

# Induction of neurite outgrowth of PC12 cells by an inhibitor of vacuolar H<sup>+</sup>-ATPase, bafilomycin A<sub>1</sub>

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Bafilomycin A<sub>1</sub>, a selective inhibitor of vacuolar H<sup>+</sup>-ATPase, induced neurite outgrowth of PC12 cells dose- and time-dependently: more than 50% of the cells extended neurite-like spikes after 24 h treatment with 100 nM bafilomycin A<sub>1</sub>. Its dose-response ran roughly parallel to that of a bafilomycin A<sub>1</sub>-induced lysosomal pH increase. It was inhibited by LiCl, an inhibitor of the phosphorylation of microtubule-associated proteins and, like nerve growth factor (NGF)-induced neurite outgrowth, it was also inhibited by cycloheximide and actinomycin D. But, unlike the NGF-effect, it was not associated with rapid induction of *c-fos*.

Bafilomycin A<sub>1</sub>; Neurite outgrowth; Nerve growth factor; Vacuolar H<sup>+</sup>-ATPase; *c-fos*; PC12 cell

## 1. INTRODUCTION

Vacuolar H<sup>+</sup>-ATPases are multi-subunit enzymes which hydrolyze ATP, generating a proton gradient that is used for acidification of vacuolar compartments within cells [1] as lysosomes, endosomes, the Golgi complex and secretory granules including synaptic vesicles. The acidification of such organelles has been considered to play numerous and critical roles in maintaining the normal function of the endocytic and exocytic pathways [2]. However, it remains unclear whether the acidification of these organelles is responsible for other cellular processes such as cell proliferation and differentiation.

Recently, Bowman et al. reported that bafilomycins, which are macrolide antibiotics with a 16-membered lactone ring isolated from *Streptomyces* spp. [3], selectively inhibited the vacuolar H<sup>+</sup>-ATPases of *Neurospora crassa* vacuoles, bovine chromaffin granules and *Zea mays* vacuoles [4]. Bafilomycin A<sub>1</sub> is a powerful reagent for exploring the possible functions of acidic organelles in the regulation of cellular processes. Using bafilomycin A<sub>1</sub> we investigated the possible roles of acidic organelles in the PC12 cell differentiation. PC12 is a cloned pheochromocytoma cell line isolated from the

*Abbreviations:* NGF, nerve growth factor; Bt<sub>2</sub>cAMP, dibutyryl adenosine 3':5'-cyclic monophosphate; MAP, microtubule-associated protein.

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rat adrenal medulla, which is induced to differentiate into sympathetic neuron-like cells by NGF and other inducers [5,6]. During the course of the study we found that bafilomycin A<sub>1</sub> unexpectedly induced neurite outgrowth of PC12 cells.

Here we report the characterization of the bafilomycin A<sub>1</sub>-induced neurite outgrowth of PC12 cells and discuss a possible mechanism of action.

## 2. MATERIALS AND METHODS

### 2.1. Materials

Bafilomycin A<sub>1</sub> and mouse 7S nerve growth factor (NGF) were kindly provided by Professors K. Altendorf (University of Osnabrück, Germany) and K. Hayashi (Gifu College of Pharmacy, Japan), respectively. Plasmid pc-fos1 was supplied by the JCRB gene bank and dibutyryl adenosine 3':5'-cyclic monophosphate (Bt<sub>2</sub>cAMP) was obtained from Sigma. Fluorescein dextran (Mr 70,000, anionic) was obtained from Molecular Probes.

### 2.2. Cell culture

Rat pheochromocytoma PC12 cells were cultured in Dulbecco's modified Eagle's medium (Flow Laboratories) supplemented with 10% fetal calf serum (M. A. Laboratory) and 5% horse serum (Cosmo Bio, Japan) at 37°C in 5% CO<sub>2</sub>.

### 2.3. Neurite outgrowth assay

Cells were plated in 24-well culture dishes at a density of 5 × 10<sup>4</sup> cells/ml and assayed on the following day. After incubation with or without additives for the period indicated in the figure legends, the extent of neurite outgrowth was estimated as follows: a neurite was defined as a process whose length was more than 1.5-times of that of the cell body judged from the photographs taken under phase contrast microscopy (Olympus, Tokyo). About 150 cells were counted per well, and the percentage of cells with neurites was calculated.

