

The *Kluyveromyces lactis* *KEX1* gene encodes a subtilisin-type serine proteinase

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KEX1 is a chromosomal gene required for the production of the killer toxin encoded by the linear DNA plasmid pGKL-1 of *Kluyveromyces lactis*. The nucleotide sequence of the cloned *KEX1* gene has been determined. The deduced structure of the *KEX1* protein, 700 amino acids long, indicated that it contained an internal domain with a striking homology to the sequences of the subtilisin-type proteinases, and a probable transmembrane domain near the carboxyl terminus. The results confirm the hypothesis that the product of the gene *KEX1* of *K. lactis* is a proteinase involved in the processing of the toxin precursor.

Killer toxin; Protein processing; Nucleotide sequence

1. INTRODUCTION

The killer toxin of *Kluyveromyces lactis* is encoded by the linear DNA plasmid pGKL-1 and secreted to the culture media. The *kex1* mutation of the host chromosome [1] leads to a non-killer phenotype. We have previously cloned the *KEX1* gene, and demonstrated, by in vivo complementation, that this gene was functionally related to the *KEX2* gene [2] of *Saccharomyces cerevisiae* [3]. The latter is known to code for a proteinase which converts the precursor proteins of M1 toxin and α -factor into mature molecules which are secreted [4–6]. Comparison of the amino acid sequence of the *K. lactis* toxin subunits [7] and the killer plasmid gene sequence [8,9] also suggested that a *KEX2*-type enzyme should be involved in the processing of the toxin precursor in *K. lactis*. The sequence of the *KEX1* gene confirmed this expectation.

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The nucleotide sequence presented here has been submitted to the EMBL/GenBank database under the accession number X07038

2. MATERIALS AND METHODS

2.1. Strains

The *K. lactis* (*K. marxianus* var. *lactis* [10]) strains used have been previously described [3,11].

2.2. DNA sequencing

DNA fragments of the *KEX1* gene region were subcloned into the multi-functional vectors pTZ18R and pTZ19R (Pharmacia). Serial deletions of cloned DNA were generated according to Lin et al. [12]. Single-stranded DNA was sequenced by the dideoxy chain termination method [13].

2.3. RNA analysis

Total RNA was extracted from the strain 2359/152. Poly(A)⁺ RNA was isolated by two passages over an oligo(dT)-cellulose (BRL) column, essentially as described in Maniatis et al. [14]. RNA was electrophoresed through an agarose gel containing formaldehyde, transferred to a nitrocellulose membrane, then hybridized with probe according to Maniatis et al. [14].

3. RESULTS AND DISCUSSION

3.1. Nucleotide sequence of the *KEX1* gene of *K. lactis*

The *KEX1* gene has previously been identified in a *K. lactis* DNA bank by complementation of the *kex1* mutation [3]. The complementing region of the cloned fragment contained a single large open reading frame of 2100 bp. A radioactive probe

