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## **EFFECT OF SOME NATURAL PREPARATIONS ON CONTENTS OF PROTEIN AND DRY MATTER IN SWEET PEPPER (*CAPSICUM ANNUUM*L.)**

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### **ABSTRACT**

The aim of study conducted in 2010–2012 was to evaluate the influence of some natural preparations on the protein and dry matter content in the sweet pepper fruits. Fruits of sweet pepper (cv. Roberta F1) were the object of research. The natural preparations from garlic extract (Bioczso Plynny), grapefruit extract (Biosept 33 SL), sea algae (Bio-algeen S90 Plus) and blastospores of *Aureobasidium pullulans* (Boni Protect Forte) were used for protection of pepper plants. Determination of protein and dry matter in pepper fruits was made during the middle of the harvest period.

The presented results indicated small differences in protein and dry matter content between the fruits from control combinations and those protected with natural preparations. It belongs to ascertain, that none of the applied natural preparations had a significant effect on the content of protein and dry matter in pepper fruits. Therefore, it should be stated that the application of tested preparations cannot decrease the quality of produced pepper and that the application of natural products does not result in the reduce the nutritional value of pepper, regarding proteins and dry matter, therefore they can be used in the practice.

**Key words:** *Capsicum annuum*, biological control, garlic extract, grapefruit extract, sea algae, *Aureobasidium pullulans*, protein, dry matter.

### **INTRODUCTION**

Recently, the consumers are more conscious of quality and safety of food. The management of pepper diseases relies in a high degree on using the fungicides. However, the chemical control results in the residues of fungicides not only in treated plants but also in the soil what causes the disturbance of biological balance. Additionally, the fungicides, regarded earlier as relatively safe, proved to be more dangerous to people's health than insecticides and herbicides together. Their residues in food might be carcinogenic [32]. Their resistance of pathogens to fungicides also is a problem.

From 1 January 2014 the crop production in all member countries of European Union has to be based on the rules of integrated plant protection according to the Council Directive 2009/128/CE on the sustainable use of pesticides and the later regulations of the European Parliament and the Council [6, 28].

The yield of crops depends on many factors, among them one of the most important is the health status of plants. Recent activities focus on developing natural products that can be effective, long lasting and safe to human beings and the environment. Biological and biotechnical products show antifungal and antibacterial activity against many pathogens [7, 14, 15, 21, 33, 34] and are accepted by the consumers as environmentally friendly.

There are many reports on the efficacy of natural products in plant protection but there is no full information on the effect of

these preparations on the content of chemical compounds and nutrients in pepper fruits. The aim of the study was the answer to the question: is nutritional value of pepper fruits treated with natural preparations higher or lower than of those cultivated traditionally?

In the presented study we compared the contents of protein and dry matter in sweet pepper fruits treated with the natural products to those protected with the fungicide.

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## MATERIAL AND METHODS

### Field experiment

The experiment was carried out in 2010–2012 in the horticultural farm in Zezulin in Lublin province (N51°20', E22°49'). The objects of research were the plants of sweet pepper (*Capsicum annuum* L.) cv. 'Roberta F<sub>1</sub>'.

Pepper seedlings were planted in the field in the second decade of May in the spacing of 0.67 × 0.35 m. Twenty seedlings were planted in each plot of 4.69 m<sup>2</sup>. In each year of investigations the experiment was set up in randomized blocks in 4 replications. The investigated factors were the natural preparations which types, application methods and concentrations are described in Table 1. There were two control combinations: absolute (with plants without any treatment) and relative (with plants treated with the fungicide Amistar 250 SC). Around experiment plots one row of pepper plants was planted. Those plants were not used in the experiment.

Table 1. Combinations of field experiment in 2010–2012

Experimental combination	Concentration of preparation [%]	Number of treatments
C – control, plant without treatments	lack	lack
A – control, plants protected with azoxystrobin (Amistar 250 SC)	0.1	2
BCZ – plants protected with garlic pulp extract (biotechnical preparation Bioczos Plymny)	2.0	6
BS – plants protected with grapefruit extract (biotechnical preparation Biosept 33SL)	0.2	6
BA – plants protected with sea algae <i>Ascophyllum nodosum</i> (biotechnical preparation Bio-algeen S90 Plus)	0.3	6
BP – plants protected with blastospores fungi <i>Aureobasidium pullulans</i> strains (biopreparation Boni Protect Forte)	0.1	6

### Analysis of protein and dry matter content

The evaluation of protein content in pepper fruits was conducted with Kjeldahl's method, while the estimation of dry matter content was done with dryer method. Standard uncertainty ( $\pm U$ ) for the determination of protein content was 9.08%, for dry matter standard uncertainty was 1.64%. These uncertainties were estimated taking into account the coverage factor  $k = 2$ , which provides a level of confidence of approximately 95%.

