

The mouse genome encodes a single homolog of the antimicrobial peptide human β -defensin 1

Kenneth M. Huttner^{a,*}, Christine A. Kozak^b, Charles L. Bevins^c

^aJoint Program in Neonatology, Children's Hospital, Boston, MA 02115, USA

^bLaboratory of Molecular Microbiology, National Institute of Allergy and Infectious Diseases, Bethesda, MD 20892-0460, USA

^cThe Departments of Immunology, Gastroenterology and Colorectal Surgery, The Cleveland Clinic Foundation, Cleveland, OH 44195, USA

Received 23 June 1997

Abstract The cysteine-rich β -defensin peptides are broad-spectrum bactericidal agents expressed in epithelial and myeloid tissues. The human β -defensin-1 (hBD-1) gene maps adjacent to the human α -defensin cluster and is expressed in the respiratory, gastrointestinal and genitourinary tracts. Here, we characterize a mouse β -defensin gene (mBD-1) which is: (1) closely related to hBD-1 both in sequence and gene organization; (2) expressed at high levels in the mouse kidney and at lower levels in brain, heart, lung, uterus, spleen, skeletal muscle, stomach, and small intestine; and (3) maps to mouse chromosome 8 at or near the location of the mouse α -defensin genes. These data indicate that mBD-1 is a close homolog of hBD-1, and suggest that analysis of its role in mouse host defense may provide significant insights into human epithelial innate immunity.

© 1997 Federation of European Biochemical Societies.

Key words: Antimicrobial peptide; Defensin; Mouse

1. Introduction

Beta-defensins are cationic, cysteine-rich antimicrobial peptides isolated from both myeloid and epithelial tissues [1]. The bovine β -defensin family includes at least fifteen related peptides with two expressed in epithelial tissues [2,3] and thirteen in neutrophils [4]. In marked contrast, a single human β -defensin, termed hBD-1, was isolated from plasma [5] and subsequently shown to be expressed at highest levels in the kidney and lower levels in epithelial tissues including the lung, pancreas, and genitourinary tract [6,7]. Evidence from studies of both cattle and human β -defensins demonstrates their involvement in epithelial host defense [3,8–10]. Impaired function of hBD-1 may play a prominent role in the pathophysiology of recurrent bacterial lung infections in the disease cystic fibrosis [10,11].

Several mouse cDNA sequences reported recently in the Sequence Tagged Site database encode a putative β -defensin. Using this sequence information, we have begun a molecular characterization of the mouse β -defensin gene family as an animal model for studying the role of β -defensin peptides in host immunity.

2. Materials and methods

2.1. Southern and Northern blots

Mouse genomic DNA was isolated from the liver of an adult FVB/NJ male, digested with restriction endonucleases, subjected to electro-

phoretic separation on a 1% agarose gel, and transferred to a Gene Screen Plus nylon membrane (NEN, Boston, MA) according to standard protocols [12]. Total RNA was isolated from adult FVB/NJ mouse tissues, subjected to electrophoresis in 1.3% agarose, and transferred to nylon membranes as described [13]. Northern and Southern blot hybridizations were overnight at 42°C in 50% formamide, 5×SSC, 1% SDS; 100 mcg/ml Salmon DNA, and 10% dextran sulfate. Post-hybridization washes were 2×10 min in 2×SSC at room temperature, followed by 2×15 min in 0.2×SSC, 0.5% SDS at 50°C.

2.2. Hybridization probes

A plasmid containing the mBD-1 cDNA was obtained from the ATCC (#974407 corresponding to GenBank #AA065510). A 261-bp PCR fragment encompassing the entire coding sequence was generated using the oligonucleotide primer pair: mBD-F1, 5'-GGCTGCC-ACCACATATGAAAACCTC-3' and mBD-R1, 5'-CCATCGCTCGTCTTTATGCTCA-3'. A 140-bp PCR fragment encoding the predicted 3' mature peptide was generated using the primer pair: mBD-F2: 5'-TGCCTTCAACATGGAGGATTCTG-3' and mBD-R1. In each case, the PCR product was gel purified and then used as template for generation of radioactive probe using either random primed DNA labeling or PCR amplification in the presence of radioactive nucleotides (NEN, Boston, MA). A 1.3-kb genomic Hind III fragment encoding the mBD-1 exon 1 sequence was labeled by random priming and used as a probe to identify exon 1 and 5' sequences.

