

PAPER • OPEN ACCESS

## The Effectiveness of *Derris elliptica* (Wall.) Benth Root Extract Against Temephos-resistant *Aedes aegypti* Larvae

To cite this article: Sayono *et al* 2019 *IOP Conf. Ser.: Earth Environ. Sci.* **292** 012052

View the [article online](#) for updates and enhancements.

# The Effectiveness of *Derris elliptica* (Wall.) Benth Root Extract Against Temephos-resistant *Aedes aegypti* Larvae

Sayono, A Permatasari, D Sumanto

Faculty of Public Health, Universitas Muhammadiyah Semarang

Corresponding author: say.epid@unimus.ac.id

**Abstract.** *Derris elliptica* (Wall.) Benth is a wild plant or weed in agricultural fields that was traditionally used for poisoning fish and plant pests. Several studies showed the low lethal concentration of *Derris elliptica* root extract to control the laboratory strain *Aedes aegypti* larvae. This study aimed to evaluate the effectiveness of *Derris elliptica* root extract against the temephos-resistant *Aedes aegypti* larvae from the Dengue endemic areas. This experimental study used post-test only control group design with five different concentrations of methanolic *Derris elliptica* root extract, namely 0.09%, 0.13%, 0.17%, 0.21%, and 0.25%, respectively. Twenty-five of F1 temephos-resistant of wild-caught *Aedes aegypti* larvae were subjected to each group of the five different concentrations, and two control groups with aquadest. The temephos-resistant strain of *Aedes aegypti* larvae was determined by bioassay test with temephos -0.02 ppm compound. Mortality rate and lethal concentration (LC50 and LC90) of the larvae were calculated after 24-h contact. Average of mortality rate in each concentration after 24-h contact with *Derris elliptica* root extract were 32.8%, 49.6%, 72.0%, 95.2%, and 100% with LC50 and LC90 were 1,600 and 2,040 ppm, respectively. The methanolic *Derris elliptica* root extract have larvicidal potential to control the temephos-resistant *Aedes aegypti* larvae. Further research to obtain secondary metabolites is needed.

**Keywords:** *Derris elliptica*, *Aedes aegypti* larvae, temephos-resistant, Dengue vector

## 1. Introduction

*Derris elliptica* (Wall.) Benth or tuba is one of the wild plants found in farmland of South and Southeast Asia [1]. The root of the tuba plant was traditionally used as the fish poison and crop pest insecticide [2,3]. This plant contains many kinds of toxic chemical compounds [4], mainly flavonoid group [5] including rotenone, toxicarol, elliptone, sumatrol, tephrosin, and deguelin [6,7]. Recently, using tuba root extract has developed in various bioactive compounds exploration not only for the agricultural field but also public health intervention, especially in Dengue control programs.

Chemical compounds of tuba root extract have been tried to control the diseases vector mosquitoes including the Dengue vector, *Aedes aegypti* and *Aedes albopictus* [8,9] since this disease affected a half of world population, mainly in the tropic and subtropical regions [10]. Dengue prevention is done by combining the vector control program and vaccination [11], although the vaccine is still under researched [12], and the chemical methods use are preferred by people in dengue endemic areas rather than others [13]. Temephos is one of the chemical compounds that world widely used to control *Aedes aegypti* larvae since the last three decades [14-17]. This phenomenon caused the emergence of temephos-resistant strains of *Aedes aegypti* world widely.

The resistance of *Aedes aegypti* larvae to temephos was reported from several countries in the world, such as Brazil [18,19], Columbia [20], Argentina [21], India [22], French West Indies [23]. In Indonesia, temephos resistant populations of this Dengue vector were reported from Jakarta [24], West Java Province [25,26], Surabaya [27], Tanjung Emas Harbor of Semarang [28], and Kendari – Southeast Sulawesi [29]. The emergence of temephos-resistant strains of *Aedes aegypti* larvae inhibits



Dengue control program in the endemic areas. Trial in using herbal compounds is an alternative effort for finding a new solution to control the resistant-strains of *Aedes aegypti* larvae.

