



Chemosystematic markers for the essential oils in leaves of *Mentha* species cultivated or growing naturally in Estonia

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Abstract. The content and composition of essential oils in leaves of *Mentha × piperita* L., *M. spicata* L., *M. arvensis* L., and *M. longifolia* (L.) Huds. from Estonia were investigated using hydrodistillation and GC–MS analyses. Some chemosystematic markers for the identification of *Mentha* species are presented. The essential oil yield of the studied species was 2.4–3.0%, 0.9–2.3%, 0.9–1.4%, and 0.7–2.5%, respectively. From the isolated essential oils, 75, 89, 67, and 69 compounds were identified and quantitatively evaluated, representing 92.2–99.5% of the total oil. The main constituents of *M. × piperita* oils were menthol (26.4–47.7%) and menthone (13.6–31.9%); the ratio of their content was usually between 1 and 3. Four chemotypes of *M. spicata* were determined: rich in carvone (62.1–67.4%), rich in piperitenone oxide (61.9%), rich in α -terpinyl acetate + *trans*- β -caryophyllene (61.9% and 11.4%, respectively), and rich in acetic acid,1-methyl-1-(4-methyl-5-oxo-cyclohex-3-enyl) ethyl ether (6.5%). The last two chemotypes were determined for the first time. Only the essential oils of *M. arvensis* were found to contain *trans*-1-octen-3-ol (3.8–7.7%) and neryl propionate (2.4% in both samples). Good chemosystematic markers for *M. arvensis* were also *trans*- (16.0–19.1%) and *cis*- β -ocimene (16.0–20.5%). The ocimene-rich chemotype was found by us for the first time. The chemosystematics of *M. longifolia* was found to be the most complicated, as many chemotypes have been determined by us and several other authors. In this study, a high concentration of carvone (51.4–57.6%) was characteristic of all analysed *M. longifolia* samples.

Key words: *Mentha × piperita*, *Mentha spicata*, *Mentha arvensis*, *Mentha longifolia*, essential oil, leaves, GC–MS.

INTRODUCTION

Several species of the genus *Mentha* are cultivated worldwide for the pharmaceutical, perfume, cosmetic, and food industries. Mints are characterized by great morphological variation, reflected in the high number of different taxonomic rank names. Recent studies conducted on the basis of morphological, cytological, and chemical markers have revealed that the genus *Mentha* consists of 18 species and includes 11 named hybrids (Krzyzanowska et al., 2011). The findings of our earlier study showed that the qualitative and quantitative content of different groups of terpenoids could be used in the chemosystematics of plant species (Orav et al., 2011a). Commercially, the most important mint species are *Mentha × piperita* L. (peppermint), *Mentha spicata* L.

(spearmint), and *Mentha arvensis* L. var. *canadensis* (cornmint) (Hajlaoui et al., 2009).

For centuries, mints have been used as a folk remedy or in alternative medicine. They have been used to treat nausea, flatulence, vomiting in pregnancy, ulcerative colitis, jaundice, bronchitis, anorexia, dental caries, rheumatism, hysteria, infantile troubles, dysmenorrhea, amenorrhea, skin diseases, and fever. In addition, they have been consumed as a vermifuge and antidote (Jagetia and Baliga, 2002; Gursoy et al., 2009; Hajlaoui et al., 2009).

According to the European Pharmacopoeia (EP), *M. × piperita* leaf as a whole drug should contain not less than 12 mL/kg (1.2%) and as a cut drug, not less than 9 mL/kg (0.9%) of essential oil (European Pharmacopoeia, 2010). The EP postulates that terpenes in *M. × piperita* essential oil used in the pharmaceutical industry in Europe should be within the following ranges:

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limonene 1.0–5.0%, cineole 3.5–14.0%, menthone 14.0–32.0%, menthofuran 1.0–9.0%, isomenthone 1.5–10.0%, methyl acetate 2.8–10.0%, isopulegol – maximum 0.2%, menthol 30.0–55.0%, pulegone – maximum 4.0%, and carvone – maximum 1.0%.

The content of *M. × piperita* oil and its composition have been widely studied also during the last three years (Schmidt et al., 2009; Sokovic et al., 2009; Bassole et al., 2010; Dolzhenko et al., 2010; Hussain et al., 2010a; Kizil et al., 2010; Zheljazkov et al., 2010b). The chief components of oil are menthone (11–46%), menthol (2–40%), isomenthone (1–16%), methyl acetate (0.3–9%), pulegone (0.1–13%), piperidone (1–6%), limonene (1–6%), 1,8-cineole (0.4–6%), etc. (Orav et al., 2004).

