

Calcium-sensitive non-muscle α -actinin contains EF-hand structures and highly conserved regions

Angelika Noegel, Walter Witke and Michael Schleicher*

Max Planck Institutes for Biochemistry and *Psychiatry, 8033 Martinsried, FRG

Received 21 July 1987

The F-actin crosslinking molecule α -actinin from the slime mould *Dictyostelium discoideum* carries two characteristic EF-hand structures at the C-terminus. The calcium-binding loops contain all necessary ligand-binding oxygens and most likely form the structural basis for the calcium sensitivity of strictly calcium-regulated non-muscle α -actinins. Furthermore, the sequence exhibits at the N-terminal site of the molecule a high degree of homology to chicken fibroblast α -actinin. This stretch of amino acids appears to have remained essentially constant during evolution and might represent the actin-binding site. The findings have led us to propose a model for the inhibitory action of Ca^{2+} on non-muscle α -actinins.

Actin-binding protein; Cytoskeleton; EF-hand structure; Evolution; Sequence homology; (*Dictyostelium discoideum*)

1. INTRODUCTION

α -Actinin, an actin filament crosslinking protein, can be found in a wide variety of organisms in both muscle and non-muscle cells. It is a homodimer whose subunits assemble in an anti-parallel fashion to form a rod-like structure. The crosslinking activity of non-muscle α -actinin is completely inhibited by calcium at the micromolar level. This calcium sensitivity distinguishes muscle and non-muscle α -actinin [1,2]. Using a partial genomic clone of the *Dictyostelium discoideum* α -actinin gene, we have previously shown that *D. discoideum* harbours a single α -actinin gene that codes for a mRNA of approx. 3.0 kb. This mRNA is present during all developmental stages of the *D. discoideum* life cycle [3]. Screening of a cDNA library [4] with this clone led to the isolation of

cDNA clones carrying the complete sequence for *D. discoideum* α -actinin. Here, we present the first complete sequence of a non-muscle α -actinin, that of *D. discoideum*. Two sequences at the C-terminus are characteristic EF-hand structures and most likely form the structural basis for the calcium sensitivity. Furthermore, the *D. discoideum* sequence exhibits a high degree of homology with a recently reported partial sequence of chicken fibroblast α -actinin [5]. The most striking homology is located at the N-terminal site of the molecule. It is possible that during evolution the conserved structure of actin forced a similarly constant complementary region in the actin-binding site of α -actinins.

2. MATERIALS AND METHODS

α -Actinin from *D. discoideum* strain AX2-214 was purified essentially as in [6]. The isolation of cDNA clones containing α -actinin-specific sequences is described by Noegel et al. [4]. Since the clones did not contain a poly(A) sequence, the cDNA library was rescreened with an *EcoRI* fragment that was located near the carboxy-terminus.

Correspondence address: A. Noegel, Max Planck Institute for Biochemistry, 8033 Martinsried, FRG

The nucleotide sequence presented has been submitted to the EMBL/GenBank database under the accession number Y00689

