVVLGVYLFKNLILNIHNFKNEGDRF
QLAYAITFSGLFALISTHCRGLLRVLRTLITYSTTYHVLPEDRVKYLQGCIKLQQKIQKI
CNELNLSYRIPALAMFLVSCLVICLLTFYATVDGGNDISTIVKVILFISGAYFEVAIFCFN
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SSLSYFLFLQECM

>MdOr52

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ELIDSANIINDITQILNESWIDIKRQLLLLRTFGICSWYTGHSLVSNIYLYISDENDKE
KLEFPFPASFPVWYSNVNSLWHFYLEYFVVTMQIYLATVASITCSGLFSVISVHCLTML
RVLRTLITYSTSEHVPQSQHRTKYLEACVRLHQNLLSFCSRNRVYQKPSLGLFISCCLLI
CLLTFKASVDLGKDISHSIKVCLYLLAAFYELLIFCLNGQRTSESERLPQAIYSSLWFDE
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>MdOr53

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>MdOr54

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RYIKYLEHTSLIKRAVFSLNIYSIQTIGVLA VPLFYHLLGQQIDIALLVPGIDKHTDF
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VDRTEPLLREIFQWHQKSTMFAQNCTDTFFWVIFVQIFASTLAIICIMVCQFLGVWPAA
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>MdOr55CTE

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ERSGELLRDIFEWHQRYMKFISIVKENYFWVILVEMATIFLCLALSLS

>MdOr56

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RDCLQKSINTAVKCIKMAFFHVAITVGLIGVVPFFRFVNERIFVMQFQLPGVDGDTE
YGYLIMNCMHSICIIFGAFGNFAADLCFFTFSHFPLFKGILSCKFHDLNDVLEGSDDAK
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>MdOr57

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YFVMNCMHCICIIFGSFGNFAADLCFFFVVSHVPMFKDILTCKFHDLNDLLEEEVADNE
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FCLILGTWPGGQSAYCFIMLYIYCGLGTVEVTNDGFIDSCYTEIIWYKLPVSQRKM
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>MdOr58

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CCLQKSINTVNRFIKCMAIIHFSITMSLIAVVPFHRVVFNERIFVMQFLLPGIDPNTAYGY
LMMNCMHCICIIFGSFGNFAADLCFFTIVSHVPLFKDLLRCKCQDLNDILEEGKDVEQE
GIGDCQILLKDIQWHQKYMRYITTVKDNYFWVLIEMGTVALSLASTLFCLILGTWPG
GLTYLAYCLMMMLYIYCGLGTVEVTNDGFIDSGYTDVIWYKLPMVERKMIQMMVMM
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>MdOr59

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EYELKSCDYARHLKKGCHLLSYFMKLCAVINVMMICGLILVAAAINVIFQKRDLYVG
DVIGIDPSTTSGFYVTFMVQACFLLVGGFGLYAGDMAFFTPISQVPTLKEILRCKFKDIN
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FCIFLGTWPGGYIYLLYCFVMMFVYCGVGTMDIANEGFIDACYNDILWYKLTASDR
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>MdOr60

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VYLNKGIGHLLANFMKVCAFINFMLVLGMTFTIFYNLIFGTNETLVYGYCPWVSLETT
GGLWTTNMVQALLIAVGGFGLYSGDMSVLTPISQIPTFKGIIQCKFRELNLLDDDHES
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>MdOr61PSE

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VYLSKGIGHLLANFMKVCAFINFMLVLGMTFTIYNLIFGTNETLVYGYCPWVSIETTG
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>MdOr62NTE

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LIFLKVSREASYWSVFAKVGTCIIGIVGALFCIMLGSPAGYIYMLYCFVMMQVFCVM
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>MdOr63INT

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EYELKSSDYAVYLKKGCDLLSYFMKLCAVINVIMICGLILVAAAINVIFQKRQLIVYGQI
FGIDPSTSTGFFVTFSVQAGFLLVGGFGLYAGDMAFFTPISQISTLKEILRCKFKEINEAM
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>MdOr64CTE