### 2.4. Fluorescent measurement of intralysosomal pH

The PC12 cell lysosomes were loaded with fluorescein dextran by incubating the cells for 48 h in medium containing fluorescein dextran

(5 mg/ml). The cells were then washed with fresh medium and incubated in media with and without bafilomycin A<sub>1</sub>, respectively. The fluorescence from cells was viewed under an IMT-2-21-CA3 inverted fluorescence microscope (Olympus, Tokyo) through a SIT-camera after excitation at 450 and 490 nm and analyzed by an ARGUS-100/pH image processor (Hamamatsu Photonics, Hamamatsu). The fluorescence intensity ratios (490–450 nm) were converted to pH values using a calibration curve obtained *in vitro* as described previously [7,8].

#### 2.5. Detection of *c-fos* expression in PC12 cells

Total RNA was prepared from  $1 \times 10^6$  cells treated with additives by a previously described method [9], electrophoresed and analyzed by RNA blot hybridization using <sup>32</sup>p-labeled *c-fos* DNA as described [10].

### 3. RESULTS

#### 3.1. Bafilomycin A<sub>1</sub> induced neurite outgrowth of PC12 cells.

When PC12 cells were cultured in the presence of bafilomycin A<sub>1</sub>, neurite-like spikes extended as shown in Fig. 1B. The length of some neurites was more than 10-times the cell body after 24 h incubation with 100 nM bafilomycin A<sub>1</sub>. The morphology of those induced by bafilomycin A<sub>1</sub> resembled that of NGF- or Bt<sub>2</sub>cAMP-induced neurites as shown in Fig. 1 C and D.

The bafilomycin A<sub>1</sub>-mediated neurite outgrowth occurred time- and dose-dependently as shown in Fig. 2. Fluorescein dextran fluorescence measurements indicated a dose-response quite similar to that of the bafilomycin A<sub>1</sub>-induced lysosomal pH increase; the apparent lysosomal pH increased from a typical pH value of  $5.0 \pm 0.2$  to 5.6, 5.9 and 6.6 after 1 h incubation with 20, 100 and 500 nM bafilomycin A<sub>1</sub>, respectively (data not shown). The lag time (1–2 days) that is usually associated with NGF-mediated neurite outgrowth, was barely evident before the emergence of neurite-like spikes (Fig. 2) resembling Bt<sub>2</sub>cAMP-mediated neurite outgrowth induction (data not shown).

These results were not due to a toxic effect, if any, of bafilomycin A<sub>1</sub> on PC12 cells, since none of the other metabolic inhibitors tested induced neurite outgrowth: they included actinomycin D, cycloheximide (inhibitors of macromolecule synthesis), azide and/or 2-deoxyglucose (inhibitors of ATP generation), cytochalasin B and colchicine (inhibitors of cytoskeletal protein polymerization) (data not shown). Furthermore, no induction was caused by any other H<sup>+</sup>-ATPase inhibitors, including *N,N'*-dicyclohexylcarbodiimide ( $\leq 0.5$  mM), *N*-ethylmaleimide ( $\leq 0.1$  M) (for vacuolar-type ATPases), azide ( $\leq 10$  mM), oligomycin ( $\leq 5$  mM) (both