1	TTTTTTGTATATTAAATTTTTTATTATTATTTTGTAAATTTTTTATTTTTTTGTAAATTAATAACCAACACATAAAAAACAAA																												84	
85	ATG	TCA	GAA	GAA	CCA	ACC	CCA	GTT	TCA	GGT	AAT	GAC	AAA	CAA	CTC	TTG	AAC	AAA	GCT	TGG	GAA	ATT	ACC	CAA	AAA	AAA	ACT	TTC	168	
1	M	S	E	E	P	T	P	V	S	G	N	D	K	Q	L	L	N	K	A	W	E	I	T	Q	K	K	T	F	28	
169	ACA	GCA	TGG	TGT	AAT	TCA	CAT	TTA	CGT	AAA	CTT	GGA	TCA	TCA	ATT	GAA	CAA	ATT	GAT	ACA	GAT	TTT	ACT	GAT	GGT	ATT	AAA	TTA	252	
29	T	A	W	C	N	S	H	L	R	K	L	G	S	S	I	E	Q	I	D	T	D	F	T	D	G	I	K	L	56	
253	GCT	CAA	TTA	TTA	GAA	GTT	ATT	TCA	AAT	GAT	CCA	GTA	TTT	AAA	GTA	AAC	AAA	ACA	CCA	AAA	TTA	AGA	AGA	ATT	CAT	AAT	ATC	CAA	336	
57	A	Q	L	L	E	V	I	S	N	D	P	V	F	K	V	N	K	T	P	K	L	R	I	H	N	I	Q	84		
337	AAT	GTT	GGT	CTC	TGT	TTA	AAA	CAT	ATT	GAA	TCA	CAT	GGT	GTT	AAA	TTG	GTT	GGT	ATT	GGT	GCT	GAA	GAG	TTA	GTT	GAT	AAA	AAC	420	
85	N	V	G	L	C	L	K	H	I	E	S	H	G	V	K	L	V	G	I	G	A	E	E	L	V	D	K	N	112	
421	TTA	AAG	ATG	ACT	TTG	GGT	ATG	ATT	TGG	ACA	ATC	ATT	CTT	CGT	TTT	GCC	ATT	CAA	GAT	ATT	TCA	ATT	GAA	GAA	TTG	AGT	GCC	AAA	504	
113	L	K	M	T	L	G	M	I	W	T	I	I	L	R	F	A	I	Q	D	I	S	I	E	E	L	S	A	K	140	
505	GAA	GCC	CTT	TTA	CTT	TGG	TGT	CAA	AGA	AAG	ACC	GAA	GGT	TAT	GAC	CGT	GTT	AAA	GTT	GGT	AAT	TTC	CAT	ACC	TCA	TTC	CAA	GAT	588	
141	E	A	L	L	L	W	C	Q	R	K	T	E	G	Y	D	R	V	K	V	G	N	F	H	T	S	F	Q	D	168	
589	GGT	CTT	GCC	TTT	TGT	GCT	CTC	ATC	CAT	AAA	CAT	AGA	CCA	GAT	TTA	ATC	AAC	TTT	GAC	TCT	TTA	AAC	AAA	GAT	GAT	AAA	GCT	GGT	672	
169	G	L	A	F	C	L	I	H	I	H	I	H	I	H	I	H	I	H	I	H	I	H	I	H	I	H	I	H	196	
673	AAC	TTA	CAA	TTG	GCT	TTT	GAT	ATT	GCC	GAA	AAA	GAA	TTG	GAT	ATC	CCA	AAG	ATG	TTG	GAT	GTT	TCC	GAT	ATG	CTC	GAT	GTC	GTT	756	
197	N	L	Q	L	A	F	D	I	A	E	K	E	L	D	I	P	K	M	L	D	V	S	D	M	L	D	V	V	224	
757	CGT	CCA	GAT	GAA	AGA	TCA	GTC	ATG	ACC	TAC	GTC	GCT	CAA	TAC	TAC	CAT	CAC	TTT	TCT	GCC	TCT	AGA	AAA	GCT	GAA	ACC	GCC	GGT	840	
225	R	P	D	E	R	S	V	M	T	Y	V	A	Q	Y	Y	H	H	F	S	A	S	R	K	A	E	T	A	G	252	
841	AAA	CAA	GTT	GGT	AAA	GTT	TTA	GAT	ACC	TTT	ATG	TTG	TTA	GAA	CAA	ACC	AAA	TCT	GAT	TAT	CTT	AAA	AGA	GCC	AAT	GAA	CTC	GTT	924	
253	K	Q	V	G	K	V	L	D	T	F	M	L	L	E	Q	T	K	S	D	Y	L	K	R	A	N	E	L	V	280	