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>MdOr65

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LEKNTNRVNRFIKFATMHLVITITLIAVVPFYRVVFNERILIMQFLPGVDPNTAYGYT
IITTIHCICILFGSGFNFAADVCFFNIVSHVPLFRDLLRCKCQDLNEILEEERASEE^EEGFAEI
ELLLKDIFQWHQKYMRYITTVENYFWVVLVEMGTVALSIASTLFCLILGKWPGGLTY
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>MdOr66

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RRISILLQELEEIFPDEKEQFAHPTLYRVRHFAQTSERLMGRRTKFFIAFCFYNSLPIAEL
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FMLCFFITQMQMHDYLTNALRHDAASVRANEKLKYLIYHTKLLRYSKEINEIFNISF
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AMD

>MdOr67

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LHPTIFKNIRSQLMPIFVLSSLYLSALISYVILPIYFLSIGSRELMYKMIPAFDYSPLWIYLL
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GNQHIAAEYNKILVETLQENVALNTFAQEIQREYSFRLFVIVAFMAASLCGLGFKVYTS
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>MdOr68CTE

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SLRPDIHAGNLRSYYPVLAFLSILYLCALLSYIVFTIYGLAVGEKPLPYKMIPPFDYN^SWYI
YTPLVLSLWVGIVASTIVGESYALTMFVHNLDGRYQMMGERLNMGVENILKFSSND
SEAIEKFHRILIATLKENIRLNKFAQEIQREFSFRIFIIFSFLAATLCVLVFKVYTSPVNSIPY
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>MdOr69

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YNIETYLDIQRKLLPTKMC^SYAMLT^VTYFLVPVLGFFSNAHLVPFKTIFHYDLDI^W
FYLP^LCLTLWIGVAVVSQLAAE^NLLATIILHLNARYLHLQSDLKE^LQTRLASDMKLS
TDKVLGEYRREFIEIVKR^NVEYNDFAQKFQNQYSFCIFVMMMAFSAVLLCVLAFKAATL
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>MdOr70

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EV CYI YTGL VTL SVMTY AIAIDGLFISFTINLVGHLKTLQHFIQS KSFEQN DEDVHKQISF
YIRYHN LILHLYQE VRQIYSPIVFGQFLITSLQVCVIVYQMVT HINTFLVFVINCTFLSIL
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>MdOr71CTE

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>MdOr72PSE

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CFVGFQMTIGVEPDTLVKLFLFLSSTAQVYLICHYSQMZMDASLNVADAVYNQNWSI
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>MdOr73

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GSFKTCIIMQKKSHLTTYARDMNQIFPNASIAVQRELNVRKYLKYSKFFSIMFSTMCLA
MLVFFNFEAITEWLIATELRGDQNAAQHLPYFMYAPWDWTGNHWSYYLLYGIQCWA
GHTSVVAQFSSDLLLYAFIGQLIMHFEAITKDVSNYRLSCTADMDFLRNIVFKHSILLE
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>MdOr74CTE

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GFVINSSWKIYMIWKKRPLIESLICDFHDIFPTKMLQQDYDVQVYLRKCHRKSFKMSL
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AGHTTAMGNVSN DMLLYSLISQLIMHDFVANTMESYEIGSGSKGVAKMEGRENGND
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>MdOr75

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>MdOr76

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>MdOr77

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>MdOr78

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>MdOr79

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>MdOr80
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>MdOr81
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>MdOr82
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>MdOr83
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LISI LSFSFAAGAAVASATIPLFLEQRSLPYDMYIPFYDHLKSPMYEILYFMQGFISMPFC
VL TYV PFTNL FIAWLT FG ISLLQIL RYKLES LPHENDE EMLKQLIELIRFH RIMNFGQTL
ESL VS FV CL VEL VLFT LMLCVLLASFLVMDNVMSKIATCIYIFCILYALFIPYWHANEFS
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>MdOr84A
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>MdOr84B

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81 CcapGr proteins in FASTA format

>CcapGr45

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>CcapGr46

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QRLSAEVLGMRSELSELRRVLQVHEELQVINTGVNNLYRFHITWLLLIFFLSNTTAFYV
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>CcapGr1.1

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FVFTVQTTVVVLVLRERVNNFLNDTDRFDEAIYNVIFISLLFTNFLLPVASWRHGPQV
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>CcapGr1.2FIX