Also the composition of *M. spicata* essential oil has been studied well by several authors, resulting in identification of different chemotypes (Edris et al., 2003; Chauhan et al., 2009; Mkaddem et al., 2009; Sokovic et al., 2009; Hussain et al., 2010a, 2010b; Kizil et al., 2010; Koliopoulos et al., 2010; Telci et al., 2010; Zheljazkov et al., 2010a, 2010b; Šarer et al., 2011). The major components identified in the oil of *Mentha arvensis* are menthol, *p*-menthone, isomenthone (Rao, 1999; Rao et al., 1999, 2000; Pandey et al., 2003; Hussain et al., 2010a, 2010b; Prasad et al., 2010). Menthone, menthol, pulegone, menthofuran, and other terpenes have been found to be principal constituents of *Mentha longifolia* (L.) Huds. (horse mint) from different countries and their chemo-geographical variation has been described by several authors (Viljoen et al., 2006;

Asekun et al., 2007; Hajlaoui et al., 2008, 2009; Singh et al., 2008; Gursoy et al., 2009; Kakhky et al., 2009; Mkaddem et al., 2009; Hussain et al., 2010a; Koliopoulos et al., 2010).

Most *Mentha* species are characterized by great morphological variation, reflected in the high number of different taxonomic rank names. In spite of the rather stable qualitative oil composition of cultivated mints, in most wild-growing *Mentha* species a great chemical diversity is observed (Mimica-Dukic and Bozin, 2008). The findings of our earlier study showed that the qualitative and quantitative content of different groups of terpenoids could be used in the chemosystematics of plant species (Orav et al., 2011a).

The aim of the present study was to find chemosystematic markers for the identification of *Mentha* species.

EXPERIMENTAL

Plant material

The content and composition of essential oils hydro-distilled from leaves of *Mentha × piperita*, *Mentha spicata*, *Mentha arvensis*, and *Mentha longifolia* cultivated or growing naturally in Estonia was studied. All leaves of cultivated or wild *Mentha* species studied were collected from July to August 2011 from different locations in Estonia (Table 1). A herbarium of the collected plant species was made. The plant material

Table 1. *Mentha* spp. samples studied from Estonia

No.	<i>Mentha</i> spp.	Origin	Comments
1	<i>Mentha spicata</i> L.	Lääne-Viru county, Vihula rural municipality, Karepa village	Cultivated in Karepa herb farm
2	<i>Mentha spicata</i> L.	Tartu county, Võnnu rural municipality, Võnnu village	Cultivated in home garden
3	<i>Mentha spicata</i> L.	Jõgeva county, Puurmani rural municipality, Puurmani village	Cultivated in home garden
4	<i>Mentha × piperita</i> L.	Tartu county, Kambja rural municipality, Pangodi village	Cultivated in home garden
5	<i>Mentha longifolia</i> (L.) Huds.	Harju county, Kuusalu rural municipality, Pudisoo village	Cultivated in home garden
6	<i>Mentha × piperita</i> L.	Tartu county, Võnnu rural municipality, Võnnu village	Cultivated in home garden
7	<i>Mentha arvensis</i> L.	Tartu county, Võnnu rural municipality, Võnnu village	Growing wild
8	<i>Mentha arvensis</i> L.	Tartu county, Võnnu rural municipality, Võnnu village	Growing wild
9	<i>Mentha spicata</i> L.	Tartu county, Võnnu rural municipality, Võnnu village	Cultivated in home garden
10	<i>Mentha longifolia</i> (L.) Huds.	Botanical Garden of the University of Tartu	Cultivated in herb garden
11	<i>Mentha longifolia</i> (L.) Huds.	Botanical Garden of the University of Tartu	Cultivated in herb garden
12	<i>Mentha longifolia</i> (L.) Huds.	Botanical Garden of the University of Tartu	Cultivated in herb garden, different than No. 11
13	<i>Mentha × piperita</i> L.	Saare county, Saaremaa island	Cultivated by order of Vadi herb company, dried leaves obtained from pharmacy in Tartu, package labelled as ‘Peppermint leaves’
14	<i>Mentha spicata</i> L.	Tartu county, Alatskivi rural municipality, Ronisoo village	Cultivated in home garden

was dried in a dark room at room temperature ($20\pm2^\circ\text{C}$) for ten days. Each sample was labelled, packed in a paper bag, and stored in the dark at room temperature until assayed in September 2011. Mint species were identified using the herbarium by botanist Dr Ülle Reier from the Department of Botany, University of Tartu. Samples Nos 13 and 14 were obtained from retail pharmacies in Tartu, Estonia; their packages were labelled as *Menthae piperitae folium*. The voucher specimens (No. Lamiaceae/Men1-13) are deposited in the Institute of Pharmacy, University of Tartu, Estonia.