925	CAA	TGG	ATT	AAC	GAT	AAA	CAA	GCA	TCA	CTT	GAA	TCA	CGT	GAT	TTT	GGT	GAT	TCC	ATC	GAA	TCT	GTT	CAA	AGT	TTC	ATG	AAC	GCT	1008	
281	Q	W	I	N	D	K	Q	A	S	L	E	S	R	D	F	G	D	S	I	E	S	V	Q	S	F	M	N	A	308	
1009	CAT	AAA	GAA	TAT	AAA	ACC	GAA	AAA	CCA	CCA	AAG	GCT	CAA	GAA	GTC	TCT	GAA	TTG	GAA	GCT	ATC	TAC	AAT	TCA	TTA	CAA	ACT	1092		
309	H	K	E	Y	K	K	T	E	K	P	P	K	G	Q	E	V	S	E	L	E	A	I	Y	N	S	T	Q	T	336	
1093	AAA	TTA	CGT	TTA	ATT	AAA	CGT	GAA	CCA	TTT	GTT	GCA	CCA	GCT	GGT	CTC	ACT	CCA	AAT	GAA	ATC	GAT	TCC	ACT	TGG	TCC	GCT	TTA	1176	
337	K	L	R	L	I	K	R	E	P	F	V	A	P	A	G	L	T	P	N	E	I	D	S	T	W	S	A	L	364	
1177	GAG	AAA	GCT	GAA	CAA	GAA	CAT	GCT	GAA	GCC	CTC	CGT	ATT	GAA	CTC	AAA	CGT	CAA	AAG	AAA	ATT	GCA	GTT	CTC	TTA	CAA	AAA	TAC	1260	
365	E	K	A	E	Q	E	H	A	E	A	L	R	I	E	L	K	R	Q	K	K	I	A	V	L	L	Q	K	Y	392	
1261	AAT	CGT	ATT	CTC	AAG	AAA	CTC	GAA	AAC	TGG	GCC	ACC	ACC	AAA	TCT	GTC	TAC	CTC	GGT	TCC	AAT	GAA	ACC	GGT	GAC	AGT	ATC	ACT	1344	
393	N	R	I	L	K	K	L	E	N	W	A	T	T	K	S	V	Y	L	G	S	N	E	T	G	D	S	I	T	420	
1345	GCT	GTT	CAA	GCT	AAA	TTA	AAG	AAT	TTA	GAA	GCT	TTT	GAT	GGT	GAA	TGT	CAA	TCA	TTG	GAA	GGT	CAA	TCA	AAC	TCT	GAT	CTC	CTC	1428	
421	A	V	Q	A	K	A	L	K	N	L	E	A	F	D	G	E	C	Q	S	L	E	G	Q	S	N	S	D	L	L	448
1429	AGC	ATT	CTT	GCT	CAA	TTA	ACT	GAA	CTC	AAC	TAC	AAT	GGT	GTA	CCA	GAA	CTC	ACT	GAA	CGT	AAA	GAT	ACA	TTC	TTT	GCT	CAA	CAA	1512	
449	S	I	L	A	Q	L	T	E	L	N	Y	N	G	V	P	E	L	T	E	R	K	D	T	F	F	A	Q	Q	476	
1513	TGG	ACT	GGT	GTT	AAA	TCA	TCT	GCT	GAA	ACC	TAC	Y	AAA	ACC	ACT	CTT	TTA	GCT	GAA	CTT	GAA	AGA	CTC	CAA	AAG	ATT	GAA	GAC	TCA	1596
477	W	T	G	V	K	S	S	A	E	T	Y	K	A	N	T	L	A	E	L	E	R	L	Q	K	A	I	G	D	S	504
1597	TTG	GTC	GAA	TTC	GCC	AAG	AGA	GCC	GCT	CAA	TTA	AAT	GTT	TGG	ATT	GAA	GCT	GCC	GAT	GAT	CAT	GTA	TTT	GAT	CCA	ATC	AAT	GTT	1680	
505	L	V	E	F	A	K	R	A	A	Q	L	N	V	W	I	E	A	A	D	D	H	V	F	D	P	I	N	V	532	
1681	GAC	TCT	GTT	CAA	GGT	GTC	CAA	GAA	ATT	CAA	GAG	AAA	TTT	GAC	GCT	ATC	CTC	CAC	GAT	CAA	TCA	CAA	CAA	TTC	GCT	GAA	TTG	GAA	1764	
533	D	S	V	Q	G	V	Q	E	I	Q	E	K	F	D	A	F	L	H	D	Q	S	Q	Q	F	A	E	L	E	560	
1765	GCC	CTC	GCT	GCT	TTA	ACT	CAA	CAA	CTC	CGT	GAA	CTC	GGT	CGT	TCT	GAA	AAC	GAT	TAT	TCA	GTC	ATT	TCA	TAC	GAT	GAA	CTC	TCT	1848	
