

## Review

# Arsenic-induced Cancers: A Review with Special Reference to Gene, Environment and Their Interaction

Mayukh Banerjee, Pritha Bhattacharjee and Ashok K. Giri<sup>1</sup>

Molecular and Human Genetics Division, Indian Institute of Chemical Biology (A Unit of Council of Scientific and Industrial Research, New Delhi), Kolkata, India

(Received August 29, 2011; Revised October 10, 2011; Accepted October 11, 2011)

Chronic exposure to the toxic metalloid arsenic, either through occupational or environmental route results in the development of a plethora of dermatological as well as non-dermatological health effects, multi-organ cancers being the most severe ones. Several epidemiological studies have reported the occurrence of different types of cancers in populations exposed chronically to arsenic from different corners of the globe, but the spectrum of symptoms and severity vary widely. Several genetic and environmental factors might play an important causal role for such observed differences, including genetic polymorphisms, dietary habits and socio-economic status of the population. Unfortunately, the paradoxical nature of this carcinogen makes the use of animal model systems unsuitable, since they do not reflect the exact *in vivo* condition of arsenic exposure in human beings. Hence, in this present review, we have attempted to collate the isolated pieces of information available in existing literature depicting the various forms of arsenic-induced cancers, variations and etiology thereof. We have attempted to depict the entire spectrum of carcinogenic outcomes of chronic arsenic exposure and the interactions among several components like, genes (polymorphisms and mutations), environmental factors and dietary factors in the process. The literature reviews are primarily based on the human beings (*in vivo*), but to understand the underlying mechanism, we have also collated the information based on cell line models, and animal models where human data is scanty or unavailable. Thus, here we present a comprehensive review on the current state of research in the field of arsenic-induced cancer and explore the underlying mechanism of arsenic-induced carcinogenicity, along with relevant historical perspective.

**Key words:** arsenic, diet, gene-environment interaction, cancer

## Introduction

Arsenic has been used by mankind since ages immemorial both for constructive as well as destructive purposes. Besides its widespread use in traditional medicine, arsenic has often been the poison of choice for silent killing in case of several cases of royal feuds.

However, in the present days, it is the chronic form of arsenic toxicity, rather than the willful acute poisoning, which plagues the world. Occurrences of chronic arsenic toxicity from different sources have been reported from 70 countries spanning all the 5 continents and renders an imponderable 150 million individuals at risk (1,2). In fact, arsenic has been assigned the status of group I carcinogen by IARC (3) in human beings and ranks 1 on the ATSDR-CERCA priority list of hazardous substances (4).

Chronic exposure to arsenic for long time periods can occur through both occupational as well as environmental routes. In many instances, chronic exposure to this deadly toxicant is a result of working (or from industrial wastes) in mining and smelting industries (5–9). Occupational exposure to arsenic can also occur from working at or near several other kinds of factories where arsenic is used as a raw material or generated as a sludge or waste as in glass factories (10,11), semiconductor factories (12–17), power plants (18,19) and so on. This may also stem from being in constant touch with arsenical pesticides (20–26). However, in majority of the cases, the exposure is environmental in origin, either through drinking of groundwater contaminated from geogenic sources (27–38) or burning of arsenic rich coal (39,40).

Whatever be the source of chronic exposure to arsenic, the results are nearly always detrimental in the extreme. Chronic arsenic exposure leads to the development of classical skin lesions like raindrop pigmentation, palmo-plantar hyperkeratosis, hypopigmentation. In addition, arsenic exposure has also been proved to give rise to a plethora of non-dermatological disease outcomes including peripheral neuropathy, conjunctivitis, respiratory problems, liver enlargements and opportunistic infections (41–45). However, the most dreaded outcome of chronic arsenic exposure are undoubtedly

<sup>1</sup>Correspondence to: Ashok K. Giri, Molecular and Human Genetics Division, Indian Institute of Chemical Biology, 4 Raja S. C. Mullick Road, Jadavpur Kolkata – 700 032, India. Tel: +91-33-2499-5724, Fax: +91-33-2473-5197, E-mail: giri@csiriicb.in; ak giri15@yahoo.com

the multi-organ cancer induced by this toxicant. Since the first reports associating arsenic exposure with cancerous outcomes in 1887 (46), numerous epidemiological studies have proved beyond doubt that chronic arsenic exposure (be it environmental or occupational) gives rise to a wide variety of cancers, including those of skin and also of internal organs like lungs, liver, urinary bladder and kidney. Interestingly, arsenic is regarded as a “paradoxical carcinogen” because of its ability to induce cancer in human beings on its own, but not in any other animal model. This makes the study of arsenic-induced carcinogenesis particularly difficult and cumbersome. Most of the data regarding arsenic-induced carcinogenesis are derived from epidemiological studies. Although there are a few reviews reflecting the role of arsenic in specific cancers, there is none to our knowledge, which looks at arsenic exposure with respect to the development of cancers as a whole (including both internal and external cancers). Also, in spite of multifarious studies being performed to elucidate the role of several genes and environmental factors in arsenic susceptibility and carcinogenicity, there has been no adding up of all those work yet which could lead to the spelling out of meaningful conclusions with respect to carcinogenicity of arsenic. Hence, in the present text, we aim to present data from a wide variety of studies to show that arsenic and its metabolites induce a wide variety of cancers in human beings and then to highlight on the role of gene-environment interactions, especially diet and methylation in arsenic-induced cancers.

### Arsenic-induced Skin Cancers

Skin is the major site of arsenic toxicity. Prolonged exposure to arsenic manifests itself in the form of several well characterized skin lesions including both non-malignant and malignant forms. These different types of skin cancers, ranging from Bowen's disease to highly metastasizing squamous cell carcinoma, are however, not exclusive to arsenic exposure and may occur under several conditions even in absence of arsenic exposure. In the present section we will review the different types of skin cancers induced by arsenic exposure.

**Bowen's disease:** Bowen's disease (BD), first described by the American dermatologist Dr. J. T. Bowen in 1912, is an early and mild form of *in situ* carcinoma. It is characterized by reddish undulated skin patches with irregular edges and distinct demarcation lines and usually occurs in sun covered areas in case of arsenic exposure. The epidermis exhibits a striking disorganization, with loss of normal cell polarity and disruption of normal progressive maturation of the epidermal cells and vacuolization of cells are common (47). That arsenic ingestion might lead to BD was suggested as early as 1961 by Graham *et al.* (48). It had also been reported by several other groups such as Imamura *et al* (49) and Yeh

*et al.* (50,51). Arsenic-induced BD has been reported sporadically from several populations including Japanese (49,52-55), Japanese-American population in Hawaii (56), Thai (57), Taiwan (58,59), Finlandish (60), Turkey (61,62), Korea (63,64), West Bengal, India (65-67), Nepal (68) and South Africa (69).

At the genetic level, arsenic-induced BD (As-BD) is characterized by high levels of genomic instability like most other cancers and exhibit enhanced induction of both chromosomal aberrations (66) as well as micronuclei (67) and also significantly higher sister chromatid exchange and replicative index (70). Interestingly, most of the chromosomal aberrations induced are of chromosome type, i.e., involves both strands of a chromosome, whereas, the incidence of chromatid type of aberrations are similar to that of arsenic-induced non-cancerous skin lesion patients (66). Chromosome type aberrations are extremely potent in activating the protooncogenes and/or suppression of tumor suppressor genes and thus are harbingers of carcinogenic outcomes, and the similar findings are encountered in case of As-BD patients. This point is further illustrated by the fact that considerable number of As-BD patients has a wide range of unique *p53* mutations (in target tissues), mostly in exons 5 and 8 (71). Generation of reactive oxygen species (ROS) is expected to play a significant role in the etiology of As-BD as seen from the elevated levels of 8-hydroxy-2'-deoxyguanosine in the bowen's tissue.

As-BD is different from non-arsenical BD in having unique histology. Studies using immunostaining technique reveal that Langerhans cells are significantly decreased in As-BD along with loss of dendrites compared to adjacent normal skin from the same patients (72,73). Again keratin 16 (marker for hyperproliferation of skin cells) was also found to be highly expressed in the basal and suprabasal cell layers of Bowen's tissue of As-BD patients, but not in case of non-arsenic BD patients (74). Keratin 6 expression was also found to be altered in case of As-BD (75). Additionally, at the molecular level, pathogenesis of As-BD was also found to be different from non-arsenical BD in terms of *p53* expression. Enhanced *p53* expression was found to be necessary for proliferative potential in case of As-BD (59). Also, the early dysplastic cells of As-BD showed overexpression of Bcl2 compared to normal skin tissue (76). Immunological characterization of As-BD shows that there might be an enhanced recruitment of helper T cells to the site of tumor (73). It is commonly observed that As-BD patients exhibit both cutaneous and systemic immune dysfunctions (77). Studies revealed that As-BD patients had significantly elevated CD4<sup>+</sup> cells in the cancerous tissues but significantly decreased in the lymphocytes, along with concomitant induction of Fas expression in CD4<sup>+</sup> cells (77). Furthermore, As-BD patients were found to have subnormal cell-mediated

immunity, due to impairment of IL2 receptor expression in lymphocytes via the tumor necrosis factor receptor 1 signaling pathway (78,79).

**Basal cell carcinoma:** Besides being the most common of all human cancers, basal cell carcinoma (BCC) is also a very common outcome of chronic arsenic exposure. Sporadic BCC occurs predominantly in sun exposed areas of the body, whereas, arsenic-induced BCC is found mostly in the sun covered trunk parts (80) and are usually multiple in number (81). Although, under normal circumstances, fair skinned individuals are more prone to develop BCC, incidences of arsenic-induced BCC have been reported from several countries of the world, encompassing both fair skinned and dark skinned populations. Development of multiple BCC has been reported in arsenic exposed individuals from Canada (82), Oregon, USA (83), Spain (84), China (73), Taiwan (73), Singapore (76,85,86), Australia (87,88), West Bengal, India (45,89), New Hampshire, USA (90), Argentina (91) etc.