2.3. RT-PCR

Total RNA from adult mouse tissues was reverse transcribed using SuperScript according to manufacturer's protocol (Gibco BRL, Gaithersburg, MD). PCR amplification of first strand product used the following profile: 95° for 5 min then 35 cycles of 94° for 0.5 min, 58° for 0.5 min, 72° for 0.5 min. PCR products were analyzed on 2% NuSieve/1% SeaKem LE agarose gels (FMC, Rockland, ME).

2.4. Screening of genomic library

The 261-bp mBD-1 PCR product was used as a probe for filter hybridization to a mouse 129/SvJ genomic library (Stratagene, La Jolla, CA). Duplicate positives were plaque purified, their phage DNA isolated and analyzed by restriction endonuclease digestion, and subfragments cloned for sequencing as described [14].

2.5. DNA sequencing

Purified plasmid DNA was sequenced from both strands using a thermal cycling method with fluorescent dye-labelled dideoxynucleotide terminators and Taq DNA polymerase. Sequence data were analyzed using the DNA and protein analysis software MacVector (IBI, New Haven, CT).

2.6. Chromosomal mapping

The mBD-1 locus (referred to as *Defb1*) was mapped by analysis of two sets of multilocus crosses: (NFS/N or C58/J×M. m. *musculus*)×M. m. *musculus* [15] and (NFS/N×M. *spretus*)×M. *spretus* or C58/J [16]. Progeny of these crosses have been typed for over 800 markers distributed over the 19 autosomes and X chromosome including the Chr 8 markers *Col4a* (collagen 4a), *Plat* (plasminogen activator, tissue), *Polb* (DNA polymerase beta), *Raml* (replication of amphotropic virus 1), and *Plcd1* (phospholipase C, delta 1) as described previously [17,18]. Recombination was determined according to Green [19] and loci were ordered by minimizing the number of recombinants.

*Corresponding author. Fax: (1) (617) 355-7677.

E-mail: huttner_k@al.tch.harvard.edu

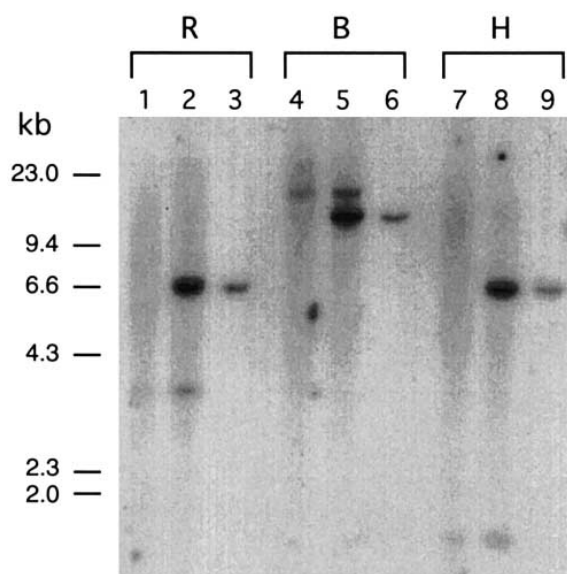


Fig. 1. Compilation of Southern blot hybridization of mouse genomic DNA. R, *EcoRI*; B, *BamHI*; H, *HindIII*. Probes used were mBD-1 entire coding sequence (lanes 2, 5, 8), mBD-1 5' sequences (lanes 1, 4, 7), and mBD-1 3' sequences (lanes 3, 6, 9) (see Section 2).

3. Results

3.1. mBD-1 genomic sequence

Nearly identical cDNA sequences encoding a putative β -defensin peptide were found in mouse kidney, heart, and macrophage cDNA libraries (GenBank #AA065510, AA104376 and AA071757, respectively). A 261-bp PCR fragment encompassing the entire open reading frame of this mouse β -defensin (mBD-1) cDNA was used as a probe in Southern hybridization (Fig. 1). Two bands, one intense and one weak, were identified in each of three different restriction enzyme digestions of mouse genomic DNA (lanes 2, 5 and 8).

