

Etiologies of sperm oxidative stress

Parvin Sabeti Ph.D.Candidate, Soheila Pourmasumi Ph.D.Candidate, Tahereh Rahiminia Ph.D.Candidate, Fatemeh Akyash Ph.D.Candidate, Ali Reza Talebi Ph.D.

Research and Clinical Center for Infertility, Shahid Sadoughi University of Medical Sciences, Yazd, Iran.

Corresponding Author:

Ali Reza Talebi, Research and Clinical Center for infertility, Bouali Ave., Yazd, Iran.
Email: prof_talebi@ssu.ac.ir;
prof_talebi@hotmail.com
Tel: (+98) 35 38247085

Received: 23 August 2015
Revised: 3 December 2015
Accepted: 31 January 2016

Abstract

Sperm is particularly susceptible to reactive oxygen species (ROS) during critical phases of spermiogenesis. However, the level of seminal ROS is restricted by seminal antioxidants which have beneficial effects on sperm parameters and developmental potentials. Mitochondria and sperm plasma membrane are two major sites of ROS generation in sperm cells. Besides, leukocytes including polymorphonuclear (PMN) leukocytes and macrophages produce broad category of molecules including oxygen free radicals, non-radical species and reactive nitrogen species. Physiological role of ROS increase the intracellular cAMP which then activate protein kinase in male reproductive system. This indicates that spermatozoa need small amounts of ROS to acquire the ability of nuclear maturation regulation and condensation to fertilize the oocyte. There is a long list of intrinsic and extrinsic factors which can induce oxidative stress to interact with lipids, proteins and DNA molecules. As a result, we have lipid peroxidation, DNA fragmentation, axonemal damage, denaturation of the enzymes, over generation of superoxide in the mitochondria, lower antioxidant activity and finally abnormal spermatogenesis. If oxidative stress is considered as one of the main cause of DNA damage in the germ cells, then there should be good reason for antioxidant therapy in these conditions.

Key words: Oxidative stress, Sperm, Reactive oxygen species, Etiology.

Introduction

Male fertility depends on spermatogenesis process which produces the large numbers of cell by the testis known as spermatozoa (1). Oxygen is important for aerobic metabolism of spermatogenic cells, but this molecule may have detrimental effects on cells via production of ROS. Significant positive correlation has reported between levels of reactive oxygen species (ROS) and percentage of spermatozoa with many kinds of abnormalities like, abnormal heads, acrosome abnormalities, mid piece anomalies, cytoplasmic droplets and tail defects (2). In fact, enhancement of ROS level in semen produced by abnormal spermatozoa can be a main cause of sub-fertility and even infertility. For example, teratozoospermic men who have a high level of ROS in their semen, show subfertility or even infertility (3, 4). ROS production rate is a main parameter in oxidative stress condition, which is the imbalance between ROS level and body's antioxidant capacity (5-7).

Sperm, as well as critical phases of spermiogenesis are particularly susceptible to

ROS induced damages for several reasons including; a) The period of sperm chromatin condensation is very susceptible, b) Sperm cells don't have DNA repair mechanisms, c) Sperm membrane contain high levels of poly unsaturated fatty acids, d) Sperm themselves produce ROS, especially during passage through epididymis, e) Sperm possess low levels of cytoplasmic antioxidant enzymes (because most of the antioxidant enzymes are lost in spermiogenesis) and f) Sperm spend long periods as isolated cells in both male and female genital tracts (8-12). However, the level of seminal ROS is restricted by seminal antioxidants such as β -mercaptoethanol, protein, vitamin E, vitamin C, cysteamine, cycteine, taurin and hypotaurin (13-24). In fact, these compounds have beneficial effects on sperm parameters and developmental ability of embryos (25).

In this article, we reviewed the free radicals actions in male reproductive system and the effects of antioxidants on oxidative stress and fertility potential of spermatozoa.

