

PIXE and ICP-MS Analysis of *Andrographis Paniculata* Medicinal Plant

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Abstract: The concentrations of elements Li, Be, Al, Cl, K, Ca, Ti, V, Cr, Mn, Fe, Co, Ni, Cu, Zn, As, Se, Br, Rb, Sr, Ag, Cd, Ba, Pb and U in *Andrographis Paniculata* medicinal plant used in the treatment of Diabetes Mellitus were determined by using Particle Induced X-ray Emission (PIXE) and Inductively Coupled Plasma Mass Spectroscopy (ICP-MS) techniques. This plant was collected from four different geographical locations in Andhra Pradesh, India in order to assess the regional variation of elemental concentrations. Appreciable levels of K, Ca, Cr, Mn, Cu and Zn determined in this plant can be correlated to the antidiabetic property of *Andrographis Paniculata* since these elements are known to regulate and potentiate insulin action. Presence of toxic elements As, Cd and Pb necessitates the adoption of precautionary measures while prescribing dosage of the herbal medicine prepared from this plant for the treatment diabetes mellitus.

Keywords: PIXE, ICP-MS, Trace elements, Medicinal Plants.

1. INTRODUCTION

A plethora of well known analytical techniques with remarkable sensitivities and detection limits have made it possible to analyze a wide variety of samples for their elemental content from every conceivable field of scientific or technical interest. Particle Induced X-ray Emission (PIXE) is a relatively simple yet powerful analytical technique used for the multi-elemental analysis of a wide range of samples [1]-[5]. This technique serves as an excellent tool for elemental analysis, particularly trace elemental analysis because of its capability to detect simultaneously several elements present at very low concentrations and the ease of sample preparation and analysis. Inductively Coupled Plasma Mass Spectroscopy (ICP-MS) is another reliable and effective technique used to quantify several elements with 1pg/ml sensitivity in a variety of samples [6]-[8]. In this method, initially the samples are ionized with inductively coupled plasma. Consequently a mass spectrometer is used to separate and quantify those ions. ICP-MS allows determination of elements with atomic mass from 7 to 250. This encompasses Li to U. This technique has the capability to scan all the elements simultaneously. The other advantage of using ICP-MS lies in the fact that it allows isotopic speciation.



Since times immemorial, in every culture throughout the world herbal drugs constituting medicinal plant preparations have been used for healing purposes and maintaining good health. The earliest recorded evidence of their use in Indian, Chinese, Egyptian, Greek, Roman and Syrian texts dates back to about 5000 years. The herbal medicines have been derived from rich traditions of ancient civilizations and scientific heritage. Initially with the onset of industrial revolution and the upsurge of allopathic medicine, herbal drugs suffered a setback. Concerns over the iatrogenic effects of conventional medicine, advances in phytochemistry and the desire for more self-reliance led to extensive research on identification of plant compounds effective in curing certain diseases.

In recent years, considerable attention is being paid in assessing the therapeutic effect of medicinal plants from their elemental content point of view by using different analytical techniques. In a study on total concentration of selected trace elements in neem powder and in neem tea by ICP-MS, Novotnik et al [9] observed substantial amounts of essential elements and negligible amounts of toxic elements. Mineral elemental concentration of some Chinese medicinal herbs used to improve kidney function was determined by flame atomic absorption spectroscopy [10]. Regional variation in essential and toxic elemental content of *Murrayakoenigii* (curry leaves) used as a spice and medicinal herb was studied by INAA [11]. Naga Raju et al [12], [13] determined elemental concentration in the medicinal plants used in the treatment of diabetes and cancer using PIXE.

In the present study, PIXE and ICP-MS techniques are used to identify and quantify the elements that are present in the medicinal plant *Andrographis Paniculata*, which is widely used in the treatment of diabetes mellitus (DM). An attempt has also been made to study the variation of elemental concentration with respect to geographical location.