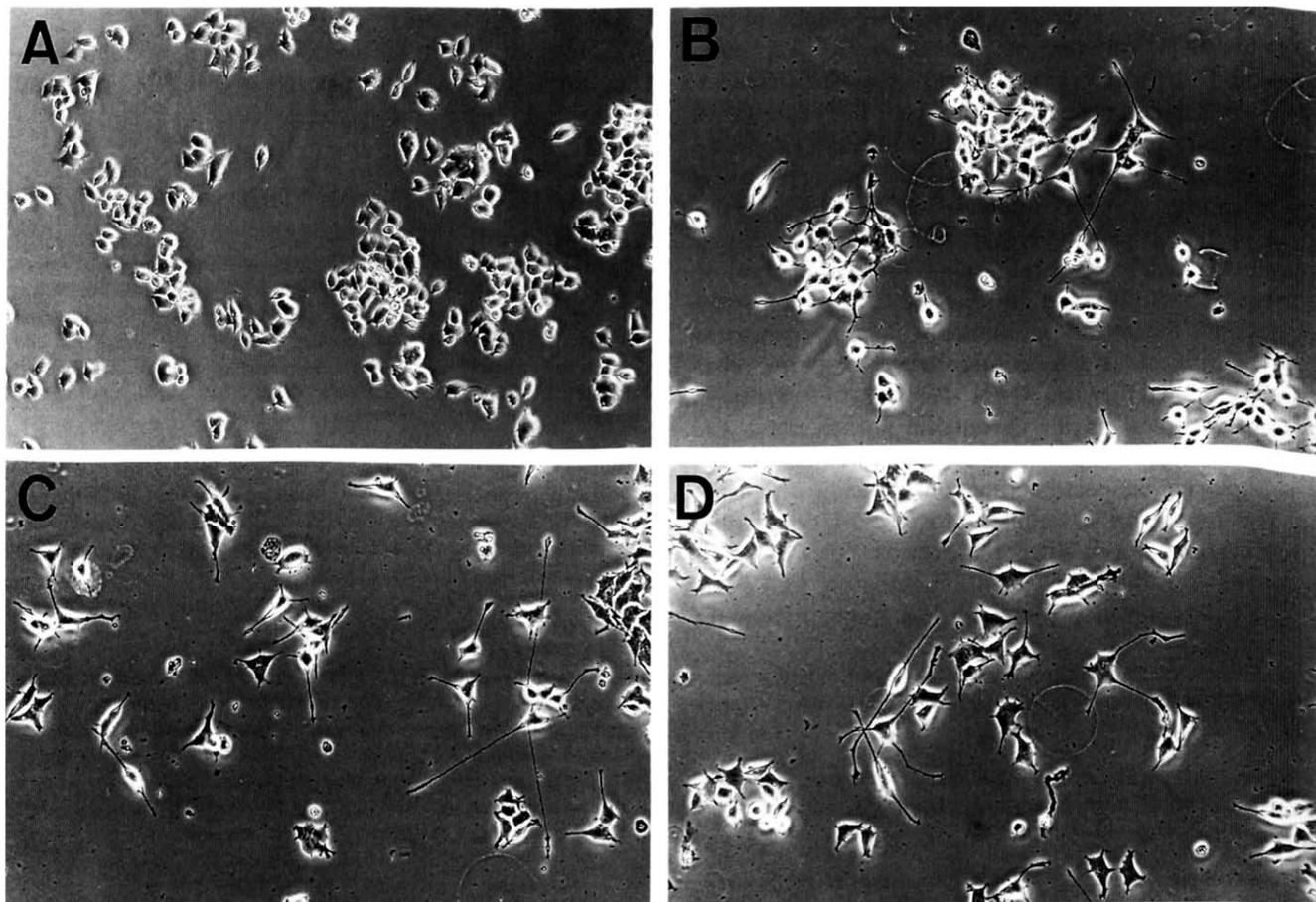


Fig. 1. Neurite outgrowth of PC12 cells induced by bafilomycin A<sub>1</sub>, NGF and Bt<sub>2</sub>cAMP. (A) control; (B) bafilomycin A<sub>1</sub> (100 nM); (C) NGF (500 ng/ml); (D) Bt<sub>2</sub>cAMP (0.5 mM). Cells were treated for 16 h (A,B and D) or 2 days (C) (×200).

