

Review Article

L-Arginine and Alzheimer's Disease

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Abstract: Alzheimer's disease (AD), the most common form of dementia, is characterized by progressive neurodegeneration and loss of cognitive and memory functions. Although the exact causes of AD are still unclear, evidence suggests that atherosclerosis, redox stress, inflammation, neurotransmitter dysregulation, and impaired brain energy metabolism may all be associated with AD pathogenesis. Herein, we explore a possible role for L-arginine (L-arg) in AD, taking into consideration known functions for L-arg in atherosclerosis, redox stress and the inflammatory process, regulation of synaptic plasticity and neurogenesis, and modulation of glucose metabolism and insulin activity. L-arg, a precursor of nitric oxide and polyamine, exhibits multiple functions in human health and may play a prominent role in age-related degenerative diseases such as AD.

Key Words: L-arginine; nitric oxide synthase; nitric oxide; arginase; polyamines; neurogenesis, stem cells, Alzheimer's disease

Introduction

Alzheimer's disease (AD) is an age-related neurodegenerative disease with an insidious onset, characterized by memory impairment and cognitive disturbances that become increasingly more severe with disease progression. It is a debilitating and dehumanizing illness, inflicting immense suffering on its victims and their families, and on society. Approximately 4.5 million Americans are currently affected by AD [1]. However, if there are no effective strategies to treat or prevent AD [2], it is projected to affect up to 9 million people by 2040 as the elderly population grows.

The neuropathology of AD is characterized by senile plaques, neurofibrillary tangles (NFT), and neuronal loss [3-6]. Although the exact causes of AD are still unknown, studies suggest that the genesis of sporadic AD is associated with atherosclerosis, redox stress, inflammatory processes, and/or abnormal neurotransmission and brain glucose metabolism. Current treatment strategies are

limited to altering cholinergic and NMDA neurotransmission and show only modest efficacy. No treatments are currently available to target the underlying mechanism of the disease.

L-arginine (L-arg) is a semi-essential, proteinogenic amino acid [7] that was discovered in mammalian protein by Hedin in 1895 [8], and since 1886 it has been recognized as a naturally occurring molecule [9]. It is involved in two major metabolic pathways as shown in **Figure 1**. One of them is the nitric oxide synthase (NOS) pathway where L-arg is converted to NO and L-citrulline [10, 11]. The other pathway is the arginase pathway that will be discussed further below.

There are three isoforms of NOS that have been discovered so far. They are named according to the cell types from which they were first isolated: neuronal NOS (nNOS), inducible NOS (iNOS) and endothelial NOS (eNOS) [10, 12]. These NOSs have different functions [10-13]. The expression of nNOS and eNOS are constitutive and regulated by

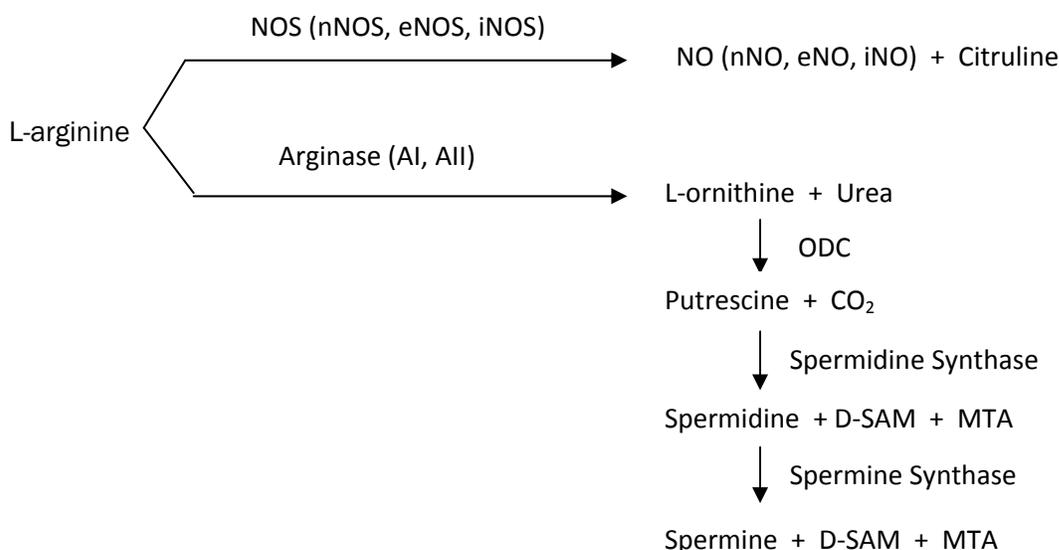


Figure 1 The major two metabolic pathways of L-arginine. ODC, ornithine decarboxylase; D-SAM, decarboxylase SAM; MAT, methylthioadenosine.

calcium/calmodulin. Neuronal NO (nNO) and endothelial NO (eNO) are produced at low rates by nNOS and eNOS, respectively [14]. The relationship of L-arg to the isoforms of NOS is intricate. Noticeably, the intracellular L-arg concentration (about 1-2 mM), taken up and maintained by endothelial cells through the transport system, is so much higher than the K_m value of purified eNOS ($\approx 2.9 \mu\text{mol/L}$) that eNO should not be increased further by addition of extracellular L-arg. However, the “L-arginine paradox” that synthesis of eNO can be enhanced as a response in a concentration-dependent manner to the increase of extracellular L-arg concentration has been observed [13, 15]. This reaction plays a crucial role in the vascular homeostasis associated with L-arg [16]. In terms of iNOS, its expression is induced in inflammatory cell types by cytokine stimulation, and its activity is independent of calcium, and production rate of inducible NO (iNO) is high [17].

L-arg and NO affect the cardiovascular system as endogenous antiatherogenic molecules that protect the endothelium, modulate vasodilatation, and interact with the vascular wall and circulating blood cells [18-22]. Together, they can function in the brain as noradrenergic, noncholinergic neurotransmitters in learning and memory, synaptic plasticity, and neuroprotection [23, 24]. They can influence the immune system too by

playing a key role in regulating inflammatory processes [25] and redox stress. They can also modulate the metabolism of glucose and insulin activity as natural constituents from diets [26] and regulate neurogenesis. Since L-arg and its product, NO, exert such a range of critical roles in regulating physiological functions of brain and other organs, we hypothesize that L-arg can possibly affect the AD pathogenesis. The other metabolic pathway that involves L-arg is the arginase pathway where L-arg is broken down into urea and L-ornithine and genesis of polyamines including putrescine, spermidine, and spermine [27, 28]. Two isoforms of arginase (AI and AII) [29] were discovered in 1973 [30, 31], identified positively in 1983 [32], and confirmed subsequently in 1989 [33]. They are encoded by different genes, distributed in different tissues, cell types and intracellular locations, and, have different biochemical properties [28, 34, 35]. AI, called liver-type arginase, was first found as a component of the urea cycle. It is expressed at high level in livers as a cytosolic enzyme and at a low level in central nerve system (CNS). It is also induced to express at a high level when exposed to multiple cytokines and factors in various tissues and cells [28, 34-36]. AII is called kidney-type arginase and is expressed at a low level in the mitochondrion, and it too can be induced by cytokines. Like AI, it is also expressed in the germinal zones, hippocampus, spinal, and

other motor neurons of mice [37, 38]. Loss of AI leads to potentially fatal hyperammonemia and hyperargininemia, states characterized by a series of stereotypic clinical disorders such as growth retardation, increased mental impairment, and spasticity [39-40]. However, these symptoms can be partially attenuated through enhancing the expression and activity of AI1 to compensate for the deficiency of AI [41-44]. Based on the distribution and expression of these isoforms, we postulate that AI and AI1 might participate in many physiological processes, including inflammation, neurogenesis and apoptosis.

Polyamines are the major products of L-arg metabolized by arginase. Ornithine acts as a starting substrate to be converted into putrescine, spermidine and spermine. There are three main polyamines that can be identified with their different lengths of carbon chains [45, 46]. They act as variably functional molecules that are essential for cell regeneration, tissue growth, and development [47-51].

In this review, we explore a possible role for L-arg in AD, taking into consideration the known functions of L-arg in atherosclerosis, oxidative stress and the inflammatory process, regulation of synaptic plasticity and neurogenesis, and modulation of glucose metabolism and insulin activity.

The Possible Effects of L-Arg on AD via Anti-atherosclerosis

The Relationship between AD and Atherosclerosis

Increasing evidence suggests a strong relationship between AD and atherosclerosis. Indeed, some investigators have proposed that AD is a primary neurovascular disease [52].

First, AD and atherosclerosis have many risk factors in common [53-55]. Numerous studies have shown that established risk factors for vascular disease, including diabetes mellitus, smoking, and atherosclerosis, also predispose individuals to AD [56-61].

Second, autopsy series have provided evidence of links between atherosclerosis and AD [62, 63]. Seward *et al* found that the atherosclerotic lesions and the degree of stenosis of Circle of Willis are significantly

more severe in AD brains than in age-matched controls. Additionally, the index of stenosis apparently relates to the total plaque score, neuritic plaque score, NFT score, Braak stage score, and white matter rarefaction score, all of which are measures for AD neuropathological lesion [62]. Beach *et al* also reported that increase in the atherosclerotic grade increased the odds ratios for the diagnoses of AD and vascular dementia (VaD) [63]. Furthermore, studies suggest that the possible mechanism through which atherosclerosis influences the development of AD is hypoperfusion in the brain [62, 63]. Additionally, based on previous points, Torre *et al* and other researchers found it possible to clinically diagnose AD earlier through neuroimaging techniques such as single-photon emission computed tomography (SPECT) because the presence of microvascular abnormalities precedes cognitive impairment and neurodegeneration [52, 64, 65, 66]. Hirao *et al* reported that subjects with reduced regional cerebral blood flow in the bilateral temporo-parietal areas and the precuneii will finally become AD cases [67].

Third, some studies have shown that treatment of atherosclerosis may also benefit AD. Sparks *et al* suggested that administration of atorvastatin to patients with AD may attenuate cognitive decline and generally slow down the progression of mild-to-moderate AD [68]. That study agrees with others in which statins were used as the treatment for AD [69-71]. Petanceska *et al* even found that administration of atorvastatin can significantly reduce A β amyloid deposition in an animal model [72].

In summary, increasing evidence suggests that atherosclerosis is associated with the AD progression. Interdicting atherosclerosis might therefore delay the onset or slow the progression of AD.

L-Arg Affects AD via Anti-atherosclerosis

L-arg exerts its function in the cardiovascular system mainly through the increase of NO production [73-75]. Lack of L-arg in vascular endothelium may result in the deficiency of NO [16], a key feature in the development of atherosclerosis (18). Thus, abnormalities in L-arg availability and metabolism are proposed in the pathogenesis of atherosclerosis, especially in hypercholesterolemia [76].

Creager *et al* discovered that forearm vasodilatation is markedly improved through administration of L-arg in an endothelium-dependent manner [77]. Similar results were seen in other studies [78, 79]. In fact, the effect is more profound than that observed after lipid-lowering therapy [80-82]. Other studies obtained parallel results in patients with hypercholesterolemia [78, 83]. From previous studies, hypercholesterolemia as a risk factor of atherosclerosis is well known to cause early endothelial dysfunction, abnormal interactions between vascular cells, platelets and monocytes [84, 85], and disability of L-arg [76]. However, extra dietary supplements of L-arg may decrease platelet aggregation [82, 86] and mononuclear cell adhesiveness in hypercholesterolemic patients [87, 88]. Furthermore, thiobarbituric acid reactive substances (a marker of lipid peroxidation) are decreased after L-arg infusion in hypercholesterolemic subjects [89]. Recent studies showed that chronic oral supplementation with L-arg may block the progression of atherosclerotic plaques via restoration of NOS substrate availability and decrease of vascular stress [90, 91].

Hypertension, an established risk factor for atherosclerosis is strongly associated with AD [92, 93]. Therefore, through its effect on hypertension, L-arg may affect AD. Siani *et al* reported that oral administration of L-arg as an enriched diet in healthy volunteers caused a reduction in arterial blood pressure [94]. Rector *et al* showed that arterial blood pressure dropped in patients with heart failure after treatment with L-arg [95]. The study also reported that acutely oral L-arg improves brachial artery flow-mediated dilation in patients with essential hypertension [78].

Cigarette smoking, another salient risk factor for atherosclerosis may also be affected by L-arg and be linked to AD. An association between smoking and an increased risk of dementia has been reported [59, 96, 97], although not always [98, 99, 100]. Smoking raises oxidative stress to degenerate NO through increasing oxygen-derived free radicals and lipid peroxides [101]. It also accelerates monocyte adhesion and the vulnerability of low density lipoprotein (LDL) to be oxidized [102]. L-arg can affect atherosclerosis through attenuating the effects of smoking. Using treatment with extra L-arg, Adams *et al* reported that adhesion of

monocyte and endothelial cells and the expression of intercellular adhesion molecule in endothelial cells are decreased [103]. Other studies also showed that the microcirculation is improved by L-arg supplementation in smokers [76, 104, 105].

The mechanisms through which L-arg affects atherosclerosis are not fully understood, and a number of possible mechanisms have been proposed, including the "L-arginine paradox". Excess L-arg can enhance NOS activity through NO production, especially when battling with the deficiency of eNO in the presences of LDL cholesterol [106], by acting as (i) a relaxing factor in the regulation of vasodilatation [107]; (ii) an inhibitor to attenuate platelet aggregation [108], and monocyte and leukocyte adhesion [109]; (iii) an inhibitor to depress the proliferation of smooth muscle cells [110]; and (iv) reducer of vascular oxidative stress and the expression of redox-regulated genes [111]. It is worth mentioning that only eNO is helpful to anti-atherosclerosis, whereas iNO accelerates atherogenesis through synthesis of the cytotoxic NOO⁻ radical [112]. Further, exertion of its function by L-arg upon the cardiovascular system is concentration-dependent. At lower plasma concentrations, L-arg can selectively improve endothelial function so that patients with elevated asymmetric dimethylarginine (ADMA) levels have diminished NOS activity; at middle concentration levels, it can perform direct vasodilatation through the endocrine effects of secreting insulin and growth hormone; at higher concentration levels, it can produce vasculature unspecific vasodilatation [113]. Moreover, chronic supplement of L-arg may have anti-hypertensive effect through the reduction of renal vascular resistance and the depression of angiotensin-converting enzyme [114, 115].

In conclusion, L-arg has multiple direct and indirect effects on human vasculature, and might play an important role in the pathogenesis of both atherosclerosis and AD.

L-Arg, as a Precursor of NO, Affects AD via Influencing Oxidative Stress

The Relationship between AD Pathology and Brain Oxidative Stress

Brain oxidative damage is prevalent in AD due to high cerebral energy demand and oxygen

consumption that are required for brain functions and possible failure of brain antioxidant defenses [116]. Numerous experimental data, as indicated by different markers for oxidative damage of DNA, protein, lipid and glucose, shows that oxidative stress plays an important role in AD pathogenesis, and is highly associated with brain A β amyloidosis [117-123]. Much experimental evidence also implies that increased oxidative damage may not just be the consequence but a primary cause of AD pathogenesis [124]. Indeed, A β amyloidogenesis promotes generation of free radicals, oxidative damages, and inflammation in AD brain [125].

In summary, oxidative stress contributes to the progress of AD and there may be a vicious cycle between brain oxidative stress and Alzheimer's A β amyloidogenesis.

L-Arg Affects AD via Influencing Oxidative Stress

NO derived from L-arg is a potential source of redox stress. It can be quickly cleared through reacting with superoxide (O $_2^-$) to generate peroxynitrite (ONOO $^-$) with a half-life of <1 sec while cells are in a pro-oxidative state. As a highly reactive species, ONOO $^-$ can react via homolytic or heterolytic cleavage and, generate secondary constituents of nitroxidative stress and highly reactive oxygen/nitrogen species (ROS/RNS) including NO $_2^+$, NO $_2$, and OH radical. The high nitroxidative stress acts to initiate the redox reaction, thereby inducing apoptosis and overall damage to neurons and endothelial cells [126]. The toxic constituents that are generated from the reaction of NO under oxidative stress are the property of a family called "reactive nitrogen oxidative species (RNOS)," of which peroxynitrite and nitrogen oxide are the main constituents [127, 128]. Furthermore, the term "nitroxidative stress" has been used to indicate the cellular damage that is elicited by excess NO and RNOS [129, 130]. Wang *et al* supported these assertions when they reported neuronal apoptosis induced in a concentration- and time-dependent manner while ONOO $^-$ increased, H $_2$ O $_2$ rapidly decayed, and ROS slowly decreased [131]. Other studies also suggest that NO and ROS are involved in the pathogenesis of AD by synergistically inducing neuronal damage and death [127, 132, 133].