BCC is characterized by tumor of basaloma cells of epithelial origin and is non-invasive in nature. Histologically, arsenic-induced BCC and BCC of other origin are similar and consist of transformed basaloma cells with large nucleus and poorly differentiated cytoplasm and are usually associated with elongation of epidermis into the dermal layer (80). This is also associated with decrease in the number of Langerhans cells and disruption of the Langerhans cell dendrites (73). Biochemical analyses showed that at the molecular level, BCC is characterized by a heightened expression of cytokeratins K6, K16 and K17 and a total absence of K1 (75). As in the case of As-BD, BCC also showed elevated expression of p53 and Bcl2. Additionally, Ki-67 was also found to be overexpressed in case of BCC (71,76) compared to normal skin tissue. However, in a study by Castren *et al.* (60) it was shown that mutations in p53 are not required for this overexpression in case of BCC. Contradictory results were obtained in another study from Taiwan; it was found that almost 30% of the BCC tissues had p53 mutations, mostly in exons 5 and 8, and these mutations were different from those encountered in case of UV-related skin cancers (71). Interestingly, the level of p53 expression was significantly lower in arsenic-induced BCC tissue when compared to BCC tissue of non-arsenical origin, perhaps due to hypermethylation (87). The risk of development of arsenic-induced BCC has been shown to be genetically modulated as well. In a study on arsenic-exposed population from Europe, XRCC3 codon 241 T variant was proved to confer elevated protection against the development of BCC, whereas, NBS1 codon 185 CC genotype imparted heightened risk of BCC development in men only (92). In another study, the risk of development of BCC was found to be depressed if an individual had at least one A

allele at codon 23 of XPA gene and at least one Asp allele at codon 312 of XPD gene (93).

**Squamous cell carcinoma:** Squamous cell carcinoma (SCC) is the severest form of skin cancer caused by chronic arsenic ingestion. It is the only invasive form of skin cancer brought about by arsenic exposure which can metastasize and have lethal outcomes. Arsenic-induced SCC can arise *de novo* or from pre-existing BD, and usually develop at the extremities of the body. Unlike sporadic SCC, arsenic-induced SCC often develops at sun protected areas of the body. That prolonged arsenic exposure could lead to the development of SCC was speculated as early as 1948 (94). Since then, occurrences of SCC in cases of chronic arsenic exposure has been reported from several areas of the globe including Canada (82), Oregon, USA (83), Sweden (95), Taiwan (71,75,96), Singapore (76,85,86), Turkey (61), France (97), China (98), West Bengal, India (45), New Hampshire, USA (93), etc. SCC usually appears as small papule at the early stage which, in the later stages might often become crusty and ulcerated (99). It is basically a malignancy of the surface epidermal cells, which gets transformed into poorly differentiated elongated mass of epidermal cells, proliferate, invading the underlying dermal layers and at an advanced stage metastasize into other organs. SCC is characterized by considerable decrease in the number of Langerhans cells as also disruption of the Langerhans cell dendrites and an elevated recruitment of T helper cells to the tumor infiltrates (73). Histologically, there is no difference between arsenic-induced SCC and sporadic SCC (100).

At the biochemical level, SCC is characterized by overexpression of cytokeratins K6, K16 and K17 similar to that in BCC. In addition there is a progressive impairment of K1 and K2 phosphorylation, which is thought to play a key role in the carcinogenic outcome (75). Again, like in BCC, there is also overexpression of p53, Bcl2 and Ki67 in case of SCC (76). This result is also supported by the fact that in a study from Taiwan (71), it was shown that 44% of the arsenic-induced SCC tumor samples had novel p53 mutations. However, in another study, contrasting results were obtained and p53 overexpression was shown to be a non-requisite step in the pathogenesis of SCC (59). Also, no p53 mutations were found in SCC tissues from two arsenic-exposed individuals in a study by Castren *et al.* (60), although overexpression of p53 protein was detected, which supports the view that tissue specific p53 mutations are perhaps not important as far as SCC development is concerned. Single nucleotide polymorphisms in the DNA repair pathway genes were however, shown to be associated with conferring protection to SCC development upon arsenic exposure in a New Hampshire population. Individuals with at least one A allele of XPA codon 23 and those with a combination of at least one

Asn allele at codon 312 and at least one Gln allele at codon 751 of XPD were shown to have significantly lesser risk of development of arsenic-induced SCC compared to those having the wild genotypes (93). The process of SCC development upon chronic arsenic exposure may involve the activation of Nrf2 and subsequent up-regulation of MMP-9, keratin-1, keratin-9, involucrin and loricrin expression, as shown by studies on arsenic transformed HaCaT cell line and animal studies (101). Aberrant expression of several cytokeratin genes, including CK1, CK13 and CK15 were also found to be operative during malignant transformation of HaCaT cell line by chronic arsenic treatment (102).

### **Arsenic-induced Non-dermatological Cancers**

Although skin lesions are considered to be the hallmarks of arsenic toxicity, yet, the fact that chronic exposure to this deadly toxicant leads to the development of several varieties of internal cancers is now widely accepted. Several epidemiological studies over the last fifty years and more have repeatedly linked chronic arsenic exposure to malignancies of internal organs, especially lungs, liver and bladder. This section of the review will summarize the literature exploring the relationship between arsenic exposure and non-dermatological cancerous outcomes.

**Lung cancer:** Although arsenic exposure has long been speculated to be causal to lung cancer, debate regarding its actual role and importance in etiology had been a long lasting one (103). By the middle of the 1970s, a plethora of epidemiological studies were conducted in different parts of the world, which yielded unequivocal results and established firmly the causal role of occupational arsenic exposure towards the development of lung cancer and elevated mortality thereof (104–106). Similar results were obtained from other subsequent studies (107–119). Furthermore, it has been demonstrated that increased lung cancer risk due to chronic arsenic exposure is independent of the exposure route; exposure via ingestion and inhalation leading to similar increment in lung cancer predisposition in the population (120). Several studies have explored the potential synergistic/interactive effect of smoking and arsenic exposure towards induction of lung cancer. Most of these studies have shown that smokers, exposed chronically to arsenic have a very high risk of developing lung cancer (108,113,121–125). Additionally, it has also been demonstrated that smoking can affect lung cancer propensity in arsenic exposure status, depending upon the GST gene polymorphic status of the individual (125). Chance of development of lung cancer is further increased to a great extent by early childhood or *in utero* exposure to inorganic arsenic (126).

The role of arsenic exposure in lung cancer etiology is not clearly understood. Some evidences indicate that the

length of exposure, rather than the intensity might be more important in lung carcinogenesis (107) and that arsenic might play an important role in the later stages of lung cancer development, although, a possibility of its involvement in the early stages was not ruled out (127). The basic mechanism by which arsenic induces carcinogenic outcomes in lung cells seems to be via the generation of reactive oxygen species (ROS). Studies utilizing cultured lung epithelial cells showed significant increase ROS biomarkers. There was also a significant increment in the expression of gamma-glutamylcysteine synthetase ( $\gamma$ -GCS) on arsenic exposure with concomitant increase in the expression of c-fos, c-jun and in DNA binding and transactivation efficacy of AP-1, perhaps in a NF- $\kappa$ B dependant manner (128). The validity of these observations are further strengthened by a later study which demonstrated that cultured human bronchial epithelial cells, BEAS-2B, when exposed to low concentration sodium arsenite, resulted in enhanced cell growth and cyclin D1 expression, mediated by p52-Bcl3 complex formation (129). This induction of ROS in the cellular milieu is hypothesized to be responsible for DNA damage in the form of DNA strand breaks, and formation of DNA-protein crosslinks as demonstrated by using human fetal lung fibroblast cells (130–131). There are some evidences from rodent model which demonstrate that chronic arsenic exposure might also interfere with the methylation status of certain genes and bring about lung cancer (132), although it is yet to be validated in human system. Recent evidences show that DNA losses at chromosomes 1q21.1, 7p22.3, 9q12, and 19q13.31 are frequently associated with arsenic-induced lung tumors (133).

**Bladder cancer:** Bladder cancer has been associated with chronic arsenic exposure for quite some time. The first report came out as early as 1986, when the results of an epidemiological retrospective study carried out in arsenic endemic region of Taiwan demonstrated that incidence of bladder cancer is significantly associated with ingestion of arsenic laced water (29) and was soon confirmed by another study in 1988 (134). Soon a plethora of studies were conducted in different parts of the world, exploring the relationship between incidence of bladder cancer and chronic arsenic exposure, and the results were unequivocal (111,135–142).