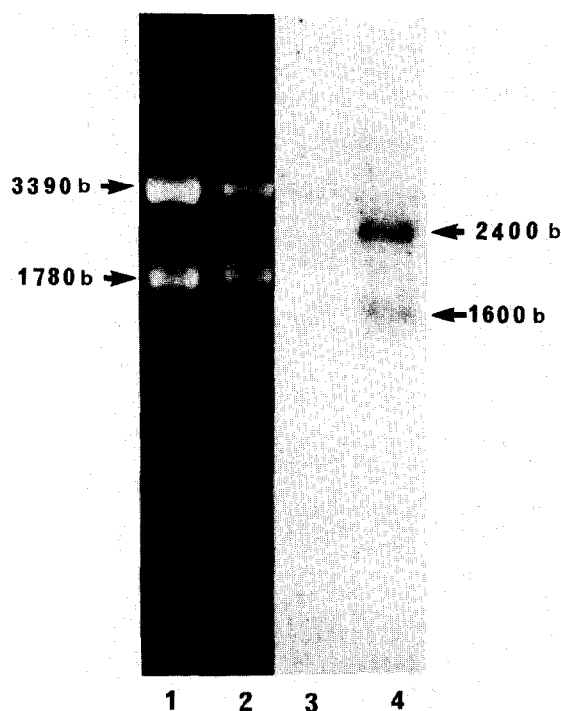


Fig.1. Northern blot analysis of the *KEX1* gene mRNA. Molecular mass markers are cytoplasmic ribosomal RNAs of *K. lactis*. Lane contents: lanes 1 and 3, 15 μ g total RNA; lanes 2 and 4, 10 μ g *K. lactis* poly(A⁺) enriched RNA. Lanes 1 and 2 show ethidium-stained RNA; lanes 3 and 4, hybridised RNA.

containing 80% of the cloned *KEX1* gene and a flanking 1 kb fragment was used to detect the *KEX1* transcript (fig.1). Hybridization with poly(A)⁺ enriched RNA detected a major RNA of 2400 bases, consistent with the length of the open reading frame. A weak signal of a 1600 base-long RNA was also detected, which corresponded to a second gene starting 800 bp downstream of the *KEX1* gene (not studied).

The DNA sequence of the *KEX1* gene and its flanking regions is shown in fig.2. The deduced protein sequence contained three methionine codons in the N-terminal region. If we assume that the first one is the translational start codon, the 5'-leader region of the gene shows a few typical features of a yeast gene: a TATA box (−188 to −185), an almost canonical CAAT box (−115 to −106) and a purine at −3. The 3'-flanking region is AT rich and has several termination codons in frame.

3.2. Amino acid sequence of the *KEX1* product

The open reading frame can code for a protein of 700 amino acids. The main features of this putative protein are summarized in fig.3. The hydrophobicity profile [15] reveals three major hydrophobic regions: the 18 amino acid residues at the amino-terminus, the 14 amino acids long central segment and the largest hydrophobic region near the carboxyl-terminus. This last region presents several characteristics of the membrane-spanning domain found in transmembrane proteins [15,16]. This sequence is immediately followed by a short stretch of positively charged amino acids. This structure is supposed to play a role in the anchoring of the protein in the membrane [17]. The supposed transmembrane domain is also preceded by clusters of serine and threonine residues (603–643). This domain is thought to serve as attachment sites for O-linked carbohydrate chains [16]. There are also five potential N-glycosylation sites (Asn-X-Thr/Ser) in the *KEX1* protein.

3.3. The *KEX1* encoded protein has a domain highly homologous to the subtilisin-type proteinases

The deduced amino acid sequence of the *KEX1* protein showed a region of striking homology with a family of serine proteases: subtilisin BPN' from *Bacillus amyloliquefaciens* [18], thermitase from *Thermoactinomyces vulgaris* [19] and the alkaline extracellular protease AEP from *Yarrowia lipolytica* [20]. Fig.4 is an optimized sequence comparison of the four proteins. Maximum of homology was observed around the essential amino acids of the active site (corresponding to Asp-164, His-202, Ser-373 of the *KEX1* sequence). The residues of the active site triad are situated at about the same relative positions in all four proteins. In addition, the *KEX1* sequence contained a cysteine (Cys-206) localized 5 residues after the active site His, as in the case of thermitase and proteinase K from *Tritirachium album* [19].

The *KEX1* sequence is much longer than the other three enzymes compared, but the homology is localized exclusively within the 155–388 region, without significant expansion or deletions.

