

# Kinetic analysis on the substrate specificity of 3-isopropylmalate dehydrogenase

Kentaro Miyazaki<sup>a,\*</sup>, Katsumi Kakinuma<sup>b</sup>, Hiroaki Terasawa<sup>b</sup>, Tairo Oshima<sup>a</sup>

<sup>a</sup>Department of Life Science, Tokyo Institute of Technology, Nagatsuta 4259, Midori-ku, Yokohama 227, Japan

<sup>b</sup>Department of Chemistry, Tokyo Institute of Technology, O-okayama, Meguro-ku, Tokyo 152, Japan

Received 18 June 1993

Substrate specificity of 3-isopropylmalate dehydrogenase is analyzed using a series of synthetic (2*R*,3*S*)-3-alkylmalates. Each analog with hydrogen, methyl, ethyl, isopropyl, isobutyl, *tert*-butyl, and isoamyl group on C-3 functions as a substrate, implying a broad substrate specificity of the enzyme toward alkylmalates. The incremental binding energy of the isopropyl group of 3-isopropylmalate to the enzyme is estimated to be 3.55 kcal/mol, the rather small value supporting the broad specificity. Although the enzyme shows a broad specificity toward the alkylmalates, it does not show activity with isocitrate which has a negatively charged carboxymethyl group instead of the alkyl groups.

Analog; Hydrophobicity; 3-Isopropylmalate dehydrogenase; Substrate specificity

## 1. INTRODUCTION

3-Isopropylmalate dehydrogenase (EC 1.1.1.85, IPMDH) catalyzes the oxidative decarboxylation of (2*R*,3*S*)-3-isopropylmalate to 2-oxoisocaproate in the leucine biosynthetic pathway. We have cloned [1] and sequenced [2] the gene from an extreme thermophile, *Thermus thermophilus* HB8, and characterized the enzymatic properties [3]. X-ray crystallographic analysis was also conducted at the high resolution of 2.2 Å [4]. A stereospecific synthesis of the substrate has recently been developed [5] and the reaction mechanisms of the enzyme have been elucidated using chemically isotope-labeled isopropylmalates. The hydride-accepting site of the nicotinamide ring during the enzyme reaction has been identified as A (or proR position) by using deuterated isopropylmalate [6]. It was also confirmed that the configuration at the C-3 position of the substrate is retained throughout the decarboxylation by using another labeled isopropylmalate [7].

In this paper, we describe the kinetic analysis of the substrate specificity of *T. thermophilus* IPMDH using a series of (2*R*,3*S*)-3-alkylmalates.

## 2. EXPERIMENTAL

*T. thermophilus* IPMDH was prepared as previously described [3]. (2*R*,3*S*)-3-Alkylmalates were synthesized as described elsewhere (Kakinuma, K. et al., submitted for publication). Kinetic constants were determined in 50 mM sodium *N*-2-hydroxyethylpiperazine-*N'*-2-ethanesulfonate (pH 7.8), containing 100 mM KCl, 5.0 mM MgCl<sub>2</sub>, and 5.0 mM NAD in a total volume of 500 μl at 60°C. The concentration of analogs were 400–2,000 μM for malate; 3.33–10 μM for meth-

ylmalate, isoamylmalate, and *tert*-butylmalate; and 2.86–6.67 μM for ethylmalate, isobutylmalate, and isopropylmalate, respectively. Initial velocities were determined by monitoring the formation of NADH at absorbance 340 nm on a spectrophotometer. Values of  $k_{cat}$  were the means of three experiments.

## 3. RESULTS AND DISCUSSION

All the alkylmalates were active as a substrate of IPMDH, implying a broad substrate specificity of the enzyme toward alkylmalates. The kinetic constants  $K_m$ ,  $k_{cat}$ , and  $k_{cat}/K_m$  of various alkylmalates are summarized in Table I. The  $K_m$  value decreases up to isopropylmalate and increases afterwards; the  $k_{cat}$  value decreases depending on the increase of the hydrophobicity; and  $k_{cat}/K_m$  increases up to ethylmalate and decreases afterwards. These results suggest that the reaction generally occurs (Fig. 1) independently of the hydrophobicity of the alkyl group.