Several previous studies about using a bioactive compound of tuba root extract to control *Aedes aegypti* larvae were done in different origin and with different extraction solvents. The studies used laboratory strain mosquito and showed various results. Our study in Semarang using four different of ethanolic extract of tuba root ranged from 0.5 to 4 percent showed that mortality rate of *Aedes aegypti* larvae ranged from 86 – 100 percent, and the effective concentration was 2% above [8]. Another study from Manado, North Sulawesi also used ethanol solvent and laboratory strain of *Aedes aegypti* larvae and showed the lethal concentration of 50% (LC50) was 44.75 ppm [9]. In Malaysia, a similar study use four solvent variations of methyl chloride and methanol comparison showed that the 0.05 mg/ml of tuba root extract caused 83.33% of larvae mortality [30].

The use of different extraction solvent of tuba root resulted in different lethal concentration of laboratory strain of *Aedes aegypti* larvae mortality. Another study showed that methanol solvent results the highest extractable solid rather than hexane, ethanol, and acetone [31]. This study aimed to study the effectiveness of methanolic extract of tuba root to control the field strain *Aedes aegypti* larvae which resistant to temephos 0.02 mg/L.

## 2. Material and methods

### 2.1. *Derris elliptica* origin, determination, and extraction.

Root of tuba plant was obtained from Jepara district, Central Java Province, Indonesia. Determination test for this plant was conducted at the Chemical Laboratory of Mathematical and Natural Sciences Faculty of Universitas Padjadjaran, Bandung, Indonesia. Extraction process based on the previous method [32] by using methanol solvent was done at the Chemical Laboratory of Mathematical and Natural Sciences Faculty of Universitas Negeri Semarang, Semarang, Indonesia.

### 2.2. *Aedes aegypti* larvae collection and rearing.

Larvae of *Aedes* mosquitoes were collected from perimeter areas of Tanjung Emas Harbor, Semarang, Indonesia. Based on the previous study [28], the *Aedes aegypti* population was resistant to temephos 0.02 mg/l. Larvae were reared into adult stage and subjected to the morphological species identification based on the Walter Reed Biosystematics Unit procedure [33]. The eggs of *Aedes aegypti* mosquito resulted before from the identification test were reared in the cage and allowing the mating and resulting the first generation (F1). The 3-5 days of F1 larvae were subjected to temephos 0.02 mg/l bioassay test for determining the recent resistance status [34] of the population to the temephos compound. The temephos bioassay test resulted from the 73% of mortality of *Aedes aegypti* larvae; indicating the resistance status to the compound.

### 2.3. Experimental design.

The posttest-only control group design experiment was conducted at the Epidemiology and Tropical Diseases Laboratory of Public Health Faculty of Universitas Muhammadiyah Semarang. Preliminary bioassay test using the tuba root extract was designed based on the previous study [9] based on concentration ranged from 10, 50, 100, 500, and 1,000 ppm, and resulting the concentration 1,000 ppm (0.1%) causing the 52% of mortality. Based on the results, experiments were performed with concentration ranged from 0.09 %, 0.13 %, 0.17 %, 0.21 %, dan 0.25 %. As many as the 25 of third instar of *Aedes aegypti* larvae (temephos-resistant strain from Tanjung Emas, Semarang) in each group were subjected to the five groups of experiment with five replications, and accompanied by the 25 larvae of control in the cup with tap water. Larvae were contacted for 24 hours to the solution of tuba root extract, and the knockdown larvae were recorded at the 30, 60, 120, 240, 480, and 1440 minutes. Mortality rate of *Aedes aegypti* larvae were recorded after 24 hours contacting with the tuba root extract solution.

### 2.4. Data analysis.

Descriptive and analytical data analysis was performed by the SPSS statistical software using the one-way analysis of variance and probit test. Results of the data analysis were presented in the tables and figures.

### 2.5. Ethical consideration.

Ethics approval was obtained from the Ethic Committee of Health Research of Public Health Faculty of Universitas Muhammadiyah Semarang with registration number 125/KEPK-FKM/UNIMUS/2018.

## 3. Results

*Derris elliptica* root extract with the tested-concentration range has the larvicidal effect to the temephos-resistant strain of *Aedes aegypti* larvae. Average of the knockdown *Aedes aegypti* larvae increased based on the concentration of the extract, and one hundred percent of knockdown rate was reached at the 24 hours exposure by the concentration of 0.25% (Figure 1).