### Statistical analysis

The data were analyzed by the analysis of variance (Tukey's test) at 5% significance level using the SAS statistical system (SAS Version 9.1, SAS Inst., Cary, N.C., USA).

## RESULTS AND DISCUSSION

The biological value and chemical composition of pepper fruits depend on many factors, like the cultivar, cultivation method, variable environment conditions, harvest time and the type of agrichemicals [3–5, 9, 12, 13, 16, 17, 19, 22, 23]. The content of nutrients in the fruits changes depending on their ripening stage [18, 20, 23].

The fruits of sweet pepper are the source of many vitamins and minerals (magnesium, calcium, potassium, phosphorus, iron) that have a great importance in the diet of each human being [2, 3, 10, 12, 13, 17, 20, 24–27]. They contain also many antioxidants (vitamins C and E, beta-carotene) that play an important role in human nourishments [27, 32]. The epidemiologic researches indicate that these compounds increase the immunity system protecting the human organisms against the diseases of civilization [11, 25]. The fruits of sweet pepper are not an important source of proteins but the content of these compounds have a great biological importance [31, 32]. Schuphan [29] presented the studies on protein content in pepper where the amount of raw protein varied between 1.2% and 1.5% in fresh fruits. Somos [31] describes several studies undertaken in Hungary on this group of chemical compounds. The results of those researches showed that the amount of protein in pepper pericarp was 16–17% of dry matter. According to the results of investigations conducted by Simonovska et al. [30], the protein content in the pericarp of pepper cultivated in Macedonia amounted to 14.13% of total dry matter.

The analysis of obtained results indicates that the content of protein in the fresh fruits of sweet pepper was rather low: from

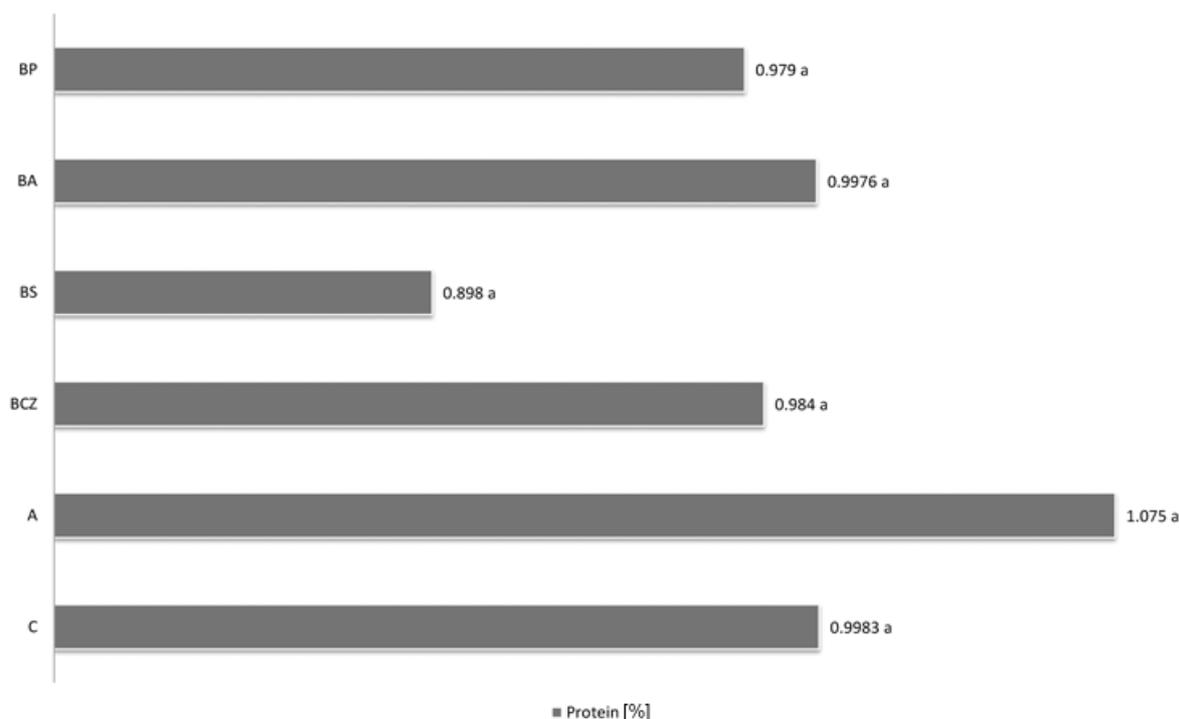
0.633% (in 2010) for the fruits protected with Biosept to 1.15% (in 2012) for the combination with Boni Protect (Tab. 2) and was similar to the results obtained by some researchers [29–31].