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>CcapGr2

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LTTCFQKEIVSSHSDVISKYRMLWL YLSELLQSLGNAYARTYSTYSIFMFINIVIAVYG
AFAEIVDNTDVSRDSYKEVGLIVDGLYCSTLLFIFCDCSHNATLGVAKGIQKVLLIEDV
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>CcapGr3

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VFVAYVAKNRISIVTSLGPFEAAVIYLFLVNILPLLIPLLWWEARKIARLWNDWDD
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CYTFTFINLYLFFFVITLSVYGLMSQLSEGFGIKDIGLAITAIWNVFLFYICDKAHYASF N
VRTNFQKKLLMVELNMNSDAQTEINMFIRATEMNPSNINC GFFF DVNRNLFKGLLT
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>CcapGr4

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NNTTVHFTFEDTYKNYDYVYELLPTTAVGSFILVGNFFCTFVWN YM DLFIMMV GK
GIA YRFEQIKMRINKLLDKEVPESIFMEIRDHYIKLIELLEYVDEDLSGIILLSCANNLYF
VCYQLLNIFNKLRWPINYVYFWFSLLFLIGRTAFVFLT AAA INDEAKEALGVRRVSDK
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>CcapGr5

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LYRRERPHLSRVLPFSIWYLPVVEWINLTAYPRSFTDAFIICVSVGLAARFHQLYLRIE
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>CcapGr6

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FICVQIFHSFNNRDTFIKEFYFWSSLLVLLRILTMMMSASAVHDEANKIMSTMYEIPK
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>CcapGr7

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NFNDIFVMAVASVALSARFRQLNEHMLRAAKQPTTEKFWMMDNRINYRNMC KLCEATD
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>CcapGr8

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FNGVEPLIFRLSII VVCSSAIVLARKWPALMLDWHEVECDLPEYLTQIEK GRLAYKLKM
VTVVAMALSLAEHLLNITSIIYSNSCPQTNDRIRDF FILTNQHLFELFPY SIYLALWGKT
QNILCTFIWNYMDV FVMIVSIGLA AKFKQLNDNL YKF KGMR MPEEY WSTRRK QYRNL
CELCTRVDAAISVITMISFSNNLYFICVQLLRSLNK MPSFAQAA YFYFSFFF LIGRTLA VS
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>CcapGr9

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YSV VYVILTSIDTGLTMNMVFR GAMEVKNISPLVFHV NALLVAICFLRAV KWPQLML
KWQRVERQLPPHQ MWRDREAL ALRVHKVTFVLITLALTEHLLSVVSGIHF AHCSP
NSDPIK SFFIAVSPHTFLIFNYSTWLAWCGKLLNVNTFGWSYMDVFLMIIGLGLSS LG
QVQNSLNRVKGKTMPEAYWTRTRLQYRLICDLIEQVDGVVSGMIMLSFANNLFFVCIQ
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>CcapGr10

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>CcapGr11

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>CcapGr12

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>CcapGr13

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>CcapGr14

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>CcapGr15

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>CcapGr16

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>CcapGr17a

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>CcapGr17b

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NMDKIIAILNRLKIDQFISELPTSDAAPQKSFKRTSGNKYIMWMFCYICLITIPIVHRLFE
TLSFEIVILAIAYQLENYTSYAFIVFISSLHELTMRFQYVNHQIEKYNRKFPKSSEQANIS
SPRRLEHKTTQDILNMSRFAENSVVIYSLYNDLLKMVNQLADLALLTFLLYSCTG
ILTC
TYTSTLFNFHKRNESFSLMWNVSWIVNFSAVVILLALRCDRVTKLEANRTSQLARVYG
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DN

>CcapGr18

MQDETWSLNILHTKRQRYKLATAHTLVKDLKWILLYLKVLGLPIFKITHTYQMAMPN
NRSCAALYTHATHVFGFLIIISYIYIVCSPTANSVIFIYGEMDNATTAFQILLSITCYAIVF

WKSANKAQPFLAIINQLSVDQELQQYPRAPAALENKCDFYKGYLLMLLAHISTFWSM
KWMNSHECNSSMICVCIFFIYLYMHAVINA YVVVAILLHLLTMRFYLNFIGIYAAK
RNAVGAGVSTQCGDGTWAGCGGERTAEDVDFVVFSQDMIFFYRLHNHLLVIFETLN
DYVHIALLVFIGYFLYGSTIAVYYFYFAILRDDIQIISVIWCALFLCLHGPTAAVLMRK
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>CcapGr19