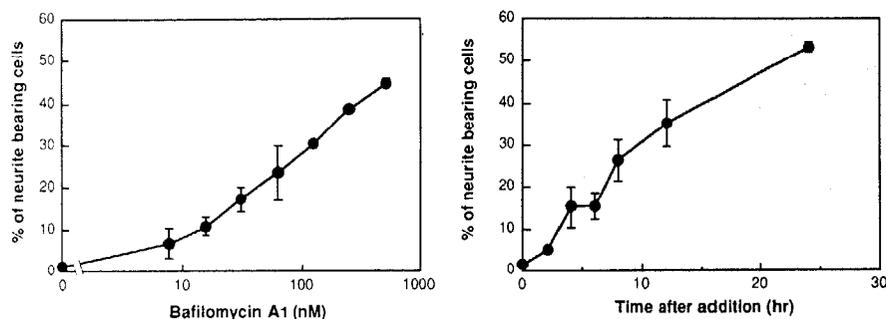


Fig. 2. Dose response (Left) and time-course (Right) of bafilomycin A<sub>1</sub>-induced neurite outgrowth of PC12 cells. (Leftpanel) Cells were scored for neurite outgrowth as described in Materials and Methods after a 16 h incubation with bafilomycin A<sub>1</sub>. Without the drug, the extent of neurite outgrowth was below 1%. The bafilomycin A<sub>1</sub> solvent, DMSO, did not induce any neurite outgrowth at 1% (v/v). Means  $\pm$  SD of duplicates are shown. (Rightpanel) Cells were scored for neurite outgrowth at the indicated times after addition of bafilomycin A<sub>1</sub> (100 nM). Means  $\pm$  SD of duplicates are plotted.

for mitochondrial F<sub>0</sub>F<sub>1</sub>-type ATPases) as well as vanadate ( $\leq 100$  mM) (for E<sub>1</sub>E<sub>2</sub>-type ATPases) (data not shown).

Cells treated with 100 nM bafilomycin A<sub>1</sub> for more than 2 days became detached from the dish. However, the detached cells were still viable judging from the trypan blue dye exclusion test and could be re-differentiated with NGF (data not shown).

### 3.2. LiCl inhibited the neurite outgrowth induced by bafilomycin A<sub>1</sub>

NGF-induced neurite outgrowth was inhibited by LiCl, an inhibitor of the phosphorylation of some microtubule-associated proteins (MAPs) [11]. As shown in Fig. 3, the bafilomycin A<sub>1</sub>-induced neurite outgrowth was also markedly inhibited by LiCl in a dose-dependent manner. The inhibitory profile was quite similar to that of NGF- or Bt<sub>2</sub>cAMP-mediated neurite induction. These results suggest that the bafilomycin effect was mediated by MAP phosphorylation and that the terminal responses of bafilomycin A<sub>1</sub>-induced neurite outgrowth are probably the same as those of Bt<sub>2</sub>cAMP or NGF. In support of this hypothesis, the bafilomycin effect was synergistically enhanced with Bt<sub>2</sub>cAMP or NGF (data not shown), suggesting that bafilomycin A<sub>1</sub>-

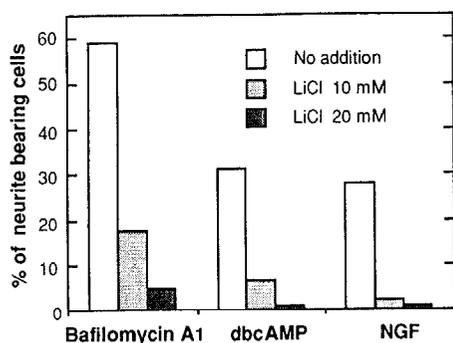


Fig. 3. The effect of LiCl on the neurite outgrowth induced by bafilomycin A<sub>1</sub>, Bt<sub>2</sub>cAMP or NGF. The neurite outgrowth was estimated after incubating PC12 cells with bafilomycin A<sub>1</sub> (100 nM, 20 h), Bt<sub>2</sub>cAMP (0.5 mM 20 h) and NGF (500 ng/ml, 2 days) in the absence or presence of LiCl (10 mM or 20 mM). Bars represent the mean values of duplicates. Deviations were less than 5%

induced neurite outgrowth is involved in the same cellular pathway as NGF- or Bt<sub>2</sub>cAMP-mediated responses.