Hydrodistillation of essential oil

The essential oil was isolated from dried plant material using the distillation method described in the European Pharmacopoeia (2010). To take up the essential oil 20 g of the whole drug and 0.50 mL of xylene were used. Distillation time was 2 h at a rate of 3–4 mL/min.

Analysis of essential oils

Capillary gas chromatography (GC-FID)

The essential oils were analysed using a Chrom-5 chromatograph (Laboratori Pristroje Praha, Czech Republic) with a flame ionization detector (FID) on two fused silica capillary columns with two stationary phases: poly(5% diphenyl-95% dimethylsiloxane) (SPBTM-5, 30 m × 0.25 mm, Supelco, Switzerland) and polar polyethylene glycol (SW-10, 30 m × 0.25 mm, Supelco, Switzerland). The film thickness of both stationary phases was 0.25 µm. Carrier gas helium (purity >99.999%), with a split ratio of 1 : 150 and flow rate of 30–35 cm/s, was applied. The temperature program was from 50 to 250°C at 2°C/min, the injector temperature was 250°C. A Clarity Lite chromatography station (DataApex Ltd, Czech Republic) was used for data processing.

The components of the oils were identified by comparing their retention indices (RI) on two columns to the RI values of reference standards (from Sigma), to our RI data, and to literature data (Davies, 1990; Zenkevich, 1996, 1997, 1999). The results obtained were confirmed by GC-MS.

The percentage composition of the oils was calculated using the normalization method without correction factors. The relative standard deviation of the percentages of oil components in three repeated GC analyses of the single oil sample did not exceed 5%.

Gas chromatography/mass spectrometry (GC-MS)

GC-MS analysis of samples was carried out using a GCMS-QP2010 (Shimadzu, Japan) on a fused silica capillary column (30 m × 0.32 mm) with a bonded

stationary phase: poly(5%-diphenyl-95%-dimethyl) siloxane (ZB-5, Zebron). For further details, see Raal et al. (2010).

RESULTS AND DISCUSSION

Content of essential oils

The yield of essential oil from different *M. × piperita* samples varied in the range of 0.1–3.0% (Table 2) and corresponded to the EP standard. As a comparison, in our earlier study, peppermint cut drugs from several countries contained 0.8–3.3% essential oil (Orav et al., 2004). The content of essential oil in *M. spicata* was 0.9–2.3%, in *M. arvensis* 0.9–1.4%, and in *M. longifolia* 0.7–2.5%. The oil yield from the leaves of other mints can be similar or even higher than that of *M. × piperita*, for example 2.9–3.2% in *M. spicata* (Edris et al., 2003; Sarer et al., 2011). According to Hussain and co-authors (2010a), *M. × piperita* contains 1.1–1.2%, *M. spicata* 1.0–1.2%, *M. arvensis* 0.9–1.7%, and *M. longifolia* 0.7–1.1% of essential oil.

Previously we found much more significant differences between the minimum and maximum yields of essential oils: 17-fold in *Coriandrum sativum* (Orav et al., 2011b), 16-fold in *Levisticum officinale* (Raal et al., 2008), 14-fold in *Thymus serpyllum* (Paaver et al., 2008), and 10-fold in *Chamomilla recutita* (Orav et al., 2010), collected or obtained as commercial samples from different retail pharmacies in various countries. Over years, the variance of oil yield in *Juniperus communis* branches was 14-fold (Raal et al., 2010). Therefore, instead of the content of essential oil, the yield of terpenoids should be the basis of the chemosystematics of the genus *Mentha*. Moreover, the content of oil can be affected by several factors such as climatic and vegetation variations.

Compositon of essential oils

Up to 75, 89, 67, and 69 compounds were identified and quantified in the oils isolated from *M. × piperita*, *M. spicata*, *M. arvensis*, and *M. longifolia*, representing 92.2–99.5% of the total oil (Table 2).