Sources of ROS

Different cell types in human ejaculate including seminal leukocytes and abnormal

spermatozoa have been shown to be the main sources of ROS generation (26). It has been demonstrated that the morphologically abnormal spermatozoa are very active in ROS productions (27). Between different sperm anomalies, residual cytoplasm or cytoplasmic droplet may be considered as most important one in ROS production (28). Normally, these excessive cytoplasm are omitted by Sertoli cells during spermiogenesis (29, 30). However, the spermatozoa with this abnormality are immature and functionally retarded (31, 32). Studies have shown that ROS generation in cytoplasmic droplets is mediated by cytosolic enzyme, called "glucose-6-phosphate dehydrogenase" (G6PD), which induces high levels of ROS production during two pathways, I) nicotinamide adenine dinucleotide phosphate (NADPH) located in sperm plasma membrane and II) NADPH-dependent oxido-reductase known as diphorase located in middle piece at mitochondrial respiratory level (33-37). In fact, mitochondria and sperm plasma membrane are two major sites of ROS generation in sperm cells (38).

There are large numbers of mitochondria in middle piece of sperm cells which supply energy for their motility. Mitochondrial dysfunction is related to ROS production and ROS can affect mitochondrial integrity in spermatozoa, that is a mutual cycle; ROS causing injury to mitochondrial membrane and injured mitochondrial membrane causing an enhancement in ROS production (35, 39). On the other hand, in a recent study, it was shown that NADPH oxidase 5 (NOX5) of spermatozoa plays an important role in ROS production (40).

Another source of ROS in seminal fluid is Peroxides-positive leukocytes including polymorphonuclear (PMN) leukocytes (50-60% of all seminal leukocytes) and macrophage (40-50% of the rest) (41, 42). It has shown that activated leukocytes in response to different inducers like infection and inflammation can produce up to 100 fold higher levels of ROS compared with non-activated leukocytes (8, 43). The World Health Organization (WHO) defines "Leukocytospermia" as the increased leukocytes infiltration in semen with concentration more than $1 \times 10^6/\text{ml}$ (44).

Types of ROS

Reactive oxygen species represent a broad category of molecules including: a) Oxygen free radicals, such as superoxide anion (O_2^-), hydroxyl radical (OH) and hyperoxyl radical (HOO^\bullet). b) Non radical species, such as hypochlorous acid (HOCl) and hydrogen peroxide (H_2O_2). c) Reactive nitrogen species and free nitrogen radicals such as nitroxyl, nitrous oxide, peroxynitrite, etc. (1, 45, 46).

Physiological role of ROS in male reproductive system

Aitken and his group were the pioneers of research and study in the field of physiological role of ROS, in male reproductive system (47). Several study, have indicated that spermatozoa needs small amounts of ROS, to acquire the ability of fertilizing the oocytes (48-51). It is also demonstrated that spermatozoa need a small amounts of ROS, for capacitation, hyperactivation, motility, acrosome reaction and fertilization (52, 53). In fact, ROS, along with several factors in spermatozoa, increase intracellular cAMP which then activates protein Kinase A. These changes, in turn, increase tyrosine phosphorylation, that is the major driving force for capacitation (54).

In addition, increase in cAMP levels, leading to an increase in sperm motility or hyper activation. Also, as a result of capacitation, acrosome membrane becomes unstable, then several hydrolytic enzymes like acrosin are released during acrosomal reaction and allowing sperm to binds oocyte (55, 56). It has been suggested that, ROS, take part in the regulation of nuclear maturation in spermatozoa. According to this, these reactive agents produce the lipid peroxides, and they probably provide a substrate for GPX4 (Phospholipid Hydroperoxide Glutathione Peroxidase), and so cause the oxidation of nuclear proteins and they facilitate nuclear condensation (57). In fact GPX4 is able to use thiols in nuclear protein, as an alternative reductant to glutathione (58).

