

## RESEARCH NOTE

# High frequency of *BRCA1/2* and *p53* somatic inactivation in sporadic ovarian cancer

MICHAL ZIKAN<sup>1,2\*</sup>, MARKETA JANATOVA<sup>1</sup>, DAVID PAVLISTA<sup>2</sup> and PETR POHLREICH<sup>1</sup>

<sup>1</sup>Department of Biochemistry and Experimental Oncology, 1st Faculty of Medicine, Charles University, Prague, U Nemocnice 5, 128 53 Prague 2, Czech Republic

<sup>2</sup>Clinic of Obstetrics and Gynaecology, 1st Faculty of Medicine, Charles University and General Teaching Hospital, Prague, Apolinarska 18, 128 53 Prague 2, Czech Republic

## Introduction

Our knowledge about the role of *BRCA* genes in hereditary breast and ovarian cancer is rapidly expanding (Zikan *et al.* 2005). However, we know very little about their role in sporadic breast ovarian cancer. In epithelial ovarian cancer, allelic losses of the *BRCA1* gene are found in 23%-44% and of *BRCA2* in 25%-40% of samples, respectively (Gras *et al.* 2001; Geisler *et al.* 2002; Hilton *et al.* 2002; McCoy *et al.* 2003). Somatic mutations of *BRCA1/2* genes are typically extremely rare, but interestingly are found especially in clear cell carcinomas (Takahashi *et al.* 1996; Hilton *et al.* 2002). Other possible means of somatic inactivation of *BRCA* genes have been described: occasionally through gross genomic rearrangements, but most commonly by hypermethylation of promotor sequences or related (or independent) reduced expression of *BRCA* genes (Esteller *et al.* 2000; Miyamoto *et al.* 2002).

Here, we report on a study of the frequency and mechanisms of *BRCA1*, *BRCA2* and *p53* inactivation in 30 sporadic epithelial ovarian cancer samples using intragenic and flanking microsatellite markers. We examined loss of heterozygosity (LOH) for each gene, as well as the presence of somatic mutations and loss of expression, and attempted to relate the results to tumour histotype.

## Materials and methods

### Tumour samples

For analysis, we used blood and tissue of fresh-frozen tumour samples of ovarian epithelial cancer patients operated upon, at the Clinic of Obstetrics and Gynecology, First

Faculty of Medicine, Charles University, Prague, and the General Teaching Hospital in Prague. All analysed tumours were sporadic i.e. no other tumours were referred to in the family history and all the patients were more than 40 years old at the time of diagnosis. All participants gave their written informed consent.

All freshly frozen samples were examined by a pathologist to evaluate the proportion of tumourous and nontumourous tissue. Only samples with over 50% tumourous tissue were used for our study. The histopathological characteristics of analysed tumours are given in table 1. Majority of patients (21/30 : 70%) were diagnosed in stage III International Federation of Gynecology and Obstetrics (FIGO). Stages I and IV were similarly represented (4/30 and 5/30: 13.3% and 16.7%, respectively), and none of the analysed tumours belong to stage II patient.

**Table 1.** Histopathological characterisation of examined tumours.

serous carcinoma	17/30 (56.7%)
G1	1
G2	5
G3-4	11
mucinous carcinoma	6/30 (20%)
G1	4
G2	1
G3-4	1
anaplastic	5/30 (16.6%)
G3-4	5
endometrioid carcinoma	2/30 (6.7%)
G1	0
G2	1
G3-4	1

\*For correspondence. E-mail: michal.zikan@lf1.cuni.cz.

**Keywords.** *BRCA1*; *BRCA2*; *p53*; sporadic ovarian cancer; loss of heterozygosity.

### Genetic material

DNA and RNA were isolated from freshly frozen tumour tissue and from peripheral venous blood by standard procedures, described in detail by Pohlreich *et al.* (2003). DNA from blood was used as a reference DNA for LOH experiments. In addition, mutation analysis of both *BRCA* genes was carried out on these blood samples to confirm the sporadic origin of analysed tumours. No hereditary mutations was found in the analysis of DNA from blood in these samples.