Although the nucleotide sequence of the *KEX2* gene of *S. cerevisiae* has not been published, the structure of the *KEX1* protein deduced from the

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-221 -211
TTCTATCG AATCGGGTGC

-201 -191 -181 -171 -161 -151 -141
TCTTCATGTG TTACACGTCT TTTATACAGC ATAAAAATAA AGGCCATTCC AAAAAAGTTGT ACAATACTAA

-131 -121 -111 -101 -91 -81 -71
GGGCTAGTA CAGCTAGACA AATTAGGTGC AATCTCTAAA TCAGGATATC AGCTCTACGC CGGGCAAGTC

-61 -51 -41 -31 -21 -11 -1
ATTGAATAAG ATTTTCCACT TACTATTAC CTTTTCCCT TAATATTCCT TAATTTTCAG AACGATAGTT

15 30 45 60
MET Ile Leu Ser Ser Gln Leu MET Leu Ala Leu Ile Ala Val Ser Gly Tyr Gly Lys Ala
ATG ATC CTA TCG TCG CAG CTC ATG CTA GCT TTA ATA GCA GTG TCA GGA TAC GGT AAA GCA

75 90 105 120
MET Gln Val Pro Lys Lys Asp His Glu Asn Arg Gln Tyr Phe Ala Ile Glu Ser Tyr Asp
ATG CAA GTT CCT AAA AAA GAC CAC GAA AAT AGG CAG TAT TTT GCA ATT GAA TCT TAT GAT

135 150 165 180
Asp Val Gly Asn Leu Leu Ala Glu His Ser Asp Trp Ser Phe Glu His Asp Val Arg Gly
GAT GTA GGT AAT CTA CTA GCG GAA CAC AGT GAC TGG AGT TTC GAG CAC GAT GTT CGA GGC

195 210 225 240
Leu Ala Asn His Tyr Val Phe Ser Lys Pro Leu Gln Ser Leu Gly Lys Arg Asp Ala Ile
CTT GCC AAT CAC TAT GTG TTC TCG AAA CCG TTG CAG AGT TTG GGT AAA CGA GAT GCG ATT

255 270 285 300
Asp Thr Gly Tyr Ser Glu Asn Ile Ile Asp Phe His Asp Leu Pro Pro Val Gln Leu His
GAC ACA GGA TAT TCA GAA AAC ATC ATT GAT TTC CAC GAT CTA CCC CCC GTT CAG TTA CAC

315 330 345 360
Lys Arg Leu Pro Ile Gly Asp Ser Ser MET Glu Gln Ile Gln Asn Ala Arg Ile Leu Phe
AAA AGA TTG CCT ATT GGG GAT TCT AGT ATG GAA CAA ATC CAG AAC GCT AGA ATT CTT TTC

375 390 405 420
Asn Ile Ser Asp Pro Leu Phe Asp Gln Gln Trp His Leu Ile Asn Pro Asn Tyr Pro Gly
AAT ATT TCT GAT CCA TTG TTT GAT CAG CAG TGG CAC TTG ATC AAT CCA AAC TAC CCT GGA

435 450 465 480
Asn Asp Val Asn Val Thr Gly Leu Trp Lys Glu Asn Ile Thr Gly Tyr Gly Val Val Ala
AAT GAC GTT AAC GTA ACT GGT TTA TGG AAA GAA AAC ATC ACT GGC TAT GGT GTA GTG GCA

495 510 525 540
Ala Leu Val Asp Asp Gly Leu Asp Tyr Glu Asn Glu Asp Leu Lys Asp Asn Phe Cys Val
GCA TTG GTG GAT GAT GGA TTG GAT TAT GAG AAC GAA GAT TTA AAA GAC AAT TTC TGT GTT

555 570 585 600
Glu Gly Ser Trp Asp Phe Asn Asp Asn Asn Pro Leu Pro Lys Pro Arg Leu Lys Asp Asp
GAA GGT TCT TGG GAT TTT AAT GAC AAC CCA TTG CCG AAG CCA AGG CTA AAA GAT GAT

615 630 645 660
Tyr His Gly Thr Arg Cys Ala Gly Glu Ile Ala Ala Phe Arg Asn Asp Ile Cys Gly Val
TAC CAT GGT ACC CGC TGC GCA GGT GAA ATA GCG GCT TTC CGT AAT GAT ATT TGT GGG GTT