Etiologies of oxidative stress

Although, there is a long list of intrinsic and extrinsic factors which can induce oxidative

stress, but the main generally accepted etiologies are the followings:

Alcohol consumption

Several studies have shown that alcohol consumption, can increase the abnormalities in nucleous and plasma membrane of spermatozoa (59, 60). In an experimental study, it was shown that ethanol consumption increases the sperm cells percentage with chromatin abnormalities (61). In alcohol metabolism pathway, NADH and acetaldehyde are produced which, NADH increases the activity of respiratory chain in mitochondria and acetaldehyde interacts with lipids and proteins to produce ROS (62, 63).

Cigarette smoking

Cigarette smoking can decrease the sperm motility, number of normal sperm and produce reactive oxygen species by lipid peroxidation (64-67). In addition, it can decrease the antioxidants level, for example: vitamin E, vitamin C and increase in ROS level of seminal plasma (61, 68-71). Also, cigarette smoking can cause the inflammatory reaction and increase in leukocytes number in testicles (72, 73). The other changes that have been observed in smokers are including DNA fragmentation, axonemal damage, and reducing the sperm number (74-77).

Varicocele

Several studies have shown, that the oxidative stress in varicocele men could be the result of both increase in the level of ROS, and decrease in the total antioxidant capacity (78-80). In fact, high levels of ROS, and low levels of total antioxidant capacity, lead to impairment of cell membrane structure and DNA integrity of spermatozoa (30, 81). Talebi *et al* indicate that the varicocele patients have more spermatozoa with abnormal chromatin condensation than fertile controls (82). In addition, Ha *et al* showed that ROS may cause damage to blood-testis barrier in varicocele patients (83). Other studies have shown the presence of NO, in spermatic vein of varicocele men. NO, is a lipophilic molecule and it has cytotoxic effects on adjacent sperm cells. In addition, NO and superoxide which released by monocytes form peroxynitrite cause more spermatozoa damage (84, 85). Fisher *et al* stated that, sperm cells of varicocele patients have high cytoplasmic

droplets which produce the high levels of ROS (86).

Obesity

Anthony *et al* demonstrated that ROS overproduction, and abnormal hormonal regulation in obese men lead to suboptimal semen quality (87). It is suggested that dysregulation of adipocytokine and ROS generation, are the causes of oxidative stress in these patients (88). Over production of ROS, could be due to increase the metabolic rates, and maintain hemostasis in obese men. Besides, increase in ROS production and temperature, in testicles, may denature the enzymes, implicated in spermatogenesis. Hjollund *et al* stated that, increasing the temperature of scrotal skin was associated with decrease in sperm concentration (89). Another study, indicate the negative correlation between sperm density and total count (90).

Diabetes

For the first time, Baynes linked the diabetes mellitus to ROS (91). In diabetic patients, we have the increase levels of ROS, and impairment of antioxidant defense capacity (92). However, oxidative stress which is related to hyperglycemia is due to over generation of superoxide in mitochondria (93, 94). Agbaje *et al* stated that, the level of sperm DNA fragmentation in diabetic men is higher than normal men (95). On the other hand, there are some data indicating the effects of experimentally-induced diabetes on sperm chromatin quality and DNA integrity (96-99).

Physical exercise

An interesting issue is that too much exercises, due to muscle aerobic metabolism, produce large amounts of ROS, which lead to oxidative stress (100). Manna *et al* observed that the rates of exercises are related to reduction in sperm motility and count which may be the result of testicular oxidative stress (101).

Psychological stress

Several studies have indicated that, psychological stresses increase the level of ROS in seminal plasma and decrease the antioxidant protection capacities which in turn lead to reduction in sperm quality. It is

suggested that these negative effects of psychological stresses may be related to central destruction of gonadotropin drive (102-104).