In contrast, David *et al* drew a totally opposite conclusion reporting that NO provided protection against ROS by way of cell culture [134, 135]. They also found that neurons expressing NOS survived under ischemia reperfusion, whereas neurons surrounding the ischemia area and not expressing NOS died [136]. The possible mechanism that NO can attenuate the toxic effects of ROS might be that NO can directly react with O $_2^-$ to form ONOO $^-$; thereby rapidly rearranging nitrate at physiological pH 4.0 before it interacts with cells [135].

Whether NO is neuroprotective or neurotoxic also depends on the different functions of its isoforms, the stage of treatment with corrective drugs [137], the local concentration of NO, especially at different ischemia stages [138, 139], and the concentration of ROS [140]. Glebov *et al* used L-arg and its inhibitor by intravenous injection separately after inducing oxidative stress in rats. They found that iNOS inhibitor improves antioxidant protection, whereas L-arg and the nonselective inhibitor do not [141]. They further suggested that iNO produced by iNOS enhances oxidative stress. Another study showed that NOS activities and the expression of markers for oxidative stress are increased in cell culture and that the use of nNOS inhibitor cannot rescue the cells from dying [142]. The finding suggested that nNOS might not be toxic. It was also reported that iNOS is a mediator of neuroprotection induced by preconditioning with oxidative stress such as H $_2$ O $_2$ at low concentration in a cell culture [140].

In addition, some studies showed that ischemia/reperfusion in the brain possibly causes AD [143, 144]. L-arg can protect it through exerting its anti-oxidant functions. If lacking L-arg and NO, the brain would have an increase of superoxide anion formation [147]. Administration of L-arg may be associated with the antiradical and antioxidant effects of NO, inhibiting the effects of inositol-1,2,5-triphosphates, and inhibiting the accumulation of leukocytes in the reperfused tissue [145, 146]. Maksimovich *et al* suggested that the antioxidant property of L-arg in brain ischemia/reperfusion might be because of activation of NO synthesis, involving eNOS which acts as a radical trap, and facilitating the removal of radical and reductions in their toxicity [148]. However, inhibiting the activity of nNOS and iNOS resulted in improvements in

brain circulation and reduction of the ischemic zone [149]. eNO affects vessel walls by inhibition of lipoxygenase-dependent lipid and lipoprotein oxidation [147, 150]. Further, it affects vessels by its ability to enhance the perfusion of brain tissues via NO-dependent dilation of vessels [151], and neurons by suppression of the N-methyl-D-aspartate (NMDA) receptor activity [150, 152]. Also, eNO affects the prooxidant-antioxidant equilibrium by inducing a shift associated not only with its potentially high levels that can react with the multitude of target molecules responsible for the development of oxidative stress, but also with its decrease to contributions of other factors to the antioxidant potential of the body, especially changes in the oxygen affinity of hemoglobin [153].

Even in patients with hyperlipidemia-hyperglycemia, administration of L-arg can decrease the oxidative stress [154]. Supplementation with L-arg improves oxidative stress by inducing postprandial hypertriglyceridemia [155-157], preventing the depletion of serum plasma glutathione peroxidase that is a serum antioxidant enzyme, and preventing endothelial dysfunction [157, 158].

In conclusion, L-arg and NO can have a dual role in AD under oxidative stress. Their neuroprotective or neurotoxic roles are limited by isoforms and the concentration of ROS.

The Effects of L-Arg on AD via Influencing Inflammation

The Relationship between AD and Inflammation

Increasing evidence shows that chronic inflammatory processes of the central nervous system (CNS) are neurotoxic and may contribute to AD pathogenesis [159]. For example, during inflammation, elevated pentraxins, increased pro-inflammatory cytokines, chemokine alterations and microglial activation trigger functional impairment and structural damage to the CNS [160].

On the other hand, A β as a central mediator in AD pathogenesis [161, 162] may also promote neurodegeneration by inducing the activation of microglial cells and astrocytes. The induction results in the acceleration of

inflammation through releasing various inflammatory mediators [163, 164]. In addition, some epidemiological studies strongly support that non-steroidal anti-inflammatory agents may have therapeutic value in AD [165-168]. We conclude that there is a great potential that improvement in the immune system may prevent CNS inflammation, and hence, AD pathology.

L-Arg Regulates Inflammation

Over the last two decades, increasing evidence suggests that L-arg plays important roles in immunological processes.

L-arg is a potent modulator of immune cell functioning [25]. Kirk *et al* fed mice with 1% arginine HCL and found an increase in thymic weight due to increased number of total thymic T lymphocytes [169]. In the athymic mice, arginine supplementation increased the total number of T cells and, amplified delayed-type hypersensitivity responses. In humans, dietary supplementation has been shown to enhance T-cell-mediated function and speed up wound healing by increasing reparative collagen synthesis [170].

The ability of L-arg to regulate immune cell-mediated function depends on its concentration. Albina *et al* found that low concentrations of L-Arg (<0.1 mM) in culture media enhance activation-associated functions in rat resident peritoneal macrophages, including cytotoxicity against tumor cells, superoxide production, and phagocytosis. On the contrary, higher concentrations of L-arg (about 0.1 mM to 1.2 mM) suppress superoxide production, cytotoxicity, phagocytosis and protein synthesis. They also revealed that low concentrations of L-arg enhance phagocytosis probably due to macrophage-derived arginase activity. Probably due to NO⁻ production induced by L-arg/NO pathway [171, 172], higher, non-physiological concentrations of L-arg produce more prominent decreases of phagocytic activity compared with controls - a result that agrees with Potenza *et al* [25]. In summary, L-arg can be a modulator regulating inflammation.

The Effect of L-Arg and NO on AD via Influencing Inflammation

Scott *et al* used L-arg to revise free radical

production and the development of experimental allergic encephalomyelitis (EAE) in a rat model. They found that L-arg can suppress the development of neurological symptoms and the formation of inflammatory lesions in the CNS of diseased animals, eventually efficiently delaying disease onset. They also found that superoxide and hydrogen peroxide are markedly decreased and the level of nitrite, a breakdown of NO formation, is significantly increased in the CNS [173]. In conclusion, they recommended L-arg is a protective molecule, modulating oxidant-mediated neuroinflammation by the production of NO [173]. However, other studies reported that iNO's effect on neurons contributed to neurodegenerative disease [174, 175]. Vodovotz *et al* found that NFT-bearing neurons express iNOS in the brain regions influenced by AD [176]. Others found that nitrotyrosine staining is increased in AD brains tissue [177]. Still other studies suggested that high generation of iNO may contribute to pathogenesis in AD due to sustained exposure and oxidative damage by peroxynitrite - an intermediate iNO reaction product [143, 144]. These results agreed with a prior study [178]. In addition, iNO, as a free radical, activated cyclooxygenase II (COX-2) that in turn activated the arachidonic acid cascade that is known to be pro-inflammatory [179, 180]. All in all, these findings seem to suggest that overproduction of iNO is harmful by inducing the inflammatory process and possibly AD. The discrepancies about the role of NO under oxidative stress have already been elaborated above.

In conclusion, L-arg and NO, as modulators, may play a role in AD by influencing inflammatory processes. Regulating the level and the metabolic pathway of L-arg, and selectively producing different isoforms of NO may produce therapeutic effects. Further investigations are necessary, however, to confirm or comprehend these effects and potentials.

The Effect of L-Arg on AD through Production of the Neurotransmitter NO

NO is a Neurotransmitter

The first evidence that NO acts as a neurotransmitter is reported by Garthwaite *et al*. They showed that stimulation of cerebellar NMDA receptors by glutamate releases NO

[181] that then acts as a neurotransmitter in CNS to regulate the synaptic plasticity involved in cognitive processes, memory, long-term potentiation (LTP) and long-term depression (LTD) [182]. Some evidence has shown that NO, produced presynaptically or in interneurons postsynaptically, acts during cerebellar and striatal LTD. On the other hand, the postsynaptic generation of NO presynaptically acts in hippocampal and cortical LTP [183]. Furthermore, Thomas *et al* found that NO, as a transmitter, modulated synaptic efficacy at the neuromuscular junction. They also demonstrated that NO regulates transmitter release and adenosine-induced depression via a cGMP-dependent mechanism which occurs after Ca²⁺ entry [184-186]. The results agree with Nickels *et al* [187].

The Effects of NO on AD

Since it was found immunohistochemically in rats [151] that NO and neurons are strongly linked via localized NOS protein, researchers supposed that NO as a transmitter is related with AD. Thus, they started further investigations to observe the concentration of NO in the brain with AD and later showed that the concentration of NO is decreased through examining the concentration of transmitters related with NO in cerebrospinal fluid (CSF). Barford *et al* reported that tetrahydrobiopterin (BH4), which is a co-enzyme of NOS [188], is decreased significantly in the AD brain [189]. The reduction of BH4 might induce a diminished NOS activity that then might deteriorate neuronal function and lead to a decrease of NO production in AD [190]. Toghi *et al* reported, which agreed with Lowe *et al* [191], that L-glutamate that is released through stimulation by NO is decreased in the CSF in the AD brain [134]. Kuiper *et al* further confirmed this result and even found that the reduction of the level of glutamate is linked with the increasing age in the patients with AD [192]. The decrease of L-glutamate might therefore contribute to memory impairment in patients with AD [193]. Kuiper *et al* also reported that the nitrate content that is rapidly oxidized from NO is decreased in CSF in AD [194]. The findings suggested that the development of AD might be due to a decrease of NO synthesis [192]. Pazzo *et al* used an NO donor and inhibitor in animal models with AD and suggested that A β -impaired NO generation resulted from reducing NMDA receptor signal

transduction via subtracting NADPH availability to NOS [195, 196]. They and others also found that NO had a protective effect on A β -induced damage of the nervous system [195, 197]. In addition, it was reported that administration of NOS inhibitors did not protect against A β -induced neurotoxicity but that administration of NO donors did exert a neuroprotective effect [198].

On the other hand, Manh *et al* gave chronic intravenous injection of A β 1-40 into the hippocampus in rat models. Then they found that the expression of iNOS and the production of iNO are increased, while the release of acetylcholine (ACh) and dopamine is decreased, a situation believed to be one of the primary causes of cognitive deficits in patients with AD. The rats were then treated with iNOS inhibitors. As a result, the inhibitor of iNOS restored the impairment of ACh and dopamine release and prevented memory impairment. The study indicated the toxic effect of A β on brain function due to NO synthesized by iNOS via dysfunction of cholinergic signaling and that, if treated with iNOS inhibitors, cholinergic dysfunction and memory performance could improve [199]. As an essential transmitter, iNO may contribute to the generation and development of AD.

Why are there so many different results about whether NO is beneficial or harmful to AD? Some studies revealed that NO is a neurotoxic factor in A β -induced synaptic dysfunction and cell death through stimulating iNOS, but not eNOS and nNOS [196, 200-203]. Furthermore, an increase in hippocampal iNOS and a decrease in nNOS in aged rats were observed [204]. So these effects might explain the conflicts about synaptic dysfunction due to activation of iNOS and the lack of synaptic plasticity for downregulation of NO production [205].

In conclusion, eNO and nNO, but not iNO, as transmitters, may have a neuroprotective effect against A β -induced impairment of LTP and ameliorate cognition in patients with AD, though additional studies are warranted.

The Effects of L-Arg on AD via Regulating Glucose Metabolism and Insulin Activity

The Relationship between Glucose Metabolism, Insulin Activity and AD

Converging evidence has confirmed that a potential association exists among metabolism of glucose, insulin activity and AD [206].

Metabolism of glucose appears to play a role in memory. Patients with AD have showed particular abnormalities of glucose homeostasis [207, 208], such as decreased glucose metabolism in the hippocampus, superior and middle temporal gyri and the cingulate gyrus [209, 210] via CMRglc or PET [211-217]. Craft *et al* examined the effects of acute glucose administration on memory in patients with AD and age-matched controls. Glucose administration temporarily improved memory function in both AD patients and controls. However, as compared with controls, it took the AD patients much longer for their glucose levels to return to baseline. The study suggested that patients with AD have less efficient gluco-regulation as compared with controls and that efficient gluco-regulation improves memory in patients with AD [218]. The same results were found in other studies [219-222]. Furthermore, it was investigated that acutely raising plasma or cerebral glucose levels facilitated non-contextual and contextual verbal memory, visual memory, and produced beneficial effects in a variety of learning paradigms. The same effects occurred in patients with AD who accepted acute administration of glucose [218, 223-225].

Administration of glucose with optimal doses might modulate ACh release related with cognition and learning [226]. It was also found out that administration of glucose could reverse deficits induced by cholinergic blockade [227-230] and even directly interact with other neurotransmitter systems including the gamma-aminobutyric acid (GABA) system [231]. The effects of glucose were dose-dependent with an inverted U-shaped function [226, 229]. Specifically, acute hyperglycemia can facilitate memory, whereas chronic hyperglycemia may impair memory, at least in older adults [232]. On the other hand, some investigators found that DM might be associated with an increased risk of developing AD and might affect cognitive systems differently [233-235].

Mild-to-moderate cognitive dysfunction in patients with type I and type II diabetes mellitus (DM1, DM2) may be caused by chronic hyperglycemia [236-238] or insulin

resistance syndrome [239]. Hoyer *et al* established an animal model that mimics the abnormal cerebral glucose/energy metabolism through inhibiting the neuronal insulin receptor to show that oxidative/energy metabolism, phospholipids composition of membranes, cholinergic and catecholaminergic functions, learning memory, and cognition are abnormal as seen in AD [240]. Those findings agree with other studies [241, 242]. Patients with moderate-to-severe AD have also had elevated true plasma insulin levels and decreased CSF insulin levels [243]. Studies showed that AD might be associated with reduced insulin sensitivity [244]. Other clinical studies showed that induced hyperinsulinemia while maintaining euglycemia could facilitate memory for patients with AD and normal adults [245-247]. All of the previous studies revealed that peripheral insulin abnormalities are associated with AD [248].

Raising peripheral insulin levels can improve memory when the level of plasma glucose is normal as insulin might modulate LTP through increasing the cell membrane expression of NMDA receptors [249]. After activity of NMDA receptor, neuronal Ca^{2+} influx is increased to activate α -calcium/calmodulin-dependent-kinase II (aCaMK II) and other Ca^{2+} dependent enzymes, and, finally to boost synaptic associations between neurons [250].

In summary, it is possible that abnormal glucose metabolism and impaired insulin activity contribute to cognitive decline in patients with AD. Regulating glucose metabolism and insulin activity may have positive impacts on these patients.

L-Arg might have Therapeutic Potential in AD through Regulating Glucose Metabolism and Insulin Activity

In DM, impaired production of NO results in impaired NO activity because of the uncoupling of receptor-mediated signal transduction [251-253], a deficiency of the NOS substrate L-arg [254-256], or a reduced availability of one or more cofactors essential for optimal functioning of NOS [257-259]. Excitingly, it was found that L-arg can modulate the glucose metabolism via increasing NO synthesis [260, 261] to normalize plasma glucose levels [262] and attenuate hyperglycemia [263].

Some observations of possible mechanisms about L-arg and NO to regulate metabolism of glucose and insulin activity are as follows.

First, NO normalizes metabolism of glucose via increasing glucose transport. NO donors have increased glucose transport in skeletal muscle, while inhibition of NOS activity blunted contraction-stimulated glucose transport and had no effect on insulin-stimulated glucose transport [264]. Similar results were found from a human vascular smooth muscle cell culture and adipose tissues [265, 266]. These studies showed that NO is capable of stimulating glucose transport through glucose transporter 4 translocation via insulin signaling pathway and the other mechanisms [264-266].

Second, NO increases glucose uptake in various cells. Acute infusion of NO donor resulted in greater glucose uptake, as studies have reported [267, 268]. However, NO has been implicated as an important signaling molecule in the contraction-mediated glucose uptake pathway at low concentrations, and, as an inhibitory molecule at higher concentrations [269, 270].

Third, L-arg regulates insulin release. L-arg stimulates glucose-induced insulin secretion via the NO pathway [271, 272]. It is assayed by the demonstration of expression and production of NOS in insulinoma and primary β -cells, and the insulinotropic action of NO [271]. In addition, L-arg stimulates glucose-induced insulin secretion from pancreatic islets that could occur independently of NO. The secretion of insulin by L-arg is mediated by membrane depolarization via protein kinase A- and C- activation and L-arg-induced Ca^{2+} influx [273]. It was also reported that liver cells can be engineered to produce insulin, and insulin secretion can be induced through treatment with L-arg via the production of NO [274], actions that happen when hepatic NOS are involved in the secretion of a hepatic insulin sensitizing substance that mediates peripheral insulin sensitivity [275].

Fourth, L-arg and NO enhance insulin sensitivity. Guarino *et al* confirmed from that study that insulin sensitivity is enhanced in a dose-dependent manner by co-administration of NO and glutathione (GSH) to the liver [276]. NOS protein expression that is enhanced by chronic exercise implied that NO may play a role in the improved glucose tolerance and

increased insulin sensitivity characteristic of a trained state [264]. However, some studies showed that deficiency of NO increases insulin sensitivity via modified insulin binding capacity and downregulates the expression of gene encoding resistin [277]. Finally, a study showed systemic NOS inhibition could increase human insulin sensitivity [278].

