

Effect of benzyl amino purine and indole-3-acetic acid on propagation of *Sterculia foetida* in vitro

E Yuniastuti, C E Widodo, Samanhudi and M N I Delfianti

Study Program of Agrotechnology, Faculty of Agriculture, Universitas Sebelas Maret Surakarta, Indonesia

E-mail: yuniastutisibuea@staff.uns.ac.id

Abstract. *Sterculia foetida* is an oval seed plants that can be used as biofuel, which is one of the environmental friendly fuels. This plant is quite hard to find because not many peoples cultivate the plants. An in vitro propagation is one way to preserve the plant. This research aimed to determine optimum concentration of benzyl amino purine (BAP) and indole-3-acetic acid (IAA) to propagate *S. foetida* in vitro. The results showed that woody plant medium (WPM) added by 4 mg L BAP⁻¹ and 0.5 mg L IAA⁻¹ was able to produce complete plantlet, whereas those added by 4 mg L BAP⁻¹ and 1 mg L IAA⁻¹ generated the best growth of shoot and leaves.

1. Introduction

Sterculia foetida is a type of potential plant to be developed as a source of biofuel which has not been cultivated [1]. This plant can be grown in the tropics and sub-tropics (30°N-35°S). Oval-shaped seed is white with yellow seed coat [2]. The main composition of dry beans is the fats (51.78%), protein (21.61%), starch (12.1%), sugar (5%), cellulose (5.51%) and ash (3.9%) [3].

In this research, propagation of plant through tissue culture. Propagation by tissue culture is expected to provide seed mass, uniforms and all year round. Tissue culture also produces pathogen-free plant and germplasm storage [4]. Tissue culture media used, are woody plant medium (WPM) with plant growth regulator (PGR) benzyl amino purine (BAP) and indole-3-acetic acid (IAA). The BAP is a class of cytokines with function for cell division in meristematic tissue, stimulates the differentiation of the cells generated in meristem and encourage the growth of side shoots, leaves apical dominance and expansion. The IAA is a commonly used class of auxin in tissue culture serves to spur the process of cell elongation. This research aimed to determine optimum concentration of BAP and IAA to propagate *S. foetida* by in vitro.

2. Methods

Research was held at Laboratory of Plant Physiology and Biotechnology, Faculty of Agriculture Universitas Sebelas Maret Surakarta. The explants were maintained in the green house for three months. Materials used in the study were explant from stem of *S. foetida*, BAP (0, 2, 4, and 6 mg L⁻¹), IAA (0, 0.5 and 1 mg L⁻¹) and some nutrients used in the manufacture of WPM medium. The data were analyzed using descriptive methods that describe the results of observational studies.



3. Results and discussion

3.1. Callus and shoot emergence

Callus induction from petiole explant was readily obtained within 2 weeks [5]. Formation of callus culture media is very important to observe because it is hopeful to promote callus differentiation. In this research, callus that first appeared in the initiation medium was white translucent with crumb texture. The results of BAP and IAA with varying concentrations to produce callus growth is uneven (Fig. 2a). The emergence of the fastest callus was found in the treatment of 4 mg L BAP⁻¹ and 4 mg L BAP⁻¹ + 1 mg L IAA⁻¹ are two weeks after planting, while largely the result of interaction of BAP and IAA capable of producing callus on the 4th week after planting. The IAA concentration of 0.5 mg L⁻¹ and 1 mg L BAP⁻¹ can accelerate emergence of a callus. According Rao and Purohit [6] high concentrations of auxin stimulates callus formation and morphogenesis pressing.