Aging

Desai *et al* stated that, ROS production has a central role in age-related reduction of male fertility by affecting on aging biomarkers (105). In addition, several studies have shown, increasing sperm DNA damage with age in fertile and infertile men (106-108). An experimental study revealed that spermatozoa from older animals produce more free radicals compared with younger ones and the former have lower antioxidant activity, too (109, 110).

Environment factors

Environment pollutants may be considered as one of the main source of ROS production. De Rosa *et al*, indicated that NO and lead can diminish the seminal quality, and motor vehicles by releasing NO, have a negative effect on male fertility (111). It is shown that Pb can decrease sperm normal morphology, count and viability (112). Also, it was shown that, butyl benzyl phalate has toxic effect on testis and reduces the testosterone level in serum (113, 114). Lee *et al* in a study on rats, indicated that oral administration of phalate esters, increased the ROS production and reduce the level of antioxidant in testis and finally it causes disturbances in spermatogenesis (115).

Electro-magnetic waves from cell phones, by ROS production, have deleterious effect on sperm. In one study, the semen samples were exposed to radiofrequency electromagnetic waves and a significant reduction in sperm motility, viability, and an increase in the level of ROS, with reduction in ROS-TAC score were found (116-118).

Infections

ROS has endogenous and exogenous sources. The endogenous ROS produces by immature sperm and leukocytes of semen (50). The male reproductive ducts infections and inflammatory reactions can be considered as exogenous sources (119). A few studies demonstrated the elevation of ROS concentration in infectious diseases. Mazilli *et al* demonstrated that, in patients with sperm culture-positive for aerobic bacteria, the

superoxide anion production was high (120). Also, a high ROS level in chronic non-bacterial inflammation was seen (121).

The major sources of seminal ROS are polymorphonuclear leukocytes. In fact, the bacterial products and cytokines, can increase the ROS generation in these cells (122, 123). In a variety of human infectious, ROS, are produced generally by viruses besides bacteria and parasites (124). When a virus enters the cell, it impairs the cellular functions and it leads to an inequality in ROS system (125). It has been shown that urinary tract infections cause an elevation in ROS level in sperm fraction after percoll separation (126). There is a strong relation between inflammation of male genital system and infertility (127, 128). Actually, ROS that are produced in testis infection and epididymis are dangerous for sperm, because, antioxidant protection in sperm is low and ROS can affect these cells in a long period (129, 130).

Antioxidant and male fertility

There is a balance between ROS production and antioxidant activity in male reproductive system in normal conditions. But overproduction of ROS in semen can affect sperm or seminal plasma antioxidant defense mechanisms and cause oxidative stress (50, 131, 132). The body has developed antioxidant defense system by scavenging and minimizing the formation of oxygen derived radicals to protect itself from oxidative damage (133, 134). Although, ROS have both physiological and pathological function, to keep this level in confident range, the human body has a system against ROS (135, 136). In fact, when the free radicals level increases pathologically, antioxidant activity activated to prevent ROS oxidative damage (137).

Seminal plasma has endogenous antioxidant for protecting spermatozoa from oxidative damage (138, 139). These antioxidants are divided to enzymatic and non-enzymatic antioxidant and male reproductive system has both antioxidant (1). Enzymatic antioxidant contains superoxide dismutase, catalase, and peroxidase that catalytically remove reactive oxygen species from biological systems. Sperm themselves

predominantly process this enzymatic antioxidant (140).

Total seminal antioxidant activity is also supplemented by non-enzymatic antioxidants in semen. Non-enzymatic antioxidants in semen are usually present in the form of vitamin C, vitamin E, beta carotenes, carotenoids, flavonoids and metal binding proteins such as albumin, ferritin, and myoglobin, that act as antioxidant by inactivating pro-oxidant transition metal ions (141-143). Seminal plasma has the main antioxidant role, defending the spermatozoa from the ROS produced by the immature sperm cells and leukocytes through non-enzymatic scavengers existing in semen (144, 145). However, the spermatozoa have minimum antioxidant enzymes. In addition, antioxidant enzymes in sperm cannot prevent tail and acrosome membranes from lipid peroxidation. In other word, the sperm cells also need extra antioxidant defense system (146).