The *M. × piperita* oil samples studied contained 26.4–47.7% menthol. Menthol is mentioned as the main constituent of peppermint oil also by many other authors (Stojanova et al., 2000; Rohloff et al., 2001; Pino et al., 2002; Schmidt et al., 2009; Sokovic et al., 2009; Bassole et al., 2010; Dolzhenko et al., 2010; Hussain et al., 2010a; Kizil et al., 2010; Zheljazkov et al., 2010b). In our previous study, the essential oil composition of peppermint samples sold in pharmacies was determined. Samples from Estonia ($n=2$) contained 31.6–35.8% menthol. Herbs from France, Hungary, Belgium,

Table 2. Content and composition (%) of essential oils of *Mentha* spp. Boldface highlights the main components (not less than 2%); tr – traces (<0.05%)

No.	Compound	RI	M. spicata												<i>M. spicata</i> M. × piperita <i>M. longifolia</i>		
			SPB-5	SW-10	1	2	3	4	5	6	7	8	9	10	11		
1	α -Thujene	924	1024	0.1	tr	0.1	0.1	–	0.1	0.2	0.1	0.1	0.1	tr	0.1	tr	
2	α-Pinene	927	1020	0.7	0.7	0.8	0.5	0.8	2.8	3.1	0.3	0.7	0.3	0.6	0.7	0.7	
3	Camphene	940	1069	tr	–	0.1	tr	tr	–	1.0	0.8	tr	–	tr	tr	tr	
4	Sabinene	968	1120	0.5	0.6	0.6	0.4	0.7	1.3	0.5	1.7	0.5	0.2	0.4	0.4	0.6	
5	β-Pinene	970	1107	1.0	1.0	1.2	1.2	0.7	1.3	9.1	13.0	0.5	0.8	0.3	0.7	1.1	0.9
6	1-Octen-3-ol	980	1450	tr	–	0.1	0.1	–	–	2.8	4.7	tr	–	–	tr	–	
7	Myrcene	990	1165	2.3	2.4	5.5	0.4	0.6	0.3	2.2	1.8	5.8	0.9	0.6	0.6	0.3	3.3
8	3-Octanol	995	1398	0.9	0.4	0.3	0.2	0.5	0.4	1.4	1.3	–	0.6	0.5	1.1	0.2	0.5
9	α -Phellandrene	1000	1068	0.1	0.1	–	–	0.1	–	tr	–	0.1	0.1	–	0.1	–	0.2
10	Isobutyl isovalerate	1005	1195	–	–	0.1	–	–	–	–	–	0.1	–	–	–	–	–
11	α -Terpinene	1012	1177	tr	–	0.1	0.2	0.1	0.2	0.1	0.1	0.1	tr	0.1	–	0.2	–
12	<i>p</i> -Cymene	1020	1264	–	–	0.1	–	0.1	–	tr	–	tr	–	–	0.1	–	–
13	<i>trans</i> -2-Heptenyl acetate	1020	1405	–	–	–	–	–	–	0.1	0.3	–	–	–	–	–	–
14	Limonene	1025	1200	10.4	11.8	1.3	8.9	10.3	9.8	0.7	0.5	16.2	20.5	1.3	0.6	8.2	2.6
15	1,8-Cineole	1028	1209	5.8	1.6	0.7	0.2	0.6	7.6	7.9	4.1	–	–	1.5	4.8	2.8	13.9
16	<i>cis</i>-β-Ocimene	1035	1237	0.5	0.7	1.6	0.7	0.2	16.2	20.5	1.3	0.4	0.3	0.2	0.4	0.8	–
17	<i>n</i> -Butyl-2-methyl butanoate	1037	1240	0.1	0.1	0.1	0.1	0.1	–	–	0.2	0.1	–	0.1	tr	–	
18	<i>trans</i>-β-Ocimene	1044	1250	0.2	0.3	0.2	0.2	0.1	16.0	19.1	0.5	0.1	0.1	0.1	0.1	0.3	–
19	γ -Terpinene	1052	1243	0.1	0.1	0.1	0.3	0.1	0.4	0.3	0.2	0.2	tr	–	tr	0.3	–
20	2-Methyl benzaldehyde	1058	1613	–	0.5	–	–	–	–	0.7	–	–	–	–	–	–	–
21	<i>trans</i> -2-Octen-1-ol	1058	1588	0.2	0.2	–	0.1	0.3	0.1	0.5	0.3	–	1.0	0.1	0.2	0.1	0.4
22	<i>cis</i>-Sabinene hydrate	1063	1466	1.3	0.1	1.1	2.6	0.1	2.7	–	–	0.2	0.1	–	0.1	3.2	tr
23	Unidentified 1	1063	1558	–	–	–	–	–	–	0.2	1.3	–	–	–	–	–	–
24	Terpinolene	1083	1274	0.1	0.1	0.2	0.1	0.1	0.1	0.2	0.2	0.3	0.1	–	0.1	0.1	0.1
25	<i>trans</i> -Sabinene hydrate	1098	1540	–	–	0.3	–	0.5	–	–	–	–	–	–	–	0.7	–
26	Linalool	1100	1554	0.1	tr	0.4	–	tr	–	0.5	0.4	tr	–	–	tr	–	tr
27	Isoamyl isovalerate	1102	1300	tr	tr	tr	–	0.1	–	tr	0.3	0.1	0.1	0.2	–	0.1	0.1
28	Benzyl alcohol	1105	–	–	tr	–	–	tr	–	tr	0.1	–	–	–	–	–	–
29	Amyl isovalerate	1106	1300	–	–	0.1	0.4	–	0.2	–	0.4	0.1	0.2	–	0.1	0.1	0.1
30	Octen-1-ol acetate	1113	1373	–	–	–	–	–	–	–	–	–	–	2.7	–	–	–