2. Materials and Methods

2.1. Sample Collection and Preparation

Whole plant samples of *Andrographis Paniculata* medicinal plant were collected from Vizianagaram, Visakhapatnam, Vijayawada, and Tadepalligudem regions of Andhra Pradesh, India. In each region 10 samples were chosen from different sites for elemental analysis. These samples were washed thoroughly with tap water followed by double distilled water to remove the surface contamination and were then oven dried at 60°C. The dried plant samples were ground to a fine powder by using an agate mortar. A known quantity of yttrium with known concentration (100 ppm) was added as an internal standard to the samples to check the reliability of the experimental results obtained in the present study.

For ICP-MS analysis, 50 mg of each powdered sample was taken into a high-pressure closed Teflon decomposition vessel and 1 mL of concentrated HNO₃ (Trace metal grade, Fisher Make) was added to it. After 30 minutes of pre-digestion, 4 mL of Milli-Q water was added to 1 mL of each acid mixture and the vessels were slowly shaken and sealed. Consequently, decomposition of the samples was carried out in a programmed Microwave Accelerated Reaction System (MARS5, SEM, USA). Ten replicate digestions were made for each sample. Finally, the digested samples were transferred to a 25 mL volumetric flask and made up the volume with Milli-Q water. Blanks were prepared in same manner by excluding the sample. For PIXE analysis, a homogeneous mixture of 90 mg of powdered sample with 60

mg of high purity graphite powder was prepared and compressed into a pellet using a pelletizing machine.

2.2. ICP-MS analysis

Inductively Coupled Plasma Mass Spectroscopy (ICP-MS) analysis has been carried using Agilent 7700 series ICP-MS system available at Centre for Study on Bay of Bengal, Andhra University, Visakhapatnam, India. The optimal operating conditions for ICP-MS analysis is given in Table 1.

2.3. PIXE analysis

Present PIXE measurements were carried out by irradiating the samples with 3 MeV proton beam from the 3 MV Pelletron Accelerator facility available at Ion Beam Laboratory, Institute of Physics, Bhubaneswar, India. The detailed procedure for PIXE experiment can be found elsewhere in our earlier works [14],[15]. The obtained PIXE spectra of *Andrographis Paniculata* medicinal plant corresponding to the four different regions are shown in Fig. 1. Guelph PIXE (GUPIXWIN) software package[16] was used to analyze the obtained PIXE spectra and thereby identify and quantify the elements present in the studied samples. The precision and accuracy of the results were checked by analyzing NIST Certified Reference Material – Peachleaves (Sample No.1547) in same experimental conditions.

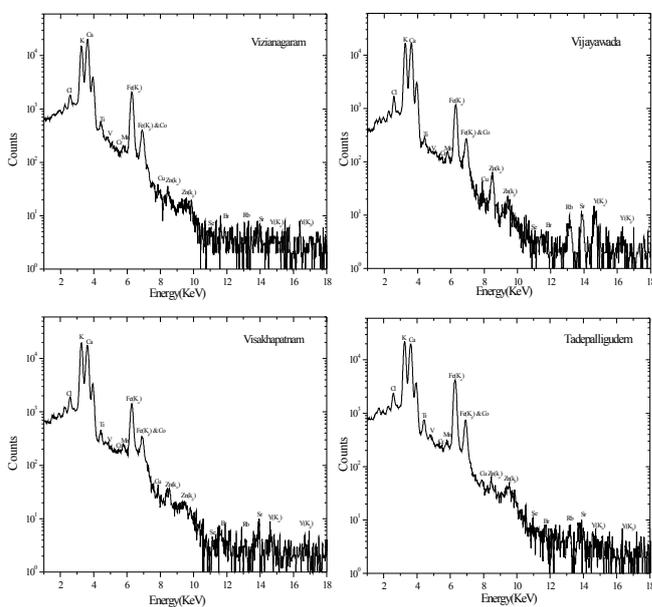


Fig.1. PIXE spectra of *Andrographis Paniculata* medicinal plant from four regions.