Immunohistopathological studies showed that arsenic-induced bladder tumors were pathobiologically and prognostically identical to non-arsenical bladder tumors (143). At the cytological level, it was found that level of arsenic exposure was directly correlated with the induction of chromosomal instability and that tumors from arsenic exposed individuals might behave much more aggressively than bladder tumors of non-exposed individuals (144,145). There are also some evidences which show that males are more prone to arsenic-in-

duced bladder cancer than females, irrespective of the smoking status (146). Studies on cell line have provided convincing evidence that inorganic arsenic exposure leads to a dose dependant increase of cells in the S phase along with reduction in cell proliferation and hampered DNA replication. This deviation from normal cell cycle was brought about by temporal alterations in the expression of p53 and p21 and their failure to control downstream Cdk2-cyclin E activity (147). It was also shown that Bcl2 and c-fos might be important players in the induction of arsenic-induced bladder cancer, but not for that of non-arsenical bladder cancers (148). Arsenic methylation status of individuals was also found to be associated with the propensity of developing bladder cancers as the ones excreting maximum amount of monomethyl arsenous acid (MMA) were found to be at greatest risk (149). Similar results were obtained from another study which showed that individuals with a high primary methylation index had a higher risk of developing bladder cancer (150). Further studies on human bladder cell lines demonstrated that chronic exposure to MMA leads to hyperproliferation and concomitant reduction in doubling time followed by anchorage independent growth and consequent tumorigenicity (151,152). It was also shown that this MMA-induced transformation was brought about by a complex interplay of several intricately interwoven pathways, including external signal regulated kinase, phosphoinositide 3-kinase, cyclooxygenase 1 and cyclooxygenase 2 (153). Again, level of cytokines IL-1, IL-6 and IL-8 were found to be increased in case of chronic arsenic exposure in the same cell line (UROtsa) and was demonstrated to be important from malignant transformation following exposure (154). All these effects are perhaps brought about by the generation of ROS and subsequent lipid peroxidation, protein carbonylation and oxidative DNA damage by trivalent arsenicals (155). The involvement of ROS in the process was further demonstrated by the strong correlation between bladder cancer status and induction of urinary 8-hydroxydeoxyguanosine (156). Epigenetic mechanisms also play an important part in the etiology of arsenic-induced bladder cancer as the promoter hypermethylation of death associated protein kinase (DAPK) gene was shown to be instrumental in the process (157).

**Liver cancer:** Liver being the seat of arsenic metabolism retains inorganic as also methylated forms of arsenic perhaps for the longest time period inside the body and is thus an ideal target for arsenic-induced carcinogenesis. It has been well documented for a long time that there is a strong association between chronic arsenic ingestion and induction of liver cancer. Literature survey shows that as early as 1968, ingestion of arsenic containing Fowler's solution (used as a medicine) for prolonged periods had been demonstrated to give rise to

hemangioendothelial sarcoma of liver (158). Soon after, it was found out that chronic arsenic exposure (be it from medicinal or environmental) could also give rise to angiosarcoma of the liver (159–162). These sporadic observations necessitated large scale well designed epidemiological studies to link liver cancer incidence with chronic arsenic intake. Several studies were conducted in different parts of the world and bear testimony to this association (29,30,80,109,110,141,163–165).

The mechanisms by which chronic exposure to arsenic brings about malignant transformation in the liver tissue are difficult to study in human beings. Consequently, most of our current ideas are based on experiments carried out in rodent models using unrealistically high doses of arsenic like 85 ppm (166), which are not to be found under normal ambient conditions in the nature. Hence, the results obtained from such studies are subject to close scrutiny as far as carcinogenesis in human liver is concerned. However, a few studies have been carried out in exposed human liver tissues and human liver cell lines, which could be more reliable in elucidating the role of arsenic exposure to such carcinogenic outcomes. In one study, using transcriptomic technique on biopsy samples from liver tissues from arsenic exposed individuals, it was shown that transcription of around 60 genes are aberrantly altered compared to unexposed controls. These include genes encoding for cell-cycle regulation and proliferation, the apoptosis-related genes, the DNA damage related genes, and the genes encoding for cellular regulators and growth factors (167). Perhaps the most potent mechanism involved in such arsenic induced malignant changes in hepatocytes is the generation of oxidative stress and consequent oxidative DNA damage as visualized by studies on HepG2 cell line (168).

**Kidney cancer:** Several epidemiological studies have demonstrated the association of chronic arsenic intake with heightened incidence of kidney cancer. Dose response relationship between induction of kidney cancer in both exposed males and females were shown in Taiwan population in 1989 (30). The results were corroborated from similar studies on chronically exposed population from Japan (54), Taiwan (109,110,164,169), Chile (137) and India (170).

Study regarding the mechanism of arsenic-induced kidney carcinogenesis in human system is almost non-existent and the meager amount of evidence we have is all based on studies in non-human mammalian systems. It has been shown that chronic exposure to arsenic leads to the expression of several stress proteins in the kidney (171) and that mitochondria might be the major target in kidney carcinogenesis owing to the oxidative stress generated by chronic arsenic exposure. It was further shown that human embryonic kidney cell line when treated with low dose of inorganic trivalent arsenic leads

to aberrant alteration in the expression of a plethora of genes including those involved in stress response, proto-oncogene, signaling molecules, transcription factors, chemokine receptors and proteolytic enzymes, leading to complex cellular injury (172).

### Role of Diet in Arsenic Carcinogenicity

Of the several probable environmental factors that might have an impact on arsenic toxicity, susceptibility and carcinogenicity, the role of diet is perhaps the most important. Several studies have probed into different aspects of diet on arsenic toxicity and the results show beyond a trace of doubt that nutritional status of the individual often plays a key role in determining the degree of proneness to the toxic effects of chronic arsenic exposure. Malnutrition was demonstrably associated with a higher prevalence of arsenic-induced skin cancer (173). Again, fasting was shown to have a profound effect on arsenic metabolism and speciation as a significantly higher amount of methylarsonate was found to be excreted in the urine of fasting individuals (174). In fact, the interaction of arsenic exposure and diet can act at two different stages as has been elegantly summarized in a review by Vahter (175); viz., by modulating arsenic-induced oxidative stress and by modulating arsenic metabolism. Epidemiological studies have been conducted which shows that dietary factors, especially those involved in one-carbon metabolism, such as folate, methionine and homocysteine are important modulators of arsenic methylation, toxicity and carcinogenicity (176–178). Presence of these factors in diet helps in alleviating the toxic effects or arsenic exposure. Folate has been shown to increase arsenic-induced genomic DNA methylation in peripheral blood leukocytes in exposed population (179). Interestingly and anomalously, however, in one study, it was demonstrated that the arsenic methylation pattern was normal in pregnant woman from arsenic exposed population in Bangladesh, in spite of the prevalent malnutrition (180).

Generation of reactive oxygen species (ROS) is considered to be one of the most potent mechanisms for arsenic-induced toxicity and carcinogenesis (reviewed in 181,182). Hence, it is expected that a diet rich in antioxidants would be helpful in combating the toxic and carcinogenic effects or chronic arsenic exposure. Several antioxidative agents have been tried out to modulate the toxic effects of arsenic exposure, although, most of the studies have used rodent models. However, only a few studies have been conducted using human cell lines and human subjects and the results show that antioxidative agents are effective in alleviating arsenic toxicity. Vitamin A could alleviate the genotoxic effects of arsenic exposure in peripheral blood lymphocytes (183,184). Ascorbic acid treatment was shown to protect HL66 and U266 cells for arsenic-induced toxicity as measured by

cell viability, colony formation and apoptosis assays (185). Similarly, treatment with plant flavonoids silymarin and quercetin were found to reduce arsenic-induced heat shock protein 70 kDa expression, lipid peroxidation, and gamma-glutamyl-transpeptidase activity, which are the hallmarks of arsenic-induced oxidative stress (186). On the other hand, natural anthraquinone derivatives can enhance the cytotoxic effects of arsenic as seen on a study using human esophageal carcinoma derived cell line EC/CHUK1 (187). Aqueous extract of garlic reduced intracellular reactive oxygen species (ROS) level in human malignant melanoma cells (A375), human keratinocyte cells (HaCaT) and in cultured human normal dermal fibroblast cells and consequently could attenuate arsenic-induced cytotoxicity (188). Similar alleviating effects were found for other antioxidants including black tea polyphenols (189), curcumin (190,191), folate (192), melatonin (193), plasma alpha tocopherol (194) etc.

Selenium plays a special role in arsenic toxicity and susceptibility; the two metals show an antagonistic behavior in the body. Selenium was shown to have protective effects against arsenic-induced clastogenicity in cultured human lymphocytes (195). These results were further corroborated by a study on smelter workers exposed chronically to arsenic, where, the chromosomal aberration rate of cultured lymphocytes was lowered about 46.1% after treatment with selenium 150 g/d for 21 days (196). Such observations have been reinforced by epidemiological studies. Nail selenium content of individuals chronically exposed to arsenic was inversely associated with arsenic-induced OGG1 expression, which is a hallmark of oxidative stress (197). In addition, it was also found that at any given arsenic exposure level, the risk of premalignant skin lesions was consistently greater among participants with blood selenium lower than the average level (198). The mechanisms for such mutual detoxification are not perfectly understood although, it has been hypothesized that arsenic and selenium increase the biliary excretion of each other and consequent excretion from the system (reviewed in 199). Also, it has been shown that low selenium status is associated with retention of higher amount of inorganic arsenic in the body and a lower biotransformation into dimethylarsinic acid (200).

### Conclusion

Thus, to sum up, chronic exposure to arsenic in human beings leads to a host of different types of dermatological and non-dermatological cancers. The individual susceptibility to such cancers are however controlled by a plethora of several low penetrant genes, mutations and polymorphisms thereof and their complex interplay with several environmental factors, notable of them being the dietary factors, which modulate arsenic

biotransformation and the oxidative stress induced by inorganic arsenic and its methylated metabolites.