675 690 705 720
Gly Val Ala Tyr Asn Ser Lys Val Ser Gly Ile Arg Ile Leu Ser Gly Gln Ile Thr Ala
GGT GTC GCC TAT AAC TCT AAG GTA TCC GGT ATC AGA ATT TTG TCA GGC CAG ATC ACA GCC

735 750 765 780
Glu Asp Glu Ala Ala Ser Leu Ile Tyr Gly Leu Asp Val Asn Asp Ile Tyr Ser Cys Ser
GAA GAT GAG GCT GCT TCA TTA ATT TAT GGA CTA GAC GTT AAT GAT ATT TAC TCT TGC TCG

795 810 825 840
Trp Gly Pro Ser Asp Asp Gly Lys Thr MET Gln Ala Pro Asp Thr Leu Val Lys Lys Ala
TGG GGT CCA TCT GAT GAC GGT AAA ACT ATG CAA GCG CCG GAT ACA TTA GTA AAA AAG GCA

855 870 885 900
Ile Ile Lys Gly Val Thr Glu Gly Arg Asp Ala Lys Gly Ala Leu Tyr Val Phe Ala Ser
ATC ATA AAA GGT GTA ACA GAA GGA CGA GAT GCA AAA GGT GCA CTA TAT GTA TTT GCG AGT

915 930 945 960
Gly Asn Gly Gly MET Phe Gly Asp Ser Cys Asn Phe Asp Gly Tyr Thr Asn Ser Ile Phe
GGG AAT GGT GGT ATG TTT GGC GAC AGC TGC AAC TTT GAC GGC TAC ACA AAC TCT ATA TTT

975 990 1005 1020
Ser Ile Thr Val Gly Ala Ile Asp Trp Lys Gly Leu His Pro Pro Tyr Ser Glu Ser Cys
TCT ATC ACT GTA GGT GCC ATT GAT TGG AAG GGC CTA CAT CCT CCA TAT TCT GAA TCA TGT

1035 1050 1065 1080
Ser Ala Val MET Val Val Thr Tyr Ser Ser Gly Ser Gly Asn Tyr Ile Lys Thr Thr Asp
TCT GCT GTA ATG GTT GTT ACT TAT TCT TCG GGA TCA GGA AAT TAC ATA AAA ACA ACA GAT

1095 1110 1125 1140
Leu Asp Glu Lys Cys Ser Asn Thr His Gly Gly Thr Ser Ala Ala Ala Pro Leu Ala Ala
TTA GAC GAA AAA TGT TCC AAT ACG CAT GGA GGC ACT TCA GCT GCA GCT CCT CTT GCA GCT

1155 1170 1185 1200
Gly Ile Tyr Thr Leu Val Leu Glu Ala Asn Pro Asn Leu Thr Trp Arg Asp Val Gln Tyr
GGT ATA TAT ACT TTA GTG CTG GAA GCT AAC CCG AAC TTA ACA TGG CGA GAT GTA CAA TAC