In conclusion, L-arg and NO can regulate the metabolism of glucose and insulin activity that affects AD. Further studies are needed.

The Effects of L-Arg on AD via Neurogenesis

The Relationship between Neurogenesis and AD

One of the characteristics of AD is the loss of neurons [1-4]. Recent studies provide new therapeutic strategies in the treatment of neurodegenerative diseases such as AD including the use of drugs and the transplant of tissues from the ventral mesencephalon [279-285]. An alternate approach is to target neurogenesis.

Neurogenesis in the adult brain of most mammals takes place from neural precursor cells that are derived from adult stem cells in the subgranular cell layer of the dentate gyrus of the hippocampus and in the subventricular zone of the lateral ventricle [286-289]. Precursors divide in the dentate gyrus, mature in the granular cell layer, migrate within the rostral migratory stream, and differentiate rapidly to functionally recruit the lost ones [290-294]. Recent studies have also shown that stem cells isolated from bone marrow or the umbilical cord differentiate into neural precursor cells and neural cell types under specific conditions [295-300]. They even engraft and partially correct a lesion when transplanted into Parkinson disease (PD) models [301-303]. These functional recruits occur and can be enhanced after neurogenesis [304], and are integrated both structurally and functionally into pre-existing neuronal networks [305, 306]. Such findings indicate that neurogenesis in the brain might have potential therapeutic use.

L-arg is attracting increasing attention as a regulator in neurogenesis and apoptosis. Many researchers show that L-arg is involved in different types of cell generation and apoptosis through the following major

metabolic pathways [307-315].

The Effects of L-Arg on Neurogenesis through the Arginase Pathway

Sara et al showed that proliferation of neural stem cells (NSCs) is increased under AD deficiency in a mouse model and that derived NSCs matured and differentiated into neurons more quickly than their counterparts [316]. In addition, it was found that overexpression of AD could accelerate the extension of neurite in older dorsal root ganglial neurons [317, 318]. Extracellular administration of arginase can be antiapoptotic under oxidative stress and the other conditions that induce neuronal apoptosis [319]. Esteve et al also found that arginase acts as a central regulator of trophic factor-deprived motor neuronal survival [320]. These primary *in vivo* and *in vitro* studies indicate that arginase plays a role in the neural cell cycle. Du et al even used arginase as a therapeutic factor to treat focal brain ischemia by combining antiexcitotoxic and antiapoptotic measures rather than using either agent alone [321, 322].

The mechanisms of arginase in neurogenesis are supposed to be as follows: 1) Arginase controls cell proliferation through modulating the number of neural cells in the S-phase of the cell cycle [35]; 2) The expression of genes in cell growth is elevated to increase proliferation but not differentiation during a deficiency of arginase [35]; 3) Arginase is increased as a response of cAMP which is a crucial downstream component of the neurotrophin-induced "regeneration" pathway [47, 323]; 4) Neuron cell survival is increased and apoptosis is decreased through the administration of arginase, a phenomenon possibly due to its clearing up of excitotoxic necrosis in cortical neuronal cultures by reducing the production of NOS [82] and thereby inhibiting NO production [35]; 5) Esch et al demonstrated that the function of arginase to antiapoptosis depended on the depletion of arginine and the inhibition of "death proteins" synthesis [319] which is similar to the findings by Sonoki et al [325]; 6) Arginase exerts its function also through its products: polyamines, which play bivalent functions in neural cell growth and death [326].

Emerging evidence has proved that polyamines are involved in the development of

the CNS [327, 328]. Depletion of polyamines during nervous system development will lead to a deficiency of neuronal morphogenesis [329]. Chu *et al* showed that polyamines are able to improve axonal regeneration of neurons after injury [330, 331]. Malaterre *et al* found that neural progenitor proliferation is significantly increased in dentate gyrus and in the subventricular zone in a rodent brain when it is given putrescine. Conversely, the reduction of polyamines decreases the proliferation of an adult neural progenitor [332]. Cayre *et al* reported that the short-chain putrescine can induce neuronal precursor cells to mitogenesis and, hence, increase proliferation, while the long-chain spermidine and spermine fail to do. In contrast, spermidine and spermine can simulate neuron differentiation and neurite elongation, whereas putrescine cannot alter any morphological character of these interneurons *in vitro*. It is believed that short-chain and long-chain polyamines play specific roles during neurogenesis [333]. Putrescine enhances neuronal proliferation through regulating proto-oncogene transcription and expression, and acting on cell cyclins [334, 335]. Spermidine and spermine enhance differentiation through affecting the major cytoskeletal elements [336], and regulating casein kinase II activity, which participates in neurogenesis [337, 338].

Polyamines are involved in neuronal survival and apoptosis in concentration-dependent manner [330]. Overproduction of polyamines and the increase of their activities can induce death of fibroblasts [339, 340]. Sparapani *et al* found that high concentrations of polyamines are toxic to granule cells in culture. This toxicity is mediated through the NMDA receptor by interaction of exogenously added polyamines with endogenous glutamate released by neurons in the medium, especially spermine and spermidine [341]. In serum-containing medium, polyamines can be cytotoxic while they oxidize to aminoaldehyde and hydrogen peroxide by polyamine oxidases [342-344]. On the other hand, lower concentrations of polyamines prevent apoptotic neuronal death and toxin-and axotomy-induced cell death of sympathetic neurons in cell culture [330]. This protective function is exerted through both NMDA receptor-dependent process that enhance the activities of glutamate and NMDA at the NMDA receptor via the allosteric mechanism [345]

and independent mechanisms [330]. These findings agree with those from other studies [346]. Furthermore, it was reported that only spermine promoted neuronal survival by its trophic effects through an ifenprodil-sensitive mechanism [331, 347, 348].

According to previous studies, suitable concentrations of polyamines are neuroprotective in neurodegenerative models [349, 350], such as ischemic stroke [351]. However, results are contradictory on whether using a polyamine synthesis inhibitor is also neuroprotective in stroke models [352, 353]. Rao *et al* showed that blood-brain barrier breakdown is more severe by putrescine, while breakdown is attenuated by spermine and spermidine after ischemia [353]. However, in stroke models, putrescine is increased, while there is no change of spermine and spermidine, and inhibitor of polyamines did not reduce spermine and spermidine [354]. Further studies are necessary to understand the exact roles of polyamines in such pathological conditions [353-355].

Collectively, in the metabolism of L-arg, arginase can decrease proliferation and differentiation in neurogenesis, whereas it can prevent neuron apoptosis and induce neuron survival. Polyamines, products of L-arg through the arginase pathway, have their specific functions in neurogenesis according to the length of carbon chain. Varying suitable concentrations of polyamines exist both in physiological and pathological conditions that can exert a positive impact on neuronal survival.

The Effect of L-Arg on Neurogenesis through the NOS Pathway

Growing evidence reveals that NO plays a critical role in regulating neurogenesis, neural survival, and apoptosis in CNS. It is reported that NO regulates both proliferation and differentiation of neural stem cells and neural precursor cells. Elisabetta *et al* showed that the effect of NO deprivation during the early cerebellar neurogenesis not only stimulates a brief increase in cell proliferation through reducing availability of cGMP, but also traces into adulthood in rats brain [356]. Torroglosa *et al* found that NO, as a negative regulator, decreased subventricular zone stem cell proliferation through inhibition of epidermal growth factor receptor and phosphoinositide-3-

kinase/Akt pathway, producing an antimitotic effect on neurosphere cells in adult mice [357]. Lopez *et al* also reported that NO physiologically inhibited neurogenesis in the adult mouse subventricular zone and olfactory bulb by controlling the size of the undifferentiated precursor pool and promoting neuronal differentiation [358]. Cheng *et al* demonstrated that the regulation of neurogenesis by NO occurs by its action in a positive feedback loop with brain-derived neurotrophic factor (BDNF) [359]. On the other hand, chronic administration of inhibitor of NOS enhanced neurosphere formation and growth [357], increased proliferation, and decreased the differentiation of precursors [358-360].

However, Zhang *et al* suggested that administration of NO can remarkably increase neuronal progenitor cell proliferation, differentiation, and migration in subventricular zone and the dentate gyrus of the hippocampus of the adult rodent brain [361]. Cheng *et al* also reported that NO induced apoptosis of neural progenitor cells through the p38 MAP kinase pathway [362]. Other studies showed that the apoptosis of neurons is due to oxidative injury induced by NO, which acts as a general trigger [363-365]. On the other hand, stem cell survival in nNOS knockdown animals was increased [366]. The discrepant results might be due to different isoforms of NOS involved in neurogenesis.

Sabrina *et al* found that nNOS has a primary regulatory role in the migration and survival of newly formed neuronal cells, whereas its effect upon stem cell proliferation is less pronounced [367]. In contrast, it is reported that nNOS slows down cell proliferation *in vitro* [368] and signals surviving cells to switch to terminal neuronal differentiation [359, 368]. Also, the administration of nNOS inhibitor enhances cell proliferation [369]. The mechanism behind this might be that nNOS cooperates with BDNF as a positive feedback loop to regulate neural progenitor cell proliferation and differentiation in the mammalian brain [359, 370]. However, a further study showed no difference in the changes of BDNF mRNA or protein in nNOS knockout mice. That suggested that the function of nNOS, when involved in neurogenesis, might be not only dependent on the manner of BDNF, but also another unclear pathway that indirectly switches the young

neural cells from survival to differentiation [367].

Andreas *et al* showed a significant decrease of neuronal progenitor cell proliferation in the dentate gyrus in eNOS knockout mice, accompanied by a reduction in vascular endothelial growth factor (VEGF), without any changes in survival rate of newly formed cells [371]. Other studies also agreed that disruption of eNOS results in significantly decreased levels of VEGF [372, 373]. It suggests that the mechanism of selective effects of eNOS on progenitor cells proliferation might be mediated by regulating the transcription of VEGF in the hippocampus [373] to activated kinase Akt so as to downstream mechanisms and multiple pathways [371, 374, 375]. Conversely, elevating VEGF stimulates the increase of eNO through enhancing eNOS expression [146], which finally results in neurogenesis and angiogenesis that each benefits the other [141, 147]. This reveals that eNOS and VEGF act in a positive feedforward loop [371]. eNOS regulates neurogenesis through the VEGF-mediated manner, while nNOS appears to regulate neurogenesis not only by a BDNF-mediated manner. It demonstrated that nNOS and eNOS exert their effects, by indirect mechanisms, as antagonists in different phases of adult neurogenesis.

Zhu *et al* found that the expression and enzymatic activities of iNOS are elevated in the dentate gyrus after cerebral ischemia [378]. Later their further studies indicated that iNOS is crucial to accelerate neurogenesis, which is associated with enhancing cell proliferation and increasing mature granule neurons in the same area after cerebral ischemia [149]. While using the inhibitor of iNOS or antagonist of NMDA receptor, no increase of neurogenesis was observed [379-381]. It was reported that iNOS is activated quickly through activation of NMDA receptors [382, 383]. It was also reported that the reduction of nNOS and eNOS activities induce iNOS expression which produces iNO to stimulate cell proliferating factors through activated nuclear factor- β in the hippocampus [384, 385]. It also suggested that nNOS and iNOS play an opposite role in regulating neurogenesis in the ischemic hippocampus [384, 385]. However, under physiological conditions, nNO that derived from nNOS depresses iNOS expression

by inhibiting nuclear factor-NFκB activation [386].

Collectively, the metabolism of L-arg through the NOS pathway produces both positive and negative effects on neurogenesis. The authors suggest that the phenomena may be explained by the different function of three isoforms of NOS on neurogenesis. However, more research is recommended on this issue.

In summary, neurogenesis therapies involving stem cells and lineage-committed precursor cells are revolutionizing the concept of neurogenerative medicine. It is being increasingly accepted that generation and transplantation of lineage-committed precursor cells are very important steps in the process. However, the environmental and neurotrophic factors including inducible signals and transmitters around precursors and stem cells are critical to the success of therapy. In this context, we elucidate that L-arg is involved in neurogenesis through its metabolic pathways and its products. Better understanding of the metabolic procedures of L-arg would allow us to selectively choose to accelerate or attenuate some of those metabolic steps so as to contribute a valuable course to neurogenerative therapies for AD.

Conclusion

L-arg is an essential amino acid, involved in diverse physiological and pathological processes, including neurotransmission, neurogenesis and neuroplasticity, cellular redox metabolism and redox stress, inflammation, and regulation of cerebral blood flow. Increasing evidence implicates L-arg in the pathogenesis of diverse age-related diseases, including Alzheimer's disease. Understanding of the precise biochemical roles of L-arg will aid to rational development of therapeutic agents for various relevant human diseases intervention.

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References

- [1] Evans DA, Funkenstein HH, Alpert MS, Scherr PA, Cook NR, Chown MJ, Herbert LE, Hennekens CH and Taylor JO. Prevalence of Alzheimer's disease in a community population of older persons. *JAMA* 1989;262: 2551-2556.
- [2] U.S. Government Printing Office, Washington, DC. National Institute on Aging (1995) Progress Report on Alzheimer's Disease. NIH Publication 95-3994.
- [3] Selkoe DJ. Alzheimer's disease: genes, proteins, and therapy. *Physiol Rev* 2001;81: 741-766.
- [4] Yaari R and Corey-Bloom J. Alzheimer's disease. *Semin Neurol* 2007;27:32-41.
- [5] Goedert M and Spillantini MG. A century of Alzheimer's disease. *Science* 2006;314:777-781.
- [6] Mattson MP. Pathways towards and away from Alzheimer's disease. *Nature* 2004;430: 631-639.
- [7] Rose WC. The nutritional significance of the amino acids. *Physiol Rev* 1938;18:109-136.
- [8] Hedin SG. Eine methode das lysin zu isolieren, nebst einigen Bemerkungen uber das lysatinin. *Z Physiol Chem* 1895;21:297-305.
- [9] Schulze E and Steiger E. Uber das Arginin. *Z Physiol Chem* 1886;11:43-65.
- [10] Moncada S and Higgs A. The L-arginine-nitric oxide pathway. *N Engl J Med* 1993;329: 2002-2012.
- [11] Sidney M and Morris Jr. Regulation of enzymes of the urea cycle and arginine metabolism. *Annu Rev Nutr* 2002;22: 87-105.
- [12] Forstermann U, Closs EI, Pollock JS, Nakane M, Schwarz P, Gath I and Kleinert H. Nitric oxide synthase isozymes. Characterization, purification, molecular cloning, and functions. *Hypertension* 1994;23:1121-1131.
- [13] Boger RH. The pharmacodynamics of L-arginine. *J Nutr* 2007;137(Suppl 2):1650S-1655S.
- [14] Malinski T. Nitric oxide and nitroxidative stress in Alzheimer's disease. *J Alzheimers Dis* 2007;11:207-218.
- [15] Loscalzo J. What we know and don't know about L-arginine and NO. *Circulation* 2000; 101:2126-2129.
- [16] Siasos G, Tousoulis D, Antoniadis C, Stefanadi E and Stefanadis C. L-Arginine, the substrate for NO synthesis: An alternative treatment for premature atherosclerosis? *Int J Cardiol* 2007;116:300-308.