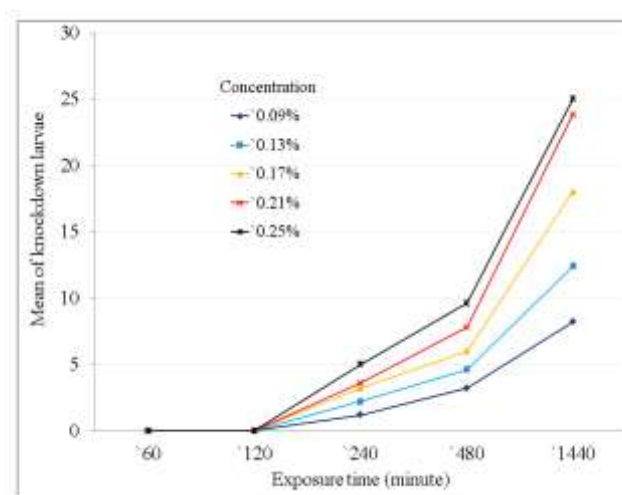


Figure 1. The knockdown of *Aedes aegypti* larvae based on exposure time and tuba root extract concentration.

Different concentration of tuba root extract caused significantly difference of the temephos-resistant of *Aedes aegypti* larvae mortality after 24-h exposure (Figure 2). The lowest and highest mortality rates (32.8% and 100%) of *Aedes aegypti* larvae were showed by the 0.09% and 0.25% of concentrations (Table 1). Probit analysis showed that the effective concentrations (LC50 and LC90) were 0.160 % (0.142 % - 0.170 %) and 0.204% (0.194 % – 0.218 %).

Table1 Mortality rate (%) of the temephos-resistant *Aedes aegypti* larvae after 24 hours exposed to tuba root extract

Concentration (%)	Minimum (%)	Maximum (%)	Average (%)
0,09	20	44	32.80
0,13	40	60	49.60
0,17	64	84	72.00
0,21	88	100	95.20
0,25	100	100	100.00
0 (control)	0	0	0

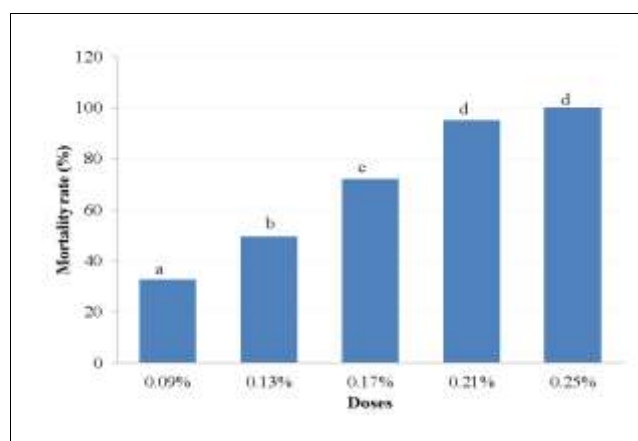


Figure 2. Mortality rate difference of *Aedes aegypti* larvae after 24 hours exposure of tuba root extract concentrations

#### 4. Discussion

Larvacidal potency of tuba root extract for control the primary Dengue vector, *Aedes aegypti* mosquito has been studied and reported in the last decades with various results [7-9]. Two previous studies using laboratory strain of *Aedes aegypti* larvae with different solvent extraction, ethanol and methanol showed the equal result [7,8], but a study from North Sulawesi using ethanol solvent resulted in the lowest LC50, namely 44.75 ppm. The result arose a new hypothesis that the chemical compound containing of the tuba plant is not influenced by solvent, but different geographic habitat. It is similar to the previous report that the different geographic result different chemical compound quantity [35].

This study is the first report that this trial was conducted to increase the efficacy of larvacidal effect of tuba root extract against temephos-resistant *Aedes aegypti* larvae. Knockdown effect of each concentration level of tuba root extract was started at the 2<sup>nd</sup> hour of observation showing the slow toxicity mechanism. The main result showed that the effective dose of tuba root extract for control this strain is 36 folds higher than the laboratory strain. The lowest LC50 of the previous study was 44.75 ppm equal with 0.0045% [9]. It can be understood that the current study uses temephos-resistant *Aedes aegypti* larvae as the research subject so that higher larvacidal doses were needed. Although the resistant ratio of *Aedes aegypti* to organophosphate insecticide was not available yet, but a study in Semarang showed the resistance ratio of this species to pyrethroid compound was 119.75 folds [36]. A comprehensive study to test the larvacidal effect of tuba root extracts by comparing susceptible and temephos-resistant larvae at the same time and place is needed to strengthen these findings.