### 3.3. Bafilomycin A<sub>1</sub>-induced neurite outgrowth depended on the de novo syntheses of RNA and protein

Since new RNA synthesis is required for NGF-mediated neurite generation (though not for regeneration) [12], the effects of inhibitors of RNA and protein synthesis on the bafilomycin A<sub>1</sub>-induced neurite outgrowth were examined. As shown in Fig. 4, both actinomycin D and cycloheximide inhibited bafilomycin A<sub>1</sub>- and NGF-induced neurite outgrowth, suggesting that bafilomycin A<sub>1</sub>-induced effect is dependent on the de novo synthesis of RNA and protein.

The removal of cycloheximide after 8 h restored bafilomycin A<sub>1</sub>-induced neurite outgrowth (data not shown).

### 3.4. Bafilomycin A<sub>1</sub>-induced neurite outgrowth did not depend on c-fos superinduction

The rapid superinduction of *c-fos* proto-oncogene expression is one of the major early events associated with NGF-induced neuronal differentiation in PC12 cells [13] and is considered to be responsible for long-term responses to NGF including neurite outgrowth [14]. However, *c-fos* induction was not detected 2 h after the addition of bafilomycin A<sub>1</sub> (Fig. 5, lanes 2-4), contrary to NGF where *c-fos* superinduction was detected by northern blot hybridization within 30 min (Fig. 5, lane 5). This indicates that the bafilomycin A<sub>1</sub>-effect is not mediated by *c-fos* superinduction.

## 4. DISCUSSION

The mechanism of neurite extension of neuronal cells has been studied in model systems using neuroblastoma (NG108-15) or pheochromocytoma (PC12) cells and various inducers, including NGF and Bt<sub>2</sub>cAMP. Accumulating evidence suggests that protein kinase C and adenylate cyclase systems are activated in response to NGF and other inducers [15-17]. However no causal

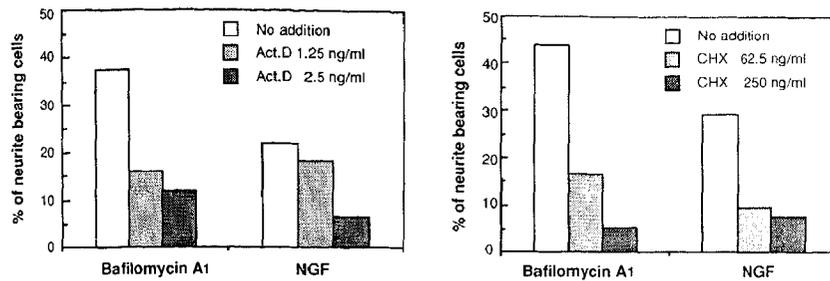


Fig. 4. Effects of actinomycin D (left) and cycloheximide (right) on the induction of neurite outgrowth by bafilomycin A<sub>1</sub> and NGF. Cells were scored for neurite outgrowth after incubation for 20 h (bafilomycin A<sub>1</sub>, 100 nM) or 2 days (NGF, 500 ng/ml). Actinomycin D (Act.D, 10 ng/ml) or cycloheximide (CHX, 250 ng/ml) added alone did not induce any neurite outgrowth. Values represent the mean values of duplicates. Deviation were less than 5% in every case.

relationships have been established yet between the activated second messenger systems and neurite outgrowth induction because of the pleiotropic effects of NGF and other factors.

The experimental system we described in this paper involving bafilomycin A<sub>1</sub> and PC12 cells may be useful to clarify the mechanism of neurite outgrowth induction during neuronal differentiation. The characteristics of the bafilomycin A<sub>1</sub>-induced neurite outgrowth were as follows: (I) as with the Bt<sub>2</sub>cAMP-effect, it was rapid, reversible (Fig. 2), and inhibited by LiCl an inhibitor of MAP-kinase (Fig. 3); (II) similar to the effect of NGF, it was dependent on the de novo synthesis of RNA and protein (Fig. 4). This is in contrast to the neurite outgrowth induced by protein kinase inhibitors such as staurosporine, which occurs independently of protein synthesis [18]; (III) unlike the NGF- and Bt<sub>2</sub>cAMP-effect [12,13], outgrowth was not associated