- [17] Forstermann U, Boissel JP and Kleinert H. Expressional control of the constitutive isoforms of nitric oxide synthase (NOS I and NOS III). *FASEB J* 1998;12:773-790.
- [18] Boger RH, Bode-Boger SM and Frolich JC. The L-arginine - nitric oxide pathway: Role in atherosclerosis and therapeutic implications. *Atherosclerosis* 1996;127:1-11.
- [19] Cooke JP and Dzau VJ. Nitric oxide synthase: role in the genesis of vascular disease. *Annu Rev Med* 1997;48:489-509.
- [20] Cooke JP. The pivotal role of nitric oxide for vascular health. *Can J Cardiol* 2004;20(Suppl B):7B-15B.
- [21] Li XA, Everson W and Smart EJ. Nitric oxide, caveolae, and vascular pathology. *Cardiovasc Toxicol* 2006;6:1-13.
- [22] Napoli C, de Nigris F, Williams-Ignarro S, Pignalosa O, Sica V and Ignarro LJ. Nitric oxide and atherosclerosis: An update. *Nitric Oxide* 2006;15:265-279.
- [23] Bohme GA, Bon C, Lemaire M, Reibaud M, Piot O, Stutzmann JM, Doble A and Blanchard JC. Altered synaptic plasticity and memory formation in nitric oxide synthase inhibitor-treated rats. *Proc Natl Acad Sci USA* 1993;90:9191-9194.
- [24] Paakkari I and Lindsberg P. Nitric oxide in the central nervous system. *Ann Med* 1995;27:369-377.
- [25] Potenza MA, Nacci C and Mitolo-Chieppa D. Immunoregulatory effects of L-arginine and therapeutic implications. *Curr Drug Targets Immune Endocr Metabol Disord* 2001;1:67-77.
- [26] Jobgena WS, Friedb SK, Fuc WJ, Meiningerd CJ and Wu G. Regulatory role for the arginine-nitric oxide pathway in metabolism of energy substrates. *J Nutr Biochem* 2006;17:571-588.
- [27] Wu G and Morris SM Jr. Arginine metabolism: nitric oxide and beyond. *Biochem J* 1998;336:1-17.
- [28] Iyer R, Jenkinson CP, Vockley JG, Kern RM, Grody WW and Cederbaum S. The human arginases and arginase deficiency. *J Inherit Metab Dis* 1998;1:86-100.
- [29] Sidney M and Morris SM Jr. Regulation of enzymes of the urea cycle and arginine metabolism. *Annu Rev Nutr* 2002;22:87-105.
- [30] Glass RD and Knox WE. Arginase isozymes of rat mammary gland, liver and other tissues. *J Biol Chem* 1973;248:5785-5789.
- [31] Kaysen GA and Strecker HJ. Purification and properties of arginase of rat kidney. *Biochem J* 1973;133:779-788.
- [32] Spector EB, Rice SCH and Cederbaum SD. Immunologic studies of arginase in tissues of normal human adults and arginase-deficient patients. *Pediatr Res* 1983;17:941-944.
- [33] Grody WW, Argyle C, Kern RM, Dizikes GJ, Spector EB, Strickland AD, Klein D and Cederbaum SD. Differential expression of the two human arginase genes in hyperargininemia: enzymatic pathologic and molecular analysis. *J Clin Invest* 1989;83:602-609.
- [34] Lange PS, Langley B, Lu P and Ratan RR. Novel roles for arginase in cell survival, regeneration, and translation in the central nervous system. *J Nutr* 2004;134(Suppl):2812S-2817S.
- [35] Becker-Catania SG, Gregory TL, Yang Y, Gau CL, de Vellis J, Cederbaum SD and Iyer RK. Loss of arginase I results in increased proliferation of neural stem cells. *J Neurosci Res* 2006;84:735-746.
- [36] Boeshore KI, Schreiber RC, Vaccariello SA, Sachs HH, Salazer R, Lee J, Ratan RR, Leahy P and Zigmund RE. Novel changes in gene expression following axotomy of a sympathetic ganglion: a microarray analysis. *J Neurobiol* 2004;59:216-235.
- [37] Yu H, Iyer RK, Kern RT, Rodriguez WI, Grody WW and Cederbaum SD. Expression of arginase isozymes in mouse brain. *J Neurosci Res* 2001;66:406-422.
- [38] Yu H, Iyer, RK, Yoo PK, Kern RM, Grody WW and Cederbaum SD. Arginase expression in mouse embryonic development. *Mech* 2002;115:151-155.
- [39] Terheggen HF, Schwenk A, Lowenthal A, van Sande M and Colombo JPZ. Hyperargininemia with arginase deficiency, a new familial metabolic disease: clinical aspects. *Kinderheilk* 1970;107:298-312.
- [40] Scaglia F and Lee B. Clinical, biochemical, and molecular spectrum of hyperargininemia due to arginase I deficiency. *Am J Med Genet C Semin Med Genet* 2006;142:113-120.
- [41] Cederbaum SD, Shaw KNF, Spector EB, Verity MA, Snodgrass PJ and Sugarman GI. Hyperargininemia due to arginase deficiency. *Pediatr Res* 1979;13:827-833.
- [42] Spector EB, Rice SCH and Cederbaum SD. Immunologic studies of arginase in tissues of normal human adults and arginase-deficient patients. *Pediatr Res* 1983;17:941-944.
- [43] Grody WW, Kern RM, Klein D, Dodson AE, Wissman PB, Barsky SH and Cederbaum SD. Arginase deficiency manifesting delayed clinical sequelae and induction of a kidney arginase isozyme. *Hum Genet* 1993;91:1-5.
- [44] Spector EB, Jenkinson CP, Grigor MR, Kern RM and Cederbaum SD. Subcellular location and differential antibody specificity of arginase in tissue culture and whole animals. *Int J Dev Neurosci* 1994;12:337-342.
- [45] Tabor CW and Tabor H. Polyamines. *Annu Rev Biochem* 1984;53:749-790.
- [46] Morgan DML. Polyamines. *Essays Biochem* 1987;46: 82-115.
- [47] Cai D, Deng K, Mellado W, Lee J, Ratan RR and Filbin MT. Arginase I and polyamines act downstream from cyclic AMP in overcoming inhibition of axonal growth MAG and myelin in

- vitro. *Neuron* 2002;35:711-719.
- [48] Nishioka K. Introduction to polyamines. In: Nishioka K (ed) *Polyamines in Cancer: Basic Mechanisms and Clinical Approaches*. New York Springer, 1996; pp 1-5.
- [49] Schipper RG, Penning LC and Verhofstad AA. Involvement of polyamines in apoptosis. Facts and controversies: effectors or protectors? *Cancer Biol* 2000;10:55-68.
- [50] Thomas T and Thomas TJ. Polyamines in cell growth and cell death: molecular mechanisms and therapeutic applications. *Cell Mol Life Sci* 2001;58:244-258.
- [51] Auvinen M, Jarvinen K, Hotti A, Okkeri J, Laitinen J, Janne OA, Coffino P, Bergman M, Andersson LC, Alitalo K and Holtta E. Transcriptional regulation of the ornithine decarboxylase gene by c-Myc/Max/Mad network and retinoblastoma protein interacting with c-Myc. *Int J Biochem Cell Biol* 2003;35:496-521.
- [52] de la Torre JC. Alzheimer disease as a vascular disorder: Nosological evidence. *Stroke* 2002;33:1152-1162.
- [53] Rhodin JA and Thomas T. A vascular connection to Alzheimer's disease. *Microcirculation* 2001, 8:207-220.
- [54] Breteler MM, Bots ML, Ott A and Hofman A. Risk factors for vascular disease and dementia. *Haemostasis* 1998;28:167-173.
- [55] Roher AE, Esh C, Kokjohn TA, Kalbak W, Luehrs DC, Seward JD, Sue LI and Beach TG. Circle of Willis atherosclerosis is a risk factor for sporadic Alzheimer's disease. *Arterioscler Thromb Vasc Biol* 2003;23:2055-2062.
- [56] Breteler MM. Vascular involvement in cognitive decline and dementia: epidemiologic evidence from the Rotterdam Study and the Rotterdam Scan Study. *Ann N Y Acad Sci* 2000;903:457-465.
- [57] Breteler MM. Vascular risk factors for Alzheimer's disease: an epidemiological study. *Neurobiol Aging* 2000;21:153-160.
- [58] Ott A, Stolk RP, Hofman A, van Harskamp F, Grobbee DE and Breteler MM. Association of diabetes mellitus and dementia: the Rotterdam Study. *Diabetologia* 1996;39:1392-1397.
- [59] Ott A, Slioter AJ, Hofman A, van Harskamp F and Witteman JC. Smoking and risk of dementia and Alzheimer's disease in a population-based cohort study: the Rotterdam Study. *Lancet* 1998;351:1840-1843.
- [60] Van Duijn CM, Havekes LM, van Broeckhoven C, de Knijff P and Hofman A. Apolipoprotein E genotype and association between smoking and early onset Alzheimer's disease. *Br Med J* 1995;310:627-631.
- [61] Graves AB, van Duijn CM, Chandra V, Fratiglioni L, Heyman A, Jorm AF, Kokmen E, Kondo K, Mortimer JA, Rocca WA, Shalat S, Soininen H and Hofman A. Alcohol and tobacco consumption as risk factors for Alzheimer's disease: a collaborative re-analysis of case-controlled studies. *Int J Epidemiol* 1991;20:S48-S57.
- [62] Roher AE, Esh C, Kokjohn TA, Kalbak W, Luehrs DC, Seward JD, Sue LI and Beach TG. Circle of willis atherosclerosis is a risk factor for sporadic Alzheimer's disease. *Thromb Vasc Biol* 2003;23:2055-2062.
- [63] Beach TG, Wilson JR, Sue LI, Newell A, Poston M, Cisneros R, Pandya Y, Esh C, Connor DJ, Sabbagh M, Walker DG and Roher AE. Circle of Willis atherosclerosis: association with Alzheimer's disease, neuritic plaques and neurofibrillary tangles. *Acta Neuropathol (Berl)* 2007;113:13-21.
- [64] Hirao K, Ohnishi T, Hirata Y, Yamashita F, Mori T, Moriguchi Y, Matsuda H, Nemoto K, Imabayashi E, Yamada M, Iwamoto T, Arima K and Asada T. The prediction of rapid conversion to Alzheimer's disease in mild cognitive impairment using regional cerebral blood flow SPECT. *Neuroimage* 2005;28:1014-1021.
- [65] Johnson KA, Jones K, Holman BL, Becker J, Spiers PA, Satlin A and Albert MS. Preclinical prediction of Alzheimer's disease using SPECT. *Neurology* 1998;50:1563-1571.
- [66] Johnson KA and Albert MS. Perfusion abnormalities in prodromal Alzheimer's disease. *Neurobiol Aging* 2000;21:289-292.
- [67] Matsuda H, Mizumura S, Nagao T, Ota T, Iizuka T, Nemoto K, Kimura M, Tateno A, Ishiwata A, Kuji I, Arai H and Homma A. An easy Z-score imaging system for discrimination between very early Alzheimer's disease and controls using brain perfusion SPECT in a multicentre study. *Nucl Med Commun* 2007;28:199-205.
- [68] Sparks DL, Sabbagh M, Connor D, Soares H, Lopez J, Stankovic G, Johnson-Traver S, Ziolkowski C and Browne P. Statin therapy in Alzheimer's disease. *Acta Neurol Scand Suppl* 2006;185:78-86.
- [69] Zamrini E, McGwin G and Roseman JM. Association between statin use and Alzheimer's disease. *Neuroepidemiology* 2004;23:94-98.
- [70] Miida T, Takahashi A, Tanabe N and Ikeuchi T. Can statin therapy really reduce the risk of Alzheimer's disease and slow its progression? *Curr Opin Lipidol* 2005;16:619-623.
- [71] Zigman WB, Schupf N, Jenkins EC, Urv TK, Tycko B and Silverman W. Cholesterol level, statin use and Alzheimer's disease in adults with Down syndrome. *Neurosci Lett* 2007;18; 416:279-284.
- [72] Petanceska SS, DeRosa S, Olm V, Diaz N, Sharma A, Thomas-Bryant T, Duff K, Pappolla M and Refolo LM. Statin therapy for Alzheimer's disease: will it work? *J Mol Neurosci* 2002;19:155-161.
- [73] Maxwell AJ and Cooke JP. Cardiovascular

- effects of L-arginine. *Curr Opin Nephrol Hypertens* 1998;7:63-70.
- [74] Ignarro LJ, Cirino G, Casini A and Napoli C. Nitric oxide as a signaling molecule in the vascular system: an overview. *J Cardiovasc Pharmacol* 1999;34:879-886.
- [75] Walter R, Mark M and Reinhart WH. Pharmacological concentrations of arginine influence human whole blood viscosity independent of nitric oxide synthase activity in vitro. *Biochem Biophys Res Commun* 2000; 269:687-691.
- [76] Böger RH, Bode-Böger SM, Szuba A, Tsao PS, Chan JR, Tangphao O, Blaschke TF and Cooke JP. Asymmetric dimethylarginine (ADMA): a novel risk factor for endothelial dysfunction: its role in hypercholesterolemia. *Circulation* 1998;98:1842-1847.
- [77] Creager MA, Gallagher SJ, Girerd XJ, Coleman SM, Dzau VJ and Cooke JP. L-Arginine improves endothelium dependent vasodilation in hypercholesterolemic humans. *J Clin Invest* 1992;90:1248-1253.
- [78] Cooke JP and Creager A. Endothelial dysfunction in hypercholesterolemia is corrected by L-arginine. *Basic Res Cardiol* 1991;86(Suppl 2):173-181.
- [79] Thorne S, Mullen MJ, Clarkson P and Donald AE. Early endothelial dysfunction in adults at risk from atherosclerosis: different responses to L-arginine. *J Am Coll Cardiol* 1998;32:110-116.
- [80] Stroes ESG, Koomans HA, DeBruin TWA and Rabelink TJ. Vascular function in the forearm of hypercholesterolemic patients off and on lipid-lowering medication. *Lancet* 1995;346: 467-471.
- [81] Clarkson P, Adams MR, Powe AJ, Donald AE, McCredie R, Robinson J, McCarthy SN, Keech A, Celermajer DS and Deanfield JE. Oral L-arginine improves endothelium-dependent dilation in hypercholesterolaemic young adults. *J Clin Invest* 1996;15:97:1989-1994.
- [82] Maxwell AJ, Anderson B, Zapien MP and Cooke JP. Endothelial dysfunction in hypercholesterolemia is reversed by nutritional duraproduct designed to enhance nitric oxide activity. *Cardiovasc Drugs Ther* 2000;14:309-316.
- [83] Tsao PS, McEvoy LM, Drexler H, Butcher EC and Cooke JP. Enhanced endothelial adhesiveness in hypercholesterolemia is attenuated by L-arginine. *Circulation* 1994;89: 2176-2182.
- [84] Drexler H and Hornig B. Endothelial dysfunction in human disease. *J Mol Cell Cardiol* 1999;31:51-60.
- [85] Goumas G, Tentolouris C, Tousoulis D, Stefanadis C and Toutouzas P. Therapeutic modification of the L-arginine-eNOS pathway in cardiovascular diseases. *Atherosclerosis* 2001;154:255-267.
- [86] Wolf A, Zalpour C, Theilmeyer G, Wang BY, Ma A, Anderson B, Tsao PS and Cooke JP. Dietary L-arginine supplementation normalizes platelet aggregation in hypercholesterolemic humans. *J Am Coll Cardiol* 1997;29:479-285.
- [87] Bode-Böger SM, Böger RH, Creutzig A, Tsikas D, Gutzki FM, Alexander K and Frölich JC. L-arginine infusion decreases peripheral arterial resistance and inhibits platelet aggregation in healthy subjects. *Clin Sci (Lond)* 1994;87:303-310.
- [88] Theilmeyer G, Chan JR, Zalpour C, Anderson B, Wang BY, Wolf A, Tsao PS and Cooke JP. Adhesiveness of mononuclear cells in hypercholesterolemic humans is normalized by dietary L-arginine. *Arterioscler Thromb Vasc Biol* 1997;17:3557-3564.
- [89] Kawano H, Motoyama T, Hirai N, Kugiyama K, Yasue H and Ogawa H. Endothelial dysfunction in hypercholesterolemia is improved by L-arginine administration: possible role of oxidative stress. *Atherosclerosis* 2002;161:375-380.
- [90] Dhawan V, Handu SS, Nain CK and Ganguly NK. Chronic L-arginine supplementation improves endothelial cell vasoactive functions in hypercholesterolemic and atherosclerotic monkeys. *Mol Cell Biochem* 2005;269:1-11.
- [91] Jeremy RW, McCarron H and Sullivan D. Effects of dietary L-arginine on atherosclerosis and endothelium-dependent vasodilation in the hypercholesterolemic rabbit. Response according to treatment duration, anatomic site and sex. *Circulation* 1996;94:498-506.
- [92] Verreault R, Kaltenbach G and Berthel M. Hypertension and Alzheimer's disease. *Presse Med* 2005;34:809-812.
- [93] Skoog I and Gustafson D. Update on hypertension and Alzheimer's disease. *Neurol Res* 2006;28:605-611.
- [94] Siani A, Pagano E, Iacone R, Iacoviello L, Scopacasa F and Strazzullo P. Blood pressure and metabolic changes during dietary L-arginine supplementation in humans. *Am J Hypertens* 2000;13:547-551.
- [95] Rector TS, Bank AJ, Mullen KA, Tschumperlin LK, Sih R, Pillai K and Kubo SH. Randomized, double-blind, placebo-controlled study of supplemental oral L-arginine in patients with heart failure. *Circulation* 1996;93:2135-2141.
- [96] Galanis DJ, Petrovitch H, Launer LJ, Harris TB, Foley DJ and White LR. Smoking history in middle age and subsequent cognitive performance in elderly Japanese-American men. *Am J Epidemiol* 1997;145:507-515.
- [97] Aggarwal NT, Bienias JL, Bennett DA, Wilson RS, Morris MC, Schneider JA, Shah RC and Evans DA. The relation of cigarette smoking to incident Alzheimer's disease in a biracial