**Table 2. Content of protein [%] in fresh sweet pepper fruits protected with natural preparations in 2010–2012**

Experimental combination	Protein ( $\pm$ U)		
	2010	2011	2012
Control	0.915 A* $\pm$ 0.083	0.940 BC* $\pm$ 0.085	1.140 B* $\pm$ 0.103
Amistar 250 SC	0.876 AB $\pm$ 0.0794	0.889 C $\pm$ 0.08	1.460 A $\pm$ 0.132
Bioczos Płynny	0.795 AB $\pm$ 0.072	1.017 AB $\pm$ 0.092	1.140 B $\pm$ 0.103
Biosept 33 SL	0.633 C $\pm$ 0.057	1.071 A $\pm$ 0.097	0.990 B $\pm$ 0.089
Bio-algeen S90 Plus	0,893 A $\pm$ 0.081	0.990 ABC $\pm$ 0.089	1.110 B $\pm$ 0.101
Boni Protect Forte	0.807 AB $\pm$ 0.073	0.980 ABC $\pm$ 0.088	1.150 B $\pm$ 0.104
LSD 0.05	0.1021	0.1083	0.2807

\*Values followed by the same letters within a column are not significantly different at the 0.05 level probability according to the Tukey test.

The highest content of protein was found in 2012 and the lowest one in 2010. The content of nutrients, as well as proteins, depends on genetic factors and agricultural practices (e.g. pesticides applications) [1]. The investigations on the effect of biological and biotechnical preparations on the contents of mineral and biologically active compounds in pepper fruits were conducted by only few researchers [8, 10, 15]. Among others, Jamiółkowska [15] stated that the application of garlic pulp (Bioczos Płynny), grapefruit extract (Biosept 33 SL), sea algae extract (Bio-algeen Plus) and Boni Protect Forte (*Aureobasidium pullulans*) did not have a significant effect on the content of vitamin C in fresh fruits. The presented results indicated small differences in protein content between the fruits from control combinations and those protected with natural preparations (Tab. 2). The mean values show the lack of significant differences in the content of protein between the combinations of the experiment (Fig. 1).