## References

- Mondal D, Banerjee M, Kundu M, Banerjee N, Bhattacharya U, Giri AK, Ganguli B, Sen Roy S, Polya DA. Comparison of drinking water, raw rice and cooking of rice as arsenic exposure routes in three contrasting areas of West Bengal, India. *Environ Geochem Health*. 2010; 32: 463–77.
- Polya DA, Charlet L. Rising arsenic risk? *Nature Geoscience*. 2009; 2: 383–4.
- IARC. IARC monographs on the evaluation of carcinogenic risk of chemicals to humans: some drinking water disinfectants and contaminants, including arsenic. IARC Sci Publ. 2004; 84: 1–477.
- ATSDR. CERCLA list of hazardous substances [Accessed at: <http://www.watsdrcdcgov/cercla/07listhtml>, Accessed on 17032011]
- Trepka MJ, Heinrich J, Schulz C, Krause C, Popescu M, Wjst M, Wichmann HE. Arsenic burden among children in industrial areas of eastern Germany. *Sci Total Environ* 1996; 180: 95–105.
- Hysong TA, Burgess JL, Cebrián Garcia ME, O'Rourke MK. House dust and inorganic urinary arsenic in two Arizona mining towns. *J Expo Anal Environ Epidemiol*. 2003; 13: 211–8.
- Tollestrup K, Frost FJ, Harter LC, McMillan GP. Mortality among children residing near the American Smelting and Refining Company (ASARCO) copper smelter in Ruston, Washington. *Arch Environ Health*. 2003; 58: 683–91.
- Paiva L, Martínez V, Creus A, Quinteros D, Marcos R. Sister chromatid exchange analysis in smelting plant workers exposed to arsenic. *Environ Mol Mutagen*. 2006; 47: 230–5.
- Paiva L, Martínez V, Creus A, Quinteros D, Marcos R. Evaluation of micronucleus frequencies in blood lymphocytes from smelting plant workers exposed to arsenic. *Environ Mol Mutagen*. 2008; 49: 200–5.
- Vuyyuri SB, Ishaq M, Kuppala D, Grover P, Ahuja YR. Evaluation of micronucleus frequencies and DNA damage in glass workers exposed to arsenic. *Environ Mol Mutagen*. 2006; 47: 562–70.
- Apostoli P, Bartoli D, Alessio L, Buchet JP. Biological monitoring of occupational exposure to inorganic arsenic. *Occup Environ Med*. 1999; 56: 825–32.
- Edelman P. Environmental and workplace contamination in the semiconductor industry: implications for future health of the workforce and community. *Environ Health Perspect*. 1990; 86: 291–5.
- Hwang YH, Lee ZY, Wang JD, Hsueh YM, Lu IC, Yao WL. Monitoring of arsenic exposure with speciated urinary inorganic arsenic metabolites for ion implanter maintenance engineers. *Environ Res*. 2002; 90: 207–16.
- Liao YH, Yu HS, Ho CK, Wu MT, Yang CY, Chen JR, Chang CC. Biological monitoring of exposures to aluminium, gallium, indium, arsenic, and antimony in optoelectronic industry workers. *J Occup Environ Med*. 2004; 46: 931–6.
- Hu CW, Pan CH, Huang YL, Wu MT, Chang LW, Wang CJ, Chao MR. Effects of arsenic exposure among semiconductor workers: a cautionary note on urinary 8-oxo-7,8-dihydro-2'-deoxyguanosine. *Free Radic Biol Med*. 2006; 40: 1273–8.
- Chou WC, Chio CP, Liao CM. Assessing airborne PM-bound arsenic exposure risk in semiconductor manufacturing facilities. *J Hazard Mater*. 2009; 167: 976–86.
- Park D, Yang H, Jeong J, Ha K, Choi S, Kim C, Yoon C, Park D, Paek D. A comprehensive review of arsenic levels in the semiconductor manufacturing industry. *Ann Occup Hyg*. 2010; 54: 869–79.
- Bencko V, Symon K, Stálnik L, Bátor J, Vanco E, Svandová E. Rate of malignant tumor mortality among coal burning power plant workers occupationally exposed to arsenic. *J Hyg Epidemiol Microbiol Immunol*. 1980; 24: 278–84.
- Pesch B, Ranft U, Jakubis P, Nieuwenhuijsen MJ, Hergemöller A, Unfried K, Jakubis M, Miskovic P, Keegan T. Environmental arsenic exposure from a coal-burning power plant as a potential risk factor for nonmelanoma skin carcinoma: results from a case-control study in the district of Prievidza, Slovakia. *Am J Epidemiol*. 2002; 155: 798–809.
- Wagner SL, Weswig P. Arsenic in blood and urine of forest workers as indices of exposure to cacodylic acid. *Arch Environ Health*. 1974; 28: 77–9.
- Peoples SA, Maddy KT, Peifer WR, Edmiston S. Occupational exposures to pesticides containing organoarsenicals in California. *Vet Hum Toxicol*. 1979; 21: 417–21.
- Mabuchi K, Lilienfeld AM, Snell LM. Cancer and occupational exposure to arsenic: a study of pesticide workers. *Prev Med*. 1980; 9: 51–77.
- Miller CS, Hoover WL, Culver WH. Exposure of pesticide applicators to arsenic acid. *Arch Environ Contam Toxicol*. 1980; 9: 281–8.
- Wojeck GA, Nigg HN, Braman RS, Stamper JH, Rouseff RL. Worker exposure to arsenic in Florida grapefruit spray operations. *Arch Environ Contam Toxicol*. 1982; 11: 661–7.
- Abdelghani AA, Anderson AC, Jaghabir M, Mather F. Arsenic levels in blood, urine, and hair of workers applying monosodium methanearsonate (MSMA). *Arch Environ Health*. 1986; 41: 163–9.
- Dennis LK, Lynch CF, Sandler DP, Alavanja MC. Pesticide use and cutaneous melanoma in pesticide applicators in the agricultural health study. *Environ Health Perspect*. 2010; 118: 812–7.
- Tseng WP. Effects and dose-response relationships of skin cancer and blackfoot disease with arsenic. *Environ Health Perspect*. 1977; 19: 109–19.
- Chen CJ, Chuang YC, Lin TM, Wu HY. Malignant neoplasms among residents of a blackfoot disease-endemic area in Taiwan: high-arsenic artesian well water and cancers. *Cancer Res*. 1985; 45: 5895–9.
- Chen CJ, Chuang YC, You SL, Lin TM, Wu HY. A retrospective study on malignant neoplasms of bladder,