- urban community population. *Neuroepidemiology* 2006; 26:140-146.
- [98] Ford AB, Mefrouche Z, Friedland RP and Debanne SM. Smoking and cognitive impairment: a population-based study. *J Am Geriatr Soc* 1996;44:905-909.
- [99] Launer LJ, Feskens EJ, Kalmijn S and Kromhout D. Smoking, drinking, and thinking. The Zutphen Elderly Study. *Am J Epidemiol* 1996;143:219-227.
- [100] Dickerson TJ and Janda KD. Glycation of the amyloid beta-protein by a nicotine metabolite: a fortuitous chemical dynamic between smoking and Alzheimer's disease. *Proc Natl Acad Sci USA* 2003;100:8182-8187.
- [101] Sanderson KJ, van Rij AM, Wade CR and Sutherland WHF. Lipid peroxidation of circulating low density lipoproteins with age, smoking and in peripheral vascular disease. *Atherosclerosis* 1995;118:45-51.
- [102] Weber C, Erl W, Weber K and Weber PC. Increased adhesiveness of isolated monocytes to endothelium is prevented by vitamin C intake in smokers. *Circulation* 1996; 93:1488-1492.
- [103] Adams MR, Jessup W and Celermajer DS. Cigarette smoking is associated with increased human monocyte adhesion to endothelial cells: reversibility with oral L-arginine but not vitamin C. *J Am Coll Cardiol* 1997;29:491-497.
- [104] Panza JA, Casino PR, Badar DM and Quyyumi AA. Effect of increased availability of endothelium-derived nitric oxide precursor on endothelium-dependent vascular relaxation in normal subjects and in patients with essential hypertension. *Circulation* 1993;87: 1475-1481.
- [105] Taddei S, Mattei P, Virdis A, Sudano I, Ghiadoni L and Salvetti A. Effect of potassium on vasodilation to acetylcholine in essential hypertension. *Hypertension* 1994;23:485-490.
- [106] Pritchard KA, Groszek L, Smalley DM, Sessa WC and Wu M. Native low density lipoprotein increases endothelial cell nitric oxide synthase generation of superoxide anion. *Circ Res* 1995;77:510-518.
- [107] Palmer RMJ, Ferrige AG and Moncada S. Nitric oxide accounts for the biological activity of endothelium-derived relaxing factor. *Nature* 1987;327:524-526.
- [108] Radomski MW, Palmer RMJ and Moncada S. An L-arginine/nitric oxide pathway present in human platelets regulates aggregation. *Proc Natl Acad Sci USA* 1990;87:5193-5197.
- [109] Kubes P, Suzuki M and Granger DN. Nitric oxide: an endogenous modulator of leukocyte adhesion. *Proc Natl Acad Sci USA* 1991;88: 4651-4655.
- [110] Garg UC and Hassid A. Nitric oxide generating vasodilators and 8-bromocyclic guanosine monophosphate inhibit mitogenesis and proliferation of cultured rat vascular smooth muscle cells. *J Clin Invest* 1989;83:1774-1777.
- [111] Zeiher AM, Fisslthaler B, Schray-Utz B and Busse R. Nitric oxide modulates the expression of monocyte chemoattractant protein-1 in cultured human endothelial cells. *Circ Res* 1995;76:980-986.
- [112] Hobbs AJ, Higgs A and Moncada S. Inhibition of nitric oxide synthase as a potential therapeutic target. *Annu Rev Pharmacol Toxicol* 1999;39:191-220.
- [113] Boger RH and Bode-Boger SM. The clinical pharmacology of L-arginine. *Annu Rev Pharmacol Toxicol* 2001;41:79-99.
- [114] Higashi Y, Oshima T, Ono N, Hiraga H, Yoshimura M, Watanabe M, Matsuura H, Kambe M and Kajiyama G. Intravenous administration of L-arginine inhibits angiotensin-converting enzyme in humans. *J Clin Endocrinol Metab* 1995;80:2198-2202.
- [115] Piatti P, Fragasso G, Monti LD, Setola E, Lucotti P, Fermo I, Paroni R, Galluccio E, Pozza G, Chierchia S and Margonato A. Acute intravenous L-arginine infusion decreases endothelin-1 levels and improves endothelial function in patients with angina pectoris and normal coronary arteriograms: correlation with asymmetric dimethylarginine levels. *Circulation* 2003;107:429-436.
- [116] Halliwell B. Protection against tissue damage in vivo by desferrioxamine: what is its mechanism of action? *Free Radic Biol Med* 1989;7:645-651.
- [117] Smith MA, Perry G, Richey PL, Sayre LM, Anderson VE, Beal MF and Kowall N. Oxidative damage in Alzheimer's. *Nature* 1996;382:120-121.
- [119] Sayre LM, Zelasko DA, Harris PL, Perry G, Salomon RG and Smith MA. 4-Hydroxynonenal-derived advanced lipid peroxidation end products are increased in Alzheimer's disease. *J Neurochem* 1997;68: 2092-2097.
- [120] Smith MA. Alzheimer disease. *Int Rev Neurobiol* 1998;42:1-54.
- [121] Nunomura A, Perry G, Pappolla MA, Friedland RP, Hirai K, Chiba S and Smith MA. Neuronal oxidative stress precedes amyloid-beta deposition in Down syndrome. *J Neuropathol Exp Neurol* 2000;59:1011-1017.
- [122] Nunomura A, Perry G, Aliev G, Hirai K, Takeda A, Balraj EK, Jones PK, Ghanbari H, Wataya T, Shimohama S, Chiba S, Atwood CS, Petersen RB and Smith MA. Oxidative damage is the earliest event in Alzheimer disease. *J Neuropathol Exp Neurol* 2001;60:759-767.
- [123] Zhu X, Raina AK, Lee HG, Casadesus G, Smith MA and Perry G. Oxidative stress signaling in Alzheimer's disease. *Brain Res* 2004;1000: 32-39.
- [124] Perry G, Taddeo MA, Nunomura A, Zhu X, Zenteno-Savin T, Drew KL, Shimohama S,

- Avila J, Castellani RJ and Smith MA. Comparative biology and pathology of oxidative stress in Alzheimer and other neurodegenerative diseases: beyond damage and response. *Comp Biochem Physiol C Toxicol Pharmacol* 2002;133:507-513.
- [125] Reddy PH. Amyloid precursor protein-mediated free radicals and oxidative damage: implications for the development and progression of Alzheimer's disease. *J Neurochem* 2006;96:1-13.
- [126] Malinski T. Nitric oxide and nitrosative stress in Alzheimer's disease. *J Alzheimers Dis* 2007;11:207-218.
- [127] Guix FX, Uribesalgo I, Coma M and Munoz FJ. The physiology and pathophysiology of nitric oxide in the brain. *Prog Neurobiol* 2005;76:126-152.
- [128] Pacher P, Beckman JS and Liaudet L. Nitric oxide and peroxynitrite in health and disease. *Physiol Rev* 2007;87:315-424.
- [129] Ridnour LA, Thomas DD, Mancardi D, Espey MG, Miranda KM, Paolocci N, Feelisch M, Fukuto J and Wink DA. The chemistry of nitroxidative stress induced by nitric oxide and reactive nitrogen oxide species. Putting perspective on stressful biological situations. *Biol Chem* 2004;385:1-10.
- [130] Hirst DG and Robson T. Nitrosative stress in cancer therapy. *Front Biosci* 2007;12:3406-3418.
- [131] Wang JY, Shum AY, Ho YJ and Wang JY. Oxidative neurotoxicity in rat cerebral cortex neurons: synergistic effects of H₂O₂ and NO on apoptosis involving activation of p38 mitogen-activated protein kinase and caspase-3. *J Neurosci Res* 2003;72:508-519.
- [132] Castegna A, Thongboonkerd V, Klein JB, Lynn B, Markesbery WR and Butterfield DA. Proteomic identification of nitrated proteins in Alzheimer's disease brain. *J Neurochem* 2003;85:1394-1401.
- [133] Sultana R, Poon HF, Cai J, Pierce WM, Merchant M, Klein JB, Markesbery WR and Butterfield DA. Identification of nitrated proteins in Alzheimer's disease brain using a redox proteomics approach. *Neurobiol Dis* 2006;22:76-87.
- [134] Wink DA, Hanbauer I, Krishna MC, DeGrafej W, Gamson J and Mitchell JB. Nitric oxide protects against cellular damage and cytotoxicity from reactive oxygen species. *Proc Natl Acad Sci USA* 1993;90:9813-9817.
- [135] Wink DA, Hanbauer I, Laval F, Cook JA, Krishna MC and Mitchell JB. Nitric oxide protects against the cytotoxic effects of reactive oxygen species. *Ann N Y Acad Sci* 1994;738:265-278.
- [136] Dawson VL, Dawson TM, London ED, Bredt DS and Snyder SH. Nitric oxide mediates glutamate neurotoxicity in primary cortical cultures. *Proc Natl Acad Sci USA* 1991;88:6368-6371.
- [137] Glebov AN and ZinchukVV. Prooxidant-antioxidant state of the organism during oxidative stress and correction of the L-arginine-NO system. *Bull Exp Biol Med* 2006;141:368-370.
- [138] Dawson VL and Dawson TM. Nitric oxide neurotoxicity. *J Chem Neuroanat* 1996;10:179-190
- [139] Iadecola C. Bright and dark sides of nitric oxide in ischemic brain injury. *Trends Neurosci* 1997;20:132-139.
- [140] Tang XQ, Yu HM, Zhi JL, Cui Y, Tang EH, Feng JQ and Chen PX. Inducible nitric oxide synthase and cyclooxygenase-2 mediate protection of hydrogen peroxide preconditioning against apoptosis induced by oxidative stress in PC12 cells. *Life Sci* 2006;24:79:870-876.
- [141] Pomara N, Singh R, Deptula D, Chou JC-Y, Schwartz MB and LeWitt P. Glutamate and other CSF amino acids in Alzheimer's disease. *Am J Psychiatry* 1992;149:251-254.
- [142] McFarland R, Blokhin A, Sydnor J, Mariani J and Vogel MW. Oxidative stress, nitric oxide, and the mechanisms of cell death in Lurcher Purkinje cells. *Dev Neurobiol* 2007;67:1032-1046.
- [143] Aliev G, Smith MA, Seyidov D, Neal ML, Lamb BT, Nunomura A, Gasimov EK, Vinters HV, Perry G, LaManna JC and Friedland RP. The role of oxidative stress in the pathophysiology of cerebrovascular lesions in Alzheimer's disease. *Brain Pathol* 2002;12:21-35.
- [144] Maier CM and Chan PH. Role of superoxide dismutases in oxidative damage and neurodegenerative disorders. *Neuroscientist* 2002;8:323-334.
- [145] Miliutina NP, Ananian AA and Shugalei VS. Antiradical and antioxidant effect of arginine and its action on lipid peroxidation in hypoxia. *Biull Eksp Biol Med* 1990;110:433-435.
- [146] Maksimovich NE and Maslakov DA. The amino acid L-arginine and the potential for its use in clinical practice. *Zdravookhranenie* 2003;5:35-37.
- [147] Sedlak J and Lindsay RH. Estimation of total, protein-bound, and nonprotein sulfhydryl groups in tissue with Ellman's reagent. *Anal Biochem* 1968;25:192-205.
- [148] Wink DA, Miranda KM and Espey MG. Mechanisms of the antioxidant effects of nitric oxide. *Antioxid Redox Signal* 2000;1:203-213.
- [149] Maksimovich NE, Zinchuk VV and Maslakov DA. The degree of oxidative stress in the rat brain during ischemia and reperfusion in conditions of correction of the L-arginine-NO system. *Neurosci Behav Physiol* 2006;36:373-378.
- [150] Calabrese V, Bates TE and Stella AM. NO synthase and NO-dependent signal pathways in brain aging and neurodegenerative disorders: the role of oxidant/antioxidant

- balance. *Neurochem Res* 2000;25:1315-1341.
- [151] Bredt DS, Hwang PM and Snyder SH. Localization of nitric oxide synthase indicating a neural role for nitric oxide. *Nature* 1990; 347:768-770.
- [152] Robak J and Gryglewski RJ. Nitric oxide donors as scavengers of superoxide anions. *Pol J Pharmacol* 1993;45:51-58.
- [153] Zinchuk VV. Involvement of nitric oxide in forming the oxygen binding properties of hemoglobin. *Usp Fiziol Nauk* 2003;34:33-45.
- [154] Popov D, Costache G, Georgescu A and Enache M. Beneficial effects of L-arginine supplementation in experimental hyperlipemia-hyperglycemia in the hamster. *Cell Tissue Res* 2002;308:109-120.
- [155] Kawano H, Motoyama T, Hirai N, Kugiyama K, Yasue H and Ogawa H. Endothelial dysfunction in hypercholesterolemia is improved by L-arginine administration: possible role of oxidative stress. *Atherosclerosis* 2002;161:375-380.
- [156] Steer P, Millgård J, Basu S, Lithell H, Vessby B, Berne C and Lind L. Vitamin C, diclofenac, and L-arginine protect endothelium-dependent vasodilation against elevated circulating fatty acid levels in humans. *Atherosclerosis* 2003;168:65-72.
- [157] Lin CC, Tsai WC, Chen JY, Li YH, Lin LJ and Chen JH. Supplements of L-arginine attenuate the effects of high-fat meal on endothelial function and oxidative stress. *Int J Cardiol* 2008;127:331-340.
- [158] Tsai WC, Li YH, Lin CC, Chao TH and Chen JH. Effects of oxidative stress on endothelial function after a high-fat meal. *Clin Sci* 2004; 106:315-319.
- [159] Rosenberg PB. Clinical aspects of inflammation in Alzheimer's disease. *Int Rev Psychiatry* 2005;17:503-514.
- [160] McGeer PL and McGeer EG. Local neuroinflammation and the progression of Alzheimer's disease. *J Neurovirol* 2002;8: 529-538.
- [161] Walsh DM, Klyubin I, Fadeeva JV, Rowan MJ and Selkoe DJ. Amyloid-beta oligomers: their production, toxicity and therapeutic inhibition. *Biochem Soc* 2002;30:552-557.
- [162] Walsh DM and Selkoe DJ. Deciphering the molecular basis of memory failure in Alzheimer's disease. *Neuron* 2004;44:181-193.
- [163] Tan J, Town T, Paris D, Mori T, Suo Z, Crawford F, Mattson MP, Flavell RA and Mullan M. Microglial activation resulting from CD40-CD40L interaction after beta-amyloid stimulation. *Science* 1999;286:2352-2355.
- [164] Akiyama H, Barger S, Barnum S, Bradt B, Bauer J, Cole GM, Cooper NR, Eikelenboom P, Emmerling M, Fiebich BL, Finch CE, Frautschy S, Griffin WS, Hampel H, Hull M, Landreth G, Lue L, Mrazek R, Mackenzie IR, McGeer PL, O'Banion MK, Pachter J, Pasinetti G, Plata-Salaman C, Rogers J, Rydel R, Shen Y, Streit W, Strohmeyer R, Tooyoma I, Van Muiswinkel FL, Veerhuis R, Walker D, Webster S, Wegrzyniak B, Wenk G and Wyss-Coray T. Inflammation and Alzheimer's disease. *Neurobiol Aging* 2000;21:383-421.
- [165] Ferencik M, Novak M, Rovensky J and Rybar I. Alzheimer's disease, inflammation and non-steroidal anti-inflammatory drugs. *Bratisl Lek Listy* 2001;102:123-132.
- [166] Pasinetti GM. From epidemiology to therapeutic trials with anti-inflammatory drugs in Alzheimer's disease: the role of NSAIDs and cyclooxygenase in beta-amyloidosis and clinical dementia. *J Alzheimers Dis* 2002;4:435-445.
- [167] Hoozemans JJ, Veerhuis R, Rozemuller AJ and Eikelenboom P. Non-steroidal anti-inflammatory drugs and cyclooxygenase in Alzheimer's disease. *Curr Drug Targets* 2003; 4:461-468.
- [168] Barbul A, Wasserkrug HL, Seifter E, Rettura G, Levenson SM and Efron G. Thymic stimulatory actions of arginine. *J Surg Res* 1980;29:228.
- [169] Nijima A and Meguid MM. Influence of systemic arginine-lysine on immune organ function: an electrophysiological study. *Brain Res Bull* 1998;45:437-441.
- [170] Kirk SJ, Hurson M, Regan MC, Holt DR, Wasserkrug HL and Barbul A. Arginine stimulates wound healing and immune function in elderly human beings. *Surgery* 1993;114:155-159.
- [171] Albina JE, Mills CD, Barbul A, Thirkill CE, Henry WL, Mastrofrancesco B and Caldwell MD. Arginine metabolism in wounds. *Am J Physiol* 1988;254:459-467.
- [172] Albina JE, Caldwell MD, Henry WL and Mills CD. Regulation of macrophage functions by L-arginine. *J Exp Med* 1989;169:1021-1029.
- [173] Scott GS and Bolton C. L-arginine modifies free radical production and the development of experimental allergic encephalomyelitis. *Inflamm Res* 2000;49:720-726.
- [174] Dawson VL, Brahmabhatt HP, Mong JA and Dawson TM. Expression of inducible nitric oxide synthase causes delayed neurotoxicity in primary mixed neuronal-glial cortical cultures. *Neuropharmacology* 1994;33:1425-1430.
- [175] Chao CC, Hu S, Sheng WS, Bu D, Bukrinsky MI and Peterson PK. Cytokine-stimulated astrocytes damage human neurons via a nitric oxide mechanism. *Glia* 1996;16:276-284.
- [176] Vodovotz Y, Lucia MS, Flanders KC, Chesler L, Xie QW, Smith TW, Weidner J, Mumford R, Webber R, Nathan C, Roberts AB, Lippa CF and Sporn MB. Inducible nitric oxide synthase in tangle-bearing neurons of patients with Alzheimer's disease. *J Exp Med* 1996;184: 1425-1433.