Table.1. Optimal operating conditions for ICP-MS analysis

Parameters with values

Plasma conditions

Rf frequency: 27 MHz

Rf power: 1 kW

Gas flow rate

Carrier gas: Ar 0.8 L/min

Auxiliary gas: Ar 1.1 L/min

Coolant gas: Ar 15 L/min

Sampling conditions

Sampling depth: 7 mm from work coil

Sampling cone: Nickel 1.0 mm orifice diameter

Skimmer cone: Nickel, 0.4 mm orifice diameter

Nebulizer: Cross-flow type

Sampling uptake rate: 0.4 ml/min

Data acquisition

Data point: Multi-element hope by peak hopping

Dwell time: 3 points/peak

Integration: 50 ms/point

Repetition: 1000 times

3. RESULTS AND DISCUSSION

The results of PIXE measurements for the determination of concentration of elements in *Andrographis Paniculata* medicinal plant collected from four different regions are presented in Table.2 and the ICP-MS results for the concentration of elements are presented in Table 3. From the tables, it is clear that the fifteen different elements namely Cl, K, Ca, Ti, V, Cr, Mn, Fe, Co, Cu, Zn, Se, Br, Rb and Sr were determined using PIXE whereas twenty different elements Li, Be, Al, V, Cr, Mn, Fe, Co, Ni, Cu, Zn, As, Se, Rb, Sr, Ag, Cd, Ba, Pb and U were determined using ICP-MS. Among all the determined elements Al, Cl, K and Ca are found to be most abundant elements followed by Fe. The significant levels of elements present in the analysed medicinal plant, correlate well with the traditional use of this plant in the treatment of diabetes mellitus.

The role of trace elements in various biochemical and physiological processes in humans is well understood. Elements by themselves or as component of enzymes may play a vital role in the development and management of DM as it is a disease of metabolic abnormality. Out of all the elements identified in the studied plant, the elements K, Ca, Cr, Mn, Cu and Zn have insulin potentiating property as they are responsible for the secretion of insulin from the beta cells of the islets of Langerhans.

Table.2. Concentration of elements ($\mu\text{g/g}$) in *Andrographis Paniculata* medicinal plant from four regions by PIXE.

Elements	Vizianagaram	Visakhapatnam	Vijayawada	Tadepalligudem
Cl(%)	0.822 \pm 0.057	1.21 \pm 0.068	0.890 \pm 0.045	1.48 \pm 0.085
K(%)	2.68 \pm 0.028	4.51 \pm 0.043	3.01 \pm 0.029	5.18 \pm 0.049
Ca(%)	4.14 \pm 0.034	4.82 \pm 0.043	3.53 \pm 0.030	5.96 \pm 0.049
Ti	484 \pm 37	418 \pm 40	159 \pm 26	841 \pm 49
V	14.6 \pm 2.62	9.75 \pm 1.75	11.8 \pm 2.59	13.9 \pm 2.78
Cr	9.14 \pm 1.82	27 \pm 5.13	8.22 \pm 1.23	18.3 \pm 1.57
Mn	87.9 \pm 15.4	153 \pm 21	85.2 \pm 12.6	177 \pm 24
Fe(%)	0.311 \pm 0.002	0.247 \pm 0.003	0.170 \pm 0.002	0.593 \pm 0.005
Co	5.01 \pm 0.901	0.932 \pm 0.177	2.62 \pm 0.524	1.34 \pm 0.281
Cu	18.4 \pm 7.60	7 \pm 1.47	34.5 \pm 6.49	9.20 \pm 1.74
Zn	38.5 \pm 7.89	49.2 \pm 10.2	97.9 \pm 8.66	80.4 \pm 13.2
Se	1.88 \pm 0.282	1.72 \pm 0.275	0.696 \pm 0.125	1.12 \pm 0.201
Br	5.7 \pm 1.08	11.8 \pm 2.47	11.2 \pm 7.91	14 \pm 12.9
Rb	20.4 \pm 13.9	29.5 \pm 15.5	49.2 \pm 13.8	43.3 \pm 9.09
Sr	121 \pm 22	92 \pm 12	113 \pm 20	176 \pm 23

Table.3. Concentration of elements ($\mu\text{g/g}$) in *AndrographisPaniculata* medicinal plant from four regions by ICP-MS.