Table 2. *Continued*

Continued overleaf

Table 2. Continued

No.	Compound	RI	Sample number													
			SPB-5	SW-10	M. spicata	M. × piperita	M. longifolia	M. arvensis	M. spicata	M. × piperita	M. longifolia	M. arvensis	M. spicata	M. × piperita	M. longifolia	M. arvensis
59	cis-Carveol	1227	1844	—	—	—	—	—	—	—	—	—	—	—	—	—
60	cis-3-Hexenyl valerate	1235	1512	—	—	0.6	—	—	0.9	0.7	0.5	—	—	—	—	—
61	Pulegone	1236	1635	—	—	2.2	0.2	—	—	—	—	—	—	—	0.5	—
62	Carvone	1238	1730	67.4	62.1	—	57.6	—	tr	0.1	2.3	56.8	51.4	56.4	0.2	66.5
63	Hexyl isovalerate	1246	1575	—	tr	—	—	0.1	0.1	0.1	0.3	—	—	—	—	—
64	Piperitone	1250	1715	0.4	0.4	0.7	0.7	0.1	0.2	—	2.7	2.4	1.4	0.8	0.3	—
65	Methyl citronellate	1260	1588	—	—	—	—	—	0.2	—	—	—	—	—	—	—
66	p-Mentha-(7),8(10)-dien-9-ol	1266	1266	0.1	0.2	—	—	0.2	—	—	0.3	0.1	—	0.1	—	0.1
67	Piperitone oxide	1273	1716	0.1	0.1	0.2	—	0.2	—	—	—	1.2	0.8	0.1	0.1	0.1
68	1-Decanol	1273	1770	—	—	—	—	2.2	1.5	—	—	—	—	—	—	—
69	Anethole	1280	1827	—	—	—	—	0.1	—	—	—	—	—	—	—	0.1
70	Dihydrodulcan I	1280	1506	—	—	0.3	0.1	—	—	—	0.9	—	—	—	0.2	—
71	Neomenthyl acetate	1282	1629	0.1	0.1	—	0.1	0.1	0.2	—	tr	—	—	0.1	0.1	—
72	Bornyl acetate	1282	1575	—	tr	—	—	—	tr	—	tr	—	—	—	—	—
73	4-Terpinenyl acetate	1284	1625	—	—	—	—	—	tr	—	0.4	—	—	—	—	—
74	Dihydrodulcan II	1284	1540	—	—	0.5	—	tr	—	—	—	—	—	—	—	—
75	Perillyl alcohol	1287	1890	0.1	0.1	—	tr	0.2	—	—	0.4	—	—	tr	—	tr
76	Menthyl acetate	1290	1558	tr	tr	0.6	0.1	0.9	—	—	—	—	—	tr	0.6	0.1
77	Isomenthyl acetate	1296	1566	—	—	0.1	—	0.1	—	—	—	—	—	—	0.2	—
78	Dihydrocaryyl acetate	1304	1563	—	0.2	—	—	—	—	—	0.2	0.2	—	tr	0.1	—
79	n-Nonyl acetate	1311	1580	—	—	—	—	—	—	0.1	—	—	—	—	—	—
80	β-Terpinyl acetate	1311	1648	—	—	—	—	—	—	—	—	0.5	—	—	—	—
81	Myrcenyl acetate	1315	1690	—	—	0.1	—	—	—	—	—	0.7	—	—	—	—
82	Isodihydrocaryyl acetate	1326	1642	tr	1.4	—	—	0.9	—	—	0.4	0.1	—	0.3	—	0.1
83	trans-Caryyl acetate	1335	1730	0.1	0.2	—	—	0.1	—	—	—	—	—	—	0.3	0.1
84	δ-Elemene	1335	1477	—	—	0.1	—	—	—	—	—	—	—	—	—	—
85	α-Terpinyl acetate	1345	1695	0.1	—	—	—	—	—	—	—	—	—	32.3	—	—
86	Eugenol	1354	2165	—	—	—	—	—	—	—	—	0.4	—	—	—	—
87	α-Cubebene	1354	1449	—	—	—	—	—	—	—	0.1	—	—	—	0.1	—
88	Piperitenone oxide	1358	1957	—	—	—	—	—	—	—	—	—	—	—	—	—
89	cis-Caryyl acetate	1360	1748	tr	0.3	—	—	0.1	0.3	—	—	0.7	—	0.1	0.1	0.1
90	α-Copaene	1367	1480	0.1	tr	—	—	0.1	—	0.1	0.1	0.3	0.1	0.1	—	0.1
91	Geranyl acetate	1370	1758	—	—	—	—	0.1	—	tr	—	—	—	—	—	—