561	A	L	A	A	L	T	Q	Q	L	R	E	L	G	R	S	E	N	D	Y	S	V	I	S	Y	D	E	L	S	588	
1849	GCC	AAA	TGG	AAT	AAT	TTA	TTG	GCT	GGT	ATT	GAA	GAA	CGT	AAA	GTT	CAA	CTC	GCC	AAT	GAA	CTC	ACC	ACT	CAA	ACC	AAT	AAC	GAT	1932	
589	A	K	W	N	N	L	L	A	G	I	E	E	R	K	V	Q	L	A	N	E	L	T	T	Q	T	N	N	D	616	
1933	GTT	CTT	TGC	CAA	TCA	TTC	TCT	GTT	AAA	GCA	AAT	GAA	ATT	TCA	GAT	TAT	GTC	CGT	GTT	ACT	TTA	GAT	GCC	ATC	TCA	CAA	AAC	ACT	2016	
617	V	L	C	Q	S	F	S	V	K	A	N	E	I	S	D	Y	V	R	V	T	L	D	A	I	S	Q	N	T	644	
2017	TCA	TCA	GAT	CCA	CAA	GAA	CAA	TTA	AAC	AAT	ATC	CGT	GCT	ATC	ATC	ACC	GCT	CAT	GCT	GAA	AAG	AAA	CCA	GAA	CTC	GAT	GAA	TTA	2100	
645	S	S	D	P	Q	E	Q	L	N	N	I	R	A	I	I	T	A	H	A	E	K	P	E	L	D	E	L	672		
2101	TAC	ACC	ATT	CGT	TCT	CAA	CTC	GAA	GAA	GCT	CAA	GTT	GTC	GAT	AAC	AAA	CAT	ACT	CAA	CAC	AGT	TTA	GAA	TCA	CTT	AAA	TTA	AAA	2184	
673	Y	T	I	R	S	Q	L	E	E	A	Q	V	V	D	N	K	H	T	Q	H	S	L	E	S	L	K	L	K	700	
2185	TGG	GAT	AAA	CTC	AAT	ACA	CTC	GCT	AAA	AAG	AAT	GAA	CAA	GTT	GTT	GAA	GGT	GAA	ATT	CTT	GCT	AAA	CAA	TTA	ACT	GGT	GTT	ACT	2268	
701	W	D	K	L	N	T	L	A	K	K	N	E	Q	V	V	E	G	E	I	L	A	K	Q	L	T	G	V	T	728	
2269	GCT	GAA	GAA	TTA	AGT	GAA	TTT	AAA	GCC	TGC	TTC	TCA	CAT	TTC	GAT	AAG	GAC	AAC	GAT	AAC	AAA	TTA	AAT	CGT	CTT	GAA	TTC	TCC	2352	
729	A	E	E	L	S	E	F	K	A	C	F	S	H	F	D	K	D	N	D	N	K	L	N	R	L	E	F	S	756	
2353	TCT	TGC	TTG	AAG	AGT	ATC	GGA	GAT	GAA	TTA	ACT	GAA	GAA	CAA	TTA	AAT	CAA	GTC	ATC	AGT	AAA	ATC	GAT	ACC	GAT	GGT	AAT	GGT	2436	
757	S	C	L	K	S	I	G	D	E	L	T	E	E	Q	L	N	Q	V	I	S	K	I	D	T	D	G	N	G	784	
2437	ACC	ATT	TCA	TTT	GAA	GAA	TTT	ATT	GAT	TAC	ATG	GTT	TCA	TCA	CGT	AAA	GGT	ACA	GAC	AGC	GTT	GAA	TCA	ACT	AAA	GCT	GCA	TTT	2520	
785	T	I	S	F	E	E	F	I	D	Y	M	V	S	S	R	K	G	T	D	S	V	E	S	T	K	A	A	F	812	
2521	AAA	GTT	ATG	GCT	GAG	GAT	AAA	GAT	TTT	ATT	ACT	GAA	GCT	CAA	ATT	CGT	GCT	GCT	ATC	TCT	GAT	TCT	AAA	CAA	ATT	GAT	TAT	TTA	2604	
813	K	V	M	A	E	D	K	D	F	I	T	E	A	Q	I	R	A	A	I	S	D	S	K	Q	I	D	Y	L	840	
2605	CTC	GCC	AGT	ATG	CCA	GCT	GTT	GAA	GGT	GGT	TTT	GAC	TAC	AAT	TCA	TTT	GCT	GAA	AAA	TTA	TAC	CAA	TAA	ACAAATAATTATTAAATTAT	2692					
841	L	A	S	M	P	A	V	E	G	G	F	D	Y	N	S	F	A	E	K	L	Y	Q							862	
2693	AAATACATTTTAAAAATTAAAAA***** 2725																													