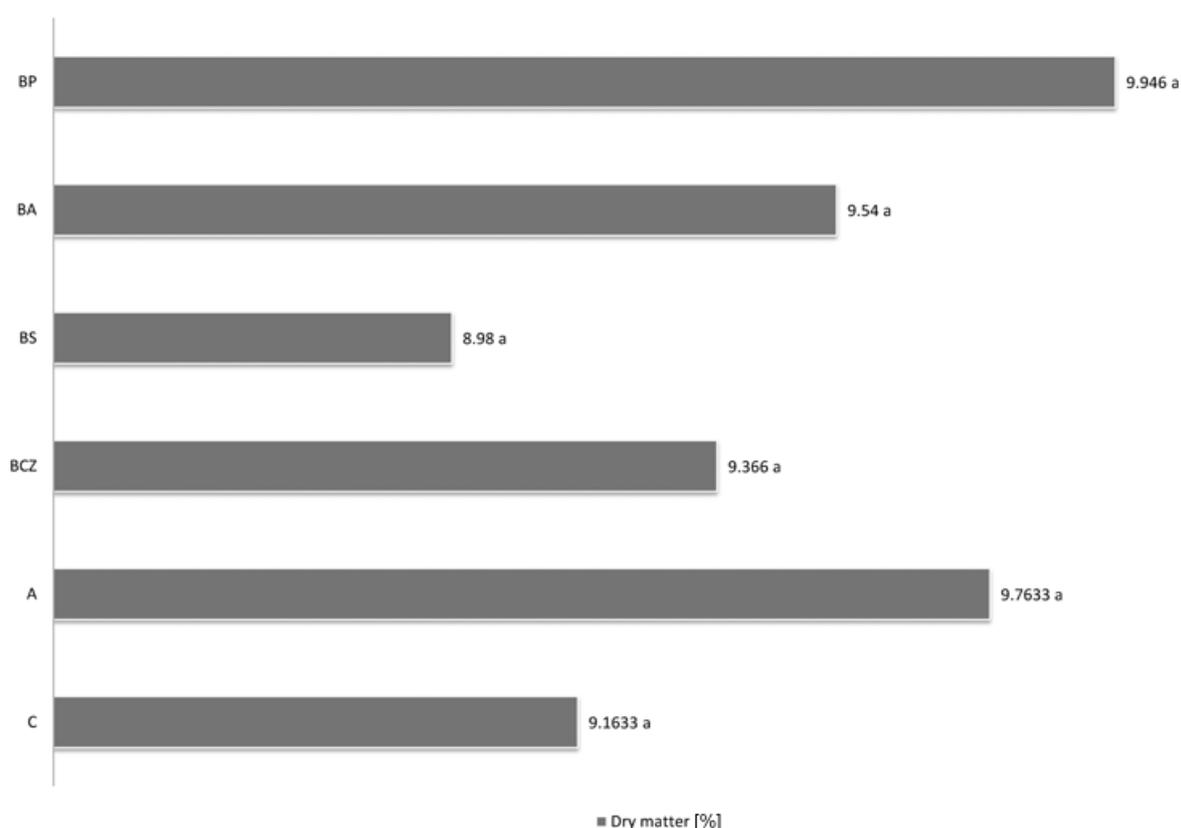
**Fig. 1. Mean content of protein [%] in fruits of sweet pepper protected with natural preparations in 2010–2012 (abbreviations as in Tab. 1), \*values followed by the same letters within a column are not significantly different at the 0.05 level probability according to the Tukey test, LSD 0.05 = 0.5606**

The content of dry matter in pepper fruits depends mainly on the cultivar and the conditions of cultivation [4]. In this study the lowest content of dry matter was observed in 2011 in the combination with Biosept and the highest one in 2012 in the combination with the fungicide (Tab. 3). The mean content of dry matter in investigated fruits ranged from 8.98% (Biosept) to 9.946% (Boni Protect Forte) but did not differ significantly (Fig. 2).

**Table 3. Content of dry matter [%] in fresh fruits of sweet pepper protected with natural preparations in 2010–2012**

Experimental combination	Dry matter ( $\pm$ U)		
	2010	2011	2012
Control	9.230 B* $\pm$ 0.151	8.900 BC* $\pm$ 0.146	9.360 D* $\pm$ 0.153
Amistar 250 SC	9.230 B $\pm$ 0.151	8.590 BC $\pm$ 0.141	11.470 A $\pm$ 0.188
Bioczys Płynny	8.970 D $\pm$ 0.146	9.320 AB $\pm$ 0.153	9.810 C $\pm$ 0.161
Biosept 33 SL	8.950 D $\pm$ 0.146	8.780 BC $\pm$ 0.144	9.210 D $\pm$ 0.151
Bio-algeen S90 Plus	9.090 C $\pm$ 0.149	8.720 C $\pm$ 0.143	10.810 B $\pm$ 0.177
Boni Protect Forte	9.880 A $\pm$ 0.162	9.880 A $\pm$ 0.162	10.080 C $\pm$ 0.165
LSD 0.05	0.0800	0.7875	0.4213

Note as Table 2



**Fig. 2. Mean content of dry matter [%] in fresh fruits of sweet pepper protected with natural preparations in 2010–2012 (abbreviations as in Tab. 1), \* values followed by the same letters within a column are not significantly different at the 0.05 level probability according to the Tukey test, LSD 0.05 = 2.4021**

Buczowska and Michałójc [4] report that the content of dry matter in the fruits of Red Knight F<sub>1</sub> cultivated in the field was significantly lower than in the fruits of this cultivar cultivated in the greenhouse. In the investigations presented by Gajc-Wolska et al. [10] the application of Göteo Goemar and BM 86 (extract of sea algae) caused the increase of dry matter, vitamin C, potassium and calcium contents in treated pepper in comparison to the control plants. Also Dobromińska and Gubarewicz [8] showed a beneficial effect of Bioalgeen on the content of dry matter and vitamin C in the fruits of cherry tomatoes. The number of treatments had also a significant effect on the content of dry matter in tomato fruits.

In the presented study none of the applied natural preparations had a significant effect on the content of dry matter in pepper fruits (Fig. 2).