Table 2. *Continued*

Continued overleaf

Table 2. Continued

No.	Compound	RI		Sample number													
		SPB-5	SW-10	M. spicata	M. × piperita	M. arvensis	M. spicata	M. longifolia	M. × piperita	M. arvensis	M. spicata	M. longifolia	M. × piperita	M. spicata	M. longifolia	M. × piperita	M. spicata
120	Caryophyllene oxide	1570	1962	0.1	0.1	0.2	0.1	0.2	0.2	0.4	0.6	0.7	0.2	0.2	0.1	0.1	—
121	Ethyl dodecanoate	1582	1830	—	—	—	—	0.3	0.2	—	—	—	—	—	—	—	—
122	Epiglobulol	1584	2080	0.2	—	0.7	0.7	0.6	—	—	—	—	0.9	1.1	0.1	—	—
123	Germancene D-4-ol	1595	2052	0.1	—	tr	—	0.5	—	—	—	—	—	0.2	—	—	—
124	Humulene epoxide I	1604	2015	—	—	—	—	—	0.8	0.3	—	—	—	—	—	—	—
125	Cubenol	1606	2000	—	—	0.2	—	—	—	—	—	—	—	—	—	—	—
126	cis-3-Hexenyl phenyl acetate	1628	2221	—	—	—	—	—	0.8	0.6	—	—	—	—	—	—	—
127	Bicyclo[3.1.1]hept-2-en-4-ol, 2,6,6-trimethyl hexanoate	1630	2241	—	—	—	—	—	—	0.5	—	—	—	—	—	—	—
128	δ-Cadinol	1634	2161	tr	—	0.1	0.1	0.3	tr	—	0.1	—	0.2	tr	0.2	0.1	tr
129	α-Cadinol	1643	2218	0.2	0.2	0.1	0.4	tr	—	—	0.5	0.3	0.4	0.2	0.1	0.1	tr
130	Isovaleryl decanoate	1645	2181	—	—	—	—	—	—	0.2	0.1	—	—	—	—	—	—
131	Selina-6-en-8-ol	1677	2267	0.1	0.1	—	—	0.2	—	—	—	0.3	0.2	0.4	0.1	—	0.1
132	Unidentified 3, MW 254	1760	2344	—	—	—	—	—	—	0.4	0.1	—	—	—	—	—	—
133	Hexahydrofarnesyl acetone	1840	2087	—	—	—	—	0.1	—	—	—	—	—	—	—	—	—
134	Palmitic acid	1972	2920	—	—	—	—	—	—	—	—	—	—	—	—	—	0.1
Total, %		98.7	97.9	97.4	99.5	96.8	98.2	98.0	98.1	97.0	99.2	95.2	98.1	99.4	98.7	2.5	3.0
Content of essential oil, %		2.1	2.1	0.9	3.0	1.4	2.4	0.9	1.4	1.1	2.2	0.7	2.5	3.0	2.5	2.3	2.3

See Table 1 for number and origin of samples.
Mass spectral data of unidentified compounds:

- (1) – m/z: 150(100), 135(85), 91(52), 79(45), 107(32), 41(30), 96(27), 82(22)
- (2) – m/z: 161(100), 105(50), 91(42), 81(25), 79(23), 55(20), 41(20), 93(18), 120(16), 133(15)
- (3) – m/z: 114(100), 57(70), 43(60), 41(43), 66(40), 67(35), 71(30), 85(25)

— Not found.