Based on the previously reported two-exon structure of the human and bovine β -defensin genes, we predicted that the 5' and 3' regions of the mBD-1 cDNA would be encoded by separate exons, and that the two bands detected represented the two exons of mBD-1. This was supported by rehybridization of the same filter with a probe specific for the 5' mBD-1 sequences (lanes 1, 4, 7), which identified only the less intense band in each lane, and a probe specific for the 3' mBD-1 sequences (lanes 3, 6, 9), which identified only the more intense band.

We used the same 261-bp mBD-1 PCR fragment as a probe to identify mBD-1 sequences in a mouse 129/SvJ phage genomic library. From approximately 10^6 pfu screened we identified fifteen duplicate positives, three of which were plaque-purified and their DNA analyzed. Phage 4E-2 contains a genomic insert of approximately 20 kb including a 2.8-kb *EcoRI* fragment encoding the mBD-1 exon 1 sequence (Fig. 2). Phages A7 and 9D-1 contain overlapping genomic inserts of 13–14 kb which encode the mBD-1 exon 2 sequence.

Sequences from the deduced exons correspond to the reported cDNA sequence, and the predicted splice site is analogous to that of hBD-1 (Fig. 3A). The mouse mBD-1 gene sequence demonstrates significant nucleotide identity to the hBD-1 sequence as shown in Fig. 3B and C. Homology search of the GenBank database using the mBD-1 genomic sequence identified the hBD-1 sequences as well as a partial cDNA clone labeled CDK4 (#89820). Interestingly, CDK4 was isolated from rat kidney and predicts a peptide similar to mBD-1 [20].

3.2. Chromosomal localization

The hybridization probe for mBD-1 identified *PvuII* fragments of 6.9 kb in *M. spretus* and 6.0 kb in NFS/N and C58/J. Southern blot analysis of the parental DNAs of the *M. m. musculus* cross failed to identify any RFLPs. Inheritance of the variant fragments was compared with inheritance of over 800 markers previously typed in the *M. spretus* crosses and linkage was observed with markers on Chr 8. As indicated in Fig. 4, the mBD-1 locus (*Defb1*) mapped near *Polb*, *Plat*, and

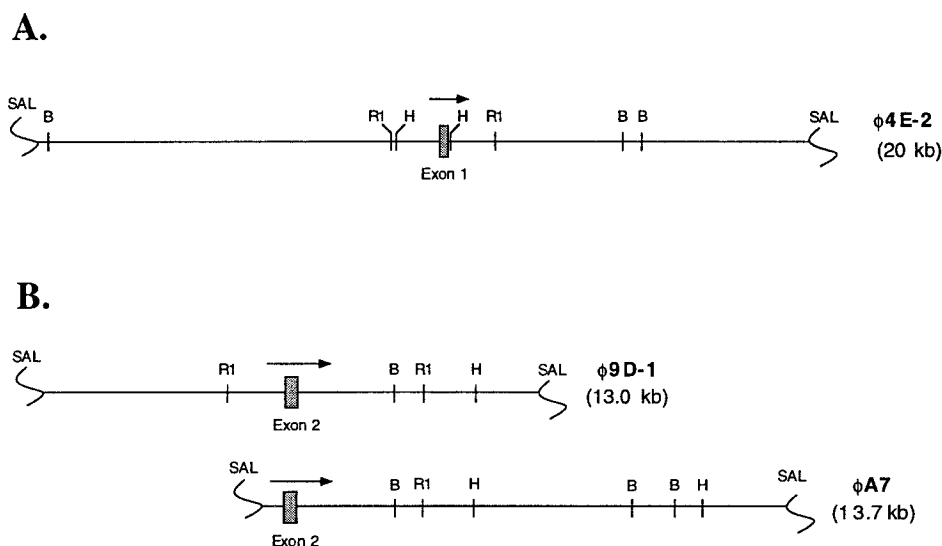


Fig. 2. Restriction enzyme maps of lambda phage clone genomic inserts encoding mBD-1 exon 1 (ϕ 4E-2) and exon 2 (ϕ A7 and ϕ 9D-1). SAL, *SalI*; R1, *EcoRI*; B, *BamHI*; H, *HindIII*. Location of additional *EcoRI* and *HindIII* sites present in ϕ 4E-2 downstream from the exon 1 coding sequence have not been determined.