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IARKKGLFASLYTTIVRTLCLLVIITNSYTLFSLTISYSFFGSVDLVNYVLHMFLNIFCF
AVIIFSCAVKSRSRHFLRAANKLLLIDVQLQRYANNTDCLENECAFQWKYFIYLGIFTVITL
QHIDFHKMYKTSYFLLFCYYIETSICNFFIIFIALCHLLFLRFRYLNRFVKSFTNGGRN
QKLKQQLRAKHSTENPIFYITLPEERVRPKHEIQQESFLSVSSYIYRTHYELLKIYKQLNE
YVGHA
LLVYFIYVFYLTSAMYRMIYRAKSSKLSKFPLYGFASLVLHMSILVLVTRCCSKLTEE
AQKIASIIANIYGRNEKSRKVTDKFLTTNLKADLQFTACKVFTIDNSTLFKLCCAVATYL
AILGQFCLEKSREH

>CcapGr20

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LINAWILALYRQHFNIISYCMYLMERLSFSAARQKVGIFQSVMIAAVMGAVALNTYCM
WQELAIPHSFLSICVYVTWQLYYISSIILSIYLCIIQIITGHLNNDTMNIVIVKNNHRQKR
VTGIDLDNIQERLVAYDKLLLLCNEDISTSYGVVFVLITFISTLDITFIVYILNMTTAVGY
LQTIGSTVKTAFFALPSIIFLCMMFLGSDIQEQANNTVRILAKLPRTGTGLDKMVDFKLL
KNMRKKPILTAYGFFQLDRSALFKLF TAIIMYIMILVQFSIDIENSLKKIN

>CcapGr21

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FIREFGTNEGTYKFKREGWILTIICFTILVTIPHTIISWTLYRTRVLKTLFWHFLPSTLT
LQFCIFDLMLTITHLNEYAAILRNDLNEMGERNIRRVFGHYLRGLATVKGSIVRVTPQK
LQPSEEEDERRLRVYKKFYGDIYNMFKAISECFGWSLLFFIIMYFIYFVVNSYYVIQSIIV
QLDESSTVLSNLGFLMIVTLLWILCWQSNSHEQSRQIGCIMFKLVKPSGNKSYNDLV
TDFSLQTLHQQQYVITAKEFFNLNLNL
GTMVASIVTYLVILIQRQMFSEETRVERNLMENNMRNCTVE

>CcapGr22

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ALVRWVRTCLSVGCAIISFCCMLETAVKWRQFAQIQQLRQQINYLISQLKAHLRGRK
RRSGYGLKTRLLPLLLILICESIKFSSVKNMTNTYFGTVTIVMGLRLRYFHALSLMCEFN
EYLCVLYETIDLLVTYNNTRPICEFNVWRPHNRWELEQLNLLRLYGRLFEEFQLINDC
AGWSMTLIAQLALSEFVCYTYWCLTYKLVNLGIGAVIFNLATIISLGCLHYQWFGLAE
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HVFVNYLVFLLQFTYIKTKA

>CcapGr23

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VT FIESICKCEHYQELLRLEREVDTLLVN KIDVQTII EVRVWHMAFYVVL IQIYDVVQL
VIAFTGYKSPVFFYYVVVILFIQRARYAQITCAIARLNARSCLITLLRVLVKANRPRHKY
SSEVWQPYAVWEFEYLNLRLIHGRLCELHRCVSNCFSWSIVLLFSTFTTVSNLFWCI
EIIRTNFQFGQFAYDALTILRLDTAAVLLFTAEQAKKHNMLLGGFLNLAKPLGNKTY
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VIYNSMANEESMKTEKFPTIN