- lung and liver in blackfoot disease endemic area in Taiwan. *Br J Cancer*. 1986; 53: 399–405.
- 30 Wu MM, Kuo TL, Hwang YH, Chen CJ. Dose-response relation between arsenic concentration in well water and mortality from cancers and vascular diseases. *Am J Epidemiol*. 1989; 130: 1123–32.
  - 31 Zaldívar R. A morbid condition involving cardio-vascular, broncho-pulmonary, digestive and neural lesions in children and young adults after dietary arsenic exposure. *Zentralbl Bakteriell B*. 1980; 170: 44–56.
  - 32 Zaldívar R, Prunés L, Ghai GL. Arsenic dose in patients with cutaneous carcinomata and hepatic hemangio-endothelioma after environmental and occupational exposure. *Arch Toxicol*. 1981; 47: 145–54.
  - 33 Saha KC. Melanokeratosis from arsenic contaminated tubewell water. *Indian J Dermatol*. 1984; 29: 37–46.
  - 34 Groundwater Studies for Arsenic Contamination in Bangladesh Phase I: Rapid Investigation Phase, Final Report, Main Report, British Geological Survey, Mott MacDonald Ltd (UK), January 1999.
  - 35 Vahter M, Concha G, Nermell B, Nilsson R, Dulout F, Natarajan AT. A unique metabolism of inorganic arsenic in native Andean women. *Eur J Pharmacol*. 1995; 293: 455–62.
  - 36 Feng Z, Xia Y, Tian D, Wu K, Schmitt M, Kwok RK, Mumford JL. DNA damage in buccal epithelial cells from individuals chronically exposed to arsenic via drinking water in Inner Mongolia, China. *Anticancer Res*. 2001; 21: 51–7.
  - 37 Mukherjee A, Sengupta MK, Hossain MA, Ahamed S, Das B, Nayak B, Lodh D, Rahman MM, Chakraborti D. Arsenic contamination in groundwater: a global perspective with emphasis on the Asian scenario. *J Health Popul Nutr*. 2006; 24: 142–63.
  - 38 Garelick H, Jones H, Dybowska A, Valsami-Jones E. Arsenic pollution sources. *Rev Environ Contam Toxicol*. 2008; 197: 17–60.
  - 39 Bencko V, Symon K. Test of environmental exposure to arsenic and hearing changes in exposed children. *Environ Health Perspect*. 1977; 19: 95–101.
  - 40 Liu J, Zheng B, Aposhian HV, Zhou Y, Chen ML, Zhang A, Waalkes MP. Chronic arsenic poisoning from burning high-arsenic-containing coal in Guizhou, China. *Environ Health Perspect*. 2002; 110: 119–22.
  - 41 Guha Mazumder DN, Haque R, Ghosh N, De BK, Santra A, Chakraborti D, Smith AH. Arsenic in drinking water and the prevalence of respiratory effects in West Bengal, India. *Int J Epidemiol*. 2000; 29: 1047–52.
  - 42 Hossain MK, Khan MM, Alam MA, Chowdhury AK, Delwar Hossain M, Feroze Ahmed M, Kobayashi K, Sakauchi F, Mori M. Manifestation of arsenicosis patients and factors determining the duration of arsenic symptoms in Bangladesh. *Toxicol Appl Pharmacol*. 2005; 208: 78–86.
  - 43 Baidya K, Raj A, Mondal L, Bhaduri G, Todani A. Persistent conjunctivitis associated with drinking arsenic-contaminated water. *J Ocul Pharmacol Ther*. 2006; 22: 208–11.
  - 44 Soto-Peña GA, Luna AL, Acosta-Saavedra L, Conde P, López-Carrillo L, Cebrián ME, Bastida M, Calderón-Aranda ES, Vega L. Assessment of lymphocyte subpopulations and cytokine secretion in children exposed to arsenic. *FASEB J*. 2006; 20: 779–81.
  - 45 Ghosh P, Banerjee M, De Chaudhuri S, Chowdhury R, Das JK, Mukherjee A, Sarkar AK, Mondal L, Baidya K, Sau TJ, Banerjee A, Basu A, Chaudhuri K, Ray K, Giri AK. Comparison of health effects between individuals with and without skin lesions in the population exposed to arsenic through drinking water in West Bengal, India. *J Expo Sci Environ Epidemiol*. 2007; 17: 215–23.
  - 46 Hutchinson J. An address on the study of skin diseases as illustrating the doctrines of general pathology. *Br Med J*. 1887; 2: 229–32.
  - 47 Rickert RR, Brodtkin RH, Hutter RV. Bowen's disease. *CA Cancer J Clin*. 1977; 27: 160–6.
  - 48 Graham JH, Mazzanti GR, Helwig EB. Chemistry of Bowen's disease: relationship to arsenic. *J Invest Derm*. 1961; 37: 317–32.
  - 49 Imamura S, Odaka T. Multiple Bowen's disease seen in a localized district, especially on etiologic significance of arsenic. *Hifuka Kiyo*. 1962; 57: 99–115.
  - 50 Yeh S, How SW, Lin CS. Arsenical cancer of skin: Histologic study with special reference to Bowen's disease. *Cancer*. 1968; 21: 312–39.
  - 51 Yeh S, Chen HC, How SW, Deng CS. Fine structure of Bowen's disease in chronic arsenicalism. *J Natl Cancer Inst*. 1974; 53: 31–44.
  - 52 Hamada T, Horiguchi S. Occupational chronic arsenical poisoning on the cutaneous manifestations. *Sangyo Igaku*. 1976; 18: 103–15.
  - 53 Koh E, Kondoh N, Kaihara H, Fujioka H, Kitamura K. Ureteral tumor with multiple Bowen's disease forty-two years after exposure to arsenic. *Eur Urol*. 1989; 16: 398–400.
  - 54 Tsuda T, Nagira T, Yamamoto M, Kume Y. An epidemiological study on cancer in certified arsenic poisoning patients in Toroku. *Ind Health*. 1990; 28: 53–62.
  - 55 Matsui M, Nishigori C, Toyokuni S, Takada J, Akaboshi M, Ishikawa M, Imamura S, Miyachi Y. The role of oxidative DNA damage in human arsenic carcinogenesis: detection of 8-hydroxy-2'-deoxyguanosine in arsenic-related Bowen's disease. *J Invest Dermatol*. 1999; 113: 26–31.
  - 56 Leong GK, Stone JL, Farmer ER, Scotto J, Reizner GT, Burnett TS, Elpern DJ. Nonmelanoma skin cancer in Japanese residents of Kauai, Hawaii. *J Am Acad Dermatol*. 1987; 17: 233–8.
  - 57 Foy HM, Tarmapai S, Eamchan P, Metdilogkul O. Chronic arsenic poisoning from well water in a mining area in Thailand. *Asia Pac J Public Health*. 1992–1993; 6: 150–2.
  - 58 Yu HS, Chang KL, Wang CM, Yu CL J. Alterations of mitogenic responses of mononuclear cells by arsenic in arsenical skin cancers. *Dermatol*. 1992; 19: 710–4.
  - 59 Kuo TT, Hu S, Lo SK, Chan HL. p53 expression and proliferative activity in Bowen's disease with or without chronic arsenic exposure. *Hum Pathol*. 1997; 28:



- 786–90.
- 60 Castrén K, Ranki A, Welsh JA, Vähäkangas KH. Infrequent p53 mutations in arsenic-related skin lesions. *Oncol Res.* 1998; 10: 475–82.
- 61 Cöl M, Cöl C, Soran A, Sayli BS, Oztürk S. Arsenic-related Bowen's disease, palmar keratosis, and skin cancer. *Environ Health Perspect.* 1999; 107: 687–9.
- 62 Yerebakan O, Ermis O, Yilmaz E, Basaran E. Treatment of arsenical keratosis and Bowen's disease with acitretin. *Int J Dermatol.* 2002; 41: 84–7.
- 63 Kim HJ, Min HG, Lee ES. Bowen's diseases and basal cell carcinomas in a patient. *J Dermatol.* 1999; 26: 695–7.
- 64 Park JY, Rim JH, Choe YB, Youn JI. Arsenic keratosis and pigmentation accompanied by multiple Bowen's disease and genitourinary cancer in a psoriasis patient. *J Dermatol.* 2002; 29: 446–51.
- 65 Rahman MM, Sengupta MK, Ahamed S, Chowdhury UK, Hossain MA, Das B, Lodh D, Saha KC, Pati S, Kaies I, Barua AK, Chakraborti D. The magnitude of arsenic contamination in groundwater and its health effects to the inhabitants of the Jalangi—one of the 85 arsenic affected blocks in West Bengal, India. *Sci Total Environ.* 2005; 338: 189–200.
- 66 Ghosh P, Banerjee M, De Chaudhuri S, Das JK, Sarma N, Basu A, Giri AK. Increased chromosome aberration frequencies in the Bowen's patients compared to non-cancerous skin lesions individuals exposed to arsenic. *Mutat Res.* 2007; 632: 104–10.
- 67 Ghosh P, Basu A, Singh KK, Giri AK. Evaluation of cell types for assessment of cytogenetic damage in arsenic exposed population. *Mol Cancer.* 2008; 7: 45.
- 68 Ahmad SA, Maharjan M, Watanabe C, Ohtsuka R. Arsenicosis in two villages in Terai, lowland Nepal. *Environ Sci.* 2004; 11: 179–88.
- 69 Leibowitz MR, Rippey JJ, Thompson P. Fungating Bowen's disease with presumed arsenic intoxication A case report. *S Afr Med J.* 1982; 62: 737–9.
- 70 Hsu YH, Li SY, Chiou HY, Yeh PM, Liou JC, Hsueh YM, Chang SH, Chen CJ. Spontaneous and induced sister chromatid exchanges and delayed cell proliferation in peripheral lymphocytes of Bowen's disease patients and matched controls of arseniasis-hyperendemic villages in Taiwan. *Mutat Res.* 1997; 386: 241–51.
- 71 Hsu CH, Yang SA, Wang JY, Yu HS, Lin SR. Mutational spectrum of p53 gene in arsenic-related skin cancers from the blackfoot disease endemic area of Taiwan. *Br J Cancer.* 1999; 80: 1080–6.
- 72 Wang BJ, Lee YY, Mak CP, Kao HF, Hsu ML, Hsien JR. Quantitative and morphological changes of Langerhans cells in Bowen's disease from patients with chronic arsenicism. *J Formos Med Assoc.* 1991; 90: 1093–8.
- 73 Yu HS, Chen GS, Sheu HM, Kao JS, Chang KL, Yu CL. Alterations of skin-associated lymphoid tissue in the carcinogenesis of arsenical skin cancer. *Proc Natl Sci Counc Repub China B.* 1992; 16: 17–22.
- 74 Sheu HM, Ko SS, Yu HS. Expression of keratin 16 in normal and lesional skin of solitary and multiple Bowen's disease using monoclonal antibody Ks 812 staining. *J Formos Med Assoc.* 1991; 90: 1252–5.
- 75 Yu HS, Chiou KS, Chen GS, Yang RC, Chang SF. Progressive alterations of cytokeratin expressions in the process of chronic arsenism. *J Dermatol.* 1993; 20: 741–5.
- 76 Chang CH, Tsai RK, Chen GS, Yu HS, Chai CY. Expression of bcl-2, p53 and Ki-67 in arsenical skin cancers. *J Cutan Pathol.* 1998; 25: 457–62.
- 77 Liao WT, Yu CL, Lan CC, Lee CH, Chang CH, Chang LW, You HL, Yu HS. Differential effects of arsenic on cutaneous and systemic immunity: focusing on CD4<sup>+</sup> cell apoptosis in patients with arsenic-induced Bowen's disease. *Carcinogenesis.* 2009; 30: 1064–72.
- 78 Yu HS, Chang KL, Yu CL, Wu CS, Chen GS, Ho JC. Defective IL-2 receptor expression in lymphocytes of patients with arsenic-induced Bowen's disease. *Yu Arch Dermatol Res.* 1998; 290: 681–7.
- 79 Yu HS, Liao WT, Chang KL, Yu CL, Chen GS. Arsenic induces tumor necrosis factor alpha release and tumor necrosis factor receptor 1 signaling in T helper cell apoptosis. *J Invest Dermatol.* 2002; 119: 812–9.
- 80 Centeno JA, Mullick FG, Martinez L, Page NP, Gibb H, Longfellow D, Thompson C, Ladich ER. Pathology related to chronic arsenic exposure. *Environ Health Perspect.* 2002; 110 Suppl 5: 883–6.
- 81 Møller R, Nielsen A, Reymann F. Multiple basal cell carcinoma and internal malignant tumors. *Arch Dermatol.* 1975; 111: 584–5.
- 82 Jackson R, Grainge JW. Arsenic and cancer. *Can Med Assoc J.* 1975; 113: 396–401.
- 83 Morton W, Starr G, Pohl D, Stoner J, Wagner S, Weswig D. Skin cancer and water arsenic in Lane County, Oregon. *Cancer.* 1976; 37: 2523–32.
- 84 Cabrera HN, Cuda G, López M, Costa JA. Basal cell epithelioma of the vulva in chronic endemic regional arsenic poisoning. *Med Cutan Ibero Lat Am.* 1984; 12: 81–5.
- 85 Tan SH, Tham SN, Goh CL. Skin cancers at Tertiary Referral Skin Hospital in Singapore. *Int J Dermatol.* 1995; 34: 770–6.
- 86 Wong SS, Tan KC, Goh CL. Cutaneous manifestations of chronic arsenicism: review of seventeen cases. *J Am Acad Dermatol.* 1998; 38 (2 Pt 1): 179–85.
- 87 Boonchai W, Walsh M, Cummings M, Chenevix-Trench G. Expression of p53 in arsenic-related and sporadic basal cell carcinoma. *Arch Dermatol.* 2000; 136: 195–8.
- 88 Boonchai W, Green A, Ng J, Dicker A, Chenevix-Trench G. Basal cell carcinoma in chronic arsenicism occurring in Queensland, Australia, after ingestion of an asthma medication. *J Am Acad Dermatol.* 2000; 43: 664–9.
- 89 Paul PC, Chattopadhyay A, Dutta SK, Mazumder DN, Santra A. Histopathology of skin lesions in chronic arsenic toxicity--grading of changes and study of proliferative markers. *Indian J Pathol Microbiol.* 2000; 43: 257–64.
- 90 Karagas MR, Stukel TA, Morris JS, Tosteson TD, Weiss JE, Spencer SK, Greenberg ER. Skin cancer risk in relation to toenail arsenic concentrations in a US