## CONCLUSIONS

1. The application of natural preparations (Biosept 33 SL, Bioczys Płynny, Bio-algeen S Plus, Boni Protect Forte) in the cultivation of sweet pepper does not have a significant effect on the content of protein in fresh fruits.
2. The preparations of natural origin (Biosept 33 SL, Bioczys Płynny, Bio-algeen S Plus, Boni Protect Forte) used in the

cultivation of sweet pepper does not have a significant effect on the content of dry matter in fresh fruits.

3. The tested natural preparations does not have a negative effect on the nutritional value of pepper fruits, therefore they can be used in plant protection.

## REFERENCES

1. Bosland P.W., Votava E.J., 2000. Peppers: Vegetable and Spice Capsicums. CABI Publishing. London, UK, 204 pp.
2. Bubicz M., Perucka I., Materska M., 1999. Content of bioelements of hot, sweet pepper fruit (*Capsicum annuum* L.). *Biul. Magnezol.*, 4, 2, 289–292.
3. Buczkowska H., Najda A., 2002. A comparison of some chemical compounds in the fruit of sweet, hot pepper (*Capsicum annuum* L.). *Folia Hort.*, 14, 2, 59–67.
4. Buczkowska H., Michałojć Z., 2012. Comparison of qualitative traits, biological value, chemical compounds of sweet pepper fruit. *J. Elem.*, 17, 3, 367–377.
5. Cardoso P.C., Tomazini A.P.B., Stringheta P.C., Ribeiro S.M.R., Pinheiro-Sant’Ana H.M., 2011. Vitamin C and carotenoids and conventional fruits grown in Brazil. *Food Chemistry*, 126, 411–416.
6. Council Directive 2009/128/EC of the European Parliament establishing a framework for Community action to achieve the sustainable use of pesticides. *Official Journal of the European Union* L 309/75 from 24.11.2009.
7. Dłużniewska J., 2004. Activity of biopreparations for controlling fungous diseases of rose shoots. *Prog. Plant Prot.*, 44, 2, 648–650.
8. Dobromińska R., Gubarewicz K., 2008. Influence of Bio-algeen S-90 on the field and quality of small-size tomato. *Biostimulators in modern agriculture: Solanaceous Crops*. Dąbrowski Z.T. (ed.). Editorial House Wieś Jutra, Warszawa, 8–13.
9. Gajc-Wolska J., Skapski H., 2002. Yield of field grown sweet pepper depending on cultivars, growing conditions. *Folia Hort.*, 14, 1, 95–103.
10. Gajc-Wolska J., Zielony T., Łyszkowska M., 2007. The effect of Göteo, BM86 on yield, fruit quality of sweet pepper (*Capsicum annuum* L.) in the field production. *Progress in Research on Capsicum & Eggplant*. Niemirowicz-Szczyt K. (ed.). Warsaw University of Life Sciences Press, 267–274.
11. Howard L.R., Talcott S.T., Brenes C.H., Villalon B., 2000. Changes in phytochemical and antioxidant activity of selected pepper cultivars (*Capsicum* species) as influenced by maturity. *J. Agric. Food Chem.*, 48, 5, 1713–1720.
12. Jadcak P., Grzeszczuk M., 2004. Content of mineral compounds in fruit some cultivars of hot, sweet pepper (*Capsicum annuum* L.). *J. Elementol.*, 9, 1, 15–23.
13. Jadcak P., Grzeszczuk M., Kosecka D., 2010. Quality characteristics, content of mineral components in fruit of some cultivars of sweet pepper (*Capsicum annuum* L.). *J. Elementol.*, 15, 3, 509–515.
14. Jamiolkowska A., 2009. The influence of bio-preparation Biosept 33 SL on fungi colonizing of sweet pepper plants (*Capsicum annuum* L.) cultivated in the field. *EJPAU*, vol. 12, 3. (<http://www.ejpau.media.pl/volume12/issue3/art-13.html>).
15. Jamiolkowska A., 2013. Preparaty biotechniczne i biologiczne w ochronie papryki słodkiej (*Capsicum annuum* L.) przed grzybami chorobotwórczymi i indukowaniu reakcji obronnych roślin [Biotechnical and biological preparations in the protection of sweet pepper (*Capsicum annuum* L.) against pathogenic fungi and in the induction of plant resistance mechanisms]. *Rozprawy Naukowe UP w Lublinie Monogr.* 379 pp. [In Polish].