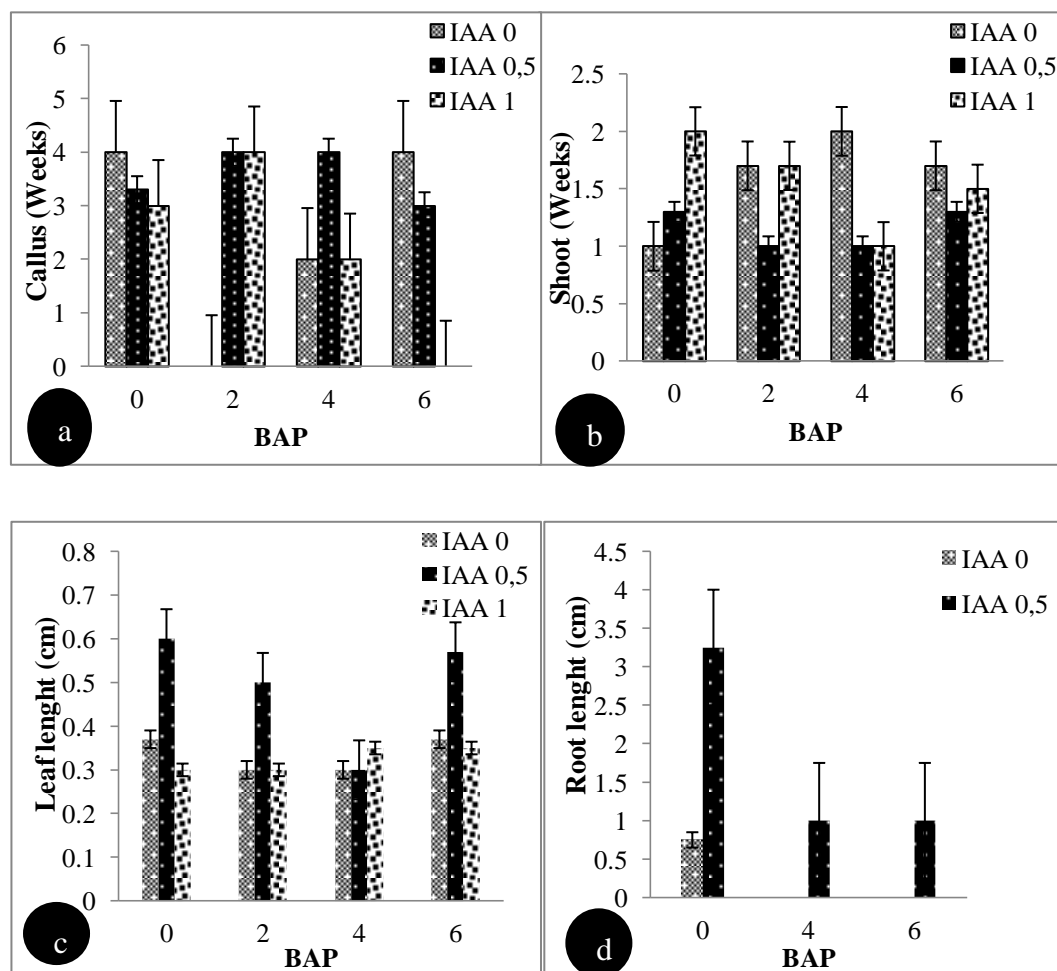


Figure 1. (a) Effect of BAP and IAA on the callus emergence of *Sterculia foetida* explant, (b) shoot emergence, (c) leaf length, (d) root length

The emergence of shoots characterized by their greenish bulge in the armpit leaves. Shoots growing from the meristem tissue composed of components promeristem or the basics of the new organs begin to form. The morphological diversity of plant organs depends on the developmental origin primordia, shoot meristem generate leaves [7]. The highest shoot regeneration potential was observed on stem and leaf segments [5]. In this research, the shoots appeared first in the treatment of 2

mg L BAP⁻¹ + 0.5 mg L IAA⁻¹; 4 mg L BAP⁻¹ + 0.5 mg L IAA⁻¹; 4 mg L BAP⁻¹ + IAA 1 mg L⁻¹; and control (Fig. 2b). The most of the four treatments shoots appear in the first week. The addition of IAA at 6 mg L BAP⁻¹ in the culture medium can accelerate the emergence of shoots, because the nature of the IAA at low concentration between 0.5-1 mg L⁻¹ combined with high concentrations of cytokines can trigger the growth of shoots [8].