It is generally accepted that antioxidant therapy can improve the sperm quality and male fertility. Vitamin C and its antioxidant property for improvement of sperm parameters is confirmed by several studies support the positive effects of vitamin C on different sperm parameters (147-149). It is shown that the addition of vitamin C and vitamin E to the sperm of normozoospermic and asthenozoospermic men reduce ROS induced DNA damage (16, 150). Treatment by vitamin E for 6 months can reduce the lipid peroxidation of spermatozoa and may increase the pregnancy rate in asthenozoospermic cases (151).

Additionally, coadministration of vitamin E and selenium increases the sperm motility in infertile men (152, 153). It is also demonstrated that the administration of zinc, vitamin C and vitamin E in asthenozoospermic patients reduces oxidative stress, apoptosis and sperm DNA fragmentation index (154-156). Many studies have indicated the improvement in sperm count and motility of asthenozoospermic and oligospermic patient after carnitine intakes (157-162). Alone or co-administration of zinc and folic acid increases the sperm count in infertile men but not in fertiles (163, 164).

Conclusion

In normal condition male reproductive system has a balance between ROS production and antioxidant activity. But, overproduction of ROS in semen can affect sperm or seminal plasma antioxidant defense mechanisms and cause oxidative stress. It is approved that oxidative stress elevates sperm chromatin/ DNA damage. A variety of several etiological factors including intratesticular, post-testicular, and external factors, such as alcohol, cigarette smoking, varicocele, diabetes and etc. have been correlated with increased levels of ROS and sperm DNA damage, and in turn, can affect the potential of male fertility. It is generally accepted that antioxidant therapy can improve the sperm quality and male fertility by reducing oxidative stresses.

Conflict of interest

All investigators disclose no conflict of interest in this study.

References

1. Sikka SC. Relative impact of oxidative stress on male reproductive function. *Cur Med Chem* 2001; 8: 851-862.
2. Kobayashi CI, Suda T. Regulation of reactive oxygen species in stem cells and cancer stem cells. *J Cell Physiol* 2012; 227: 421-430.
3. Rato L, Alves MG, Socorro S, Duarte AI, Cavaco JE, Oliveira PF. Metabolic regulation is important for spermatogenesis. *Nature Rev Urol* 2012; 9: 330-338.
4. Agarwal A, Said TM. Oxidative stress, DNA damage and apoptosis in male infertility: a clinical approach. *BJU Int* 2005; 95: 503-507.
5. Mostafa T, Anis T, El-Nashar A, Imam H, Othman I. Varicolectomy reduces reactive oxygen species levels and increases antioxidant activity of seminal plasma from infertile men with varicocele. *Int J Androl* 2001; 24: 261-265.
6. Agarwal A, Prabakaran SA, Said TM. Prevention of oxidative stress injury to sperm. *J Androl* 2005; 26: 654-660.
7. Sikka SC, Rajasekaran M, Hellstrom WJ. Role of oxidative stress and antioxidants in male infertility. *J Androl* 1995; 16: 464-468.
8. Agarwal A, Makker K, Sharma R. Clinical relevance of oxidative stress in male factor infertility: an update. *Am J Reprod Immunol* 2008; 59: 2-11.
9. Lewis S, Aitken R. DNA damage to spermatozoa has impacts on fertilization and pregnancy. *Cell Tissue Res* 2005; 322: 33-41.
10. Chenoweth P. Influence of the male on embryo quality. *Theriogenology* 2007; 68: 308-315.