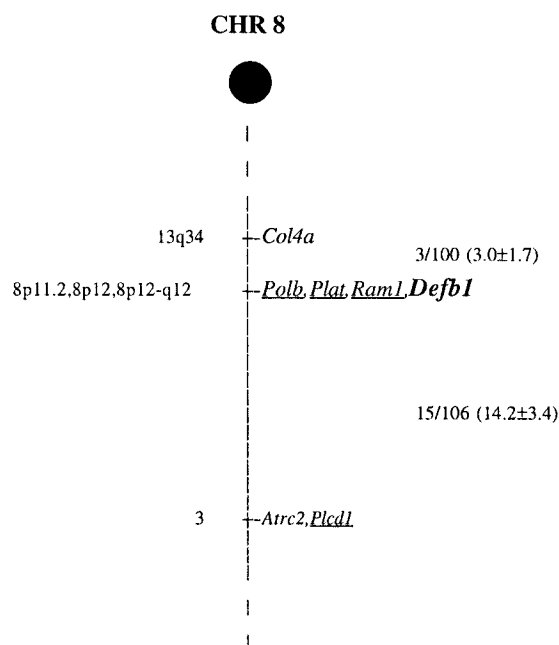


Fig. 4. Genetic map location of *Defb1* on mouse Chr 8. To the right of the map are given recombination fractions for adjacent loci. In parentheses the recombinational distances \pm S.D. are given. Map locations for the human homologs are given to the left of the map.

map location of the mouse and human β -defensins lies close to the α -defensin clusters on chromosome 8 of each species [21–23]. By Northern blot analysis, the highest level of β -defensin mRNA is found in the kidney for both the mouse and human genes. Taken together, these results suggest that unlike the α -defensin families of mice and humans, which have evolved in markedly differing patterns [1], the nearby β -defensin genes have maintained significant similarity in structure and perhaps function.

By contrast, the single intron of each of the three characterized bovine genes is short (~ 2 kb) and positioned at codon twenty (GenBank #L13373, U60447 and AF000362). The highest levels of bovine β -defensin expression were found in the bone marrow and epithelial tissues, including the trachea, tongue, and colon, while only trace β -defensin signal was found in kidney [3,4]. Although it is most probable that these bovine β -defensin genes were derived from the same ancestral precursor as both mBD-1 and hBD-1, their gene structure and

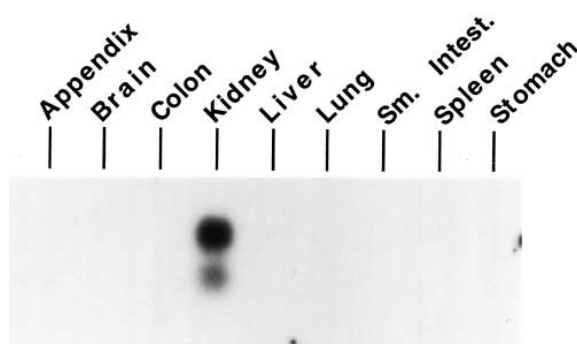


Fig. 5. Northern blot hybridization of mouse RNA using a 261-bp mBD-1 coding sequence probe. 15 μ g of total RNA per lane.

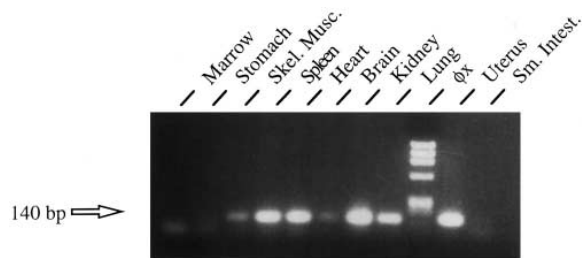


Fig. 6. RT-PCR of mouse β -defensin 1 sequences using adult mouse RNA as template. Oligonucleotide primers were mBD-F2 (forward) and mBD-R1 (reverse) (see Section 2).

expression patterns are significantly different from that of the only known mouse and human β -defensins. This suggests to us that additional mouse and human β -defensin sequences may be present which do not cross-hybridize to mBD-1 and hBD-1 sequences and which demonstrate alternative expression patterns.