Elements	Vizianagaram	Visakhapatnam	Vijayawada	Tadepalligudem
Li	0.710 \pm 0.196	ND	ND	1.05 \pm 0.30
Be	0.057 \pm 0.007	0.062 \pm 0.006	0.034 \pm 0.005	0.181 \pm 0.009
Al(%)	0.261 \pm 0.040	0.127 \pm 0.013	0.199 \pm 0.013	0.483 \pm 0.068
V	7.08 \pm 0.46	2.61 \pm 0.36	3.26 \pm 0.45	6.78 \pm 0.46
Cr	3.82 \pm 0.24	22.4 \pm 1.5	3.37 \pm 0.40	12.6 \pm 0.71
Mn	64.0 \pm 3.9	116 \pm 9	70.4 \pm 5.5	92.2 \pm 5.7
Fe(%)	0.297 \pm 0.027	0.185 \pm 0.023	0.204 \pm 0.017	0.460 \pm 0.049
Co	3.46 \pm 0.15	0.189 \pm 0.010	1.94 \pm 0.13	0.769 \pm 0.032
Ni	2.78 \pm 0.36	8.64 \pm 0.82	1.51 \pm 0.20	2.85 \pm 0.30
Cu	17.0 \pm 1.1	6.12 \pm 0.48	28.2 \pm 1.9	7.60 \pm 0.58
Zn	31.7 \pm 2.3	39.7 \pm 2.7	53.6 \pm 3.6	52.6 \pm 3.4
As	0.281 \pm 0.030	0.212 \pm 0.025	0.220 \pm 0.009	0.421 \pm 0.025
Se	0.144 \pm 0.046	0.141 \pm 0.038	0.072 \pm 0.020	0.236 \pm 0.044
Rb	19.7 \pm 1.2	21.6 \pm 1.6	23.5 \pm 1.4	39.7 \pm 2.6
Sr	91.1 \pm 9.9	78.7 \pm 9.4	66.2 \pm 14.5	136 \pm 30
Ag	0.022 \pm 0.007	0.103 \pm 0.037	0.041 \pm 0.019	0.370 \pm 0.147
Cd	0.020 \pm 0.004	0.082 \pm 0.012	0.018 \pm 0.003	0.051 \pm 0.006
Ba	34.4 \pm 0.8	127 \pm 5	95.5 \pm 2.3	79.8 \pm 5.0
Pb	1.67 \pm 0.28	1.21 \pm 0.17	1.45 \pm 0.107	5.23 \pm 0.660
U	0.090 \pm 0.020	0.021 \pm 0.005	0.067 \pm 0.015	0.451 \pm 0.104

From the obtained results, it is evident that significant levels of zinc are present in the analyzed medicinal plant. Analyses of the sample from Vijayawada region by both techniques yielded highest levels of zinc. It has been reported that zinc plays an important role in the production, storage and regulation of insulin and its deficiency is linked with several complications such as hypertension, retinopathy, thrombosis, and increased tissue resistance to insulin action in type 2 DM [17],[18]. Deficiency of intracellular Zn makes the beta cells of pancreas more vulnerable to free radical attack.

Chromium is not just a critical cofactor in insulin action but it is also an active component of glucose tolerance factor, which is required for optimal glucose utilization by the cells [19]. Some studies have shown that by supplementing Cr, blood sugar levels were decreased and also reduced the insulin requirement by the diabetic patients [20]. The highest concentration of Cr in the present study was observed in the samples collected from Visakhapatnam region. Even though very low levels of toxic elements As, Cd and Pb are determined by ICP-MS technique, this necessitates the adoption of precautionary measures while prescribing dosage of the herbal medicine prepared from this plant for the treatment diabetes mellitus.