The isolated clones were identical to those previously described and harboured in addition a poly(A) sequence. They hybridized to a single mRNA species of approx. 3.0 kb that was previously shown to be the α -actinin message [3]. Overlapping fragments of the cloned cDNAs were subcloned into vectors M13mp18/19 or into pUC18/19 [7]. The inserts were sequenced by the dideoxy chain termination method of Sanger et al. [8]. The sequences were analyzed with programs of the University of Wisconsin Genetic Computer Group (UWGCG).

3. RESULTS AND DISCUSSION

In fig.1 the cDNA sequence and derived protein sequence are shown. The protein sequence consists of 862 amino acids; the calculated molecular mass is 97.6 kDa and the N-terminus is blocked. Secondary structure prediction according to Chou-Fasman [9] suggests an α -helical content of about 50% over the entire protein. The region upstream of the methionine start codon exhibits a high AT content, a characteristic feature of non-coding DNA regions in *D. discoideum* [10].

The asterisks beneath the amino acids in regions 733–763 and 769–799 (fig.1) indicate calcium-binding loops whose functional features are in agreement with the EF-hand model for calcium-modulated proteins as proposed by Kretsinger [11]. Fig.2 aligns the sequences of the putative calcium-binding loops of *D. discoideum* α -actinin (rows e,f) with the corresponding regions of typical members from the calmodulin, troponin C, myosin light chain and parvalbumin families. Both domains contain a calcium-binding loop with all the necessary liganding oxygens and are close enough that their hydrophobic surfaces could impart mutual stability [11]. The search of the protein sequence database of the protein identification resource (PIR) showed in both cases the highest alignment scores for *Trypanosoma* calmodulin. *D. discoideum* calmodulin [12] does not exhibit a closer relationship to α -actinin; for EF-hand 1 in

α -actinin it scores slightly better than *Chlamydomonas* [13] or spinach [14] calmodulin, but for EF-hand 2 worse than these lower and higher plant calmodulins. The alignment scores suggest that α -actinin assumed the EF-hand structures very early in evolution, most likely before the separation of slime moulds and higher plants.

A partial amino acid sequence of chicken fibroblast α -actinin that was recently published [5] shows strong homology to the N-terminal third of *D. discoideum* α -actinin (residues 52–250). These two proteins are nearly identical between residues 97 and 151, where out of 55 amino acids 48 residues are identical and 5 residues are conservative replacements. It is possible that the complete sequence of the chicken fibroblast α -actinin will show the entire N-terminal portions of the molecules to be highly homologous. This strong homology of two α -actinins from unrelated organisms is also evident at the DNA level although to a lower extent (nucleotides 350–750). Using a probe from the *D. discoideum* α -actinin gene containing the highly conserved DNA stretch to test cross-hybridization to chicken DNA, we found three bands of about 1.35, 1.75 and 3.0 kb. These may represent different isoforms of chicken α -actinin or other crosslinking proteins with homologous F-actin-binding regions. Similar conclusions were drawn from the characterization of an actin-binding peptide that seems to be conserved in rat hepatic actinogelin, rat skeletal muscle and chicken gizzard α -actinins [15,16]. Limited similarities between *D. discoideum* α -actinin (residues 485–560) and non-erythroid α -spectrin [17] (residues 395–470) were found and possibly reflect functional sites for dimerization rather than common actin-binding sites. No significant sequence homologies to the actin-severing protein gelsolin [18], myosins or actins (taken from the Computer Genbank, EMBL, Feb.1987) were detected. These proteins apparently bind to sites different from the α -actinin-binding site on the actin molecule.