The change of binding energy ( $\Delta\Delta G_B$ ) of an alkyl group R of an alkylmalate, RM (relative to the hydrogen at C-3 of malate, HM), can be estimated by comparing the values of  $k_{cat}/K_m$  for the enzyme-catalyzed reactions using the relationship [8]:

$$\Delta\Delta G_B = -RT \ln(k_{cat}/K_m)_{HM}/(k_{cat}/K_m)_{RM}$$

(where  $R$  is the gas constant and  $T$  the absolute temperature). Application of this equation gives a value of 3.55 kcal/mol for the reaction of isopropylmalate. This small value supports the broad specificity of the enzyme.

In some microorganisms, the oxidation of alkylmalates have been reported to be associated with the activity of IPMDH for its nature of the broad substrate specificity [9–13]. *T. thermophilus* IPMDH also shows

\*Corresponding author. Fax: (81) (45) 922 2432.

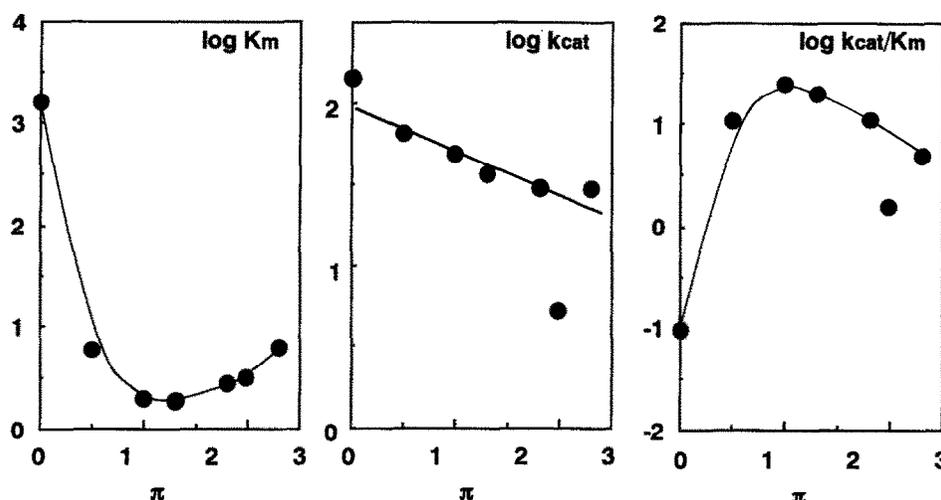


Fig. 1. The relationship between the hydrophobicity of the alkyl group and  $K_m$ ,  $k_{cat}$ , and  $k_{cat}/K_m$  for the catalysis of alkylmalate by IPMDH replotted from the values in Table I. Energies are in kcal/mol. The  $\pi$  values are the Hansch constants for the alkyl substituents [15].

a broad specificity toward alkylmalates *in vitro*, and consequently, the enzyme is expected to react not only to isopropylmalate but also to methylmalate or ethylmalate *in vivo*.

Isocitrate has a structural similarity with isopropylmalate,  $\text{HOOC}(\text{HO})\text{CHCH}(\text{X})\text{COOH}$ , in which X represents the  $\text{CH}_2\text{COOH}$  of isocitrate and the  $\text{CH}(\text{CH}_3)_2$  of isopropylmalate. Isocitrate dehydrogenase (EC 1.1.1.42), which is involved in the tricarboxylic acid cycle and also plays a role in glutamate biosynthesis, catalyzes a chemically equivalent reaction to IPMDH. In spite of the similarities of the substrates and the enzymes [14], isocitrate was completely inactive against IPMDH ( $k_{cat} < 0.2 \text{ s}^{-1}$ ). IPMDH showed a broad substrate specificity toward the alkylmalates, however, it does not show activity with isocitrate which has a negatively charged carboxymethyl group instead of the alkyl group. This may imply a significant chemical difference in the recognition site of the two enzymes towards the substituent at C-3 of malate.