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				1215				1230				1245				1260						
Leu	Ser	Ile	Leu	Ser	Ser	Glu	Glu	Ile	Asn	Pro	His	Asp	Gly	Lys	Trp	Gln	Asp	Thr	Ala			
CTC	TCA	ATA	TTG	AGC	TCT	GAG	GAA	ATA	AAT	CCG	CAC	GAT	GGA	AAG	TGG	CAG	GAT	ACA	GCT			
				1275				1290				1305				1320						
MET	Gly	Lys	Arg	Tyr	Ser	His	Thr	Tyr	Gly	Phe	Gly	Lys	Leu	Asp	Ala	Tyr	Asn	Ile	Val			
ATG	GGA	AAG	CGT	TAT	TCT	CAC	ACA	TAT	GGA	TTT	GGA	AAA	CTT	GAT	GCA	TAT	AAC	ATT	GTC			
				1335				1350				1365				1380						
His	MET	Ala	Lys	Ser	Trp	Ile	Asn	Val	Asn	Pro	Gln	Gly	Trp	Leu	Tyr	Leu	Pro	Thr	Ile			
CAT	ATG	GCA	AAA	AGT	TGG	ATC	AAT	GTA	AAC	CCA	CAA	GGT	TGG	CTT	TAC	CTT	CCT	ACA	ATC			
				1395				1410				1425				1440						
Val	Glu	Lys	Gln	Ser	Ile	Ser	Asn	Ser	Asp	Glu	Val	Ile	Glu	Ser	Thr	Val	Ser	Val	Ser			
GTT	GAA	AAA	CAG	TCT	ATC	AGT	AAT	TCA	GAT	GAA	GTT	ATA	GAA	TCC	ACA	GTC	TCA	GTT	TCT			
				1455				1470				1485				1500						
Ala	Glu	Glu	Phe	Lys	Gln	Asn	Asn	Leu	Lys	Arg	Leu	Glu	His	Val	Thr	Val	Thr	Val	Asp			
GCT	GAA	GAG	TTT	AAA	CAA	AAT	AAC	CTA	AAA	AGG	TTG	GAA	CAT	GTC	ACT	GTA	ACT	GTC	GAT			
				1515				1530				1545				1560						
Ile	Asp	Ala	Pro	Tyr	Arg	Gly	His	Val	Leu	Val	Asp	Leu	Ile	Ser	Pro	Asp	Gly	Val	Thr			
ATA	GAC	GCA	CCT	TAC	CGT	GGA	CAT	GTC	TTA	GTA	GAT	CTA	ATA	TCG	CCT	GAT	GGA	GTT	ACA			
				1575				1590				1605				1620						
Ser	Thr	Leu	Ala	Thr	Ala	Arg	Arg	Leu	Asp	Lys	Asn	Arg	Tyr	Gly	Phe	Gln	Asn	Trp	Thr			
TCT	ACC	TTA	GCG	ACA	GCT	AGA	CGT	TTA	GAT	AAA	AAC	CGC	TAT	GGT	TTT	CAA	AAT	TGG	ACT			
				1635				1650				1665				1680						
Phe	MET	Ser	Val	Ala	His	Trp	Gly	Ser	Ser	Gly	Val	Gly	Ser	Trp	Lys	Leu	Lys	Val	Lys			
TTC	ATG	TCT	GTC	GCG	CAC	TGG	GCG	TCT	AGT	GGA	GTT	GGA	AGC	TGG	AAA	TTA	AAA	GTA	AAG			
				1695				1710				1725				1740						
Ser	Thr	His	Asp	Asn	Glu	Ile	Val	Thr	Leu	Lys	Ser	Trp	Arg	Leu	Lys	MET	Phe	Gly	Glu			