- [177] Smith MA, Richey Harris PL, Sayre LM, Beckman JS and Perry G. Widespread peroxynitrite-mediated damage in Alzheimer's disease. *J Neurosci* 1997;17:2653-2657.
- [178] Lee SC, Zhao ML, Hirano A and Dickson DW. Inducible nitric oxide synthase immunoreactivity in the Alzheimer disease hippocampus: association with Hirano bodies, neurofibrillary tangles, and senile plaques. *J Neuropathol Exp Neurol* 1999;58:1163-1169.
- [179] Bauer MK, Lieb K, Schulze-Osthoff K, Berger M, Gebicke Haerter PJ, Bauer J and Fiebich BL. Expression and regulation of cyclooxygenase-2 in rat microglia. *Eur J Biochem* 1997;243:726-731.
- [180] del Zoppo G, Ginis I, Hallenbeck JM, Iadecola C, Wang X and Feuerstein GZ. Inflammation and stroke: putative role for cytokines, adhesion molecules and iNOS in brain response to ischemia. *Brain Pathol* 2000;10: 95-112.
- [181] Garthwaite J, Charles SL and Chess-Williams R. Endothelium-derived relaxing factor release on activation of NMDA receptors suggests role as intercellular messenger in the brain. *Nature* 1988;336:385-388.
- [182] Dawson TM, Dawson VL and Snyder SH. A novel neuronal messenger molecule in brain: the free radical, nitric oxide. *Ann Neurol* 1992; 32:297-311.
- [183] Bon CL and Garthwaite J. On the role of nitric oxide in hippocampal long-term potentiation. *J Neurosci* 2003;23:1941-1948.
- [184] Sanders KM and Ward SM. Nitric oxide as a mediator of nonadrenergic noncholinergic neurotransmission. *Am J Physiol* 1992;262: 379-392.
- [185] Garthwaite J and Boulton CL. Nitric oxide signaling in the central nervous system. *Annu Rev Physiol* 1995;57:683-706.
- [186] Thomas S and Robitaille R. Differential frequency-dependent regulation of transmitter release by endogenous nitric oxide at the amphibian neuromuscular synapse. *J Neurosci* 2001;21:1087-1095.
- [187] Nickels TJ, Reed GW, Drummond JT, Blevins DE, Lutz MC and Wilson DF. Does nitric oxide modulate transmitter release at the mammalian neuromuscular junction? *Clin Exp Pharmacol Physiol* 2007;34:318-326.
- [188] Hevel JM and Marletta MA. Macrophage nitric oxide synthase: relationship between enzyme-bound tetrahydrobiopterin and synthase activity. *Biochemistry* 1992;31: 7160-7165.
- [189] Barford PA, Blair JA, Eggar C, Hamon D, Morar C and Whitburn SB. Tetrahydrobiopterin metabolism in the temporal lobe of patients dying with senile dementia of Alzheimer type. *J Neurol Neurosurg Psychiatry* 1984;47:736-738.
- [190] Heizel B, John M, Klatt P, Böhme E and Mayer B. Ca²⁺/calmodulin-dependent formation of hydrogen peroxide by brain nitric oxide synthase. *Biochem J* 1992;281:627-630.
- [191] Lowe SL, Bowen CM, Francis PT and Neary D. Ante mortem cerebral amino acid concentrations indicate selective degeneration of glutamate-enriched neurons in Alzheimer's disease. *Neuroscience* 1990; 38:571-577.
- [192] Kuiper MA, Teerlink T, Visser JJ, Bergmans PLM, Scheltens P and Wolters Ch. L-Glutamate, L-arginine and L-citrulline levels in cerebrospinal fluid of Parkinson's disease, multiple system atrophy, and Alzheimer's disease patients. *J Neural Transmission* 2000;107:183-189.
- [193] McEntee WJ and Crook TH. Glutamate: its role in learning, memory, and the aging brain. *Psychopharmacology* 1993;111:391-401.
- [194] Kuiper MA, Visser JJ, Bergmans PLM, Scheltens Ph and Wolters ECh. Decreased cerebrospinal fluid nitrate levels in Parkinson's disease, Alzheimer's disease and Multiple System Atrophy patients. *J Neurol Sci* 1994;121:46-49.
- [195] McCarty MF. Vascular nitric oxide may lessen Alzheimer's risk. *Med Hypotheses* 1998;51: 465-476.
- [196] Venturini G, Colasanti M, Persichini T, Fioravanti E, Ascensi P, Palomba L, Cantoni O and Musci G. Beta-amyloid inhibits NOS activity by subtracting NADPH availability. *FASEB J* 2002;16:1970-1972.
- [197] Wirtz-Brugger F, Giovanni A Guanosine 3,5-cyclic monophosphate mediated inhibition of cell death induced by nerve growth factor withdrawal and beta-amyloid: protective effects of propentofylline. *Neuroscience* 2000;99:737-750.
- [198] Troy CM, Rabacchi SA, Friedman WJ, Frappier TF, Brown K and Shelanski ML. Caspase-2 mediates neuronal cell death induced by beta-amyloid. *J Neurosci* 2000;20:1386-1392.
- [199] Tran MH, Yamada K, Olariu A, Mizuno M, Ren XH and Nabeshima T. Amyloid β -peptide induces nitric oxide production in rat hippocampus: association with cholinergic dysfunction and amelioration by inducible nitric oxide synthase inhibitors. *FASEB J* 2001;15:1407-1409.
- [200] McCann SM. The nitric oxide hypothesis of brain aging. *Exp Gerontol* 1997;32:431-440.
- [201] Wang Q, Rowan MJ and Anwyl R. Beta-amyloid-mediated inhibition of NMDA receptor-dependent long-term potentiation induction involves activation of microglia and stimulation of inducible nitric oxide synthase and superoxide. *J Neurosci* 2004;24:6049-6056.
- [202] Monsonego A, Imitola J, Zota V, Oida T and Weiner HL. Microglia-mediated nitric oxide cytotoxicity of T cells following amyloid β -peptide presentation to Th1 cells. *J*

- Immunol* 2003;171:2216-2224.
- [203] Qin L, Liu Y, Cooper C, Liu B, Wilson B and Hong JS. Microglia enhance beta-amyloid peptide-induced toxicity in cortical and mesencephalic neurons by producing reactive oxygen species. *J Neurochem* 2002;83:973-983.
- [204] Law A, O'Donnel J, Gauthier S and Quirion R. Neuronal and inducible nitric oxide synthase expressions and activities in the hippocampal and cortices of young adult, aged cognitively unimpaired, and impaired Long-Evans rats. *Neuroscience* 2002;112:267-275.
- [205] Puzzo D, Vitolo O, Trinchese F, Jacob JP, Palmeri A and Arancio O. Amyloid-beta peptide inhibits activation of the nitric oxide/cGMP/cAMP-responsive element-binding protein pathway during hippocampal synaptic plasticity. *J Neurosci* 2005;25:6887-6897.
- [206] Watson GS and Craft S. Modulation of memory by insulin and glucose: neuropsychological observations in Alzheimer's disease. *Eur J Pharmacol* 2004;490:97-113.
- [207] Meneilly GS, Cheung E, Tessier D, Yakura C and Tuokko H. The effect of improved glycemic control on cognitive functions in the elderly patient with diabetes. *J Gerontol* 1993;48:117-121.
- [208] Razay G and Wilcock GK. Hyperinsulinaemia and Alzheimer's disease. *Age Ageing* 1994;23:396-399.
- [209] Small GW, Ercoli LM, Silverman DH, Huang SC, Komo S, Bookheimer SY, Lavretsky H, Miller K, Siddarth P, Rasgon NL, Mazziotta JC, Saxena S, Wu HM, Mega MS, Cummings JL, Saunders AM, Pericak-Vance MA, Roses AD, Barrio JR and Phelps ME. Cerebral metabolic and cognitive decline in persons at genetic risk for Alzheimer's disease. *Proc Natl Acad Sci USA* 2000;97:6037-6042.
- [210] Garrido GE, Furuie SS, Buchpiguel CA, Bottino CM, Almeida OP, Cid CG, Camargo CH, Castro CC, Glabus MF and Busatto GF. Relation between medial temporal atrophy and functional brain activity during memory processing in Alzheimer's disease: a combined MRI and SPECT study. *J Neurol Neurosurg Psychiatry* 2002;73:508-516.
- [211] Squire LR. Memory and the hippocampus: a synthesis from findings with rats, monkeys, and humans. *Psychol Rev* 1992;99:195-231.
- [212] Squire LR and Zola SM. Structure and function of declarative and nondeclarative memory systems. *Proc Natl Acad Sci USA* 1996;93:13515-13522.
- [213] Manns JR, Hopkins RO, Reed JM, Kitchener EG and Squire LR. Recognition memory and the human hippocampus. *Neuron* 2003;37:171-180.
- [214] Sakamoto S, Ishii K, Hosaka K, Mori T, Sasaki M and Mori E. Detectability of hypometabolic regions in mild Alzheimer disease: function of time after the injection of 2-[fluorine 18]-fluoro-2-deoxy-D-glucose. *Am J Neuroradiol* 2005;26:843-847.
- [215] Anchisi D, Borroni B, Franceschi M, Kerrouche N, Kalbe E, Beuthien-Beumann B, Cappa S, Lenz O, Ludecke S, Marcone A, Mielke R, Ortelli P, Padovani A, Pelati O, Pupi A, Scarpini E, Weisenbach S, Herholz K, Salmon E, Holthoff V, Sorbi S, Fazio F and Perani D. Heterogeneity of brain glucose metabolism in mild cognitive impairment and clinical progression to Alzheimer disease. *Arch Neurol* 2005;62:1728-1733.
- [216] Gong CX, Liu F, Grundke-Iqbal I and Iqbal K. Impaired brain glucose metabolism leads to Alzheimer neurofibrillary degeneration through a decrease in tau O-GlcNAcylation. *J Alzheimers Dis* 2006;9:1-12.
- [217] Mosconi L, Tsui WH, Rusinek H, De Santi S, Li Y, Wang GJ, Pupi A, Fowler J and de Leon MJ. Quantitation, regional vulnerability, and kinetic modeling of brain glucose metabolism in mild Alzheimer's disease. *Eur J Nucl Med Mol Imaging* 2007;34:1467-1479.
- [218] Craft S, Zallen G and Baker D. Glucose and memory in mild senile dementia of the Alzheimer type. *J Clin Exp Neuropsychol* 1992;14:253-267.
- [219] Benton D, Owens DS and Parker PY. Blood glucose influences memory and attention in young adults. *Neuropsychologia* 1994;32:595-607.
- [220] Allen JB, Gross AM, Aloia MS and Billingsley C. The effects of glucose on nonmemory cognitive functioning in the elderly. *Neuropsychologia* 1996;34:459-465.
- [221] Lidder JK, Sunram PG and Foster SI. Glucose and memory: fractionation of enhancement effects? *Psychopharmacology (Berl)* 1998;137:259-270.
- [222] Sunram-Lea SI, Foster JK, Durlach P and Perez C. The effect of retrograde and anterograde glucose administration on memory performance in healthy young adults. *Behav Brain Res* 2002;134:505-516.
- [223] Craft S, Dagogo-Jack SE, Wiethop BV, Murphy C, Nevins RT and Fleischman S. Effects of hyperglycemia on memory and hormone levels in dementia of the Alzheimer type: a longitudinal study. *Neurosci* 1993;107:926-940.
- [224] Manning CA, Honn VJ, Stone WS, Jane JS and Gold PE. Glucose effects on cognition in adults with Down's syndrome. *Neuropsychology* 1998;12:479-484.
- [225] Stone CA, Korol WS, Gold DL and Manning PE. Glucose enhancement of 24-h memory retrieval in healthy elderly humans. *Behav Brain Res* 1998;93:71-76.
- [226] Ragozzino ME, Unick KE and Gold PE. Hippocampal acetylcholine release during memory testing in rats: augmentation by

- glucose. *Proc Natl Acad Sci USA* 1996;93:4693-4698.
- [227] Ragozzino ME, Arankowsky-Sandoval G. and Gold PE. Glucose attenuates the effect of combined muscarinic-nicotinic receptor blockade on spontaneous alternation. *Eur J Pharmacol* 1994;256:31-36.
- [228] Stone WS, Rudd RJ and Gold PE. Glucose attenuation of atropine-induced deficits in paradoxical sleep and memory. *Brain Res* 1995;694:133-138.
- [229] Kopf SR and Baratti CM. Effects of posttraining administration of glucose on retention of a habituation response in mice: participation of a central cholinergic mechanism. *Neurobiol Learn Mem* 1996;65:253-260.
- [230] Okaichi Y and Okaichi H. Effects of glucose on scopolamine-induced learning deficits in rats performing the Morris water maze task. *Neurobiol Learn Mem* 2000;74:65-79.
- [231] Degroot A, Kornecook T, Quirion R, DeBow S and Parent MB. Glucose increases hippocampal extracellular acetylcholine levels upon activation of septal GABA receptors. *Brain Res* 2003;979:71-77.
- [232] Watson GS and Craft S. Modulation of memory by insulin and glucose: neuropsychological observations in Alzheimer's disease. *Eur J Pharmacol* 2004;490:97-113.
- [233] Wilson RS, Barnes LL, Mendes de Leon CF, Aggarwal NT, Schneider JS, Bach J, Pilat J, Beckett LA, Arnold SE, Evans DA and Bennett DA. Depressive symptoms, cognitive decline and risk of AD in older persons. *Neurology* 2002;59:364-370.
- [234] Saydah SH, Brancati FL, Golden SH, Fradkin J and Harris MI. Depressive symptoms and the risk of type 2 diabetes mellitus in a US sample. *Diabetes Metab Res Rev* 2003;19:202-208.
- [235] Arvanitakis Z, Wilson RS, Bienias JL, Evans DA and Bennett DA. Diabetes mellitus and risk of Alzheimer disease and decline in cognitive function. *Arch Neurol* 2004;61:661-666.
- [236] Gispen WH and Biessels GJ. Cognition and synaptic plasticity in diabetes mellitus. *Trends Neurosci* 2000;23:542-549.
- [237] Brownlee M. Biochemistry and molecular cell biology of diabetic complications. *Nature* 2001;414:813-820.
- [238] Biessels GJ, Van der Heide LP, Kamal A, Bley RL and Gispen WH. Ageing and diabetes: implications for brain function. *Eur J Pharmacol* 2002;441:1-14.
- [239] Meigs JB. Invited commentary: insulin resistance syndrome? Syndrome X? Multiple metabolic syndrome? A syndrome at all? Factor analysis reveals patterns in the fabric of correlated metabolic risk factors. *Am J Epidemiol* 2000;152:908-911.
- [240] Hoyer S and Lannert H. Inhibition of the neuronal insulin receptor causes Alzheimer-like disturbances in oxidative/energy brain metabolism and in behavior in adult rats. *Ann N Y Acad Sci* 1999;893:301-303.
- [241] Frolich L, Blum-Degen D, Riederer P and Hoyer S. Disturbance in the neuronal insulin receptor signal transduction in sporadic Alzheimer's disease. *Ann N Y Acad Sci* 1999;893:290-293.
- [242] Hoyer S, Lee SK, Loffler T and Schliebs R. Inhibition of the neuronal insulin receptor. An in vivo model for sporadic Alzheimer disease? *Ann N Y Acad Sci* 2000;920:256-258.
- [243] Craft S, Peskind E, Schwartz MW, Schellenberg GD, Raskind M and Porte D Jr. Cerebrospinal fluid and plasma insulin levels in Alzheimer's disease: relationship to severity of dementia and apolipoprotein E genotype. *Neurology* 1998;50:164-168.
- [244] Messier C. Diabetes, Alzheimer's disease and apolipoprotein genotype. *Exp Gerontol* 2003;38:941-946.
- [245] Craft S, Newcomer J, Kanne S, Dagogo-Jack S, Cryer P, Sheline Y, Luby J, Dagogo-Jack A and Alderson A. Memory improvement following induced hyperinsulinemia in Alzheimer's disease. *Neurobiol Aging* 1996;17:123-130.
- [246] Craft S, Asthana S, Newcomer JW, Wilkinson CW, Matos IT, Baker LD, Cherrier M, Lofgreen C, Latendresse S, Petrova A, Plymate S, Raskind M, Grimwood K and Veith RC. Enhancement of memory in Alzheimer disease with insulin and somatostatin, but not glucose. *Arch Gen Psychiatry* 1999;56:1135-1140.
- [247] Craft S, Asthana S, Cook DG, Baker LD, Cherrier M, Purganan K, Wait C, Petrova A, Latendresse S, Watson GS, Newcomer JW, Schellenberg GD and Krohn AJ. Insulin dose-response effects on memory and plasma amyloid precursor protein in Alzheimer's disease: interactions with apolipoprotein E genotype. *Psychoneuroendocrinology* 2003;28:809-822.
- [248] Gasparini L, Netzer WJ, Greengard P and Xu H. Does insulin dysfunction play a role in Alzheimer's disease? *Trends Pharmacol Sci* 2002;23:288-293.
- [249] Skeberdis VA, Lan J, Zheng X, Zukin RS and Bennett MV. Insulin promotes rapid delivery of N-methyl-D-aspartate receptors to the cell surface by exocytosis. *Proc Natl Acad Sci USA* 2001;98:3561-3566.
- [250] Byrne JH, In: Squire LR, Bloom FE, McConnell SK, Roberts JL, Spitzer NC and Zigmond MJ. (Eds.). Learning and memory: basic mechanisms. Fundamental Neuroscience. Academic Press, San Diego, CA. 2003; pp1276-1298.
- [251] Davies MG, Ramkumar V, Gettys TW and Hagen PO. The expression and function of G-proteins in experimental intimal hyperplasia. *J*