3.2. Leaf Length

Leaves for the plant have an important role because it the center of the leaf photosynthesis. Although the mechanism of formation of a crop that can perform photosynthesis by itself, conditions *in vitro* have not clearly known yet [9]. Number of leaves in this research was calculated based on observations of plantlets at 12 weeks after planting. The number of leaves was affected by the addition of plant growth regulators into the media.

The average number of leaves was ranged from 1 to 3 sheets. (Fig. 2b). Number of leaves that appear in visible explants varied. Variations in the number of leaves is possible because the endogenous hormone levels are not exactly the same so its response to the addition of plant growth regulator also varies [10]. Combination of BAP and IAA can successfully leverage in tissue culture if using specific concentration. Application of auxin with cytokinin (BAP) at the appropriate concentration can spur the growth of explants, especially in the formation of leaf, shoot and intensive segment [11]. The role of auxin in the plant growth of tissue culture, especially arranged leaf expansion [12].



Figure 2. (a) Number of leaves with 0 mg L BAP⁻¹ and 0.5 mg L IAA⁻¹. (b) 4 mg L BAP⁻¹ and 0.5 mg L IAA⁻¹

In leaf morphogenesis, the control of cell proliferation seems to be related to the control of cell size [13]. Leaf growth is a process of differentiation leaf buds, with the addition of plant growth regulator such as auxin and cytokinin encourage the differentiation process. Leaf growth begins periclinal their division followed by cell growth in children and cause a bulge, is leaf primordia. Leaf surface area will increase caused by the activity of cleavage anticlinal [14].

The average length of the largest leaf obtained at 0.5 mg L IAA⁻¹ treatment is 0.6 cm and a combination of 6 mg L BAP⁻¹ and 0.5 mg L IAA⁻¹ is 0.57 cm (Fig. 1c). High concentration of BAP combined with 0.5 mg L IAA⁻¹ had leaves longer, while some combination of BAP and IAA treatment of most other produce leaf length of 0.3 cm. This is because the ability of each cell receiving a different PGR response and have optimal limit [15]. High concentrations of BAP were able to increase leaf area of explant [16].

In most of the higher plants, the root is the part that contained in the soil, the plant mainly serves the cantilever body and for the absorption of water and minerals [17]. Plantlets root formation is one

thing that is advantageous, because it can increase the growth during the process of propagation *in vitro*. Many roots that come out so much more facilitate and expedite absorption of nutrients and water to optimal plant growth.

3.3. Root Shoot

Adding BAP and IAA response for explant able to produce a single root. It also occurred in the research [18] that were capable of producing a single root from stem explant. Roots in explant representing 1. The roots can be directly formed on the explants, either from the network or callus, if the media is given with sufficient auxin. Low concentrations of auxin could stimulate root growth [19].

The roots of plants are an important part in the growth. The plant has root length also good position in the growth media. The fact that developmental stages of root cells are roughly correlated with distance from the apical meristem [20]. Root elongation process is accelerated by the presence of auxin in the media. Elongation is caused by enlargement of cells that are formed when the embryo is still developing in the mother plant [14].

The highest root length was obtained from the concentration of 0 mg/LBAP + 0.5 mg/LIAA is 3.25 cm (Fig. 1d). Control also produce roots that is 0.75 cm, but the results were higher than the results of additional treatment BAP 4 mg/L and 6 mg/L. The higher concentration of BAP generate root length. The effect of BAP on root elongation activity, concentrations of cytokines is thought to stimulate cell division effective root [21].

4. Conclusions

It can be concluded that: (a) WPM medium added by 4 ppm BAP and 0.5 ppm IAA is able to produce complete plantlet by forming buds, leaves and roots. (b) WPM medium added by 4 ppm BAP and 1 ppm IAA produce the best growth of shoot and leave. (c) Callus can be formed in almost any combination of BAP and IAA treatment.

Acknowledgements

The authors acknowledged the financial support by INSINAS RISTEK DIKTI 2016.