>CcapGr24

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ENRSLRRKMLRRGAIMVFIYMSTELLILIA YFFT KDRHFSIYWL FYLLPFCICGVRYFQIF
AAIMIIRQRLDKLVETLNE NLKAKPMQQMSEDSK TNIK FITATLPV YETTEYLKRKY
NMDNPDMKRLLIIRDLYNRLWEVTVTVNNDFGVSILT NVGND FISITSNCYWI FLNFKS
YSGTLQDFLQIASSAIWSAPHLYNVMLALLCERTIQKTTAIALGLHRMETNVWNDNH
NTVIEQFSLQLLHQKLTFSAGFFDI NCTLLYRIVGATTYLIILI QFHMSADKLTT S

>CcapGr25

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TFLRLLQEIEFSLKLRLKQDVRLDWFVSHMRWLFYLLWLSIIC YTCFVYYFTTAQYV
GYFWHTTLYNITMRLRLIQLLIYVRVLQHYLECLSMKLRQIVAHRLAPCRQLLDV NYE
KLQSLEYLVAIKEIYGLLFKATQLFNFKAGWSLFSIIVGYMLDYGCALCWSVLSWEGF
LESHNYYVPCLWWICPMTLILWHLCHLCNRCKQLDR LIANLSCRVIIMHSSHSLHTYRI
VLHQFAIQLQLQRIEISARSFFTLDMRLIMSIFTSI ALYMVILI QFINIGNKYI

>CcapGr26

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VFAVYINGLYWRKRISGIMLTWL V ATILFTSQALT NLLII ETI WKEQEHEA FLLL LQRI E
LTFKLRLRYATNKRLFAAKLRRYLYYLYTISLAGLVLFLVTTFWLQYIGYFWYGLWFII
TMRIRIIQLLVYLRVLQHYPCLALRLSGIV AYR MTPNQ QILDVNNKRLASLD LLLAIRE
IY ALLFEAFQ LLNEFAGWSLFSIITCYMLDITC NIYWSL LSLDG FARR RYY YISSI WLV
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HIEVCAKNFFILDTRLVMSIFTAVAAYLVI IRQFLNAQQIN

>CcapGr27

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LKLRLRQCTHSAALQRNL CYIIVRLVVFSLVTLILFMITSIWL NYIGFFWNGLWSILTMR
IRVIQLLLYVRMLQHYLECLCVKLQQVVEFHSPQRQL DIDYSRLTTVEYLLAIKEVY

TLIHDAFQLNYCAGWSLFGIVVCFMFDVSCNVYWTVLSDLNWQNRRYYYIAGPVAL
LPLLVIICYLCVLCGKCKELGRRIIYLLSSIKVLNSSQSAALYRQVLHDFSVQLHQQQIE
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>CcapGr28

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WRQQEHETLLRLLHEIELSLKLRLRQDVRLDWLLPRVRWLLRYLLWTSVLFVLYN
FVELQYIGYFWYSIWFAIAMRLRLIQLLIYVRVLQHYLECLSMKLSQLVAYRVAPSRQL
MDVNYARLQSLEYLVAIKEIYDLLKAFFQLND FAGWSLFSIILCYMLDYGCTLYWSL
LSWEGYLERRNYYIACFWWLLPMSLTWHICHLCSCRQLDRVIASMLCRMIIKRSSKS
LRSYRILLQQFSTQLQLQQIEVSAKRFITLDMRLIM SICTAIATYLVIIIQFLRI

>CcapGr29

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QHETFLTLLDEIEVSLKLRLRQDVQRLLACRNMR AHLVHLLVISLAAMTLFIITSWLWLN
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ERLASLEYLLAVKEIYSLHRCFRLLNNFAGWSLFSIITCYMFDFSCNVYWTLLSFDFG
ARRRYYIAGPAAMFPLIALICHLCYLCDNCMKLAQRAAHLLCRIQIMSNAAPLRSHR
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>CcapGr30

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FICTARLRTLQLLVYLRLRH YLRLGCLKLRQIMEFHMAPSCRLLDIDYGRLGTLKCLL
AVKETYTLIHEAIQLNYFAGLSLWGIISCVLDLSSNFYWVLQSFDNFHGRYYYYIDL
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>CcapGr31

