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# ISOLATION OF AN OPHIDIAN PARAMYXOVIRUS (OPMV) IN A CAPTIVE RATTLESNAKE (*Crotalus durissus* *terrificus*) FROM BOTUCATU, SÃO PAULO STATE, BRAZIL

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**ABSTRACT:** This study reports the isolation of an Ophidian Paramyxovirus (OPMV) in sputum of a captive rattlesnake (*Crotalus durissus terrificus*) kept in a serpentarium located in Botucatu, São Paulo State, Brazil. Polymerase chain reaction (PCR) and nested-PCR were performed for the identification of the isolated virus.  
**KEY WORDS:** *Crotalus durissus terrificus*, rattlesnake, Paramyxovirus, Ophidian Paramyxovirus, hemagglutination, polymerase chain reaction.

## INTRODUCTION

Captive snake breeding has become an important activity; first to obtain venom for antiserum production, and second to study venom pharmacological properties (6,14,15).

Infectious diseases are the major problems affecting captive snakes (2,10,11,12), especially paramyxozirosis, the agent being known as ophidian paramyxovirus (OPMV) (3,5,7,8,9). OPMV causes respiratory and neurological disorders in snakes, resulting in up to 100% mortality (4,13).

This study reports the isolation of an OPMV from the sputum of a captive rattlesnake (*Crotalus durissus terrificus*) with signs of pneumonia. The snake was kept in a serpentarium located in Botucatu, São Paulo State, Brazil.

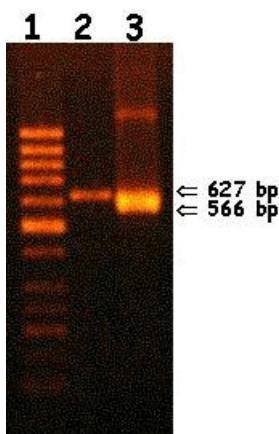
The sputum sample was inoculated in a monolayer of Vero cells and incubated under agitation for 6 days at 30°C. A second passage was made under the same conditions. The pure secretion and cell culture supernatants were submitted to hemagglutination tests with positive results.

Polymerase chain reaction (PCR) and nested-PCR were performed with cell culture samples for the identification of the isolated virus, using the technique described by Ahne *et al.* (1) with modifications. RNA was extracted from cell culture supernatants using TRIZOL LS (Gibco BRL) as recommended by the manufacturer. cDNA was obtained by reverse transcription using Superscript RTII (Gibco BRL). In PCR and nested-PCR, the primers used were those for a partial sequence of polymerase (L) gene (1) in a reaction with a final volume of 25 µl containing: 2.5 units Taq DNA Polymerase (Gibco BRL); 2.5 µl PCR Buffer 10X; 10 pmols of each primer; 1.5 mM MgCl<sub>2</sub>; 0.2 mM dNTP; and 5 µl of cDNA (PCR) or 2 µl of the PCR product (Nested-PCR). Conditions for amplification are shown in [Table 1](#).

**Table 1.** PCR and Nested-PCR conditions for amplification of polymerase (L) gene partial sequence.

| Steps | Temperature (°C) | Duration | Nº of cycles |
|-------|------------------|----------|--------------|
| 1     | 94               | 3'       | 1            |
| 2     | 94               | 1'       | 9            |
| 3     | 52 (-1°/cycle)   | 1'       | 9            |
| 4     | 72               | 1'30"    | 9            |
| 5     | 94               | 1'       | 25           |
| 6     | 44               | 1'       | 25           |
| 7     | 72               | 1'30"    | 25           |
| 8     | 72               | 4'       | 1            |

The positive results from hemagglutination, PCR, and nested-PCR ([Figure 1](#)) are effective to confirm that this virus is a Paramyxovirus. This is the first report about Ophidian Paramyxovirus (OPMV) in Brazil.



**Figure 1.** 627 bp (line 2) and 566 bp (line 3) bands from PCR and nested-PCR, respectively, compared with a ladder of 123 bp (line 1) of the first passage of sputum sample cell culture.