Table I

Kinetic parameters for the catalysis of (2*R*,3*S*)-alkylmalate by IPMDH

Alkyl group	$\pi^a$	$K_m$ ( $\mu\text{M}$ )	$k_{cat}$ ( $\text{s}^{-1}$ )	$k_{cat}/K_m$ ( $\text{s}^{-1} \cdot \mu\text{M}^{-1}$ )
Hydrogen <sup>b</sup>	0	1,562	142	0.0907
Methyl	0.50	5.94	62.7	10.6
Ethyl	1.00	1.99	48.6	24.4
Isopropyl	1.30	1.84	36.0	19.6
Isobutyl	1.80	2.71	30.4	11.2
<i>tert</i> -Butyl	1.98	3.18	5.10	1.61
Isoamyl	2.30	6.13	29.7	4.84

<sup>a</sup> The  $\pi$  values are the Hansch constants for the alkyl substituents [15].

<sup>b</sup> Malate has only one asymmetric carbon atom.

*Acknowledgements:* This work was supported in part by Grants-in-Aid for Scientific Research from the Ministry of Education Science and Culture, the Japanese Government (Nos. 02403029 and 02250215).

## REFERENCES

- [1] Tanaka, T., Kawano, N. and Oshima, T. (1981) *J. Biochem.* 89, 677–682.
- [2] Kagawa, Y., Nojima, H., Nukiwa, N., Ishizuka, M., Nakajima, T., Yasuhara, T., Tanaka, T. and Oshima, T. (1984) *J. Biol. Chem.* 259, 2956–2960.
- [3] Yamada, T., Akutsu, N., Miyazaki, K., Kakinuma, K., Yoshida, M. and Oshima, T. (1990) *J. Biochem.* 108, 449–456.
- [4] Imada, K., Sato, M., Tanaka, N., Katsube, Y., Matsuura, Y. and Oshima, T. (1991) *J. Mol. Biol.* 222, 725–738.
- [5] Yamada, T., Kakinuma, K. and Oshima, T. (1987) *Chem. Lett.* 1745–1748.
- [6] Yamada, T., Kakinuma, K., Endo, K. and Oshima, T. (1987) *Chem. Lett.* 1749–1752.
- [7] Kakinuma, K., Ozawa, K., Fujimoto, Y., Akutsu, N. and Oshima, T. (1989) *J. Chem. Soc. Chem. Commun.* 1190–1191.
- [8] Fersht, A.R. (1985) *Enzyme Structure and Mechanism*, 2nd Edn., W.H. Freeman and Company, New York.
- [9] Rabin, R., Salamon, I.I., Bleiweis, A.S. and Ajl, S.J. (1968) *Biochemistry* 7, 377–388.
- [10] Stern, J. and O'Brien, R.W. (1969) *J. Bacteriol.* 98, 147–151.
- [11] Nakano, H., Sasaki, K., Kurokawa, Y. and Katsuki, H. (1971) *J. Biochem.* 70, 429–440.
- [12] Sasaki, K., Nakano, H. and Katsuki, H. (1971) *J. Biochem.* 70, 441–449.
- [13] Kisumi, M., Komatsubara, S. and Chibata, I. (1977) *J. Biochem.* 82, 95–103.
- [14] Miyazaki, K., Eguchi, H., Yamagishi, A., Wakagi, T. and Oshima, T. (1992) *Appl. Environ. Microbiol.* 58, 93–98.
- [15] Fujita, T., Iwasa, J. and Hansch, C. (1964) *J. Am. Chem. Soc.* 86, 5175–5180.