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MMREAWCKQAQHDLFLQILEKIETALRLRLRHNVQSAQLLCDLRLIVYLSVISALGF
ALFALTMMWSYVDYFWFGWLIIRVRVIQLIVYLCILRHFLRCLCAQLRLIVAYHTA
RGHTILDVDGVHRLSLKCLLA VKEIYELLHREFQLLNEFAGWSLFTSISSYLLDAICTLY
WMLLSLDGFVRRRNYEIAGIFVLLPLALFLWYLMFLCDNCKQLGRTVAHLLSKLILMS
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>CcapGr32

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NETFLQLLAEIEVSLQLRLSCDTRQHALRQSLRRWLLGLVTLGVFVIFTSVSVLLNDI

GYLWHAAFSLLTRMRILQLLYACILRHMECLCFKLRQLVAYRTAPERRVLDVN
YEKLESLTFLLAFKENYALIFKAVGLFNNFAGWSLFGIIFGYMLDFTCHVYWSLLGLDG
YGSRYTFVVGPAVLPSVIVWQLCFVCDQCKELGDNISHLVSRLVTSTKPALKRYC
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>CcapGr33

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LAYTRLHNKWSSHTFRPEYAIYRKYVWRSSICIVIVIIDLAYIQEISKNRSWLAFFIPFVP
SGLICHRLSLQIMYFMDMLRIEVMQLNRNVERLVTFSERHCVENSQRSEYFVRIVCAEL
QILMECYQNIYELSMLKKAVGISLTCNYIKEYVMILSECYWSYWMVYNGQDVIEYSL
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>CcapGr34a

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YMVHLRNMEIIFHIAIRHELEKLKDALLADYTRFSRRVAPFVGESFVRQKLAEKQ
LTFQRIYEMYYYFQQSGVSTIAVLLMTYTRLVVDAYFILYTYHQKEPEFIENLLMLP
AYLEIPSLLTSQKCMNEVKFIAFELHNIRSSVDNSLISIQIQNFSLQILHQKIRIDGLGISA
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>CcapGr34b

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EKLTPPKPHWPLQLCTYRRFLCIFYGITIFDIIIIEILFHAMPAGEDTMIVMRFWFMFPTY
VYMAHWRNMQIILHIDVIRHELEKLRNDIALLAEYTRFARRVVPFVGESFVRQKLAE
KQLVFQHIYEMLYFFQRAFSVSTMMAVLLMIYVRVVVDTYFMFFNDAVGWRFLENLLM
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>CcapGr35

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RIWNLLNQVERYFCTPMLMLFFYNGFAIIQAIHWAYINFLDDDLRLGRIVHTVMLIV
ALL
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>CcapGr36

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RVLTVLFCITVQLKGVEYCMNAQLVQELLHLVQKQLLHRLRRCEDVERRRIYTD
LQTQNQNLSSRIWDLLNQVERYFCIPMLTFFYNGFSITQTIHWGYINFELDDNLRLCRI
AFAVVIITTLFIPCYFSQCCIDEYNRGTMHLKLKTVGIDELLAMRLQEYSLQLMHQQM
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>CcapGr37

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LLLQKFEMEFDFPKLKSCIKALRNHLAIYCTFLLFFVILLALQPIPTVRFLVCLHAEIVLKI
KIFEEMYFTVVFIVMLGDLCRAAKQCRLLETARFENTFQQREYLLGFIALQDLHALL
WENVQLLTDYFEWSLPDIVSSLIKYYKMLFIFVHAYFIHKDLIVIWLIRLSTLTMACYLC
TMCERRDRELRSYLRNCLKDRRNQPLMKCFHRISMQLWQEPIKFDAGNFVTINMETLG
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>CcapGr38

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RIRAVDEQLLRENIVVDISKTSQRILIMLALTVICEVTLVSTYIILVDYTKWISLLWIFSC
FPTFYNSLDKIWFANTLNALRQRRFMINTALDGMVESHEKLKWANTGGDSLLSTRKP
SIANVSIDASIEYLYKDFTHMEA VKKFNMPRNKITPIAHSLNFYGESFETPKPYKLSIQP
PSFNMVYESELNRDIEKVEENNNLCQLHDEICEIGKQLNELWSYPILVLMAYGFLIFT
AQLYFLYCATQKQNIPLFRSAKDPMITTILCYTAGKCIYLIYLSWKSLESKRTGICL
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>CcapGr39