## ACKNOWLEDGEMENTS

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## REFERENCES

- 1 AHNE W., BATTI WN., KURATH G., WINTON JR. Comparative sequence analyses of sixteen reptilian paramyxoviruses. *Virus Res.*, 1999, 63, 65-74. [ [Links](#) ]
- 2 BELLUOMINI HE., SALIBA AM., ABE AS. Inquérito anatomo-patológico em serpentes dos gêneros *Crotalus* e *Bothrops* [Serpentes, Viperidae, Crotalinae]. *Mem. Inst. Butantan*, 1976/77, 40/41, 123-8. [ [Links](#) ]
- 3 CLARK HF., LIEF FS., LUNGER PD., WATER D., LELOUP P., FOELSCH DW., WYLER RW. Fer-de-lance virus (FDLV): a probable Paramyxovirus isolated from a reptile. *J. Gen. Virol.*, 1979, 44, 405-18. [ [Links](#) ]
- 4 CRANFIELD MR., GRACZYK TK. Ophidian Paramyxovirus. In: MADER MR. *Reptile Medicine and Surgery*. Philadelphia: W. B. Saunders Co., 1996: 392-4. [ [Links](#) ]
- 5 FOELSCH DW., LELOUP P. Fatale endemische infektion in einem serpentarium. *Tierarztl. Prax.*, 1976, 4, 527-36. [ [Links](#) ]
- 6 GIORGI R., BERNARDI MM., CURY Y. Analgesic effect evoked by low molecular weight substances extracted from *Crotalus durissus terrificus* venom. *Toxicon*, 1993, 31, 1257-65. [ [Links](#) ]
- 7 JACOBSON ER. Implications of infectious diseases for captive propagation and introduction programs of threatened/endangered reptiles. *J. Zoo Wildl. Med.*, 1993, 24, 245-55. [ [Links](#) ]
- 8 JACOBSON E., GASKIN JM., SIMPSON CF., TERRELL TG. Paramyxo-like virus infection in a rock rattlesnake. *J. Am. Vet. Med. Assoc.*, 1980, 177, 796-9. [ [Links](#) ]

9 JACOBSON ER., GASKIN JM., WELLS S., BOWLER BS., SCHUMACHER J. Epizootic of ophidian paramyxovirus in a zoological collection: pathological, microbiological, and serological findings. *J. Zoo Wildl. Med.*, 1992, 23, 318-27. [ [Links](#) ]

10 KANEENE JB., TAYLOR RF., SIKARSKIE JG., MEYER TJ., RICHTER NA. Disease patterns in the Detroit Zoo: a study of reptilian and amphibian populations from 1973 through 1983. *J. Am. Vet. Med. Assoc.*, 1985, 187, 1132-3. [ [Links](#) ]

11 LEINZ FF., JANEIRO-CINQUINI TRF., ISHIZUKA MM., LANG LV. Sobrevivência de *Bothrops jararacussu* (Serpentes, Viperidae, Crotalinae) mantidas em cativeiro. *Mem. Inst. Butantan*, 1989, 51, 33-8. [ [Links](#) ]

12 MARCUS LC. Infectious Diseases of Reptiles. *J. Am. Vet. Med. Assoc.*, 1971, 159, 1626-31. [ [Links](#) ]

13 MURRAY MJ. Pneumonia and Normal Respiratory Function. In: MADER MR. *Reptile Medicine and Surgery*. Philadelphia: W. B. Saunders Co., 1996: 396-405. [ [Links](#) ]

14 SARTORI FILHO R., PRESTES NC., THOMAZINI IA., MENDES-GIANNINI MJ., TOSCANO E., CANAVESSI AMO., BARRAVIEIRA B. Use of fibrin glue derived from snake venom in testicular biopsy of rams. *J. Venom. Anim. Toxins*, 1998, 4, 23-35.

15 SILVA RJ., FECCHIO D., BARRAVIEIRA B. Effect of *Crotalus durissus terrificus* (LAURENTI, 1768) venom on the evolution of Ehrlich ascites tumor. *J. Venom. Anim. Toxins.*, 1997, 3, 324-41.

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