An important question is how the actin-binding

Fig.1. Nucleotide and deduced amino acid sequence of *D. discoideum* α -actinin cDNA. The underlined amino acids denote the peptides which were isolated from a tryptic digest of α -actinin and sequenced by Edman degradation [3]. At the 3'-end of the coding region a polyadenylation signal (underlined nucleotide sequence) overlaps with the last codon followed by a poly(A) stretch 36 nucleotides downstream from the stop codon. The EF-hand regions are indicated with stars.

A)

a)		0	1	2	3	4	5	6	7	8	9	0	1	2	3	4	5	6	7	8	9	0												
b)			E	L'			L'L'			L'D'	D'	D'G	I'D'		E	L'			L'L'		L'													
c)			*	*			*			*	*	*	*		*	*			*		*													
d)										X	Y	Z	-Y	-X		-Z																		
e)	E	E	L	S	E	F	K	A	C	F	S	H	F	D	K	D	N	D	N	K	L	N	R	L	E	F	S	S	C	L	K	S	I	G
CaM	E	Q	I	S	E	F	K	E	A	F	S	L	F	D	K	D	G	D	G	T	I	T	T	K	E	L	G	T	V	M	R	S	L	G
TnC	E	M	I	A	E	F	K	A	A	F	D	M	F	D	T	D	G	G	G	D	I	S	T	K	E	L	G	T	V	M	R	M	L	G
MLC	E	Q	Q	D	E	F	K	E	A	F	L	L	Y	D	R	T	G	D	S	K	I	T	L	S	Q	V	G	D	V	L	R	A	L	G

B)

b)					E	L'				L'L'				L'D'	D'		D'G			I'D'			E	L'				L'L'			L'			
c)					*					*	*			*	*		*	*		*	*		*	*		*	*		*	*				
d)														X	Y		Z	-Y		-X			-Z											
f)	L	T	E	E	Q	L	N	Q	V	I	S	K	I	D	T	D	G	N	G	T	I	S	F	E	E	F	I	D	Y	M	V	S	S	R
CaM	L	T	D	E	E	V	D	E	M	I	R	E	A	D	V	D	G	D	G	Q	I	N	Y	E	E	F	V	K	M	M	M	S	K	*
TnC	P	T	K	E	E	L	D	A	I	I	E	E	V	D	E	D	G	S	G	T	I	D	F	E	E	F	L	V	M	M	V	R	Q	M
S-100	K	E	Q	E	V	V	D	K	V	M	E	T	L	D	S	D	G	D	G	E	C	D	F	Q	E	F	M	A	F	V	A	M	I	T
Parv.	L	T	D	K	E	T	K	D	L	L	I	K	G	D	K	D	G	D	G	K	I	G	V	D	E	F	T	S	L	V	A	E	S	*

Fig.2. Alignment of EF-hand regions from α -actinin and known calcium-modulated proteins. According to the EF-hand hypothesis [11] there are 16 characteristic positions (row b) building an EF-hand structure. A true EF-hand domain scores 12 or better out of 16 [20]. Row a shows the recommended numbering [11], the stars (rows c and boxed residues) indicate the correct alignment and the liganding oxygens that correspond to the octahedral vertices (rows d). Panel A aligns residues 730–763 from *D. discoideum* α -actinin (row e) with calmodulin (*Trypanosoma brucei gambiense* [21], residues 7–40), skeletal muscle troponin C (*Rana esculenta* [22], residues 17–50) and skeletal muscle myosin A1 and A2 catalytic light chains (rabbit [23], residues 47–80). Panel B shows the alignment of α -actinin (residues 766–799; row f) with the same calmodulin (residues 116–148) and troponin C (residues 53–86) as in panel A, and in addition with S-100 protein (β -chain, bovine [24], residues 48–81) and parvalbumin (*Amphiuma means* [25], residues 77–109).