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YVDEDLQLGITVNNKRVYRRIWLMIVLTFVCEFYMFFAMMWLIVDEFEFWFNVLWIF
TSIPTIYNSLDKIWFLGILLGLRDRFEAINTALDDITEEIEKNNLQKRKGLVYEQSVG
LMLHTSLSVGSERISDIKERLVCNAFGELVCERESNKSAAVIVSYENSSYNFAALQERF
ISLCQLHDSICRIAQQLNQLWAYPLLVLMAGFVFTSLLYFAYCATWNQSIPLIFSPA
NIYTSFIVIGYIAGKCVSLMFFSWQTTQASRRAGICLHKCGVAADTNEVYEIVHLSLK
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>CcapGr40

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TRNKLFPVKKKRLYYFTWIIIATFAFAIFYDINEMSQYYGPYCFISNMVFTFPYVAGSI
VQGMFASYVSVISERFSTLNILFEKINHESDKRNLPIAVMDIENDAHKDNTYSGPA
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NLPSLFKLHDKILSLSVLVNAEFGAQCVPYMAACFVITIFSIFLETKVMFIVGGKNHVM
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LTPKVFSTESN

>CcapGr41a

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VLETLLGKSKRESKFKTGEFVTFFSCQMVLYLIAIISIVEGSNRAIKKSEKTGGIVHTLL
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>CcapGr41b

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NKYFTYQLLTIIISIAFLIIVFDAYYVLETLLGKSKRESKFKTGEFVTFFSCQMVLYLIAIISI
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>CcapGr41c

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SRFADTMQIVSGLIGITVIYFTAILPKHRLQHSLRIMQDMDVLLHSGVGKILYSKLLRYS
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MVLYLIAIISIVEGSNRAIKKSEKTGGIVHTLLNKAKNAEVKEKLQQFSMQLLHLKINFT
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>CcapGr41d

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>CcapGr41e
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>CcapGr42
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>CcapGr43
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KFNALMSLRQKLQRFAVLRVNRLLGQPIAGVMLALYCYNVGVCYIISLMKPHEASLLD
ISAWNMVVLVFGTVVYIADVTEFLVISQRLRQNFGLCMPITLQLLELQQLEKNWDRSIE
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>CcapGr44
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VLFLFFYMLWHVRRRIWRLNARLIELFHDLKQLQRNARDMLAPKSRTLQQLSGLAAE
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>CcapGr47
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EIMMISKIHERLSTLTERLNADFKLQILAGLLTQLNCICYGYYGFLNEGFLKIKVSTP
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>CcapGr48

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GSDLRQGRKQLALEAVDILRGIAQTHACLSHLERLNNEGYLQILVULLASVIESITFGY
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>CcapGr49

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FSRFTVAAPAQRHFEIKIVRNILSGLMIDVSGFIMILFAYSEQASSEFILSSIMFTFVSNILY
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>CcapGr50

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RRGTVPHFCGLLCDRLDTLVQRYQQINRLYQDSLTMHQFQILGMVFITLISNLTSFILF
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>CcapGr51

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FEKCMPSRVKAHCRQYYSNLLWLKYFSVISQLWTLYNMFFYLPDKSVISLFYCVYV
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AYQRLGQELMEVVRVHYQLTGFAERITNCYRFQIITVLFSKVINNISITYLGLKGNSF
LKSLGVTYHIFSTSFFIITLSDSYLLDILCERVVGASHEAAEVLKRFDELFELEDAVEYA
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>CcapGr52

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ERRFFEKCVPQTVKAYCRQLHGNRLLWLKYFSVVTQIVYTLVNLVYFMPKISITWLLY

SAHMFLISMLLHMILHYFLAMWYLWKRFSWLNAQLRRIFNFLRLLEAHRLLGDLA
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NSYLKSLSVVFQMFSLMAIITLSDSYLLDILCERVGASHEAAEVLKRFDELFELEDDAV
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>CcapGr53

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RNDINVV

>CcapGr54

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WSVFWRLERRYERILPTDDGNTTMHTRWLLLYYFWTQHIVTPLTSIYVLLQFPRQTNA
HIVMTFVSWALSTVQVMLAYYTLC LHCHLLQKFEAINCELQRVLTLLRNLASMERTRL
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>CcapGr55