Ukraine, Greece, and Russia contained menthol respectively 31.3%, 37.2%, 17.6%, 24.4%, 39.5%, and 1.5% (Orav et al., 2004). Thus, it may be concluded that other mint species are sometimes sold as *M. × piperita*.

Menthone, the second main constituent of *M. × piperita* oil, can also be used in the chemosystematics of peppermint. The oil of the *M. × piperita* sample from Pangodi village contained menthone even above the EP limit (32%). Menthone and menthol were the typical components of peppermint oils and can be used as chemical markers. The ratio between their content according to our current study was 0.05–3.4, but usually 1–3; the same ratio was found in our previous (Orav et al., 2004) and other studies (Pino et al., 2002; Schmidt et al., 2009; Sokovic et al., 2009). According to Rohloff and co-authors (2001), changes in menthone–menthol metabolism are related to reduced menthone content from the vegetative stage to full bloom.

Four chemotypes of *M. spicata* were found: rich in carvone (samples 1, 2, and 14; see Table 2), rich in piperitenone oxide (sample 3), rich in α -terpinyl acetate and *trans*- β -caryophyllene (both in sample 9). Acetic acid,1-methyl-1-(4-methyl-5-oxo-cyclohex-3-enyl) ethyl ether was determined only in sample 9. The studied oils of *M. × piperita* contained only low concentrations of carvone, piperitone oxide, and germacrene D, while α -terpinyl acetate was not present in these oils. Carvone was found to be one of the main constituents of *M. spicata* oils also in other studies: carvone alone or carvone and limonene by Edris et al. (2003), Chauhan et al. (2010), Hussain et al. (2010a, 2010b), Zheljazkov et al. (2010a); carvone and 1,8-cineole by Mkaddem et al. (2009) and Sarer et al. (2011). In addition, piperitone oxide and limonene (Koliopoulos et al., 2010) or pulegone and piperitone (Telci et al., 2010) were established as their main constituents. Only traces of menthone and menthol were present to a maximum of 0.1% in the spearmint oils analysed by us. However, *M. spicata* oil may sometimes contain remarkably high amounts of menthone or menthol but not both simultaneously. Hence, we found two new chemotypes of spearmint: rich in α -terpinyl acetate + *trans*- β -caryophyllene and rich in acetic acid,1-methyl-1-(4-methyl-5-oxo-cyclohex-3-enyl) ethyl ether. In addition, the *M. spicata* oils were found to contain more myrcene than other mint oils studied by us.

The presence of *trans*-1-octen-3-ol and neryl propionate was specific only to essential oils from *M. arvensis* (Table 2). Good chemosystematic markers for *M. arvensis* were also *trans*- and *cis*- β -ocimene, both being present in the concentration of 16% and more. This mint species contained also significantly more β -pinene and α -pinene, as well as 1-octen-3-ol than other mint species studied. In other studies (Rao, 1999; Rao et al., 1999; Pandey et al., 2003; Hussain et al., 2010a; Pandey et al., 2010; Prasad et al., 2010)

menthol was determined as a principal compound of *M. arvensis* oil but *cis*- β -ocimene was not found. The ocimenes-rich *M. arvensis* chemotype was determined by us for the first time.

The present study showed that carvone should not be considered as a specific chemosystematic marker for *M. spicata* carvone-chemotype because carvone was found also in the oils of *M. longifolia*, whose chemosystematics is most complicated. In two samples out of four (Nos 10 and 11, collected from the same place) rather high concentrations of germacrene D with α -muurolene, limonene with 1,8-cineole (samples 5, 10, and 12), and *trans*- β -caryophyllene (samples 10 and 11) and less *trans*-dihydrocarvone (samples 5 and 12) and piperitone (samples 10 and 11) were found. Other authors identified the mentofuran-rich (Viljoen et al., 2006), piperitone oxide-rich (Hussain et al., 2010a), *cis*-piperitone oxide and piperitenone oxide-rich (Viljoen et al., 2006; Singh et al., 2008; Kakhky et al., 2009), menthone-rich (Asekun et al., 2007), menthone, menthol, and pulegone-rich (Hajlaoui et al., 2008), pulegone-rich (Mkaddem et al., 2009), carvone and limonene-rich, piperitenone oxide and 1,8-cineole-rich (Koliopoulos et al., 2010) chemotypes of *M. longifolia*. The oils of *M. spicata* contained much more myrcene (2.3–5.5%) than *M. longifolia* (0.6–0.9%). Quite large amounts of menthofuran were found in *M. × piperita* oils and a very low concentration in one *M. longifolia* sample (No. 11) from four. Hence, the chemosystematics of *M. longifolia* based on the terpenes composition seems to be rather complicated. Phenolic compounds may probably be better chemical markers for *M. longifolia* and *M. × piperita*. Krzyzanowska and co-authors (2011) showed that all their samples of *M. × piperita* had higher concentrations of ester of caffeic acid and 3,4-dihydroxyphenyllactic acid than *M. longifolia* samples.