- population-based case-control study. *Am J Epidemiol.* 2001; 153: 559–65.
- 91 Cabrera HN, Gómez ML. Skin cancer induced by arsenic in the water. *J Cutan Med Surg.* 2003; 7: 106–11.
  - 92 Thirumaran RK, Bermejo JL, Rudnai P, Gurzau E, Koppova K, Goessler W, Vahter M, Leonardi GS, Clemens F, Fletcher T, Hemminki K, Kumar R. Single nucleotide polymorphisms in DNA repair genes and basal cell carcinoma of skin. *Carcinogenesis.* 2006; 27: 1676–81.
  - 93 Applebaum KM, Karagas MR, Hunter DJ, Catalano PJ, Byler SH, Morris S, Nelson HH. Polymorphisms in nucleotide excision repair genes, arsenic exposure, and non-melanoma skin cancer in New Hampshire. *Environ Health Perspect.* 2007; 115: 1231–6.
  - 94 Robertson JW, Clement KW. Squamous cell carcinoma following prolonged use of arsenic; case report. *Ohio Med.* 1948; 44: 280.
  - 95 Gustavsson P, Gustavsson A, Hogstedt C. Excess of cancer in Swedish chimney sweeps. *Br J Ind Med.* 1988; 45: 777–81.
  - 96 Guo HR, Yu HS, Hu H, Monson RR. Arsenic in drinking water and skin cancers: cell-type specificity (Taiwan, ROC). *Cancer Causes Control.* 2001; 12: 909–16.
  - 97 Fumal I, Piérard-Franchimont C, Piérard GE. Arsenical keratoses and carcinomas. *Rev Med Liege.* 2005; 60: 217–21.
  - 98 Zhang A, Feng H, Yang G, Pan X, Jiang X, Huang X, Dong X, Yang D, Xie Y, Peng L, Jun L, Hu C, Jian L, Wang X. Unventilated indoor coal-fired stoves in Guizhou province, China: cellular and genetic damage in villagers exposed to arsenic in food and air. *Environ Health Perspect.* 2007; 115: 653–8.
  - 99 Gawkrödger DJ. Occupational skin cancers. *Occup Med (Lond).* 2004; 54: 458–63.
  - 100 Pershagen G. The carcinogenicity of arsenic. *Environ Health Perspect.* 1981; 40: 93–100.
  - 101 Pi J, Diwan BA, Sun Y, Liu J, Qu W, He Y, Styblo M, Waalkes MP. Arsenic-induced malignant transformation of human keratinocytes: involvement of Nrf2. *Free Radic Biol Med.* 2008; 45: 651–8.
  - 102 Sun Y, Pi J, Wang X, Tokar EJ, Liu J, Waalkes MP. Aberrant cytokeratin expression during arsenic-induced acquired malignant phenotype in human HaCaT keratinocytes consistent with epidermal carcinogenesis. *Toxicol.* 2009; 262: 162–70.
  - 103 Samuel HS. Arsenic and lung cancer. *Br Med J.* 1964; 1: 1656.
  - 104 Pershagen G, Elinder CG, Bolander AM. Mortality in a region surrounding an arsenic emitting plant. *Environ Health Perspect.* 1977; 19: 133–7.
  - 105 Axelsson O, Dahlgren E, Jansson CD, Rehnlund SO. Arsenic exposure and mortality: a case-referent study from a Swedish copper smelter. *Br J Ind Med.* 1978; 35: 8–15.
  - 106 Rom WN, Varley G, Lyon JL, Shopkow S. Lung cancer mortality among residents living near the El Paso smelter. *Br J Ind Med.* 1982; 39: 269–72.
  - 107 Taylor PR, Qiao YL, Schatzkin A, Yao SX, Lubin J, Mao BL, Rao JY, McAdams M, Xuan XZ, Li JY. Relation of arsenic exposure to lung cancer among tin miners in Yunnan Province, China. *Br J Ind Med.* 1989; 46: 881–6.
  - 108 Järup L, Pershagen G. Arsenic exposure, smoking, and lung cancer in smelter workers—a case-control study. *Am J Epidemiol.* 1991; 134: 545–51. Erratum in: *Am J Epidemiol.* 1992; 136: 1174.
  - 109 Smith AH, Hopenhayn-Rich C, Bates MN, Goeden HM, Hertz-Picciotto I, Duggan HM, Wood R, Kosnett MJ, Smith MT. Cancer risks from arsenic in drinking water. *Environ Health Perspect.* 1992; 97: 259–67.
  - 110 Chen CJ, Chen CW, Wu MM, Kuo TL. Cancer potential in liver, lung, bladder and kidney due to ingested inorganic arsenic in drinking water. *Br J Cancer.* 1992; 66: 888–92.
  - 111 Chiou HY, Hsueh YM, Liaw KF, Horng SF, Chiang MH, Pu YS, Lin JS, Huang CH, Chen CJ. Incidence of internal cancers and ingested inorganic arsenic: a seven-year follow-up study in Taiwan. *Cancer Res.* 1995; 55: 1296–300.
  - 112 Ferreccio C, González Psych C, Milosavjevic Stat V, Marshall Gredis G, Sancha AM. Lung cancer and arsenic exposure in drinking water: a case-control study in northern Chile. *Cad Saude Publica.* 1998; 14 Suppl 3: 193–8.
  - 113 Ferreccio C, González C, Milosavjevic V, Marshall G, Sancha AM, Smith AH. Lung cancer and arsenic concentrations in drinking water in Chile. *Epidemiology* 2000; 11: 673–9.
  - 114 Morales KH, Ryan L, Kuo TL, Wu MM, Chen CJ. Risk of internal cancers from arsenic in drinking water. *Environ Health Perspect.* 2000; 108: 655–61.
  - 115 Guo HR. Arsenic level in drinking water and mortality of lung cancer (Taiwan). *Cancer Causes Control.* 2004; 15: 171–7.
  - 116 Chen Y, Ahsan H. Cancer burden from arsenic in drinking water in Bangladesh. *Am J Public Health.* 2004; 94: 741–4.
  - 117 Mostafa MG, McDonald JC, Cherry NM. Lung cancer and exposure to arsenic in rural Bangladesh. *Occup Environ Med.* 2008; 65: 765–8.
  - 118 Liao CM, Shen HH, Chen CL, Hsu LI, Lin TL, Chen SC, Chen CJ J. Risk assessment of arsenic-induced internal cancer at long-term low dose exposure. *Hazard Mater.* 2009; 165: 652–63.
  - 119 Heck JE, Andrew AS, Onega T, Rigas JR, Jackson BP, Karagas MR, Duell EJ. Lung cancer in a US population with low to moderate arsenic exposure. *Environ Health Perspect.* 2009; 117: 1718–23.
  - 120 Smith AH, Ercumen A, Yuan Y, Steinmaus CM. Increased lung cancer risks are similar whether arsenic is ingested or inhaled. *J Expo Sci Environ Epidemiol.* 2009; 19: 343–8.
  - 121 Higgins I, Welch K, Oh M, Bond G, Hurwitz P. Influence of arsenic exposure and smoking on lung cancer among smelter workers: a pilot study. *Am J Ind Med.* 1981; 2: 33–41.
  - 122 Pershagen G, Wall S, Taube A, Linnman L. On the interaction between occupational arsenic exposure and