TCT	ACG	CAT	GAT	AAT	GAA	ATT	GTA	ACA	CTC	AAA	TCT	TGG	AGA	TTA	AAG	ATG	TTT	GGA	GAA			
				1755				1770				1785				1800						
Thr	Ile	Asp	Ala	Lys	Lys	Ala	Lys	Val	Ile	Ser	Tyr	Gly	Asn	Asp	Lys	Glu	Asp	Ala	Glu			
ACT	ATC	GAT	GCA	AAG	AAG	GCC	AAA	GTG	ATA	TCA	TAT	GGA	AAT	GAC	AAA	GAG	GAT	GCT	GAA			
				1815				1830				1845				1860						
Val	Lys	Ser	Thr	Glu	Ser	Lys	Thr	Thr	Thr	Pro	Thr	Ala	Gln	Thr	Ser	Ser	Phe	Thr	Thr			
GTT	AAG	AGT	ACC	GAA	TCT	AAA	ACC	ACA	ACT	CCC	ACT	GCA	CAA	ACT	TCG	TCA	TTC	ACG	ACG			
				1875				1890				1905				1920						
Thr	Ser	Gly	Glu	Glu	Thr	Ser	Gly	Ala	Asn	Lys	Leu	Pro	Arg	Pro	Glu	Gln	Ala	Ala	Gln			
ACT	TCT	GGA	GAA	GAA	ACA	TCT	GGT	GCA	AAT	AAG	TTG	CCT	CGT	CCC	GAA	CAG	GCT	GCC	CAG			
				1935				1950				1965				1980						
Leu	Tyr	Leu	Ala	Ile	Phe	Val	Ile	Gly	Ala	Ile	Val	Ile	Ile	Ile	Tyr	Tyr	Leu	Phe	Phe			
TTA	TAC	TTG	GCA	ATT	TTT	GTC	ATT	GGT	GCG	ATA	GTC	ATC	ATA	ATT	TAC	TAT	TTG	TTT	TTC			
				1995				2010				2025				2040						
Leu	Lys	Ser	Arg	Arg	Ile	Ile	Arg	Arg	Ser	Arg	Ala	Glu	Ala	Tyr	Glu	Phe	Asp	Ile	Ile			
TTA	AAA	TCA	AGA	AGA	ATA	ATC	AGA	AGG	TCT	AGA	GCA	GAA	GCT	TAT	GAA	TTT	GAT	ATC	ATT			
				2055				2070				2085				2100						
Asp	Thr	Asp	Ser	Glu	Tyr	Asp	Ala	Ser	Ile	Asn	Lys	Leu	Gln	Ser	Leu	Tyr	Leu	Val	Lys			
GAT	ACC	GAC	TCA	GAA	TAC	GAT	GCT	TCG	ATT	AAC	AAA	CTG	CAG	AGT	CTA	TAT	CTG	GTG	AAG			
				2115				2130				2145				2160						
<u>TAA</u>				ATG	ATG	ATA	ACC	TTG	AAG	ACT	TTA	ACT	TCG	ACA	<u>TAA</u>	ATG	AAG	AAG	AGC	TCT	CAC	CCC
				2175				2190				2205				2220						
GTG				AAA	GTT	CAA	GCA	ATA	ATC	CTT	CGG	GAA	<u>TGA</u>	ATC	TCT	GGA	ATC	TTT	CGA	CAT	CTC	CTG
				2235				2250				2265				2280						
ATC				ATA	CAA	GCA	ATT	TAC	<u>TAG</u>	GGC	AAA	ACT	CGA	TTC	CCA	ACA	AAT	AGA	AAG	ACA	<u>TAG</u>	CAT
				2295				2310				2325				2340						
AAC				AGT	TCA	TAA	GAG	ATA	AAT	CTC	AAA	ATA	CTG	ACG	TTT	TCA	TAT	AGA	AGA	TAG	GTT	TTT