- Clin Invest* 1994;94:680-1689.
- [252] Gilligan DM, Guetta V, Panza JA, Garcia CE, Quyyumi AA and Cannon RO. Selective loss of microvascular endothelial function in human hypercholesterolemia. *Circulation* 1994;90:35-41.
- [253] Mancusi G, Hutter C, Baumgartner-Parzer S, Schmidt K, Schutz W and Sexl V. High-glucose incubation of human umbilical-vein endothelial cells does not alter expression and function either of G-protein alpha-subunits or of endothelial NO synthase. *Biochem J* 1996;315:281-287.
- [254] Sobrevia L, Cesare P, Yudilevich DL and Mann GE. Diabetes-induced activation of γ and nitric oxide synthase in human endothelial cells: association with membrane hyperpolarization. *J Physiol* 1995;489:183-192.
- [255] Pieper GM and Peltier BA. Amelioration by L-arginine of a dysfunctional arginine/nitric oxide pathway in diabetic endothelium. *J Cardiovasc Pharmacol* 1995;25:397-403.
- [256] Pieper GM, Siebeneich W, Moore-Hilton G and Roza AM. Reversal by L-arginine of a dysfunctional arginine/nitric oxide pathway in the endothelium of the genetic diabetic BB rat. *Diabetologia* 1997;40:910-915.
- [257] Knowles RG and Moncada S. Review article: nitric oxide synthases in mammals. *Biochem J* 1994;298:249-258.
- [258] Asahina T, Kashiwagi A and Nishio Y. Impaired activation of glucose oxidation and NADPH supply in human endothelial cells exposed to H_2O_2 in high-glucose medium. *Diabetes* 1995;44:520-526.
- [259] Honing MLH, Morrison PJ, Banga JD, Stroes ESG and Rabelink TJ. Nitric oxide availability in diabetes mellitus. *Diabetes Metab Rev* 1998;14:241-249.
- [260] Ragoobirsingh D, McGrowder D, Dasgupta T and Brown P. The effect of nitric oxide on glucose metabolism. *Mol Cell Biochem* 2004;263:29-34.
- [261] Jobgen WS, Fried SK, Fu WJ, Meininger CJ and Wu G. Regulatory role for the arginine-nitric oxide pathway in metabolism of energy substrates. *J Nutr Biochem* 2006;17:571-588.
- [262] Kohli R, Meininger CJ, Haynes TE, Yan W, Self JT and Wu G. Dietary L-arginine supplementation enhances endothelial nitric oxide synthesis in streptozotocin-induced diabetic rats. *J Nutr* 2004;134:600-608.
- [263] Fu WJ, Haynes TE, Kohli R, Hu J, Shi W, Spencer TE, Carroll RJ, Meininger CJ and Wu G. Dietary L-arginine supplementation reduces fat mass in Zucker diabetic fatty rats. *J Nutr* 2005;135:714-721.
- [264] Balon TW. Role of nitric oxide in contraction induced glucose transport. *Adv Exp Med Biol* 1998;441:87-95.
- [265] Bergandi L, Silvagno F, Russo I, Riganti C, Anfossi G, Aldieri E, Ghigo D, Trovati M and Bosia A. Insulin stimulates glucose transport via nitric oxide/cyclic GMP pathway in human vascular smooth muscle cells. *Arterioscler Thromb Vasc Biol* 2003;23:2215-2221.
- [266] Tanaka T, Nakatani K, Morioka K, Urakawa H, Maruyama N, Kitagawa N, Katsuki A, Araki-Sasaki R, Hori Y, Gabazza EC, Yano Y, Wada H, Nobori T, Sumida Y and Adachi Y. Nitric oxide stimulates glucose transport through insulin-independent GLUT4 translocation in 3T3-L1 adipocytes. *Eur J Endocrinol* 2003;149:61-67.
- [267] Higaki Y, Hirshman MF, Fujii N and Laurie J. Nitric oxide increases glucose uptake through a mechanism that is distinct from the insulin and contraction pathways in rat skeletal muscle. *Diabetes* 2001;50:241-247.
- [268] Henstridge DC, Kingwell BA, Formosa MF, Drew BG, McConell GK and Duffy SJ. Effects of the nitric oxide donor, sodium nitroprusside, on resting leg glucose uptake in patients with type 2 diabetes. *Diabetologia* 2005;48:2602-2608.
- [269] McGrowder D, Ragoobirsingh D and Brown P. Acute effects of exogenous nitric oxide on glucose uptake in skeletal muscle of normoglycaemic and diabetic rats. *Med Sci Monit* 2006;12:28-35.
- [270] McGrowder D, Ragoobirsingh D and Brown P. Modulation of glucose uptake in adipose tissue by nitric oxide-generating compounds. *J Biosci* 2006;31:347-354.
- [271] Schmidt HH, Warner TD, Ishii K, Sheng H and Murad F. Insulin secretion from pancreatic B cells caused by L-arginine-derived nitrogen oxide. *Science* 1992;255:721-723.
- [272] Smukler SR, Tang L, Wheeler MB and Salapatek AMF. Exogenous nitric oxide and endogenous glucose-stimulated β -cell nitric oxide augment insulin release. *Diabetes* 2002;51:3450-3460.
- [273] Thams P and Capito K. L-Arginine stimulation of glucose-induced insulin secretion through membrane depolarization and independent of nitric oxide. *Eur J Endocrinol* 1999;140:87-93.
- [274] Muniappan L and Ozcan S. Induction of insulin secretion in engineered liver cells by nitric oxide. *BMC Physiol* 2007;17:7-11.
- [275] Guarino MP, Correia NC, Lauth WW and Macedo MP. Insulin sensitivity is mediated by the activation of the ACh/NO/cGMP pathway in rat liver. *Am J Physiol Gastrointest Liver Physiol* 2004;287:527-532.
- [276] Guarino MP and Macedo MP. Co-administration of glutathione and nitric oxide enhances insulin sensitivity in Wistar rats. *Br J Pharmacol* 2006;147:959-965.
- [277] Juan CC, Chang CL, Chuang TY, Huang SW, Kwok CF and Ho LT. Insulin sensitivity and resistin expression in nitric oxide-deficient rats. *Diabetologia* 2006;49:3017-3026.
- [278] Butler R, Morris AD and Struthers AD.

- Systemic nitric oxide synthase inhibition increases insulin sensitivity in man. *Clin Sci (Lond)* 1998;94:175-180.
- [279] Pollard HB, Adeyemo M, Dhariwal K, Levine M, Caohuy H, Markey S, Markey CJ and Youdim MB. The goldfish as a drug discovery vehicle for Parkinson's disease and other neurodegenerative disorders. *Ann N Y Acad Sci* 1993;28:679:317-320.
- [280] Barker RA. Prospects for the treatment of Parkinson's disease using neural grafts. *Expert Opin Pharmacother* 2000;1:889-902.
- [281] Reynolds GP. Antipsychotic drug use in neurodegenerative disease in the elderly: problems and potential from a pharmacological perspective. *Expert Opin Pharmacother* 2001;2:543-548.
- [282] Poulter MO, Payne KB and Steiner JP. Neuroimmunophilins: a novel drug therapy for the reversal of neurodegenerative disease? *Neuroscience* 2004;128:1-6.
- [283] Kuan WL and Barker RA. New therapeutic approaches to Parkinson's disease including neural transplants. *Nerorehabil Neural Repair* 2005;19:155-181.
- [284] Waldmeier P, Bozyczko-Coyne D, Williams M and Vaught JL. Recent clinical failures in Parkinson's disease with apoptosis inhibitors underline the need for a paradigm shift in drug discovery for neurodegenerative diseases. *Biochem Pharmacol* 2006;72:1197-1206.
- [285] Melnikova I. Therapies for Alzheimer's disease. *Nat Rev Drug Discov* 2007;6:341-342.
- [286] Gage FH. Structural plasticity: cause, result, or correlate of depression. *Biol Psychiatry* 2000;48:713-714.
- [287] Alvarez-Buylla A and Garcia-Verdugo JM. Neurogenesis in adult subventricular zone. *J Neurosci* 2002;22:629-634.
- [288] Alvarez-Buylla A, Seri B and Doetsch F. Identification of neural stem cells in the adult vertebrate brain. *Brain Res Bull* 2002;57:751-758.
- [289] Gage FH. Neurogenesis in the adult brain. *J Neurosci* 2002;22:612-613.
- [290] Eriksson PS, Perfilieva E, Bjork-Eriksson T, Alborn AM, Nordborg C, Peterson DA and Gage FH. Neurogenesis in the adult human hippocampus. *Nat Med* 1998;4:1313-1317.
- [291] Kornack DR and Rakic P. Continuation of neurogenesis in the hippocampus of the adult macaque monkey. *Proc Natl Acad Sci USA* 1999;96:5768-5773.
- [292] van Praag H, Schlinder AF, Christle BR, Toni N, Palmer TD and Gage FH. Functional neurogenesis in the adult hippocampus. *Nature* 2002;415:1030-1034.
- [293] Carleton A, Petreanu LT, Lansford R, Alvarez-Buylla A and Lledo PM. Becoming a new neuron in the adult olfactory bulb. *Nat Neurosci* 2003;6:507-518.
- [294] Belluzzi O, Benedusi M, Ackman M and LoTurco JJ. Electrophysiological differentiation of new neurons in the olfactory bulb. *J Neurosci* 2003;23:10411-10418.
- [295] Lee SH, Lumelsky N, Studer L, Auerbach JM and McKay RD. Efficient generation of midbrain and hindbrain neurons from mouse embryonic stem cells. *Nat Biotechnol* 2000;18:675-679.
- [296] Tropepe V, Hitoshi S, Sirard C, Mak TW, Rossant J and van der Kooy D. Direct neural fate specification from embryonic stem cells: a primitive mammalian neural stem cell stage acquired through a default mechanism. *Neuron* 2001;30:65-78.
- [297] Bjorklund LM, Sanchez-Pernaute T, Chung S, Anderson T, Chen I, Chen Y, McNaught K, Brownell AL, Jenkins BG, Wahlestedt C, Kim KS and Isacson O. Embryonic stem cells develop into functional dopaminergic neurons after transplantation in a Parkinson rat model. *Proc Natl Acad Sci USA* 2002;99:2344-2349.
- [298] Rathjen J, Haines BP, Hudson KM, Nesci A, Dunn S and Rathjen PD. Directed differentiation of pluripotent cells to neural lineages: homogeneous formation and differentiation of a neuroectoderm population. *Development* 2002;129:2649-2661.
- [299] Tabar V, Panagiotakos G, Greenberg ED, Chan BK, Sadelain M, Gutin PH and Studer L. Migration and differentiation of neural precursors derived from human embryonic stem cells in the rat brain. *Nat Biotechnol* 2005;23:601-606.
- [300] Ideda H, Osakada F, Watanabe K, Mizuseki K, Haraguchi T, Miyoshi H, Kamiya D, Honda Y, Sasai N, Yoshimura N, Takahashi M and Sasai Y. Generation of Rx+/Pax6+ neural retinal precursors from embryonic stem cells. *Proc Natl Acad Sci USA* 2005;102:11331-11336.
- [301] Lumelsky N, Lee S-H, Nguyen LJ, Sanchez-Pernaute R, Bankiewicz K and McKay R. Dopamine neurons derived from embryonic stem cells function in an animal model of Parkinson's disease. *Nature* 2002;418:50-56.
- [302] Kim SU, Park IH, Kim TH, Kim KS, Choi HB, Hong SH, Bang JH, Lee MA, Joo IS, Lee CS and Kim YS. Brain transplantation of human neural stem cells transduced with tyrosine hydroxylase and GTP cyclohydrolase 1 provides functional improvement in animal models of Parkinson disease. *Neuropathology* 2006;26:129-140.
- [303] Fu YS, Cheng YC, Lin MA, Cheng H, Chu PM, Chou SC, Shih YH, Ko M-H and Sung MS. Conversion of human umbilical cord mesenchymal stem cells in Wharton's jelly to dopaminergic neurons in vitro: potential therapeutic application for Parkinsonism. *Stem Cells* 2006;24:115-124.
- [304] Kee NJ, Preston E and Witowicz JM. Enhanced neurogenesis after transient global