activity of non-muscle α -actinin is regulated by calcium. The highly conserved sequence in the N-terminal portion of *D. discoideum* α -actinin is separated by 500 amino acids from the EF-hand structures and the two ends of the polypeptide are also separated in space. Based on EM data from α -actinin-antibody complexes, the rod-shaped α -actinin molecule consists of two filamentous subunits that extend from one end to the other in an anti-parallel fashion [19]. Thus, it is unlikely that the EF-hands interact with the N-terminal

region of the same subunit. However, as outlined in fig.3, the EF-hand of one subunit could be near the N-terminus of the other subunit and affect the binding to actin. To clarify this inhibitory mechanism we plan to change the structure of the putative actin-binding site or the calcium-binding domains by genetic engineering. Using a strain that does not produce α -actinin, we hope to construct transformants that express an α -actinin which is not regulated by calcium.

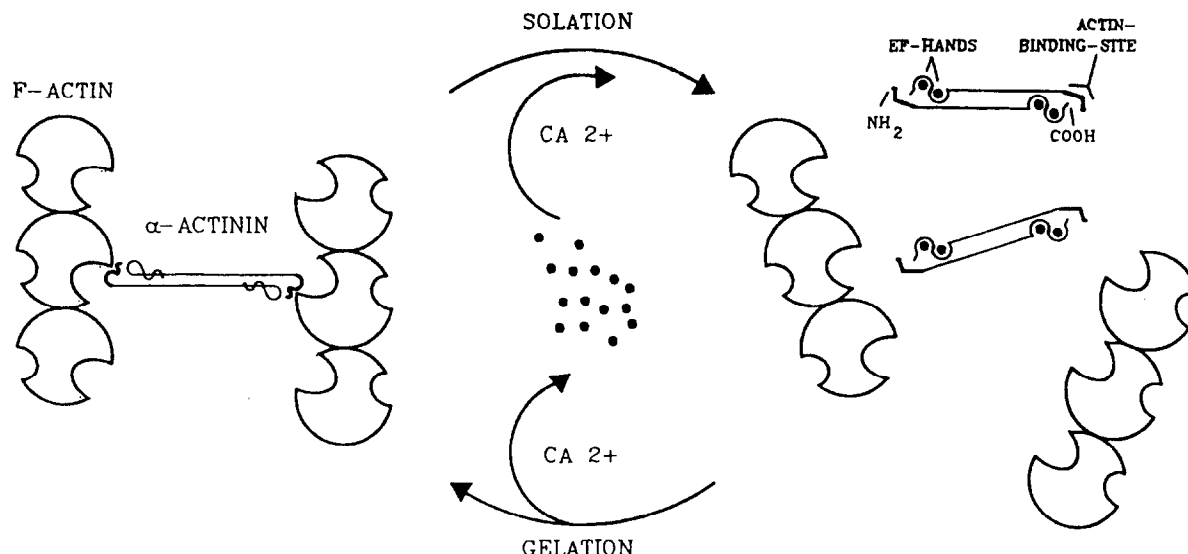


Fig.3. Proposed model for the putative interaction between the calcium- and actin-binding sites in *D. discoideum* α -actinin. The α -actinin molecule is drawn as a homodimer, whose anti-parallel subunits form a rod-shaped molecule. The binding site to F-actin resides in the N-terminal portion of the polypeptide (bold region), and the EF-hand structures at the C-terminus. In analogy to studies based on the crystal structures of troponin C [26] and calmodulin [27], the calcium-binding domains of α -actinin may wrap around the central part of the α -actinin subunit provided calcium is not present. Following an increase in the internal calcium concentration each EF-hand structure binds one Ca^{2+} and changes its conformation by folding back from the central part of the subunit, thus disturbing the interaction of the actin-binding site of the neighbouring subunit with F-actin.

ACKNOWLEDGEMENTS

We are grateful to Dr G. Gerisch for helpful discussion and advice. We thank Drs J. Segall, F. Lottspeich and G. Isenberg for critical comments on the manuscript, Drs R. Kessin and M.-L. Lacombe for generously providing the cDNA library and M. Becker for chicken DNA.

