

Hypothesis

Endogenous nitric oxide synthase inhibitors are responsible for the L-arginine paradox

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Abstract L-Arginine, the substrate of nitric oxide (NO) synthases (NOSs), is found in the mammalian organism at concentrations by far exceeding K_M values of these enzymes. Therefore, additional L-arginine should not enhance NO formation. In vivo, however, increasing L-arginine concentration in plasma has been shown repeatedly to increase NO production. This phenomenon has been named the L-arginine paradox; it has found no satisfactory explanation so far. In the present work, evidence for the hypothesis that the endogenous NOS inhibitors methylarginines, asymmetric dimethylarginine being the most powerful (IC_{50} 1.5 μ M), are responsible for the L-arginine paradox is presented. © 2000 Federation of European Biochemical Societies. Published by Elsevier Science B.V. All rights reserved.

Key words: L-Arginine paradox; Nitric oxide synthase; Endogenous inhibitor; Methylarginine; Asymmetric dimethylarginine

1. Introduction

L-Arginine is the exclusive physiological substrate for various isoforms of the nitric oxide (NO) synthases (NOSs) family (EC 1.14.13.39) which catalyzes the oxidation of L-arginine to NO and L-citrulline [1]. At least three isozymes have been identified for NO production from L-arginine [2]. These isozymes have been classified as isoform I in neuronal (nNOS) and epithelial cells, isoform II in cytokine-induced cells (iNOS) and isoform III in endothelial cells (eNOS). nNOS and eNOS are dependent on and iNOS is independent of Ca^{2+} [2]. Half-saturating L-arginine concentrations (K_M) were reported as 1.4–2.2 μ M for nNOS, 2.8–32.3 μ M for iNOS and 2.9 μ M for eNOS [2]. L-Arginine is supplied to cells by a y^+ transport system specific for cationic amino acids [3]. Freshly isolated endothelial cells have been found to contain up to 2 mM L-arginine [4]. Considering this and a K_M of 2.9 μ M for L-arginine for eNOS [2], this enzyme should be saturated in endothelial cells. It is surprising that intravenous (i.v.) or oral supplementation of L-arginine in vivo in humans augments endothelial NO production [5–18]. Supplementation of

L-arginine to hypercholesterolemic animals [5,6] and humans [7,8], in which endothelium-dependent vasodilatation is impaired, was found to improve endothelial dysfunction by increasing NO production [5–8]. Also, supplementation of L-arginine. This phenomenon, generally known as the L-arginine paradox, has found no satisfactory explanation so far. We here present evidence that concentrations of endogenous NOS inhibitors in vivo provide a satisfactory explanation for this phenomenon.

2. Proposal

The L-arginine paradox may be solved by proposing that NOS isoforms are potently inhibited in vivo and in vitro in intact cells by endogenously produced compounds. Mechanism of inhibition, inhibitor potency and intracellular concentrations of inhibitors, L-arginine and cofactors including Ca^{2+} regulate NOS activity and consequently NO production in cells capable of synthesizing NOS. Under physiological conditions, the enzyme activities of NOS isoforms are lowered to a fraction of their maximum activities (V_{max}) although the enzymes are exposed to concentrations of L-arginine which theoretically should allow the enzymes to operate at the V_{max} values of the uninhibited enzymes. Under pathological conditions, increased intracellular concentrations of the inhibitors cause additional decreases of the activity of the NOS enzymes which result in NO formation rates below the physiological levels. Under physiological and pathophysiological conditions, i.v. or oral administration of L-arginine causes an increase in circulating L-arginine concentrations which leads to an increase in intracellular L-arginine concentrations and exchange of intracellular inhibitors against extracellular L-arginine via the y^+ transport system in NOS producing cells. Antagonization of L-arginine with competitive inhibitors and decrease of intracellular concentrations of competitive and non-competitive inhibitors by extracellular L-arginine cause an increase in NOS activity and augmentation of NO production. These effects are dependent on the administered amount of L-arginine and last as long as circulating L-arginine concentrations are above the L-arginine concentrations before administration.