The variations in the elemental composition of *AndrographisPaniculata* medicinal plant collected from four different regions can be attributed to several factors like climatic

conditions, mineral composition of the soil in which the plant grows, the age of the plant, fertilizers used and environmental pollution levels.

4. Conclusion

In the present study on elemental analysis of *Andrographis Paniculata* medicinal plant collected from four different regions of Andhra Pradesh, various elements of interest were identified and quantified. Since diabetes mellitus is associated with marked alterations in the concentrations of trace elements, regulation of trace element concentrations is being proposed as a potential strategy for preventing and treating DM.

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References

- [1] M. Roumie, M. Chiari, A. Srouf, H. Sa'adeh, A. Reslan, M. Sultan, M. Ahmad, G. Calzolari, S. Nava, Th. Zubaidi, M.S. Rihawy, T. Hussein, D.-E. Arafah, A.G. Karydas, A. Simon, and B. Nsouli, "Evaluation and mapping of PM_{2.5} atmospheric aerosols in Arasia region using PIXE and gravimetric measurements," Nuclear Instruments and Methods in Physics Research Section B, vol. 371, pp. 381-386, Mar. 2016.
- [2] S. Bado, R. Padilla-Alvarez, A. Migliori, B.P. Forster, M. Jaksic, Y. Diawara, R. Kaiser, and M. Laimer, "The application of XRF and PIXE in the analysis of rice shoot and compositional screening of genotypes," Nuclear Instruments and Methods in Physics Research Section B, vol. 371, pp. 407-412, Mar. 2016.
- [3] Christopher M. Heirwegh, John L. Campbell, and Gerald K. Czamanske, "Refinement of major- and minor-element PIXE analysis of rocks and mineral," Nuclear Instruments and Methods in Physics Research Section B, vol. 366, pp. 40-50, Jan. 2016.
- [4] Jack E. Manuel, Bibhudutta Rout, Szabolcs Z. Szilasi, Gyanendra Bohara, James Deaton, Henry Luyombya, Karen P. Briski, and Gary A. Glass, "Fish gelatin thin film standards for biological application of PIXE," Nuclear Instruments and Methods in Physics Research Section B, vol. 332, pp. 37-41, Aug. 2014.
- [5] P.F.C. Jobim, C.E.I. dos Santos, N. Maurmann, G.K. Reolon, R. Debastiani, T.R. Pedroso, L.M. Carvalho, and J.F. Dias, "Analysis of memory consolidation and evocation in rats by proton induced X-ray emission," Nuclear Instruments and Methods in Physics Research Section B, vol. 332, pp. 224-228, Aug. 2014.
- [6] Enamorado-Báez, S.M, J.M. Abril, and J.M. Gómez-Guzmán, "Determination of 25 Trace Element Concentrations in Biological Reference Materials by ICP-MS following Different Microwave-Assisted Acid Digestion Methods Based on Scaling Masses of Digested Samples," ISRN Analytical Chemistry, vol. 2013, pp. 1-14, May 2013.
- [7] Şerife Tokalioğlu, "Determination of trace elements in commonly consumed medicinal herbs by ICP-MS and multivariate analysis," Food Chemistry, vol. 134, no. 5, pp. 2504-2508, Oct. 2012.
- [8] Alan R. Date and Alan L. Gray, "Determination of trace elements in geological samples by inductively coupled plasma source mass spectrometry," Spectrochimica Acta Part B, vol. 40, no. 1-2, pp. 115-122, 1985.