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>CcapGr56

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SLMEVFTQFLYAVNIYGILAILWTNSWEYTYVWKVFKEAALERSYFGKHQSLSAKC
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QVFFGIAALTAENAQNVLVLLFMLTPPAETWKIIFS AFFIAKNIVYFWLIICACELAVEA
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>CcapGr57

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MLLVNIISLVVMHFYIFILVTYRYIWIMRERLKCIAANDLGLPHSSNLQSIRYELSEITVIY
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>CcapGr58

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FNPNPNSFKYVCFTILTICVKNITLFTVSNFYLTLLYIYRLLQQLNWNFKEVVFYSLRAR
DVPDTVPIEDTTEIAFDLKATPCCKTSVECTGVDVAAIAELCRQYVRICQLIRRVC KV
YEWQVLLFLIIIFGNVMSTFYFLVYLVGKMLPKELFSPTLFLELFFINVLDLCCYMVIC
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>CcapGr59FIX

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NVVLLVATHFYL VVLHTYRLIWA NRREAIAADNIMPRLCQRRHLLSVEIDTMASIY
ARLISLCERYTRMHQLHLLL VIGSVTACNIEVLFYV RLLWSGKIPERTAFNVFAV FQIFV
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>CcapGr60FIX

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>CcapGr61

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DIECQYFAKYGLLAWKCSAYEKYIILKGLATLLRNLSFIYFLSGLA KVVTWNILLV LAL
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YLRLIKISKCFGRIYGKQLLFSFAIIACGNIQALYYLGLLWADQFVG LSLLEIFNVLHGVS
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>CcapGr62

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YGREISWYQREFQNTCNAMATDSFEILDMMRDIYFDLDTISTQLTNVFGITLLFNFLGS
CVSLSVQFFAVFKFFDTENFLNDVQAELYNRILWIIIHLGRIMTVLISNNAIIEKCRTSC
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>CcapGr63

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>CcapGr64

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VVDYFYRDCVVASSIRSLTVYLLPNLILCVSLVQYYTLLYAIAQRSIRLNIELHEEISRQK
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VWWILYSVIWTILHITKMFLILFYNYDVQRQKNSTILIINEIGRTQNEVEETVTHFVLQ
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>CcapGr65a

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RYVGETCTIRWNLWAVIYVFGLCNMCLVGAAHQLQSRVQYIRIVLAEGSARMPLV
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>CcapGr65b

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YKRNSLSGRELVHIFTPSLLCFSFFVVVICTVQLFVIYFLTC SHKAGKFRTDGTVTHRDR
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>CcapGr66

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VNLA SLYLQQCRATLQLLIMASILLTACYAQLALRLED FEQSKIVEFTYFEDLVALQK
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ELYLLIHTVYFCCV LRTVITHLFLDREVDFEDAPETSRYELTHSDVMWPQPTQFFILGF
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>CcapGr67a

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TIFVNIVNYFIKLFQRTQR LAPHKSIMCVQHFAFLIIILCMTRDIVDLLVLFDYTDYRER
AVAFAKLYLEISLIFTIHIGCIGYMSVGALYEFVN KYMREEFQPRARCLDYVLKRNHNI
RSQRYQRKLIRLTRELNECACIYNNIYRAATSFHHIRFH IFFSLIFTFSFLTTVIY SVLYL
YTFYHGF IWV WALFVCLQVFTELLIMIAAKSAVQGGFSVNKLSDSIY MAGNAEWNQ
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>CcapGr67b

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NYRLQMNRKLI RL TRELNECTSIYNDIYSAANTFHSIRYQIFFALLFEFSLFTTIMYSM
LYVRTVLHDFHWDAVVYSGQIFIGVL LILSVYSAVQSGLTTRNLSLDSVYMDNAEW
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>CcapGr68

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NNIYFVLYGLTKQFHIFRWQLIY SIYNNFVIILMVVMHSFIWQYIYAGYVDFLALFSSL
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>CcapGr69

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>CcapGr70

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>CcapGr71

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>CcapGr72

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