3. Evidence supporting this hypothesis

1. Presently, two endogenous potent inhibitors of NOS-catalyzed formation of NO from L-arginine are known, i.e. the methylated L-arginines, N^G, N^G -dimethyl-L-arginine (asymmet-

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Abbreviations: NO, nitric oxide; NOS, nitric oxide synthase; ADMA, asymmetric dimethylarginine; NMA, (mono)methylarginine; SDMA, symmetric dimethylarginine

References

- [1] Iyengar, R., Stuehr, D.J. and Marletta, M.A. (1987) *Proc. Natl. Acad. Sci. USA* 84, 6369–6373.
- [2] Förstermann, U., Closs, E.I., Pollock, J.S., Nakane, M., Schwarz, P., Gath, I. and Kleinert, H. (1994) *Hypertension* 23, 1121–1131.
- [3] Closs, E.I., Basha, F.Z., Habermeier, A. and Förstermann, U. (1997) *Nitric Oxide* 1, 65–73.
- [4] Hecker, M., Mitchell, J.A., Harris, H.J., Katsura, M., Thiemermann, C. and Vane, J.R. (1990) *Biochem. Biophys. Res. Commun.* 167, 1037–1043.
- [5] Girerd, X.J., Hirsch, A.T., Cooke, J.P., Dzau, V.J. and Creager, M.A. (1990) *Circ. Res.* 67, 1301–1308.
- [6] Rossitch, E., Alexander, E., Black, P.M. and Cooke, J.P. (1991) *J. Clin. Invest.* 87, 1295–1299.
- [7] Drexler, H., Zeiher, A.M., Meinzer, K. and Just, H. (1991) *Lancet* 338, 1546–1550.
- [8] Creager, M.A., Gallanher, S.J., Girerd, X.J., Coleman, S.M., Dzau, V.J. and Cooke, J.P. (1992) *J. Clin. Invest.* 90, 1248–1253.
- [9] Bode-Böger, S.M., Böger, R.H., Galland, A., Tsikas, D. and Frölich, J.C. (1998) *Br. J. Clin. Pharmacol.* 46, 489–497.
- [10] Bode-Böger, S.M., Böger, R.H., Löffler, M., Tsikas, D., Brabant, G. and Frölich, J.C. (1999) *J. Invest. Med.* 47, 43–50.
- [11] Bode-Böger, S.M., Böger, R.H., Alfke, H., Heinzl, D., Tsikas, D., Creutzig, A., Alexander, K. and Frölich, J.C. (1996) *Circulation* 93, 85–90.
- [12] Böger, R.H., Bode-Böger, S.M. and Frölich, J.C. (1996) *Atherosclerosis* 127, 1–11.
- [13] Kanno, K., Hirata, Y., Emori, T., Ohta, K., Eguchi, S., Imai, T. and Marumo, F. (1992) *Clin. Exp. Pharmacol. Physiol.* 19, 619–625.
- [14] Bode-Böger, S.M., Böger, R.H., Creutzig, A., Tsikas, D., Gutzki, F.-M., Alexander, K. and Frölich, J.C. (1994) *Clin. Sci.* 87, 303–310.
- [15] Böger, R.H., Bode-Böger, S.M., Gerecke, U. and Frölich, J.C. (1994) *Cardiovasc. Res.* 28, 494–499.
- [16] Böger, R.H., Bode-Böger, S.M., Gerecke, U., Gutzki, F.-M., Tsikas, D. and Frölich, J.C. (1996) *Clin. Exp. Pharmacol. Physiol.* 23, 11–15.
- [17] Böger, R.H., Bode-Böger, S.M., Mügge, A., Kienke, S., Brandes, R., Dwenger, A. and Frölich, J.C. (1995) *Atherosclerosis* 117, 273–284.