Fig.2. Nucleotide sequence of the *KEX1* gene of *K. lactis*. The amino acid sequence of the coding region, predicted from the nucleotide sequence, is given above the nucleotide sequence. Probable TATA and CAAT sequences are underlined.

gene sequence appears to have an overall organization analogous to that reported for the KEX2 protein [5,21].

3.4. The *kex1* mutation in *K. lactis* is concerned with the secretion of the killer toxin

Fig.5 illustrates the absence of the bands corresponding to the toxin subunits in the culture fluid of a *kex1* mutant, and the restoration of the

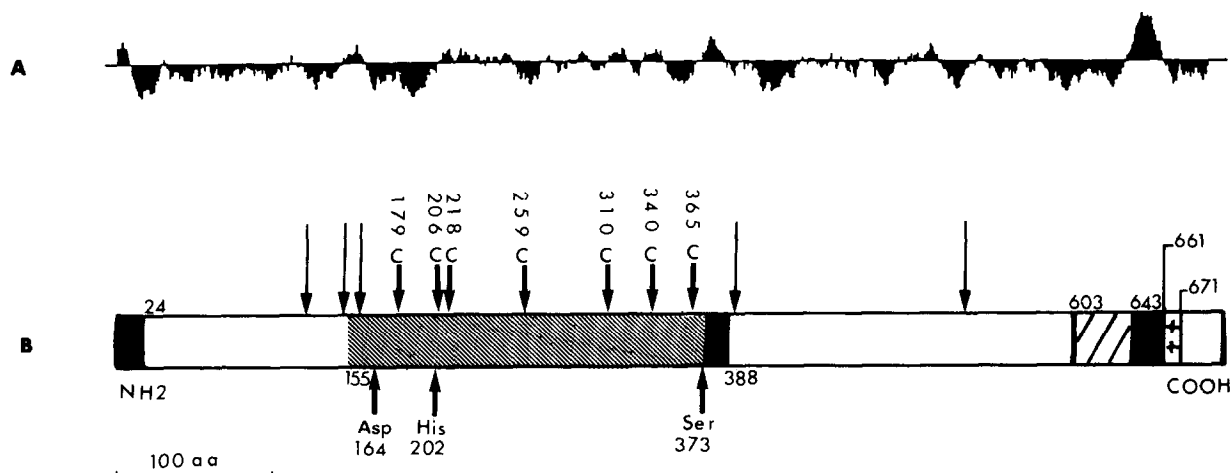


Fig.3. Main structural features of the deduced KEX1 gene product. (A) Hydropathy profile of the KEX1 product; (B) structure of the KEX1 protein. (■) Hydrophobic domains; (▨) region sharing sequence homology with subtilisin-type proteins. The three essential amino acid residues of the potential catalytic site are indicated. Putative *N*-glycosylatable sites are shown by arrows. Cysteine positions are also indicated (c). (⋄) Thr-Ser rich region; (▩) positively charged residues. Numbers indicate the amino acid positions in the deduced protein sequence.

bands in the transformant clones. The β - and γ -subunits can be seen on this gel; the α -subunit was visualized on another gel (not shown). The wild-type killer strain MW98-8C and the isonuclear plasmid-less mutant strain VD1, were used to identify the toxin subunits.

Biological roles of the *K. lactis KEX1* gene remain to be studied further. It is known that homozygous *kex1* diploids are deficient for sporulation [3].

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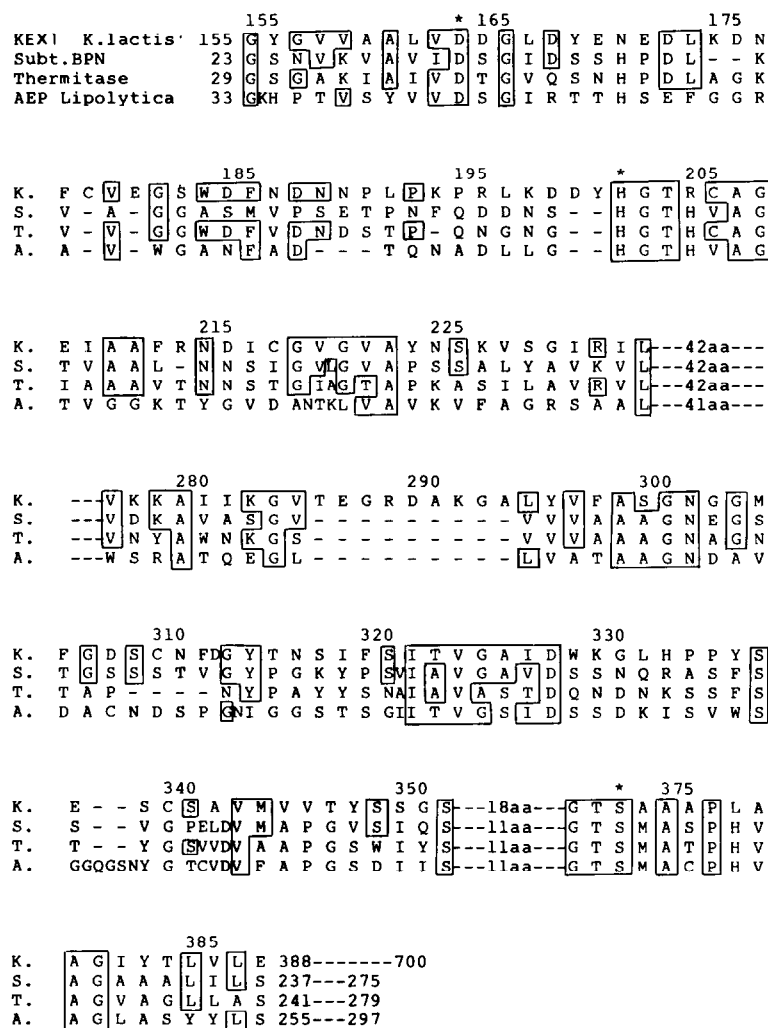