References

- [1] Sudrajat D J, Nurhasybi and Dida S 2011 Technology for improving the germination of seeded seeds (*Sterculia foetida* Linn.) *Forest Plant Res.* **8** 301–14
- [2] Yuniastuti E 2008 Intensive research program report on the identification and selection of plant pragmatism (*Sterculia foetida*) as well as plant propagation technology in vitro for the provision of biofuel raw materials (Surakarta: Faculty of Agriculture, Sebelas Maret University)
- [3] Ong H C, Silitonga U S, Masjuki H H, Mahlia T M I and Chong W T 2013 Production and comparative fuel properties of biodiesel from non-edible oils: *Jatropha curcas*, *Sterculia foetida* and *Ceiba pentandra* J. *Energy. Convers. Manag.* **73** 245–55
- [4] Gaspar T, Kevers C, Penel C and Greppin H 1996 Plant hormones and plant growth regulators in plant tissue culture *In Vitro Cell. Dev. Biol-Plant* **32** 272–89
- [5] Sujatha N and Mukta N 1996 Morphogenesis and plant regeneration from tissue cultures of *Jatropha curcas* *PCTOC* **44** 135–41
- [6] Rao M S and Purohit S D 2006 In vitro shoot bud differentiation and plantlet regeneration in *Celastrus paniculatus* Wild. *Biol. Plant.* **50** 501–506
- [7] Benkova E, Michniewicz M, Saurer M and Teichmann T 2003 Local, efflux-dependent auxin gradients as a common module for plant organ formation *Cell* **115** 591–602.
- [8] George E F and Sherring ton P D 1984 *Plant Propagation by Tissue Culture* (Eversley England: Exegetics Limited)
- [9] Wetherell D F 1982 *Introduction to In Vitro Plant Propagation* (New Jersey: Avery Publishing

- Group Inc)
- [10] Widyawati G 2010 *Effect of NAA and BAP concentration variation on Jatropha curcas L. induction of jatropha callus* Thesis Faculty of Agriculture University of March Surakarta (ID).
 - [11] Basri Z 2008 Multiplication of four varieties of chrysanthemum through tissue culture techniques *J Agroland* 15(04): 271-277.
 - [12] Teale W D, Paponov I A and Palme K 2006 Auxin in action: signalling, transport and the control of plant growth and development *Mol Cell Biol* 7: 846-859.
 - [13] Tsukaya H 2003 Organ shape and size: a lesson from studies of leaf morphogenesis *Curr Opin Biotechnol* 6: 57-62.
 - [14] Salisbury F B dan Ross C W 1995 *Plant Physiology III* (Bandung (ID): ITB).
 - [15] Santoso J, Mathius N T, Sastraprawira U, Suryatmana G dan Saodah D 2004 Propagation of quinine plants (*Cinchona ledgeriana* moens) and (*C. Succirubra* Pavon) through doubling of axillary buds *A Tower Plantation* 72(1):11-27.
 - [16] Romano A, Noronha C and Matins-loucao MA 1992 Influence of growth regulators on shoot proliferation in *Quercus suber* L. *Ann. Bot* 70: 531-536.
 - [17] Zulkarnain 2013 *Plant Crop Culture Crop Breeding Solution*. (Jakarta (ID): PT. Earth Script).
 - [18] Roostika I, Sunarlim N dan Mariska I 2005 Mikropropagasi of mangosteen plant (*Garcinia mangostana*) *J AgroBiogen* 1(1): 20-25.
 - [19] Deore A C and Johnson T S 2008 High frequency of plant regeneration from leaf-disc cultures of *Jatropha curcas* L.: an important biodiesel plant *Plant Biotechnol Rep* 2: 7-11.
 - [20] Birnbaum K, Shasha D E, Wang J Y, Jung J W, et al. 2003 A gene expression map of the *Arabidopsis* root. *Science* 302: 1956-1960.
 - [21] Pishesha P A, Mattjik N A dan Sukma D 2008 The influence of IAA, IBA, BAP, and coconut water concentration on the formation of poinsettium root (*Euphorbia pulcherrima* Wild Et Klotz) in vitro. Paper seminar of agronomy and horticulture department. Faculty of Agriculture IPB. Bogor (ID). URL: <http://repository.ipb.ac.id/handle/123456789/2640>.