- [18] Hishikawa, K., Nakaki, T., Suzuki, H., Kato, R. and Saruta, T. (1993) *J. Hypertens.* 11, 639–645.
- [19] Vallance, P., Leone, A., Calver, A., Collierr, J. and Moncada, S. (1992) *Lancet* 339, 572–575.
- [20] Kotani, K., Ueno, S.-I. and Kakimoto, Y. (1992) *J. Neurochem.* 58, 1127–1129.
- [21] Kakimoto, Y. and Akazawa, S. (1970) *J. Biol. Chem.* 245, 5751–5758.
- [22] Paik, W.K. and Kim, S. (1971) *Science* 174, 114–119.
- [23] Kakimoto, Y., Matsuoka, Y., Miyake, M. and Konishi, H. (1975) *J. Neurochem.* 24, 893–902.
- [24] Böger, R.H., Bode-Böger, S.M., Thiele, W., Junker, W., Alexander, K. and Frölich, J.C. (1997) *Circulation* 95, 2068–2074.
- [25] Tsikas, D., Junker, W. and Frölich, J.C. (1998) *J. Chromatogr. B* 705, 174–176.
- [26] Kielstein, J.T., Böger, R.H., Bode-Böger, S.M., Schäffer, J., Barbey, M., Koch, K.M. and Frölich, J.C. (1999) *J. Am. Soc. Nephrol.* 10, 594–600.
- [27] Surdacki, A., Nowicki, M., Sandmann, J., Tsikas, D., Böger, R.H., Bode-Böger, S.M., Kruszelnicka-Kwiatkowska, O., Kokot, F., Dubiel, J.S. and Frölich, J.C. (1999) *J. Cardiovasc. Pharmacol.* 33, 652–658.
- [28] Masuda, H., Goto, M., Tamaoki, S. and Azuma, H. (1999) *Br. J. Pharmacol.* 126, 211–218.
- [29] Olken, N.M., Rusche, K.M., Richards, M.K. and Marletta, M.A. (1991) *Biochem. Biophys. Res. Commun.* 177, 828–833.
- [30] Pufahl, R.A., Nanjappan, P.G., Woodard, R.W. and Marletta, M.A. (1992) *Biochemistry* 31, 6822–6828.
- [31] Feldmann, P.L., Griffith, O.W., Hong, H. and Stuehr, D.J. (1993) *J. Med. Chem.* 36, 491–496.
- [32] Klatt, P., Schmidt, K., Brunner, F. and Mayer, B. (1994) *J. Biol. Chem.* 269, 1674–1680.
- [33] Tsikas, D., Sandmann, J., Savva, A., Lueßen, P., Böger, R.H., Gutzki, F.-M., Mayer, B. and Frölich, J.C. (2000) *J. Chromatogr. B* 742, 143–153.
- [34] Sandmann, J. (2000) Ph.D. thesis, University of Hannover, Hannover.
- [35] Faraci, F.M., Brain Jr., J.E. and Heistad, D.D. (1995) *Am. J. Physiol.* 269, H1522–H1527.
- [36] Reif, D.W. and McCreedy, S.A. (1995) *Arch. Biochem. Biophys.* 320, 170–176.
- [37] Böger, R.H., Bode-Böger, S.M., Thiele, W., Creutzig, A., Alexander, K. and Frölich, J.C. (1998) *J. Am. Coll. Cardiol.* 32, 1336–1344.
- [38] Bode-Böger, S.M., Böger, R.H., Kienke, S., Junker, W. and Frölich, J.C. (1996) *Biochem. Biophys. Res. Commun.* 219, 598–603.
- [39] Böger, R.H., Bode-Böger, S.M., Szuba, A., Tsao, P.S., Chan, J.R., Tangphao, O., Blaschke, T.F. and Cooke, J.P. (1998) *Circulation* 98, 1842–1847.
- [40] Böger, R.H., Tsikas, D., Bode-Böger, S.M., Phivthong-ngam, L. and Frölich, J.C. (2000) *Naunyn-Schmiedeberg's Arch. Pharmacol.* 361, R42.