Fig.4. Amino acid homology between the KEX1 gene product of *K. lactis* (K) and a variety of subtilisin-type proteinases. All sequences are aligned with respect to the KEX1 gene product. Numbers to the left and to the right indicate respectively the amino acid in the protein sequence at which the homology starts and ends. The numbers above the sequences correspond to the KEX1 protein. Amino acids homologous to the KEX1 gene are boxed. Asterisks indicate the essential amino acids of the active site, known for subtilisin. Subt-BPN' (S), subtilisin BPN' from *Bacillus amyloliquefaciens*. Thermitase (T), thermitase from *Thermoactinomyces vulgaris*. AEP lipolytica (A), alkaline extracellular protease of *Yarrowia lipolytica*.

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(For fig.5, see overleaf.)

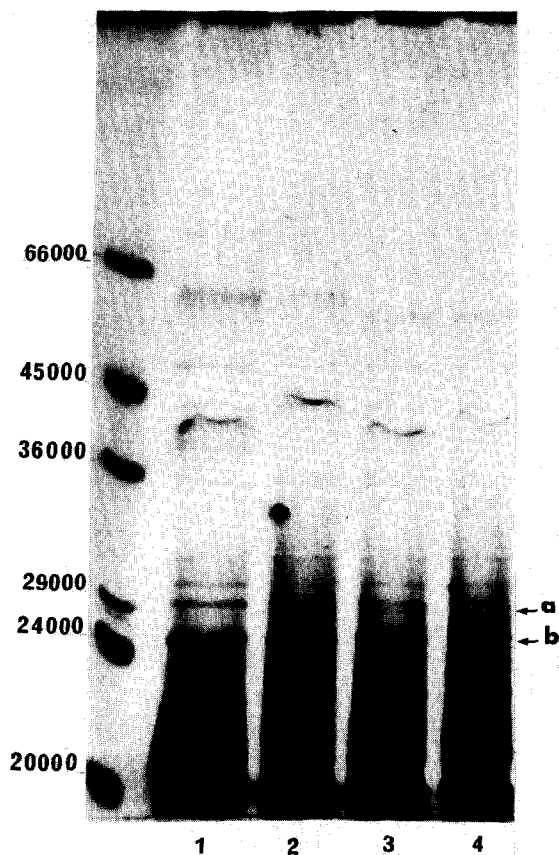


Fig.5. Secreted polypeptides in the wild-type and the *kex1* mutant strains. Polypeptides secreted into the culture medium were tested by SDS-polyacrylamide gels electrophoresis containing 11% acrylamide. Using Minicon B15 membranes (Amicon) culture supernatants were concentrated approx. 50-fold before loading 10 μ l in each gel-slot. The gel was silver stained according to Merril et al. [22]. The positions of the molecular mass markers are indicated. Lanes: 1, MW105-2D, *kex1* mutant strain; 2, MW105-2D strain, transformed with KEp6-pc7M plasmid carrying *KEX1* gene [3]; 3, MW98-8C wild-type killer strain; 4, MW98-8C without killer plasmids (VD1). a = β -subunit; b = γ -subunit.