In many antimicrobial peptide gene families examined to date there have been significant and unexpected inter-species differences which limit the generalization of biological findings [1,24]. While our data confirm that a close genetic relationship exists between the mouse and human β -defensin families, further characterization of mBD-1 and hBD-1, focusing on gene regulatory mechanisms and biological activity, will be both valuable and necessary in validating the mouse system as a model of hBD-1 function in vivo.

Acknowledgements: We would like to thank John P. Russell for technical assistance and Amy Elias for help in manuscript preparation. KMH was supported by the Child Health Research Center and CLB by AI32738.

References

- [1] Ganz, T. and Lehrer, R.I. (1994) *Curr. Opin. Immunol.* 6, 584–589.
- [2] Diamond, G., Zasloff, M., Eck, H., Brasseur, M., Maloy, W.L. and Bevins, C.L. (1991) *Proc. Natl. Acad. Sci. USA* 88, 3952–3956.
- [3] Schonwetter, B.S., Stolzenberg, E.D. and Zasloff, M.A. (1995) *Science* 267, 1645–1648.
- [4] Selsted, M.E., Tang, Y., Morris, W.L., McGuire, P.A., Novotny, M.J., Smith, W., Henschen, A.H. and Cullor, J.S. (1993) *J. Biol. Chem.* 268, 6641–6648.
- [5] Bensch, K.W., Raida, M., Magert, H.-J., Schulz-Knappe, P. and Forssmann, W.-G. (1995) *FEBS Lett.* 368, 331–335.
- [6] McCray, P.B. and Bentley, L. (1997) *Am. J. Respir. Cell Mol. Biol.* 16, 343–349.
- [7] Zhao, C., Wang, I. and Lehrer, R.I. (1996) *FEBS Lett.* 396, 319–322.
- [8] Diamond, G. and Bevins, C.L. (1994) *Chest* 105, 51S–52S.
- [9] Diamond, G., Russell, J.P. and Bevins, C.L. (1996) *Proc. Natl. Acad. Sci. USA* 93, 5156–5160.
- [10] Goldman, M.J., Anderson, G.M., Stolzenberg, E.D., Kari, U.P., Zasloff, M. and Wilson, J.M. (1997) *Cell* 88, 553–560.
- [11] Smith, J.J., Travis, S.M., Greenberg, E.P. and Welsh, M.J. (1996) *Cell* 85, 229–236.
- [12] Sambrook, J., Fritsch, E.F. and Maniatis, T. (1989) *Molecular Cloning: A Laboratory Manual*, 2nd ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY.
- [13] Diamond, G., Jones, D.E. and Bevins, C.L. (1993) *Proc. Natl. Acad. Sci. USA* 90, 4596–4600.
- [14] Huttner, K.M. and Ouellette, A.J. (1994) *Genomics* 24, 99–109.
- [15] Kozak, C.A., Peyser, M., Krall, M., Mariano, T.M., Kumar, C.S., Pestka, S. and Mock, B.A. (1990) *Genomics* 8, 519–524.

- [16] Adamson, M.C., Silver, J. and Kozak, C.A. (1991) *Virology* 183, 778–781.
- [17] Filie, J.D., Burbelo, P.D. and Kozak, C.A. (1995) *Mamm. Genom.* 6, 487.
- [18] Lyu, M.S., Park, D.J., Rhee, S.G. and Kozak, C.A. (1996) *Mamm. Genom.* 7, 501–504.
- [19] Green, E.L. (1981) *Genetics and Probability in Animal Breeding Experiments*, Oxford University Press, New York.
- [20] Page, R., Morris, C., Williams, J., von Ruhland, C. and Malik, A.N. (1997) *Biochem. Biophys. Res. Commun.* 232, 49–53.
- [21] Ouellette, A.J., Pratcheva, D., Ruddle, F.H. and James, M. (1989) *Genomics* 5, 233–239.
- [22] Liu, L., Heng, H., Zhao, C. and Ganz, T. (1996) *J. Invest. Med.* 44, 294A.
- [23] Sparks, R.S., Kronenberg, M., Heinzmann, C., Daher, K.A., Klisak, I., Ganz, T. and Mohandas, T. (1989) *Genomics* 5, 240–244.
- [24] Boman, H.G. (1995) *Annu. Rev. Immunol.* 13, 61–92.