with *c-fos* superinduction (Fig. 5). In this respect, it is similar to *N-ras*-induced neurite outgrowth which is not associated with *c-fos* superinduction [18]. As it has been suggested that *c-fos* superinduction is mediated by protein kinase C activation [13], the last feature suggests little participation of protein kinase C in bafilomycin A<sub>1</sub>-induced neurite outgrowth. In fact, K-252a, a protein kinase C inhibitor, did not affect bafilomycin A<sub>1</sub>-induced neurite outgrowth although it inhibited the NGF-effect (manuscript in preparation). Taken together, the experimental system involving bafilomycin A<sub>1</sub> and PC12 cells possesses completely new features and constitutes a unique model system which should shed light on the mechanism of neurite outgrowth in neuronal cells.

It is not clear, however, how bafilomycin A<sub>1</sub> induced the neurite outgrowth. An increase in the internal pH of acid organelles by bafilomycin A<sub>1</sub> was obvious from the decreased granular staining of PC12 cells with Acridine orange, and the dose-response of bafilomycin A<sub>1</sub>-induced pH increases in lysosomes was quite parallel to that of bafilomycin A<sub>1</sub>-induced neurite outgrowth (data not shown). These results are consistent with the explanation that the primary intracellular target of bafilomycin A<sub>1</sub> for neurite outgrowth is vacuolar H<sup>+</sup>-ATPase. However, the increased pH of acidic organelles does not seem to be sufficient for neurite induction, since NH<sub>4</sub>Cl and other basic substances that increase intra-vacuolar pH [7,20] did not induce neurite outgrowth (data not shown). In contrast to basic substances which affect only vacuolar pH, bafilomycin A<sub>1</sub>, as an inhibitor of H<sup>+</sup>-ATPase, should also decrease the membrane potential difference of acidic organelles generated by H<sup>+</sup>-ATPase [21]. However, neither the combined addition of NH<sub>4</sub>Cl and FCCP (to cancel both the pH- and membrane potential-differences) nor *N*-ethylmaleimide or *N,N'*-dicyclohexylcarbodiimide (both vacuolar H<sup>+</sup>-ATPase inhibitors) induced neurite outgrowth (data not shown). These results are incompatible with the above explanation and suggest that the primary target of bafilomycin A<sub>1</sub> for neurite outgrowth is cellular components other than vacuolar H<sup>+</sup>-ATPases. Another possibility is that vacuolar H<sup>+</sup>-ATPases have additional functions other than H<sup>+</sup>

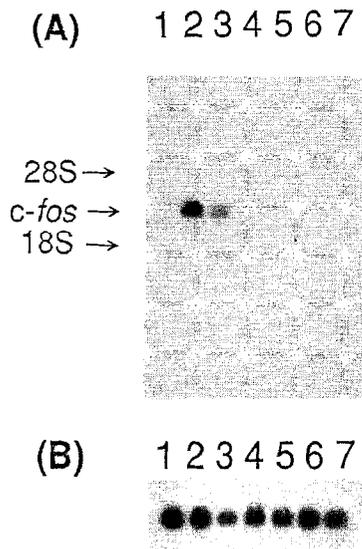


Fig. 5. Northern blot analysis of *c-fos* expression. (A) Total RNA (10 µg) isolated from PC12 cells treated with bafilomycin A<sub>1</sub> (200 nM) or NGF (500 ng/ml) was analyzed by northern blot for the expression of *c-fos* oncogene as described in Materials and Methods. (Lane 1) control; (lanes 2-4) bafilomycin A<sub>1</sub>; (lanes 5-7) NGF. (Lanes 2 and 5) 30 min; (lanes 3 and 6) 60 min; (lanes 5 and 7) 120 min. The positions of 28S rRNA, 18S rRNA and *c-fos* are shown by arrows. (B) The same filter was probed with chicken β-actin gene as an internal control.

pumping, that are sensitive to bafilomycin A<sub>1</sub> and are closely associated with neurite outgrowth. Further investigations are required to elucidate the mechanism by which bafilomycin A<sub>1</sub> induces neurite outgrowth.