- smoking and its relationship to lung cancer. *Scand J Work Environ Health*. 1981; 7: 302-9.
- 123 Hertz-Picciotto I, Smith AH, Holtzman D, Lipsett M, Alexeeff G. Synergism between occupational arsenic exposure and smoking in the induction of lung cancer. *Epidemiology*. 1992; 3: 23-31.
- 124 Chen CL, Hsu LI, Chiou HY, Hsueh YM, Chen SY, Wu MM, Chen CJ. Ingested arsenic, cigarette smoking, and lung cancer risk: a follow-up study in arseniasis-endemic areas in Taiwan. *JAMA*. 2004; 292: 2984-90.
- 125 Adonis M, Martínez V, Marin P, Berrios D, Gil L. Smoking habit and genetic factors associated with lung cancer in a population highly exposed to arsenic. *Toxicol Lett*. 2005; 159: 32-7.
- 126 Smith AH, Marshall G, Yuan Y, Ferreccio C, Liaw J, von Ehrenstein O, Steinmaus C, Bates MN, Selvin S. Increased mortality from lung cancer and bronchiectasis in young adults after exposure to arsenic in utero and in early childhood. *Environ Health Perspect*. 2006; 114: 1293-6.
- 127 Mazumdar S, Redmond CK, Enterline PE, Marsh GM, Costantino JP, Zhou SY, Patwardhan RN. Multistage modeling of lung cancer mortality among arsenic-exposed copper-smelter workers. *Risk Anal*. 1989; 9: 551-63.
- 128 Li M, Cai JF, Chiu JF. Arsenic induces oxidative stress and activates stress gene expressions in cultured lung epithelial cells. *J Cell Biochem*. 2002; 87: 29-38.
- 129 Wang F, Shi Y, Yadav S, Wang H. p52-Bcl3 complex promotes cyclin D1 expression in BEAS-2B cells in response to low concentration arsenite. *Toxicology*. 2010; 273: 12-8.
- 130 Dong JT, Luo XM. Arsenic-induced DNA-strand breaks associated with DNA-protein crosslinks in human fetal lung fibroblasts. *Mutat Res*. 1993; 302: 97-102.
- 131 Dong JT, Luo XM. Effects of arsenic on DNA damage and repair in human fetal lung fibroblasts. *Mutat Res*. 1994; 315: 11-5.
- 132 Cui X, Wakai T, Shirai Y, Hatakeyama K, Hirano S. Chronic oral exposure to inorganic arsenate interferes with methylation status of p16INK4a and RASSF1A and induces lung cancer in A/J mice. *Toxicol Sci*. 2006; 91: 372-81.
- 133 Martinez VD, Buys TP, Adonis M, Benítez H, Gallegos I, Lam S, Lam WL, Gil L. Arsenic-related DNA copy-number alterations in lung squamous cell carcinomas. *Br J Cancer*. 2010; 103: 1277-83.
- 134 Chiang HS, Hong CL, Guo HR, Lee EF, Chen TY. Comparative study on the high prevalence of bladder cancer in the blackfoot disease endemic area in Taiwan. *Taiwan Yi Xue Hui Za Zhi*. 1988; 87: 1074-80.
- 135 Cuzick J, Sasieni P, Evans S. Ingested arsenic, keratoses, and bladder cancer. *Am J Epidemiol*. 1992; 136: 417-21.
- 136 Hopenhayn-Rich C, Biggs ML, Fuchs A, Bergoglio R, Tello EE, Nicolli H, Smith AH. Bladder cancer mortality associated with arsenic in drinking water in Argentina. *Epidemiology*. 1996; 7: 117-24.
- 137 Smith AH, Goycolea M, Haque R, Biggs ML. Marked increase in bladder and lung cancer mortality in a region of Northern Chile due to arsenic in drinking water. *Am J Epidemiol*. 1998; 147: 660-9.
- 138 Kurttio P, Pukkala E, Kahelin H, Auvinen A, Pekkanen J. Arsenic concentrations in well water and risk of bladder and kidney cancer in Finland. *Environ Health Perspect*. 1999; 107: 705-10.
- 139 Chen YC, Su HJ, Guo YL, Hsueh YM, Smith TJ, Ryan LM, Lee MS, Christiani DC. Arsenic methylation and bladder cancer risk in Taiwan. *Cancer Causes Control*. 2003; 14: 303-10.
- 140 Bates MN, Rey OA, Biggs ML, Hopenhayn C, Moore LE, Kalman D, Steinmaus C, Smith AH. Case-control study of bladder cancer and exposure to arsenic in Argentina. *Am J Epidemiol*. 2004; 159: 381-9.
- 141 Chen Y, Ahsan H. Cancer burden from arsenic in drinking water in Bangladesh. *Am J Public Health*. 2004; 94: 741-4.
- 142 Marshall G, Ferreccio C, Yuan Y, Bates MN, Steinmaus C, Selvin S, Liaw J, Smith AH. Fifty-year study of lung and bladder cancer mortality in Chile related to arsenic in drinking water. *J Natl Cancer Inst*. 2007; 99: 920-8.
- 143 Chow NH, Guo YL, Lin JS, Su JH, Tzai TS, Guo HR, Su JJ. Clinicopathological features of bladder cancer associated with chronic exposure to arsenic. *Br J Cancer*. 1997; 75: 1708-10.
- 144 Moore LE, Smith AH, Eng C, Kalman D, DeVries S, Bhargava V, Chew K, Moore D 2nd, Ferreccio C, Rey OA, Waldman FM. Arsenic-related chromosomal alterations in bladder cancer. *J Natl Cancer Inst*. 2002; 94: 1688-96.
- 145 Chen CH, Chiou HY, Hsueh YM, Chen CJ, Yu HJ, Pu YS. Clinicopathological characteristics and survival outcome of arsenic related bladder cancer in Taiwan. *J Urol*. 2009; 181: 547-52.
- 146 Chen YC, Su HJ, Guo YL, Hsueh YM, Smith TJ, Ryan LM, Lee MS, Christiani DC. Arsenic methylation and bladder cancer risk in Taiwan. *Cancer Causes Control*. 2003; 14: 303-10.
- 147 Hernández-Zavala A, Córdova E, Del Razo LM, Cebrián ME, Garrido E. Effects of arsenite on cell cycle progression in a human bladder cancer cell line. *Toxicology*. 2005; 207: 49-57.
- 148 Hour TC, Pu YS, Lin CC, Huang SW, Chen J, Chiu AW, Chen CJ, Huang CY. Differential expression of molecular markers in arsenic- and non-arsenic-related urothelial cancer. *Anticancer Res*. 2006; 26: 375-8.
- 149 Steinmaus C, Bates MN, Yuan Y, Kalman D, Atallah R, Rey OA, Biggs ML, Hopenhayn C, Moore LE, Hoang BK, Smith AH. Arsenic methylation and bladder cancer risk in case-control studies in Argentina and the United States. *J Occup Environ Med*. 2006; 48: 478-88.
- 150 Pu YS, Yang SM, Huang YK, Chung CJ, Huang SK, Chiu AW, Yang MH, Chen CJ, Hsueh YM. Urinary arsenic profile affects the risk of urothelial carcinoma even at low arsenic exposure. *Toxicol Appl Pharmacol*. 2007; 218: 99-106.
- 151 Bredfeldt TG, Jagadish B, Eblin KE, Mash EA, Gan-