Toxicity of flavonoids and fatty acid compounds containing in the *Derris elliptica* root extract to mosquito larvae by a unique mechanism affecting both of acetylcholinesterase site and octopaminergic systems [37]. Another compound, saponin affect mosquito larvae by causing the change of food taste, digestive disorders, and cell damage resulting in disruption of growth and death [38,39].

In the past, the resistance of *Aedes aegypti* larvae to temephos has been reported world wide, included Indonesia [18-29]. This condition inhibits the Dengue vector control program based on chemical method, and necessary to be solved. This study provided the new promising results to obtain the new effective chemical compounds for control the Dengue vectors, especially the temephos-resistant strains in the Dengue endemic areas. Further investigation is needed to understand the kinds, role, and quantity of the secondary metabolites that have the high potency for control the resistant mosquito vectors as well as the larvacidal potency for controlling larvae of the other mosquito vectors. Several aspects are necessary to be understood such as larvacidal efficacy of tuba root extract based on degradation period, residual effect, persistence to temperature, and water replacement effect to the toxicity of the extract to *Aedes aegypti* and the other species larvae as well as the microbes in the household water container. The most important advantage of the use of *Derris elliptica* root extract is easily degradable with short residual effect.

## 5. Acknowledgment

The authors wish to thank Dean of Public Health Faculty and Head of Epidemiology and Tropical Diseases Laboratory of Universitas Muhammadiyah Semarang for their kindness in permitting us to conduct the laboratory study at the Epidemiology and Tropical Diseases laboratory; Head of Chemical Laboratory of Mathematical and Natural Sciences Faculty of Universitas Padjadjaran, Bandung, and Universitas Negeri Semarang, Indonesia. The authors wish to thank Puji Lestari Mudawamah from the Epidemiology and Tropical Diseases Laboratory of Universitas Muhammadiyah Semarang for her assistance with the bioassays experiment works.