16. Kmiecik W., Lisiewska Z., 1994. Evaluation of eight sweet pepper cultivars for field growing in the Kraków region from the aspect of requirements of the canning industry. *Folia Hort.*, 6, 2, 35–43.
17. Lee S.K., Kader A.A., 2000. Preharvest and postharvest factors influencing vitamin C content of horticultural crops. *Postharvest Biol. Tec.*, 20, 207–220.
18. Marin A., Ferreres F., Tomas-Barberan F.A., Gil M.I., 2004. Characterization and quantitation of antioxidant constituents of sweet pepper (*Capsicum annuum* L.). *J. Agric. Food Chem.*, 16, 52, 12, 3861–3869.
19. Michalik Ł., 2010. The effect of non-woven PP fabric covers on the yielding, the fruit quality of field-grown sweet peppers. *Acta Sci. Pol., Hort. Cult.*, 9, 4, 25–32.
20. Navarro J.M., Flores P., Garrido C., Martinez V., 2006. Changes in the contents of antioxidant compounds in pepper fruits at different ripening stages, as affected by salinity. *Food Chemistry*, 96, 66–73.
21. Orlikowski L.B., Skrzypczak Cz., 2003. Biocides in the control of soil-borne and leaf pathogens. *Hortic. Veget. Grow.*, 22, 426–433.
22. Orłowski M., Grzeszczuk M., Jadcak D., 2004. The estimation of the yield and content of some chemical compounds in the fruits of chosen hot pepper (*Capsicum annuum* L.) cultivars. *Folia Hort.*, 16, 2, 11–16.
23. Perez-Lopez A.J., Moises del Amor F., Serrano-Martinez A., Fortea M.I., Nunez-Delgado E., 2007. Influence of agricultural practices on the quality of sweet pepper fruits as affected by the maturity stage. *J. Sci. Food Agric.*, 87, 2075–2080.
24. Perucka I., Oleszek W., 2000. Extraction and determination of capsaicinoids in fruit of hot pepper *Capsicum annuum* L. by spectrophotometry and high-performance liquid chromatography. *Food Chem.*, 71, 287–291.
25. Perucka I., Materska M., 2003. Antioxidant activity and contents of capsaicinoids isolated from paprika fruits. *Pol. J. Food Nutr. Sci.*, 12, 53, 2, 15–18.
26. Perucka I., Materska M., 2004. Wpływ Ca<sup>2+</sup> na zawartość witaminy C, prowitaminy A i ksantofili w owocach wybranych odmian papryki ostrej [Influence of Ca<sup>2+</sup> on the vitamin C, provitamin A, xanthophyll contents in fruits of some hot pepper cultivars]. *Annales UMCS, Sec. E*, 59, 4, 1933–1939 [In Polish].
27. Perucka I., Materska M., 2007. Antioxidant vitamin contents of *Capsicum annuum* fruit extracts as affected by processing and varietal factors. *Acta Sci. Pol., Technol. Aliment.*, 6, 4, 67–74.
28. Regulation (EC) No 1107/2009 of the European Parliament and of the Council of 21 October 2009 concerning the placing of plant protection products on the market. *Official Journal of the Europe* 09/1 from 24.11.2009.
29. Schuphan W., 1948. *Gemüsebau auf ernährungswissenschaftlicher Grundlage*. Hamburg, 368 pp. [In German].
30. Simonovska J., Rafajlovska V., Kavrovski Z., Srbinska M., 2014. Nutritional and bioactive compounds in hot fruits of *Capsicum annuum* L. from Macedonia. *Macedonian Journal of Chemistry and Chemical Engineering*, 33, 1, 97–104.
31. Somos A., 1984. The paprika. *Academiai Kiado*. Budapest. 291 pp.
32. Topuz A., Ozdemir F., 2007. Assessment of carotenoids, capsaicinoids and ascorbic acid (*Capsicum annuum* L.) grown in Turkey. *J. Food Compos. Analysis*, 20, 596–602.
33. Wilson C.L., Salar J.M., El Ghaouth A., Wisniewski M.E., 1997. Rapid evaluation of plant extracts and essential oils for antifungal activity against *Botrytis cinerea*. *Plant Dis.*, 81, 2, 204–210.
34. Zydlik P. 2008. Wykorzystanie preparatów pochodzenia naturalnego w zwalczaniu niektórych chorób roślin sadowniczych [Use of natural preparations against some diseases of fruit-growing plant]. *Nauka Przyr. Technol.*, 2, 1, 3 [In Polish].

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