Although the precise mechanism of bafilomycin A<sub>1</sub> action on neurite outgrowth remains to be clarified, it is an intriguing hypothesis that a change in H<sup>+</sup>-ATPase activity is involved in cell differentiation. The lysosomal pH of *ras*-transformed fibroblasts is reportedly higher than that of normal cells by 1 pH unit or more [22]. As *ras*-transformation of PC12 cells results in cell differentiation to neuronal cells [12, 23, 24], the increase of vacuolar pH may also be associated with *ras*-induced neurite outgrowth. The possible involvement of vacuolar H<sup>+</sup>-ATPases in the regulation of cell differentiation is an issue that remains to be clarified.

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## REFERENCES

- [1] Forgac, M. (1989) *Physiol Rev.* 69, 765-796.
- [2] Mellman, I., Fuchs, R. and Helenius, A. (1986) *Annu. Rev. Biochem.* 55, 663-700.
- [3] Werner, G., Hagenmaier, H., Drautz, H., Baumgartner, A. and Zahner, H. (1984) *J. Antibiotics* 37, 110-117.
- [4] Bowman, E.J., Siebers, A. and Altendorf, K. (1988) *Proc. Natl. Acad. Sci. USA* 85, 7972-7976.
- [5] Greene, L.A. and Tischler, A. (1976) *Proc. Natl. Acad. Sci. USA* 73, 2424-2428.
- [6] Rydel, R. and Greene, L. (1987) *J. Neurosci.* 7, 3639-3653.
- [7] Ohkuma, S. and Poole, B. (1978) *Proc. Natl. Acad. Sci. USA* 75, 3327-3331.
- [8] Ohkuma, S. (1990) *Methods Enzymol.* 174, 131-154.
- [9] Chomzynski, P. and Sacchi, N. (1987) *Anal. Biochem.* 162, 156-159.
- [10] Maniatis, T., Fritsch, J. and Sambrook, J. (1982) *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Laboratory, Cold Spring Harbor, New York.
- [11] Burstein, D.E. and Greene, L. (1978) *Proc. Natl. Acad. Sci. USA* 75, 6059-6063.
- [12] Burstein, D.E., Seeley, P.J. and Greene, L. (1985) *J. Cell Biol.* 101, 862-870.
- [13] Kruijer, W., Schubert, D. and Verma, I.M. (1985) *Proc. Natl. Acad. Sci. USA* 82, 7330-7334.
- [14] Greenberg, M.E., Greene, L. and Ziff, E.B. (1985) *J. Biol. Chem.* 260, 14101-14110.
- [15] Cremins, J., Wagner, J.A. and Halegoua, S. (1986) *J. Cell Biol.* 103, 887-893.
- [16] Glowacka, D. and Wagner, J.A. (1990) *J. Neurosci. Res.* 25, 453-462.
- [17] Cho, K.-O., Skarnes, W.C., Minsk, B., Palmieri, S., Jackson-Grusby, L. and Wagner, J.A. (1989) *Mol. Cell Biol.* 9, 135-143.
- [18] Shea, T.B. and Beermann, M.L. (1991) *Cell Biol. Int. Rep.* 15, 161-168.
- [19] Guerrero, I., Pellicer, A. and Burstein, D.E. (1988) *Biochem. Biophys. Res. Commun.* 150, 1185-1192.
- [20] Poole, B. and Ohkuma, S. (1981) *J. Cell Biol.* 90, 665-669.
- [21] Ohkuma, S., Moriyama, Y. and Takano, T. (1983) *J. Biochem.* 94, 1935-1943.
- [22] Jiang, L.-W., Maher, V.M., McCormick, J.J. and Schindler, M. (1990) *J. Biol. Chem.* 265, 4775-4777.
- [23] Noda, M., Ko, M., Ogura, A., Liu, D.-G., Anamo, T., Takano, T. and Ikawa, Y. (1985) *Nature* 318, 73-75.
- [24] Bar-Sagi, D. and Feramisco, J.R. (1985) *Cell* 42, 841-848.