11. Sipinen V. Exploring co-genotoxicity of chemicals using traditional experimental methods and microarray systems. Available at: <https://www.duo.uio.no/handle/10852/11834>.
12. Prakash S, Prithiviraj E, Suresh S, Lakshmi NV, Ganesh MK, Anuradha M, et al. Morphological diversity of sperm: A mini review. *Iran J Reprod Med* 2014; 12: 239-242.
13. Kitagawa Y, Suzuki K, Yoneda A, Watanabe T. Effects of oxygen concentration and antioxidants on the in vitro developmental ability, production of reactive oxygen species (ROS), and DNA fragmentation in porcine embryos. *Theriogenology* 2004; 62: 1186-1197.
14. Luberda Z. The role of glutathione in mammalian gametes. *Reprod Biol* 2005; 5: 5-17.
15. Alkan I, Simsek F, Haklar G, Kervancioglu E, Ozveri H, Yalcin S, et al. Reactive oxygen species production by the spermatozoa of patients with idiopathic infertility: relationship to seminal plasma antioxidants. *J Urol* 1997; 157: 140-143.
16. Rolf C, Cooper T, Yeung C, Nieschlag E. Antioxidant treatment of patients with asthenozoospermia or moderate oligoasthenozoospermia with high-dose vitamin C and vitamin E: a randomized, placebo-controlled, double-blind study. *Hum Reprod* 1999; 14: 1028-1033.
17. Momeni HR, Soleimani Mehranjan M, Abnosi MH, Mahmoodi M. Effects of vitamin E on sperm parameters and reproductive hormones in developing rats treated with para-nonylphenol. *Iran J Reprod Med* 2009; 7: 111-116.
18. Sies H, Stahl W. Vitamins E and C, beta-carotene, and other carotenoids as antioxidants. *Am J Clin Nutr* 1995; 62: 1315S-1321S.
19. Fanaei H, Khayat S, Halvaei I, Ramezani V, Azizi Y, Kasaeian A, et al. Effects of ascorbic acid on sperm motility, viability, acrosome reaction and DNA integrity in teratozoospermic samples. *Iran J Reprod Med* 2014; 12: 103-110.
20. Bucak MN, Ateşşahin A, Varışlı Ö, Yüce A, Tekin N, Akçay A. The influence of trehalose, taurine, cysteamine and hyaluronan on ram semen: microscopic and oxidative stress parameters after freeze-thawing process. *Theriogenology* 2007; 67: 1060-1067.
21. Bucak MN, Tuncer PB, Sarıözkan S, Ulutaş PA, Çoyan K, Başpınar N, et al. Effects of hypotaurine, cysteamine and aminoacids solution on post-thaw microscopic and oxidative stress parameters of Angora goat semen. *Res Vet Sci* 2009; 87: 468-472.
22. Safarinejad MR, Safarinejad S. Efficacy of selenium and/or N-acetyl-cysteine for improving semen parameters in infertile men: a double-blind, placebo controlled, randomized study. *J Urol* 2009; 181: 741-751.
23. Holmes RP, Goodman HO, Shihabi ZK, Jarow JP. The taurine and hypotaurine content of human semen. *J Androl* 1992; 13: 289-292.
24. Chen Y, Foote R, Brockett C. Effect of sucrose, trehalose, hypotaurine, taurine, and blood serum on survival of frozen bull sperm. *Cryobiology* 1993; 30: 423-431.
25. Agarwal A, Gupta S, Sikka S. The role of free radicals and antioxidants in reproduction. *Current Opin Obstet Gynecol* 2006; 18: 325-332.
26. Aitken RJ, Baker HG. Andrology: Seminal leukocytes: passengers, terrorists or good Samaritans? *Hum Reprod* 1995; 10: 1736-1739.
27. Tomlinson M, White A, Barratt C, Bolton A, Cooke I. The removal of morphologically abnormal sperm forms by phagocytes: a positive role for seminal leukocytes? *Hum Reprod* 1992; 7: 517-522.