## References

- [1] Starr F, Starr K, Loope L. 2003. *Derris elliptica*: poison vine, fabaceae; 1-4. Available from: [http://www.hear.org/starr/hiplants/reports/pdf/derris\\_elliptica.pdf](http://www.hear.org/starr/hiplants/reports/pdf/derris_elliptica.pdf).
- [2] Orwa C, A Mutua, Kindt R, Jamnadass R, S Anthony. 2009 Agroforestry Database: a tree reference and selection guide version 4.0 (<http://www.worldagroforestry.org/sites/treedbs/treedatabases.asp>)
- [3] Sirichamorn Y, Adema FACB, Gavendell B, Van Welzen PC. 2012. Phylogeny of palaeotropic *Derris*-like taxa (Fabaceae) based on chloroplast and nuclear DNA sequences shows reorganization of (infra) generic classifications is needed. *American Journal of Botany* 99(11): 1793–1808. <http://doi.org/10.3732/ajb.1200390>.
- [4] Gupta RC. 2007. Rotenone. In *Veterinary Toxicology: Basic and Clinical Principles*. Academic Press: 499-501.
- [5] Musa W, Ahmad J, Lamangantjo CJ. 2016. Bioactive Compounds in Tombili Seeds and Tubile Roots as the Alternative for Synthetic Pesticide to Protect Wheats from Insects and Pests. *International Journal of ChemTech Research* 9(4):604-615. [http://www.sphinxesai.com/2016/ch\\_vol9\\_no4/3/\(604-616\)V9N4CT.pdf](http://www.sphinxesai.com/2016/ch_vol9_no4/3/(604-616)V9N4CT.pdf)
- [6] Yoon AS. 2009. Extraction and Formulation Development of *Derris elliptica* for Insect Pest Control. Thesis of Doctoral Program of Philosophy in Pharmaceutical Sciences Prince of Songkla University. <http://kb.psu.ac.th/psukb/bitstream/2010/7072/1/316615.pdf>
- [7] Zubairi SI, Sarmidi MR, Aziz RA. 2014. Bio-Active Constituents of Rotenoids Resin Extracted from *Derris elliptica* Roots: Comparison between Local Plant Extract and SAPHYR (France) Cube Resin. *Advances in Environmental Biology* 8(4): 904-909
- [8] Sayono, Suryani M, Nurullita U. 2010. Pengaruh konsentrasi flavonoid dalam ekstrak akar tuba (*Derris elliptica*) terhadap kematian larva *Aedes aegypti*. *J Kesehat Masy Indones*. 6(1):38-47. [https://www.researchgate.net/publication/292205811\\_PENGARUH\\_KONSENTRASI\\_FLAVONOID\\_DALAM\\_EKSTRAK\\_AKAR\\_TUBA\\_Derris\\_elliptica\\_TERHADAP\\_KEMATIAN\\_LARVA\\_Aedes\\_aegypti](https://www.researchgate.net/publication/292205811_PENGARUH_KONSENTRASI_FLAVONOID_DALAM_EKSTRAK_AKAR_TUBA_Derris_elliptica_TERHADAP_KEMATIAN_LARVA_Aedes_aegypti)
- [9] Komansilan A, Suriani NW, Lawalata H. 2017. Test toxic tuba root extract as a natural insecticide on larvae of *Aedes aegypti* mosquito vector of Dengue fever. *International Journal of ChemTech Research* 10(4): 522-528. . [http://sphinxesai.com/2017/ch\\_vol10\\_no4/3/\(522-528\)V10N4CT.pdf](http://sphinxesai.com/2017/ch_vol10_no4/3/(522-528)V10N4CT.pdf)
- [10] Ebi KL, Nealon J. 2016. Dengue in a changing climate. *Environmental Research* 151: 115–123. <http://dx.doi.org/10.1016/j.envres.2016.07.026>
- [11] Katzelnick LC, Coloma J, Harris E. 2017. Dengue: Knowledge gaps, unmet needs and research priorities. *Lancet Infectious Diseases* 17(3): e88-e100. doi:10.1016/S1473-3099(16)30473-X.
- [12] Rather IA, Parrav HA, Lone JB, Paek WK, Lim J, Bajpai VK, Park YH. 2017. Prevention and control strategies to counter Dengue virus infection. *Front. Cell. Infect. Microbiol.* 7:336. doi: 10.3389/fcimb.2017.00336