### LOH analysis

We used intragenic or flanking microsatellite markers. Three markers of each for *BRCA1* (D17S855, D17S1322, D17S1323) and *BRCA2* (D13S1695, D13S1699, D13S1701) region and two markers for *p53* region (TP01, TP02) were used. LOH was evaluated visually by comparison of alleles from tumour and blood, or by area under curve (AUC) computation using the formula  $(A_t \times B_n) / (B_t \times A_n)$ , where A and B are alleles, and t and n indicate tumorous and normal tissue, respectively. LOH was considered to be found if intensity of one allele in tumour was lower than 50% of normal (blood) tissue allele intensity. Microsatellite fragments were amplified by PCR and then analysed by polyacrylamide Spreadex gel electrophoresis as described in Janatova *et al.* (2003).

## Results and discussion

### LOH analysis

We analysed LOH in three tumour-suppressor genes — *BRCA1*, *BRCA2* and *p53* — using intragenic and flanking microsatellite markers. Markers heterozygous for appropriate patient were set as informative, homozygous ones as non-informative. Total heterozygosity (informativeness) ranged between 90.0% and 96.7% for the three gene regions studied.

LOH in *BRCA1* region was found in 11 out of 29 informative tumours (37.9%). Histological types were represented more or less equally in this subgroup: serous carcinoma: 4 (36.4%); mucinous carcinoma: 3 (27.2%); and anaplastic carcinoma: 4 (36.4%).

In *BRCA2* region, LOH was found in 10 out of 27 informative tumours (37.0%): 2 serous (20.0%); 4 mucinous (40.0%); 3 anaplastic (30.0%); and 1 endometroid (10.0%).

In more than half of the informative tumours (16/29: 55.1%), LOH in *p53* locus was detected, and the proportion of three major carcinoma histotypes was roughly equal: 6 serous (37.4%); 5 mucinous (31.3%); and 5 anaplastic (31.3%). There was no endometrioid carcinoma expressing LOH in *p53* (table 2).

**Table 2.** Allelic losse in informative tumours with respect to tumour histotype.

Tumour histotype	LOH		
	<i>BRCA1</i>	<i>BRCA2</i>	<i>p53</i>
serous	4/11(36.4%)	2/10 (20.0%)	6/16 (37.4%)
mucinous	3/11(27.2%)	4/10 (40.0%)	5/16 (31.3%)
anaplastic	4/11 (36.4%)	3/10 (30.0%)	5/16 (31.3%)
endometrioid	0	1/10 (10.0%)	0

In sum, out of 30 tumours at least one region of LOH was found in 18 tumours (60.0%). LOH in a sole region was present in 5 out of 18 tumours (27.8%): thrice in *p53* and twice in *BRCA*. LOH of *BRCA2* and *p53* were observed in conjunction in 2 out of 18 tumours (11.1%). Interestingly, LOH in *BRCA1* region was never present solely, but only in conjunction with either *p53* LOH (5/18 tumours: 27.8%) or *p53* and *BRCA2* LOH (6/18 tumours: 33.3%). And, in contrast, no tumours with only *BRCA1* LOH along with *BRCA2* LOH were found (table 3). These results suggest that it might be worthwhile to study the inactivation succession of such genes in sporadic ovarian epithelial tumours.

### Somatic mutations analysis

Standard mutation analysis of coding and exon-flanking sequences of *BRCA1* and *BRCA2* genes were done in tumour samples with LOH in at least one of these gene regions. Neither somatic nor hereditary mutations were found.

Considering the rarity of high frequency of repetitive sequences in both *BRCA1* and *BRCA2* regions, it is thought that gross genomic rearrangements due to inadequate homologous recombination repair are most often the events leading to somatic inactivation of these genes (Szabo and King 1997). However, such events would require complex techniques based on RNA or on MLPA methods for their detection.

**Table 3.** Conjunction of allelic losses and tumours divided to subgroups due to LOH pattern.

LOH regions	Number of tumours	Percentage of tumours with LOH	Percentage of all analysed tumours
<i>BRCA1 + BRCA2 + p53</i>	6	33.3%	20.0%
<i>BRCA1 + p53</i>	5	27.8%	16.7%
<i>BRCA2 + p53</i>	2	11.1%	6.7%
<i>BRCA2</i>	2	11.1%	6.7%
<i>p53</i>	3	16.7%	10.0%

#### **Loss of expression analysis**

We examined expression of *BRCA1* and *BRCA2* genes by RT-PCR (reverse transcription PCR). Two tumours (6.7%) lacked expression of *BRCA1* gene. In one of them, no LOH was found in *BRCA1* gene region. However, in the other tumour, allelic loss was present in all the three examined microsatellite markers for *BRCA1*, suggesting that alleles are inactivated in different ways in these two cases: by allelic loss (expressed as LOH), and lack of expression, without change in the genomic sequence.