The main results of our study are presented in Table 3.

Table 3. Some chemosystematic markers of *Mentha* species

<i>Mentha</i> species	Chemosystematic markers
<i>Mentha × piperita</i>	High concentration of both menthol and menthone, usually their ratio between 1–3. No α -terpinyl acetate
<i>Mentha spicata</i>	High chemical diversity of terpenes. Sometimes contain remarkably high amounts of menthone or menthol but not simultaneously both. 2–6% myrcene
<i>Mentha arvensis</i>	Presence of <i>trans</i> -1-octen-3-ol and neryl propionate. Rather high concentration of <i>trans</i> - and <i>cis</i> - β -ocimene. Significant amounts of β -pinene, α -pinene, and 1-octen-3-ol
<i>Mentha longifolia</i>	High chemical diversity of terpenes. 1–3% of piperitone. No germacrene D, 1-caffeoic acid, and 3,4-dihydroxyphenyllactic acid

CONCLUSION

The content and composition of essential oils hydro-distilled from four *Mentha* species cultivated or growing naturally in Estonia were studied. Some specific chemosystematic markers for their identification are presented. The markers found in the oils studied by us should also be evaluated in a larger number of mint samples and in other mint species.

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Eestis kultiveeritavate ja looduslikult kasvavate *Mentha* liikide lehtede eeterlike õlide kemosüstemaatilised markerid

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On uuritud Eestist pärieva piparmündi (*Mentha × piperita*), rohemündi (*M. spicata*), põldmündi (*M. arvensis*) ja pikalehise mündi (*M. longifolia*) vesidestillatsioonil eraldatud eeterliku õli sisaldust ning GC-MS-meetodil selle koostist. On esitatud mõningad kemosüstemaatilised markerid erinevate mündiliikide identifitseerimiseks. Eeterliku õli sisaldus uuritud mündiliikides oli vastavalt 2,4–3,0%, 0,9–2,3%, 0,9–1,4% ja 0,7–2,5%. Nimetatud liikide eeterlikes õlides identifitseeriti ja määratati kvantitatiivne sisaldus vastavalt 75, 89, 67 ning 69 ühendil, mis moodustasid 92,2–99,5% õlide koguhulgast. *Mentha × piperita* eeterliku õli põhikomponendid olid mentool (26,4–47,7%) ja mentoon (13,6–31,9%); nende sisalduse suhe oli tavaliselt vahemikus 1–3. Tehti kindlaks neli *M. spicata* kemotüüpi: karvooni (62,1–67,4%), piperitenoonoksiidi (61,9%), α -terpinüülataadi + *trans*- β -kariofüleeni (vastavalt 61,9% ja 11,4%) ning äädihkappe,1-metüül-1-(4-metüül-5-okso-tsükloheksa-3-enüül)etüüleetri (6,5%) kemotüüp, kaks nendest esmakordsest. *Trans*-1-okteen-3-ooli (3,8–7,7%) ja nerüülpropionaatti (2,4% mõlemas proovis) leiti ainult *M. arvensis*'e eeterlikust õlist. Liik *M. arvensis*'e jaoks on headeks kemosüstemaatistikaks markeriteks ka *trans*- (16,0–19,1%) ja *cis*- β -otsimeen (16,0–20,5%), kusjuures otsimeeni kemotüüp on leitud esmakordsest meie poolt. Kõige komplitseeritud on *M. longifolia* kemosüstemaatika, nii meil kui ka kirjanduses on andmeid paljudest selle taimeliigi kemotüüpidest. Kõikidele *M. longifolia* uuritud proovidele oli iseloomulik suur karvoonisaldus (51,4–57,6%).