- [13] Krianto T. 2009. Masyarakat Depok memilih fogging yang tidak dimengerti. KESMAS, Jurnal Kesehatan Masyarakat Nasional 4(1): 29-35. <http://dx.doi.org/10.21109/kesmas.v4i1.198>  
<http://journal.fkm.ui.ac.id/kesmas/article/view/198>
- [14] Donalisio MRC, Leite OF, Mayo RC, Pinheiro-Alves MJC, de-Souza A, Rangel O, Andrade VR, de-Oliveira SS, Matias VLM. 2002. Use of Temephos for Control of Field Population of *Aedes aegypti* in Americana Sao Paulo, Brazil. Dengue Bulletin 26:173-177. WHO Regional Office for South-East Asia. <http://www.who.int/iris/handle/10665/163769>
- [15] George L, Velayudhan R, Lenhart A, Toledo J, Lazaro A, Han WW, Ranzinger SR, Horstick O. 2015. Community-Effectiveness of Temephos for Dengue Vector Control: A Systematic Literature Review. PLoS Negl Trop Dis 9(9): e0004006. doi:10.1371/journal.pntd.0004006
- [16] Chaiphongpachara T, Moolrat L. 2017. Insecticide resistance of temephos on *Aedes aegypti* as dengue vector in SamutSongkhram, Thailand. *Ann Trop Med Public Health* 10:1439-42. Available from: <http://www.atmph.org/text.asp?2017/10/6/1439/222636> [cited 2019 Jan 12]
- [17] Marcombe S, Chonephetsarath S, Thammavong P, Brey PT. 2018. Alternative insecticides for larval control of the dengue vector *Aedes aegypti* in Lao PDR: insecticide resistance and semi-field trial study. *Parasites and Vectors* 11:616. <https://doi.org/10.1186/s13071-018-3187-8>.
- [18] Diniz MMCSL, Henriques ADS, Leandro DS, Aguiar DL, Beserra EB. 2014. Resistance of *Aedes aegypti* to temephos and adaptive disadvantages. *Rev Saude Publica* 48(5):775-782. doi: 10.1590/S0034-8910.2014048004649.
- [19] Bellinato DF, Viana-Medeiros PF, Araujo SC, Martins AJ, Lima JBP, Valle D. 2016. Resistance Status to the Insecticides Temephos, Deltamethrin, and Diflubenzuron in Brazilian *Aedes aegypti* Populations. *BioMed Research International*. Article ID 8603263. <http://dx.doi.org/10.1155/2016/8603263>
- [20] Grisales N, Poupardin R, Gomez S, Fonseca-Gonzalez I, Ranson H, Lenhart A. 2013. Temephos Resistance in *Aedes aegypti* in Colombia Compromises Dengue Vector Control. *PLoS Negl Trop Dis* 7(9): e2438. <https://doi.org/10.1371/journal.pntd.0002438>
- [21] Llinas GA, Seccacini E, Gardenal CN, Licastro S. 2010. Current resistance status to temephos in *Aedes aegypti* from different regions of Argentina. *Mem. Inst. Oswaldo Cruz* 105(1): 113-116. <http://dx.doi.org/10.1590/S0074-02762010000100019>.  
<http://www.scielo.br/pdf/mioc/v105n1/19.pdf>
- [22] Shetty V, Sanil D, Shetty NJ. 2015. Inheritance Pattern of Temephos Resistance, an Organophosphate Insecticide, in *Aedes aegypti* (L.). *Genetics Research International*. Article ID 181872. <http://dx.doi.org/10.1155/2015/181872>
- [23] Goindin D, Delannay C, Gelasse A, Ramdini, Gaude T, Faucon F, David JP, Gustave J, Vega-Rua A, Fouque F. 2017. Levels of insecticide resistance to deltamethrin, malathion, and temephos, and associated mechanisms in *Aedes aegypti* mosquitoes from the Guadeloupe and Saint Martin islands (French West Indies). *Infectious Diseases of Poverty* 6(38). DOI: 10.1186/s40249-017-0254-x
- [24] Prasetyowati H, Hendri J, Wahono T. 2016. Status Resistensi *Aedes aegypti* (Linn.) terhadap Organofosfat di Tiga Kotamadya DKI Jakarta. *BALABA* 12(1):23-30. <http://ejournal.litbang.depkes.go.id/index.php/blb/article/download/4454/4359>
- [25] Fuadzy H, Hendri J. 2015. Indeks entomologi dan kerentanan larva *Aedes aegypti* terhadap temefos di Kelurahan Karsamenak Kecamatan Kawalu Kota Tasikmalaya. *Vektora* 7(2):57-64. DOI : 10.22435/vk.v7i2.4504.57-64  
<http://ejournal.litbang.depkes.go.id/index.php/vk/article/view/4504>
- [26] Putra RE, Ahmad I, Prasetyo DB, Susanti S, Rahayu R, Hariani N. 2016. Detection of insecticide resistance in the larvae of some *Aedes aegypti* (Diptera: Culicidae) strains from Java, Indonesia to Temephos, Malathion and Permethrin. *International Journal of Mosquito*