- [9] Breda Novotnik, Tea Zuliani, Janez Scancar, and Radmila Milacic, "Content of trace element and chromium speciation in Neem powder and tea infusions," *Journal of Trace Elements in Medicine and Biology*, vol. 31, pp. 98-106, Apr. 2015.
- [10] Archana Kolasani, Hong Xu, and Mary Millikan, "Evaluation of mineral content of Chinese medicinal herbs used to improve kidney function with chemometrics," *Food Chemistry*, vol. 127, no. 4, pp. 1465-1471, Aug. 2011.
- [11] R.P. Choudhury and A. N. Garg, "Variation in essential, trace and toxic elemental contents in Murraykoenigii-A spice and medicinal herb from different Indian states," *Food Chemistry*, vol. 104, no. 4, pp. 1454-1463, Feb. 2007.
- [12] G. J. Naga Raju, P. Sarita, G. A. V. Ramana Murty, M. Ravi Kumar, B. Seetharami Reddy, M. John Charles, S. Lakshminarayana, T. Seshi Reddy, S. Bhuloka Reddy, and V. Vijayan, "Estimation of trace elements in some anti-diabetic medicinal plants using PIXE technique," *Applied radiation and isotopes*, vol. 64, no. 8, pp. 893-900, Feb. 2006.
- [13] G. J. Naga Raju, P. Sarita, J. Chandra Sekhara Rao, C. B. Rao, and S. Bhuloka Reddy, "Correlation of trace elemental content in selected anticancer medicinal plants with their curative ability using particle induced x-ray emission (PIXE)," *Journal of Medicinal Plants Research*, vol. 7, no. 16, pp. 1081-1086, Apr. 2013.
- [14] G. J. Naga Raju, P. Sarita, M. Ravi Kumar, G. A. V. Ramana Murty, B. Seetharami Reddy, S. Lakshminarayana, V. Vijayan, P. V. B. Rama Lakshmi, Gavarasana Satyanarayana and S. Bhuloka Reddy, "Trace elemental correlation study in malignant and normal breast tissue by PIXE technique," *Nuclear Instruments and Methods in Physics Research Section B*, vol. 2, no. 247, pp. 361-367, Jun. 2006.
- [15] G. J. Naga Raju, P. Sarita, G. A. V. Ramana Murty, M. Ravi Kumar, B. Seetharami Reddy, S. Lakshminarayana, K. Prema Chand, A. Durga Prasad, S. Bhuloka Reddy, V. Vijayan, P. V. B. Rama Lakshmi, and Gavarasana Satyanarayana, "Trace elemental analysis of normal, benign hypertrophic and cancerous tissues of the prostate gland using the particle-induced X-ray emission technique," *European journal of cancer prevention*, vol. 16, no. 2, pp. 108-115, May 2007.
- [16] J.L. Campbell, N.I. Boyd, N. Grassi, P. Bonnicks, and J.A. Maxwell, "The Guelph PIXE software package IV," *Nuclear Instruments and Methods in Physics Research Section B*, vol. 268, no. 20, pp. 3356-3363, Oct. 2010.
- [17] A. B. Chausmer, "Zinc, insulin and diabetes," *Journal of American College of Nutrition*, vol. 17, no. 2, pp. 109-115, Apr. 1998.
- [18] M. Soinio, J. Marniemi, M. Laakso, K. Pyörälä, S. Lehto, and T. Rönkä, "Serum zinc level and coronary heart disease events in patients with type 2 diabetes," *Diabetes Care*, vol. 30, no. 3, pp. 523-528, Mar. 2007.
- [19] R. A. Anderson, N. Cheng, N. A. Bryden, M. M. Polansky, N. Cheng, J. Chi, and J. Feng, "Elevated Intakes Of Supplemental Chromium Improve Glucose And Insulin Variables In Individuals With Type 2 Diabetes. *Diabetes*," vol. 46, no. 11, pp. 1786-1791, Nov. 1997.
- [20] R. A. Anderson, "Chromium, glucose tolerance, diabetes and lipid metabolism," *J. Adv. Med*, vol. 8, pp. 37-49, 1995.