REFERENCES

- [1] Stossel, T.P., Chaponnier, C., Ezzell, R.M., Hartwig, J.H., Janmey, P.A., Kwiatkowski, D.J., Lind, S.E., Smith, D.B., Southwick, F.S., Yin, H.L. and Zaner, K.S. (1985) *Annu. Rev. Cell Biol.* 1, 353–402.
- [2] Pollard, T.D. and Cooper, J.A. (1986) *Annu. Rev. Biochem.* 55, 987–1035.
- [3] Witke, W., Schleicher, M., Lottspeich, F. and Noegel, A. (1986) *J. Cell Biol.* 103, 969–975.
- [4] Noegel, A., Witke, W. and Schleicher, M. (1986) *FEBS Lett.* 204, 107–109.
- [5] Baron, M.D., Davison, M.D., Jones, P., Patel, B. and Critchley, D.R. (1987) *J. Biol. Chem.* 262, 2558–2561.
- [6] Schleicher, M., Gerisch, G. and Isenberg, G. (1984) *EMBO J.* 3, 2095–2100.
- [7] Messing, J. and Vieira, J. (1982) *Gene* 19, 269–276.
- [8] Sanger, F., Nicklen, S. and Coulson, A.R. (1977) *Proc. Natl. Acad. Sci. USA* 74, 5463–5468.
- [9] Chou, P.Y. and Fasman, G.D. (1979) *Biophys. J.* 26, 367–384.
- [10] Kimmel, A.R. and Firtel, R.A. (1983) *Nucleic Acids Res.* 11, 541–552.
- [11] Kretsinger, R.H. (1980) *CRC Crit. Rev. Biochem.* 8, 119–174.
- [12] Marshak, D.R., Clarke, M., Roberts, D.M. and Watterson, D.M. (1984) *Biochemistry* 23, 2891–2899.
- [13] Lukas, T.J., Wiggins, M.E. and Watterson, D.M. (1985) *Plant Physiol.* 78, 477–483.
- [14] Lukas, T.J., Iverson, D.B., Schleicher, M. and Watterson, D.M. (1984) *Plant Physiol.* 75, 788–795.
- [15] Mimura, N. and Asano, A. (1986) *J. Biol. Chem.*

- 261, 10680–10687.
- [16] Mimura, N. and Asano, A. (1987) *J. Biol. Chem.* 262, 4717–4723.
- [17] Birkenmeier, C.S., Bodine, D.M., Repasky, E.A., Helfman, D.M., Hughes, S.H. and Barker, J.E. (1985) *Proc. Natl. Acad. Sci. USA* 82, 5671–5675.
- [18] Kwiatkowski, D.J., Stossel, T.P., Orkin, S.H., Mole, J.E., Colten, H.R. and Yin, H.L. (1986) *Nature* 323, 455–488.
- [19] Wallraff, E., Schleicher, M., Modersitzki, M., Rieger, D., Isenberg, G. and Gerisch, G. (1986) *EMBO J.* 5, 61–67.
- [20] Kretsinger, R.H. (1980) *Ann. NY Acad. Sci.* 356, 14–19.
- [21] Tschudi, C., Young, A.S., Ruben, L., Patton, C.L. and Richards, F.F. (1985) *Proc. Natl. Acad. Sci. USA* 82, 3998–4002.
- [22] Van Eerd, J.-P., Capony, J.-P., Ferraz, C. and Pechere, J.-F. (1978) *Eur. J. Biochem.* 91, 231–242.
- [23] Frank, G. and Weeds, A.G. (1974) *Eur. J. Biochem.* 44, 317–334.
- [24] Marshak, D.R., Umekawa, H., Watterson, D.M. and Hidaka, H. (1985) *Arch. Biochem. Biophys.* 240, 777–780.
- [25] Maeda, N., Zhu, D. and Fitch, W.M. (1984) *Mol. Biol. Evol.* 1, 473–488.
- [26] Herzberg, O. and James, M.N.G. (1985) *Nature* 313, 653–659.
- [27] Babu, Y.S., Sack, J.S., Greenhough, T.J., Bugg, C.E., Means, A.R. and Cook, W.J. (1985) *Nature* 315, 37–40.