- dolfi AJ. Monomethylarsonous acid induces transformation of human bladder cells. *Toxicol Appl Pharmacol.* 2006; 216: 69–79.
- 152 Wnek SM, Jensen TJ, Severson PL, Futscher BW, Gandolfi AJ. Monomethylarsonous acid produces irreversible events resulting in malignant transformation of a human bladder cell line following 12 weeks of low-level exposure. *Toxicol Sci.* 2010; 116: 44–57.
- 153 Eblin KE, Bredfeldt TG, Buffington S, Gandolfi AJ. Mitogenic signal transduction caused by monomethylarsonous acid in human bladder cells: role in arsenic-induced carcinogenesis. *Toxicol Sci.* 2007; 95: 321–30.
- 154 Escudero-Lourdes C, Medeiros MK, Cordenas-Gonzalez MC, Wnek SM, Gandolfi JA. Low level exposure to monomethyl arsonous acid-induced the overproduction of inflammation-related cytokines and the activation of cell signals associated with tumor progression in a urothelial cell model. *Toxicol Appl Pharmacol.* 2010; 244: 162–73.
- 155 Wang TC, Jan KY, Wang AS, Gurr JR. Trivalent arsenicals induce lipid peroxidation, protein carbonylation, and oxidative DNA damage in human urothelial cells. *Mutat Res.* 2007; 615: 75–86.
- 156 Chung CJ, Huang CJ, Pu YS, Su CT, Huang YK, Chen YT, Hsueh YM. Urinary 8-hydroxydeoxyguanosine and urothelial carcinoma risk in low arsenic exposure area. *Toxicol Appl Pharmacol.* 2008; 226: 14–21.
- 157 Chen WT, Hung WC, Kang WY, Huang YC, Chai CY. Urothelial carcinomas arising in arsenic-contaminated areas are associated with hypermethylation of the gene promoter of the death-associated protein kinase. *Histopathology.* 2007; 51: 785–92.
- 158 Regelson W, Kim U, Ospina J, Holland JF. Hemangioendothelial sarcoma of liver from chronic arsenic intoxication by Fowler's solution. *Cancer.* 1968; 21: 514–22.
- 159 Lander JJ, Stanley RJ, Sumner HW, Boswell DC, Aach RD. Angiosarcoma of the liver associated with Fowler's solution (potassium arsenite). *Gastroenterology.* 1975; 68: 1582–6.
- 160 Falk H, Caldwell GG, Ishak KG, Thomas LB, Popper H. Arsenic-related hepatic angiosarcoma. *Am J Ind Med.* 1981; 2: 43–50.
- 161 Kasper ML, Schoenfield L, Strom RL, Theologides A. Hepatic angiosarcoma and bronchioloalveolar carcinoma induced by Fowler's solution. *JAMA.* 1984; 252: 3407–8.
- 162 Tsai MH, Chien RN, Hsieh SY, Hung CF, Chen TC, Sheen IS. Primary hepatic angiosarcoma: report of a case involving environmental arsenic exposure. *Chang-geng Yi Xue Za Zhi.* 1998; 21: 469–74.
- 163 Luo FJ, Luo ZD, Ma L. A study on the relationship between drinking water with high arsenic content and incidence of malignant tumour in Heihe Village, western part of Huhehot, Inner Mongolia. *Zhonghua Liu Xing Bing Xue Za Zhi.* 1995; 16: 289–91.
- 164 Tsai SM, Wang TN, Ko YC. Mortality for certain diseases in areas with high levels of arsenic in drinking water. *Arch Environ Health.* 1999; 54: 186–93.
- 165 Liaw J, Marshall G, Yuan Y, Ferreccio C, Steinmaus C, Smith AH. Increased childhood liver cancer mortality and arsenic in drinking water in northern Chile. *Cancer Epidemiol Biomarkers Prev.* 2008; 17: 1982–7.
- 166 Waalkes MP, Ward JM, Liu J, Diwan BA. Transplacental carcinogenicity of inorganic arsenic in the drinking water: induction of hepatic, ovarian, pulmonary, and adrenal tumors in mice. *Toxicol Appl Pharmacol.* 2003; 186: 7–17.
- 167 Lu T, Liu J, LeCluyse EL, Zhou YS, Cheng ML, Waalkes MP. Application of cDNA microarray to the study of arsenic-induced liver diseases in the population of Guizhou, China. *Toxicol Sci.* 2001; 59: 185–92.
- 168 Lai R, Wang Y, Li X, Yu RA. Effect of selenium and arsenic on oxidative stress, DNA oxidative damage and repair in HepG2 cells. *Wei Sheng Yan Jiu.* 2008; 37: 645–8.
- 169 Guo HR, Chiang HS, Hu H, Lipsitz SR, Monson RR. Arsenic in drinking water and incidence of urinary cancers. *Epidemiology.* 1997; 8: 545–50.
- 170 Rahman MM, Chowdhury UK, Mukherjee SC, Mondal BK, Paul K, Lodh D, Biswas BK, Chanda CR, Basu GK, Saha KC, Roy S, Das R, Palit SK, Quamruzzaman Q, Chakraborti D. Chronic arsenic toxicity in Bangladesh and West Bengal, India—a review and commentary. *J Toxicol Clin Toxicol.* 2001; 39: 683–700.
- 171 Brown IR, Rush SJ. Induction of a 'stress' protein in intact mammalian organs after the intravenous administration of sodium arsenite. *Biochem Biophys Res Commun.* 1984; 120: 150–5.
- 172 Zheng XH, Watts GS, Vaught S, Gandolfi AJ. Low-level arsenite induced gene expression in HEK293 cells. *Toxicology.* 2003; 187: 39–48.
- 173 Hsueh YM, Cheng GS, Wu MM, Yu HS, Kuo TL, Chen CJ. Multiple risk factors associated with arsenic-induced skin cancer: effects of chronic liver disease and malnutritional status. *Br J Cancer.* 1995; 71: 109–14.
- 174 Brima EI, Jenkins RO, Lythgoe PR, Gault AG, Polya DA, Haris PI. Effect of fasting on the pattern of urinary arsenic excretion. *J Environ Monit.* 2007; 9: 98–103.
- 175 Vahter ME. Interactions between arsenic-induced toxicity and nutrition in early life. *J Nutr.* 2007; 137: 2798–804.
- 176 Gamble MV, Liu X, Ahsan H, Pilsner R, Ilievski V, Slavkovich V, Parvez F, Levy D, Factor-Litvak P, Graziano JH. Folate, homocysteine, and arsenic metabolism in arsenic-exposed individuals in Bangladesh. *Environ Health Perspect.* 2005; 113: 1683–8.
- 177 Gamble MV, Liu X, Ahsan H, Pilsner JR, Ilievski V, Slavkovich V, Parvez F, Chen Y, Levy D, Factor-Litvak P, Graziano JH. Folate and arsenic metabolism: a double-blind, placebo-controlled folic acid-supplementation trial in Bangladesh. *Am J Clin Nutr.* 2006; 84: 1093–101.
- 178 Heck JE, Nieves JW, Chen Y, Parvez F, Brandt-Rauf PW, Graziano JH, Slavkovich V, Howe GR, Ahsan H. Dietary intake of methionine, cysteine, and protein and urinary arsenic excretion in Bangladesh. *Environ Health Perspect.* 2009; 117: 99–104.

- 179 Pilsner JR, Liu X, Ahsan H, Ilievski V, Slavkovich V, Levy D, Factor-Litvak P, Graziano JH, Gamble MV. Genomic methylation of peripheral blood leukocyte DNA: influences of arsenic and folate in Bangladeshi adults. *Am J Clin Nutr.* 2007; 86: 1179–86.
- 180 Li L, Ekström EC, Goessler W, Lönnerdal B, Nermell B, Yunus M, Rahman A, El Arifeen S, Persson LA, Vahter M. Nutritional status has marginal influence on the metabolism of inorganic arsenic in pregnant Bangladeshi women. *Environ Health Perspect.* 2008; 116: 315–21.
- 181 Shi H, Shi X, Liu KJ. Oxidative mechanism of arsenic toxicity and carcinogenesis. *Mol Cell Biochem.* 2004; 255: 67–78.
- 182 Hei TK, Filipic M. Role of oxidative damage in the genotoxicity of arsenic. *Free Radic Biol Med.* 2004; 37: 574–81.
- 183 Avani G, Rao MV. Genotoxic effects in human lymphocytes exposed to arsenic and vitamin A. *Toxicol In Vitro.* 2007; 21: 626–31.
- 184 Avani G, Rao MV. In vitro genotoxicity assays to evaluate the role of vitamin A on arsenic in human lymphocytes. *Ecotoxicol Environ Saf.* 2009; 72: 635–8.
- 185 Karasavvas N, Cárcamo JM, Stratis G, Golde DW. Vitamin C protects HL60 and U266 cells from arsenic toxicity. *Blood.* 2005; 105: 4004–12.
- 186 Bongiovanni GA, Soria EA, Eynard AR. Effects of the plant flavonoids silymarin and quercetin on arsenite-induced oxidative stress in CHO-K1 cells. *Food Chem Toxicol.* 2007; 45: 971–6.
- 187 Yang J, Li H, Chen YY, Wang XJ, Shi GY, Hu QS, Kang XL, Lu Y, Tang XM, Guo QS, Yi J. Anthraquinones sensitize tumor cells to arsenic cytotoxicity *in vitro* and *in vivo* via reactive oxygen species-mediated dual regulation of apoptosis. *Free Radic Biol Med.* 2004; 37: 2027–41.
- 188 Chowdhury R, Dutta A, Chaudhuri SR, Sharma N, Giri AK, Chaudhuri K. *In vitro* and *in vivo* reduction of sodium arsenite induced toxicity by aqueous garlic extract. *Food Chem Toxicol.* 2008; 46: 740–51.
- 189 Sinha D, Dey S, Bhattacharya RK, Roy M. *In vitro* mitigation of arsenic toxicity by tea polyphenols in human lymphocytes. *J Environ Pathol Toxicol Oncol.* 2007; 26: 207–20.
- 190 Biswas J, Sinha D, Mukherjee S, Roy S, Siddiqi M, Roy M. Curcumin protects DNA damage in a chronically arsenic-exposed population of West Bengal. *Hum Exp Toxicol.* 2010; 29: 513–24.
- 191 Tiwari H, Rao MV. Curcumin supplementation protects from genotoxic effects of arsenic and fluoride. *Food Chem Toxicol.* 2010; 48: 1234–8.
- 192 Xu Y, Wang H, Wang Y, Zheng Y, Sun G. Effects of folate on arsenic toxicity in Chang human hepatocytes: involvement of folate antioxidant properties. *Toxicol Lett.* 2010; 195: 44–50.
- 193 Pant HH, Rao MV. Evaluation of *in vitro* anti-genotoxic potential of melatonin against arsenic and fluoride in human blood cultures. *Ecotoxicol Environ Saf.* 2010; 73: 1333–7.
- 194 Chung CJ, Pu YS, Chen YT, Su CT, Wu CC, Shiue HS, Huang CY, Hsueh YM. Protective effects of plasma alpha-tocopherols on the risk of inorganic arsenic-related urothelial carcinoma. *Sci Total Environ.* 2011; 409: 1039–45.
- 195 Sweins A. Protective effect of selenium against arsenic-induced chromosomal damage in cultured human lymphocytes. *Hereditas.* 1983; 98: 249–52.
- 196 Hu GG. Investigation of protective effect of selenium on genetic materials among workers exposed to arsenic. *Zhonghua Yu Fang Yi Xue Za Zhi.* 1989; 23: 286–8.
- 197 Mo J, Xia Y, Wade TJ, Schmitt M, Le XC, Dang R, Mumford JL. Chronic arsenic exposure and oxidative stress: OGG1 expression and arsenic exposure, nail selenium, and skin hyperkeratosis in Inner Mongolia. *Environ Health Perspect.* 2006; 114: 835–41.
- 198 Chen Y, Hall M, Graziano JH, Slavkovich V, van Geen A, Parvez F, Ahsan H. A prospective study of blood selenium levels and the risk of arsenic-related premalignant skin lesions. *Cancer Epidemiol Biomarkers Prev.* 2007; 16: 207–13.
- 199 Zeng H, Uthus EO, Combs GF. Mechanistic aspects of the interaction between selenium and arsenic. *J Inorg Biochem.* 2005; 99: 1269–74.
- 200 Huang Z, Pei Q, Sun G, Zhang S, Liang J, Gao Y, Zhang X. Low selenium status affects arsenic metabolites in an arsenic exposed population with skin lesions. *Clin Chim Acta.* 2008 ; 387: 139–44.