- ischemia in the dentate gyrus of the rat. *Exp Brain Res* 2001;136:313-320.
- [305] van Praag H, Schinder AF, Christie BR, Toni N, Palmer TD and Gage FH. Functional neurogenesis in the adult hippocampus. *Nature* 2002;415:1030-1034.
- [306] Song HJ, Stevens CF and Gage FH. Neural stem cells from adult hippocampus develop essential properties of functional CNS neurons. *Nat Neurosci* 2002;5:438-445.
- [307] Rodriguez PC, Quiceno DG and Ochoa AC. L-arginine availability regulates T-lymphocyte cell-cycle progression. *Blood* 2007;109:1568-1573.
- [308] Hua TC and Mochhala SM. Influence of L-arginine, aminoguanidine, and NG-nitro-L-arginine methyl ester (L-name) on the survival rate in a rat model of hemorrhagic shock. *Shock* 1999;11:51-57.
- [309] Dodd F, Limoges M, Boudreau RT, Rowden G, Murphy PR and Too CK. L-arginine inhibits apoptosis via a NO-dependent mechanism in Nb2 lymphoma cells. *J Cell Biochem* 2000;77:624-634.
- [310] Holm AM, Andersen CB, Hauns S and Hansen PR. Effects of L-arginine on vascular smooth muscle cell proliferation and apoptosis after balloon injury. *Scand Cardiovasc J* 2000;34:28-32.
- [311] Erden CD, Ekmekci A, Sahin FI, Ergün MA, Oztürk G and Erbas D. L-arginine and mitomycin C-induced nitric oxide release and apoptosis in human lymphocytes. *Cell Biol Int* 2003;27:337-340.
- [312] Suschek CV, Schnorr O, Hemmrich K, Aust O, Klotz LO, Sies H and Kolb-Bachofen V. Critical role of L-arginine in endothelial cell survival during oxidative stress. *Circulation* 2003;107:2607-2614.
- [313] Hammerschmidt S, Kuhn H, Grasenack T, Gessner C and Wirtz H. Apoptosis and necrosis induced by cyclic mechanical stretching in alveolar type-II-cells—influence of captopril and L-arginine. *Pneumologie* 2004;58:222-229.
- [314] Ingram A, Parbtani A, Thai K, Ly H, Shankland SJ, Morrissey G and Scholey JW. Dietary supplementation with L-arginine limits cell proliferation in the remnant glomerulus. *Kidney Int* 1995;48:1857-1865.
- [315] Shima Y, Maeda T, Aizawa S, Tsuboi I, Kobayashi D, Kato R and Tamai I. L-arginine import via cationic amino acid transporter CAT1 is essential for both differentiation and proliferation of erythrocytes. *Blood* 2006;107:1352-1356.
- [316] Becker-Catania SG, Gregory TL, Yang Y, Gau C-L, de Vellis J, Cederbaum SD and Iyer RK. Loss of arginase I results in increased proliferation of neural stem cells. *J Neurosci Res* 2006;84:735-746.
- [317] De Jonge WJ, Marescau B, D'Hooge R, De Deyn PP, Hallemeesch MM, Deutz NE, Ruijter JM and Lamers WH. Overexpression of arginase alters circulating and tissue amino acids and guanidine compounds and affects neuromotor behavior in mice. *J Nutr* 2001;131:2732-2740.
- [318] Cai D, Deng K, Mellado W, Lee J, Ratan RR and Filbin MT. Arginase I and polyamines act downstream from cyclic AMP in overcoming inhibition of axonal growth MAG and myelin in vitro. *Neuron* 2002;35:711-719.
- [319] Esch F, Lin KI, Hills A, Zaman K, Baraban JM, Chatterjee S, Rubin L, Ash DE and Ratan RR. Purification of a multipotent antideath activity from bovine liver and its identification as arginase: nitric oxide-independent inhibition of neuronal apoptosis. *J Neurosci* 1998;18:4083-4095.
- [320] Estévez AG, Sahawneh MA, Lange PS, Bae N, Egea M and Ratan RR. Arginase 1 regulation of nitric oxide production is key to survival of trophic factor-deprived motor neurons. *J Neurosci* 2006;26:8512-8516.
- [321] Du C, Hu R, Csernansky CA, Hsu CY and Choi DW. Additive neuroprotective effects of dextrorphan and cycloheximide in rats subjected to transient focal cerebral ischemia. *Brain Res* 1996;718:233-236.
- [322] Du C, Hu R, Csernansky CA, Hsu CY and Choi DW. Very delayed infarction after focal cerebral ischemia: a role for apoptosis? *J Cereb Blood Flow Metab* 1996;16:195-201.
- [323] Cai D, Qiu J, Cao Z, McAtee M, Bregman BS and Filbin MT. Neuronal cyclic AMP controls the developmental loss in ability of axons to regenerate. *J Neurosci* 2001;21:4731-4739.
- [324] Dawson VL, Dawson TM, London ED, Bredt DS and Snyder SH. Nitric oxide mediates glutamate neurotoxicity in primary cortical cultures. *Proc Natl Acad Sci USA* 1991;88:6368-6371.
- [325] Sonoki T, Nagasaki A, Gotoh T, Takiguchi M, Takeya M, Matsuzaki H and Mori M. Coinduction of nitric-oxide synthase and arginase I in cultured rat peritoneal macrophages and rat tissues in vivo by lipopolysaccharide. *J Biol Chem* 1997;272:3689-3693.
- [326] Wallace HM, Fraser AV and Hughes A. A perspective of polyamine metabolism. *Biochem J* 2003;376:1-14.
- [327] Seiler N. Polyamine metabolism and function in brain. *Neurochem Int* 1981;3:95-110.
- [328] Slotkin TA and Bartolome J. Role of ornithine decarboxylase and the polyamines in nervous system development: A review. *Brain Res Bull* 1986;17:307-320.
- [329] Schweitzer L, Robbins AJ and Slotkin TA. Dendritic development of Purkinje and granule cells in the cerebellar cortex of rats treated postnatally with alpha-difluoromethylornithine. *J Neuropathol Exp Neurol* 1989;48:11-22.
- [330] Harada J and Sugimoto M. Polyamines

- prevent apoptotic cell death in cultured cerebellar granule neurons. *Brain Res* 1997; 753:251-259.
- [331] Chu PJ, Saito H and Abe K. Polyamines promote regeneration of injured axons of cultured rat hippocampal neurons. *Brain Res* 1995;673:233-241.
- [332] Malaterre J, Strambi C, Aouane A, Strambi A, Rougon G and Cayre M. A novel role for polyamines in adult neurogenesis in rodent brain. *Eur J Neurosci* 2004;20:317-330.
- [333] Cayre M, Malaterre M, Strambi C, Charpin P, Ternaux J-P and Strambi A. Short- and long-chain natural polyamines play specific roles in adult cricket neuroblast proliferation and neuron Differentiation in Vitro. *J Neurobiol* 2001;48:315-324.
- [334] Thomas T and Thomas TJ. Regulation of cyclin B1 by estradiol and polyamines in MCF-7 breast cancer cells. *Cancer Res* 1994;54: 1077-1084.
- [335] Thomas T, Gallo MA, Klinge CM and Thomas TJ. Polyamine-mediated conformational perturbations in DNA alter the binding of estrogen receptor to poly(dG-m5dC) z poly(dG-m5dC) and a plasmid containing the estrogen response element. *J Steroid Biochem Mol Biol* 1995;54:89-99.
- [336] Kaminska B, Kaczmarek L and Grzelakowska-Sztabert B. Inhibitors of polyamine biosynthesis affect the expression of genes encoding cytoskeletal proteins. *FEBS Lett* 1992;304:198-200.
- [337] Filhol O, Loue-Mackenbach P, Cochet C and Chambaz EM. Casein kinase II and polyamines may interact in the response of adrenocortical cells to their trophic hormone. *Biochem Biophys Res Comm* 1991;180:623-630.
- [338] Ulloa L, Diaz-Nido J and Avila J. Depletion of casein kinase II by antisense oligonucleotide prevents neuritogenesis in neuroblastoma cells. *EMBO J* 1993;12:1633-1640.
- [339] Packham G and Cleveland JL. Ornithine decarboxylase is a mediator of c-Myc-induced apoptosis. *Mol Cell Biol* 1994;14:5741-5747.
- [340] Packham G and Cleveland JL. The role of ornithine decarboxylase in c-Myc-induced apoptosis. *Curr Top Microbiol Immunol* 1995; 194:283-290.
- [341] Sparapani M, Dall'Olio R, Gandolfi O, Ciani E and Contestabile A. Neurotoxicity of polyamines and pharmacological neuroprotection in cultures of rat cerebellar granule cells. *Exp Neurol* 1997;148:157-166.
- [342] Brunton VG, Grant MH and Wallace HM. Mechanisms of spermine toxicity in baby-hamster kidney BHK cells. *Biochem J* 1991; 280:193-198.
- [343] Gilad GM and Gilad VH. Polyamines affect growth of cultured rat cerebellar neurons in different sera. *Int J De'l Neurosci* 1986;4:195-208.
- [344] Seiler N. Pharmacological properties of the natural polyamines and their depletion by biosynthesis inhibitors as a therapeutic approach. *Prog Drug Res* 1991;37:107-159.
- [345] Johnson TD. Modulation of channel function by polyamines. *Trends Pharmacol Sci* 1996; 17:22-27.
- [346] Gilad GM, Dornay M and Gilad VH. Polyamine treatment in early development leads to increased numbers of rat sympathetic neurons. *Brain Res* 1985;348:363-366.
- [347] Abe K, Chida N, Nishiyama N and Saito H. Spermine promotes the survival of primary cultured brain neurons. *Brain Res* 1993;605: 322-326.
- [348] Chu PJ, Saito H and Abe K. Polyamines promote neurite elongation of cultured rat hippocampal neurons. *Neurosci Res* 1994;19: 155-160.
- [349] Redman C, Xu MJ, Peng YM, Scott JA, Payne C, Clark LC and Nelson MA. Involvement of polyamines in selenomethionine induced apoptosis and mitotic alterations in human tumor cells. *Carcinogenesis* 1997;18:1195-1202.
- [350] Morrison B 3rd , Pringle AK, McManus T, Ellard J, Bradley M, Signorelli F, Iannotti F and Sundstrom LE. L-arginyl-3,4-spermidine is neuroprotective in several in vitro models of neurodegeneration and in vivo ischaemia without suppressing synaptic transmission. *Br J Pharmacol* 2002;137:1255-1268.
- [351] Gilad GM and Gilad VH. Polyamines can protect against ischemia-induced nerve cell death in gerbil forebrain. *Exp Neurol* 1991; 111:349-355.
- [352] Kindy MS, Hu Y, Dempsy RJ and Cereb J. Blockade of ornithine decarboxylase enzyme protects against ischemic brain damage. *Blood Flow Metab* 1994;14:1040-1045.
- [353] Rao AM, Baskaya MK, Maley ME, Prasad MR and Dempsey RJ. Ornithine decarboxylase activity and edema formation in cerebral ischemia of conscious gerbils. *J Neurochem* 1995;65:2639-2643.
- [354] Sauer DS, Martin P, Allegrini PR, Bernasconi R, Amacker H and Fagg GE. Differing effects of α -difluoromethylornithine and CGP 40116 on polyamine levels and infarct volume in a rat model of focal cerebral ischemia. *Neurosci Lett* 1992;41:131-135.
- [355] Gilad GM and Gilad VH. Polyamines in neurotrauma. *Biochem Pharmacol* 1992;44: 401-407.
- [356] Ciani E, Calvanese V, Crochemore C, Bartesaghi R and Contestabile A. Proliferation of cerebellar precursor cells is negatively regulated by nitric oxide in newborn rat. *J Cell Sci* 2006;119(Pt 15):3161-3170.
- [357] Torroglosa A, Murillo-Carretero M, Romero-Grimaldi C, Matarredona ER, Campos-Caro A and Estrada C. Nitric oxide decreases subventricular zone stem cell proliferation by

- inhibition of epidermal growth factor receptor and phosphoinositide-3-kinase/Akt pathway. *Stem Cells* 2007;25:88-97.
- [358] Moreno-Lopez B, Romero-Grimaldi C, Noval JA, Murillo-Carretero M, Matarredona ER and Estrada C. Nitric oxide is a physiological inhibitor of neurogenesis in the adult mouse subventricular zone and olfactory bulb. *J Neurosci* 2004;24:85-95.
- [359] Cheng A, Wang S, Rao MS and Mattson MP. Nitric oxide acts in a positive feedback loop with BDNF to regulate neural progenitor cell proliferation and differentiation in the mammalian brain. *Dev Biol* 2003;258:319-333.
- [360] Matarredona ER, Murillo-Carretero M, Moreno-Lopez B and Estrada C. Nitric oxide synthesis inhibition increases proliferation of neural precursors isolated from the postnatal mouse subventricular zone. *Brain Res* 2004;995:274-284.
- [361] Zhang R, Zhang L, Zhang Z, Wang Y, Lu M, Lapointe M and Chopp M. A nitric oxide donor induces neurogenesis and reduces functional deficits after stroke in rats. *Ann Neurol* 2001;50:602-611.
- [362] Cheng A, Chan SL, Milhavet O, Wang S and Mattson MP. p38 MAP kinase mediates nitric oxide-induced apoptosis of neural progenitor cells. *J Biol Chem* 2001;276:43320-43327.
- [363] Palluy O and Rigaud M. Nitric oxide induces cultured cortical neuron apoptosis. *Neurosci Lett* 1996;208:1-4.
- [364] Estévez AG, Spear N, Manuel SM, Radi R, Henderson CE, Barbeito L and Beckman JS. Nitric oxide and superoxide contribute to motor neuron apoptosis induced by trophic factor deprivation. *J Neurosci* 1998;18:923-931.
- [365] Martin LJ, Chen K and Liu Z. Adult motor neuron apoptosis is mediated by nitric oxide and Fas death receptor linked by DNA damage and p53 activation. *J Neurosci* 2005;25:6449-6459.
- [366] Zhu XJ, Hua Y, Jiang J, Zhou QG, Luo CX, Han X, Lu YM and Zhu DY. Neuronal nitric oxide synthase-derived nitric oxide inhibits neurogenesis in the adult dentate gyrus by down-regulating cyclic AMP response element binding protein phosphorylation. *Neuroscience* 2006;141:827-836.
- [367] Fritzen S, Schmitt A, Köth K, Sommer C, Lesch KP and Reif A. Neuronal nitric oxide synthase (NOS-I) knockout increases the survival rate of neural cells in the hippocampus independently of BDNF. *Mol Cell Neurosci* 2007;35:261-271.
- [368] Ciani E, Severi S, Contestabile A and Bartesaghi R. Nitric oxide negatively regulates proliferation and promotes neuronal differentiation through N-Myc downregulation. *J Cell Sci* 2004;117:4727-4737.
- [369] Park C, Kang M, Kwon YK, Chung JH, Ahn H and Huh Y. Inhibition of neuronal nitric oxide synthase enhances cell proliferation in the dentate gyrus of the adrenalectomized rat. *Neurosci Lett* 2001;309:9-12.
- [370] Xiong H, Yamada K, Han D, Nabeshima T, Enikolopov G, Carnahan J and Nawa H. Mutual regulation between the intercellular messengers nitric oxide and brain-derived neurotrophic factor in rodent neocortical neurons. *Eur J Neurosci* 1999;11:1567-1576.
- [371] Reif A, Schmitt A, Fritzen S, Chourbaji S, Bartsch C, Urani A, Wycislo M, Mössner R, Sommer C, Gass P and Lesch KP. Differential effect of endothelial nitric oxide synthase (NOS-III) on the regulation of adult neurogenesis and behaviour. *Eur J Neurosci* 2004;20:885-895.
- [372] Zhao X, Lu X and Feng Q. Deficiency in endothelial nitric oxide synthase impairs myocardial angiogenesis. *Am J Physiol Heart Circ Physiol* 2002;283:2371-2378.
- [373] Chen J, Zacharek A, Zhang C, Jiang H, Li Y, Roberts C, Lu M, Kapke A and Chopp M. Endothelial nitric oxide synthase regulates brain-derived neurotrophic factor expression and neurogenesis after stroke in mice. *J Neurosci* 2005;25:2366-2375.
- [374] Dulak J and Jozkowicz A. Regulation of vascular endothelial growth factor synthesis by nitric oxide: facts and controversies. *Antioxid Redox Signal* 2003;5:123-132.
- [375] Zhu Y, Jin K, Mao XO and Greenberg DA. Vascular endothelial growth factor promotes proliferation of cortical neuron precursors by regulating E2F expression. *FASEB J* 2003;17:186-193.
- [376] Park JS, Hong GR, Baek SW, Shin DG, Kim YJ and Shim BS. Expression and regulation of endothelial nitric oxide synthase by vascular endothelial growth factor in ECV 304 cells. *J Korean Med Sci* 2002;17:161-167.
- [377] Jin K, Zhu Y, Sun Y, Mao XO, Xie L and Greenberg DA. Vascular endothelial growth factor (VEGF) stimulates neurogenesis in vitro and in vivo. *Proc Natl Acad Sci USA* 2002;99:11946-11950.
- [378] Zhu DY, Deng Q, Yao HH, Wang DC, Deng Y and Liu GQ. Inducible nitric oxide synthase expression in the ischemic core and penumbra after transient focal cerebral ischemia in mice. *Life Sci* 2002;17:1985-1997.
- [379] Zhu DY, Liu SH, Sun HS and Lu YM. Expression of inducible nitric oxide synthase after focal cerebral ischemia stimulates neurogenesis in the adult rodent dentate gyrus. *J Neurosci* 2003;23:223-229.
- [380] Bernabeu R and Sharp FR. NMDA and AMPA/kainite glutamate receptors modulate dentate gyrus neurogenesis and CA3 synapsin-I in normal and ischemic

- hippocampus. *J Cereb Blood Flow Metab* 2000;20:1669-1680.
- [381] Arvidsson A, Kokaia Z and Lindvall O. N-methyl-D-aspartate receptor-mediated increase of neurogenesis in adult rat dentate gyrus following stroke. *Eur J Neurosci* 2001; 14:10-18.
- [382] Cardenas A, Moro MA, Hurtado O, Leza JC, Lorenzo P, Castrillo A, Bodelon OG, Bosca L and Lizasoain I. Implication of glutamate in the expression of inducible nitric oxide synthase after oxygen and glucose deprivation in rat forebrain slices. *J Neurochem* 2000;74:2041-2048.
- [383] Jander S, Schroeter M and Stoll G. Role of NMDA receptor signaling in the regulation of inflammatory gene expression after focal brain ischemia. *J Neuroimmunol* 2000;109: 181-187.
- [384] Qu XW, Wang H, De Plaen IG, Rozenfeld RA and Hsueh W. Neuronal nitric oxide synthase (NOS) regulates the expression of inducible NOS in rat small intestine via modulation of nuclear factor kappa B. *FASEB J* 2001;15: 439-446.
- [385] Luo CX, Zhu XJ, Zhou QG, Wang B, Wang W, Cai HH, Sun YJ, Hu M, Jiang J, Hua Y, Han X and Zhu DY. Reduced neuronal nitric oxide synthase is involved in ischemia-induced hippocampal neurogenesis by up-regulating inducible nitric oxide synthase expression. *J Neurochem* 2007;103:1872-1882.
- [386] Togashi H, Sasaki M, Frohman E, Taira E, Ratan RR, Dawson TM and Dawson VL. Neuronal (type I) nitric oxide synthase regulates nuclear factor kappaB activity and immunologic (type II) nitric oxide synthase expression. *Proc Natl Acad Sci USA* 1997;94: 2676-2680.