One tumour (3.3%) lacked expression of *BRCA2* gene. This tumour was heterozygous for all three *BRCA2* microsatellite markers, but no LOH was detected in *BRCA2* region. Promoter hypermethylation is thought to be a cause of lack of expression (Hilton *et al.* 2002), and such epigenetic changes are extremely important for modulation of expression of many genes during cell differentiation as well as during tumour transformation (Chan *et al.* 2002; Verma and Srivastava 2002).

#### **Conclusion**

Research of *BRCA1/2* genes role in sporadic ovarian cancer is still beginning. As our data and those from literature cited here suggest, these genes (or their defects) appear to be important in approximately one-third of sporadic ovarian epithelial cancer cases. Consequently, subsequent studies need to focus on how these tumours differ in biological features or in response to therapy.

#### **Acknowledgements**

This work was supported by the Research Project of the Ministry of Education, Youth and Sports of the Czech Republic No. MSM0021620808.

#### **References**

- Chan K. Y. K., Ozcelik H., Cheung A. N. Y., Ngan H. Y. S. and Khoo U. S. 2002 Epigenetic factors controlling the *BRCA1* and *BRCA2* genes in sporadic ovarian cancer. *Cancer Res.* **62**, 4151–4156.
- Esteller M., Silva J. M., Dominguez G., Bonilla F., Matias-Guiu X., Lerma E. *et al.* 2000 Promoter hypermethylation and *BRCA1* inactivation in sporadic breast and ovarian tumours. *J. Natl. Cancer Inst.* **92**, 564–569.
- Geisler J. P., Rathe J. A., Hattermann-Zogg M. A. and Buller R. E. 2002 *BRCA1* dysfunction is common in ovarian cancer. *J. Natl. Cancer Inst.* **94**, 61–67.
- Gras E., Cortes J., Diez O., Alonso C., Matias-Guiu X. and Prat J. 2001 Loss of heterozygosity on chromosome 13q12-q14, *BRCA2* mutations and lack of *BRCA2* promoter hypermethylation in sporadic epithelial ovarian tumours. *Cancer* **92**, 787–795.
- Hilton J. L., Geisler J. P., Rathe J. A., Hattermann-Zogg M. A., DeYoung B. and Buller R. E. 2002 Inactivation of *BRCA1* and *BRCA2* in ovarian cancer. *J. Natl. Cancer Inst.* **94**, 1396–1406.
- Janatova M., Pohlreich P. and Matous B. 2003 Detection of the most frequent mutations in *BRCA1* gene on polyacrylamide gels containing Spreadex Polymer NAB. *Neoplasma* **50**, 246–250.
- McCoy M. L., Mueller C. R. and Roskelley C. D. 2003 The role of the breast cancer susceptibility gene 1 (*BRCA1*) in sporadic epithelial ovarian cancer. *Reprod. Biol. Endocrin.* **1**, 72.
- Miyamoto K., Fukutomi T., Asada K., Wakazono K., Tsuda H., Asahara T. *et al.* 2002 Promoter hypermethylation and post-transcriptional mechanisms for reduced *BRCA1* immunoreactivity in sporadic human breast cancers. *Jpn. J. Clin. Oncol.* **32**, 79–84.
- Pohlreich P., Stribrna J., Kleibl Z., Zikan M., Kalbacova R., Petrazelka L. and Konopasek B. 2003 Mutations of the *BRCA1* gene in hereditary breast and ovarian cancer in the czech Republic. *Med. Princ. Pract.* **12(1)**, 23–29.
- Szabo C. L. and King M. C. 1997 Population genetics of *BRCA1* and *BRCA2*. *Am. J. Hum. Genet.* **60**, 1013–1020.
- Takahashi H., Chiu H. C., Bandera C. A., Behbakht K., Liu P. C. and Cough F. J. 1996 Mutations of the *BRCA2* gene in ovarian carcinomas. *Cancer Res.* **56**, 2738–2741.
- Verma M. and Srivastava S. 2002 Epigenetics in cancer: implications for early detection and prevention. *Lancet Oncol.* **3**, 755–763.
- Zikan M., Pohlreich P. and Stribrna J. 2005 Mutational analysis of the *BRCA1* gene in 30 Czech ovarian cancer patients. *J. Genet.* **84**, 63–67.

Received 21 July 2006