- Research* 3(3): 23-28. <http://www.dipterajournal.com/pdf/2016/vol3issue3/PartA/3-2-4-590.pdf>
- [27] Mulyatno KC, Yamanaka A, Ngadino, Konishi E. 2012. Resistance of *Aedes aegypti* (L.) larvae to temephos in Surabaya. *The Southeast Asian J Trop Med Public Health* 43(1):29-33. <http://www.tm.mahidol.ac.th/seameo/2012-43-1/05-5304.pdf>
- [28] Handayani N, Santoso L, Martini M, Purwantisari S. 2016. Status resistensi larva *Aedes aegypti* terhadap temephos di wilayah perimeter dan buffer Pelabuhan Tanjung Emas Kota Semarang. *Jurnal Kesehatan Masyarakat* 4(1):159-166. <http://garuda.ristekdikti.go.id/journal/article/442167>
- [29] Mubarak M, Satoto TBT, Umniyati SR. 2015. Analisis Penggunaan Insektisida Malation dan Temefos Terhadap Vektor Demam Berdarah Dengue *Aedes aegypti* di Kota Kendari Sulawesi Tenggara. *Medula* 2(2):134-142. <http://ojs.uho.ac.id/index.php/medula/article/view/2542>
- [30] Zubairi SI, Sarmidi MR, Aziz RA, Latip R, Said J. 2004. The effect of rotenone crude extract from *Derris elliptica* on the larvicidal activity (mortality) of mosquito. Proceeding of Symposium Biologi Kebangsaan ke-7, 18-20 May 2004, Awana Genting Highlands, Pahang, Malaysia. [http://eprints.utm.my/id/eprint/9800/1/ZubairiSI2004\\_the\\_effect\\_of\\_rotenone\\_crude\\_extract.pdf](http://eprints.utm.my/id/eprint/9800/1/ZubairiSI2004_the_effect_of_rotenone_crude_extract.pdf)
- [31] Ngo TV, Scarlett CJ, Bowyer MC, Ngo PD, Vuong QV. 2017. Impact of Different Extraction Solvents on Bioactive Compounds and Antioxidant Capacity from the Root of *Salacia chinensis* L. *Journal of Food Quality* 2017;1-8 ID 9305047. <https://doi.org/10.1155/2017/9305047>
- [32] Dumewa A, Syarifah, Fitriah S. 2016. Pengaruh ekstrak batang Brotowali (*Tinospora crispa*) terhadap kematian larva nyamuk *Aedes aegypti*. *Jurnal Biota* 2(2): 166-172. <http://jurnal.radenfatah.ac.id/index.php/biota/article/view/738>
- [33] Walter Reed Biosystematics Unit. <http://www.wrbu.org/>
- [34] WHO. 2005. Guidelines for laboratory and field testing of mosquito larvicides. WHO/CDS/WHOPES/GCDPP/2005.13. [http://apps.who.int/iris/bitstream/handle/10665/69101/WHO\\_CDS\\_WHOPES\\_GCDPP\\_2005.13.pdf;jsessionid=ADABDB70794E5911F960BD5AB19C365D?sequence=1](http://apps.who.int/iris/bitstream/handle/10665/69101/WHO_CDS_WHOPES_GCDPP_2005.13.pdf;jsessionid=ADABDB70794E5911F960BD5AB19C365D?sequence=1)
- [35] Moore RH. 1946. Some effects of altitude and water supply on the composition of *Derris elliptica*. *Botanical Gazette* 107(4):467-474. [https://www.jstor.org/stable/2472678?seq=1#page\\_scan\\_tab\\_contents](https://www.jstor.org/stable/2472678?seq=1#page_scan_tab_contents)
- [36] Sayono S, Hidayati APN, Fahri S, Sumanto D, Dharmana E, Hadisaputro S, Asih PBS, Syafruddin D. 2016. Distribution of Voltage-Gated Sodium Channel (Nav) Alleles among the *Aedes aegypti* Populations In Central Java Province and Its Association with Resistance to Pyrethroid Insecticides. *PLoS ONE* 11 (3): e0150577. doi:10.1371/journal.pone.0150577
- [37] Perumalsamy H, Jang MJ, Kadarkarai M, Ahn Y-J. 2015. Larvicidal activity and possible mode of action of four flavonoids and two fatty acids identified in *Millettia pinnata* seed toward three mosquito species. *Parasit Vectors* 8:237. DOI 10.1186/s13071-015-0848-8. [https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4410478/pdf/13071\\_2015\\_Article\\_848.pdf](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4410478/pdf/13071_2015_Article_848.pdf)
- [38] Geyter ED, Lambert E, Geelen D, Smagghe G. 2007. Novel advances with plant saponins as natural insecticides to control pest insects. *Pest Technology* 1(2):96-105. [http://www.globalsciencebooks.info/Online/GSBOOnline/images/0712/PT\\_1\(2\)/PT\\_1\(2\)96-105o.pdf](http://www.globalsciencebooks.info/Online/GSBOOnline/images/0712/PT_1(2)/PT_1(2)96-105o.pdf)
- [39] Jawale CS. 2014. Larvacidal activity of some saponin containing plants against the Dengue vector *Aedes aegypti*. *Trend in Biotechnology Research* 3(1):1-11. [www.sciencejournal.in](http://www.sciencejournal.in)