28. Zini A, Defreitas G, Freeman M, Hechter S, Jarvi K. Varicocele is associated with abnormal retention of cytoplasmic droplets by human spermatozoa. *Fertil Steril* 2000; 74: 461-464.
29. Gomez E, Buckingham DW, Brindle J, Lanzafame F, Irvine DS, Aitken RJ. Development of an image analysis system to monitor the retention of residual cytoplasm by human spermatozoa: correlation with biochemical markers of the cytoplasmic space, oxidative stress, and sperm function. *J Androl* 1996; 17: 276-287.
30. Moein MR, Soleimani M, Tabibnejad N. Reactive oxygen species (ROS) production in seminal fluid correlate with the severity of varicocele in infertile men. 2008; 6: 65-69.
31. Doohar GB. Differences in the rate of redistribution of receptors for concanavalin A in vivo and in vitro on spermatozoa from normal mice and from sterile mice carrying different T/t locus haplotypes. *Gamete Res* 1981; 4: 105-111.
32. Mann T, Lutwak-Mann C. Biochemistry of Spermatozoa: Chemical and Functional Correlations in Ejaculated Semen, Andrological Aspect. Male Reproductive Function and Semen, Springer; 1981: 195-268.
33. Park J, Rho HK, Kim KH, Choe SS, Lee YS, Kim JB. Overexpression of glucose-6-phosphate dehydrogenase is associated with lipid dysregulation and insulin resistance in obesity. *Mol Cell Biol* 2005; 25: 5146-5157.
34. Frederiks WM, Vreeling-Sindelárová H. Localization of glucose-6-phosphate dehydrogenase activity on ribosomes of granular endoplasmic reticulum, in peroxisomes and peripheral cytoplasm of rat liver parenchymal cells. *Histochem J* 2001; 33: 345-353.
35. Sanocka D, Kurpisz M. Reactive oxygen species and sperm cells. *Reprod Biol Endocrinol* 2004; 2: 1-7.
36. Gavella M, Lipovac V. NADH-dependent oxidoreductase (diaphorase) activity and isozyme pattern of sperm in infertile men. *Syst Biol Reprod Med* 1992; 28: 135-141.
37. Siegel D, Gibson NW, Preusch PC, Ross D. Metabolism of diaziquone by NAD (P) H:(quinone acceptor) oxidoreductase (DT-diaphorase): role in diaziquone-induced DNA damage and cytotoxicity in human colon carcinoma cells. *Cancer Res* 1990; 50: 7293-7300.
38. Marchetti C, Obert G, Deffosez A, Formstecher P, Marchetti P. Study of mitochondrial membrane potential, reactive oxygen species, DNA fragmentation and cell viability by flow cytometry in human sperm. *Hum Reprod* 2002; 17: 1257-1265.
39. Bedard K, Krause K-H. The NOX family of ROS-generating NADPH oxidases: physiology and pathophysiology. *Physiol Rev* 2007; 87: 245-313.
40. Sabour K, Ball B. Characterization of NADPH oxidase 5 in equine testis and spermatozoa. *Reproduction* 2007; 134: 263-270.
41. Ohlsson K, Bergenfeldt M, Björk P. Functional studies of human secretory leukocyte protease inhibitor. *Proteases II*: Springer; 1988: 123-31.

42. Fedder J. Nonsperm cells in human semen: with special reference to seminal leukocytes and their possible influence on fertility. *Syst Biol Reprod Med* 1996; 36: 41-65.
43. Muzio M, Bosisio D, Polentarutti N, D'amico G, Stoppacciaro A, Mancinelli R, et al. Differential expression and regulation of toll-like receptors (TLR) in human leukocytes: selective expression of TLR3 in dendritic cells. *J Immunol* 2000; 164: 5998-6004.
44. Edmund Sabanegh M, aReecha Sharma M, bDan French M, cFnu Deepinder M, aAlaa Hamada M, Ave E. The Natural History of Seminal Leukocytes in Men Seeking Infertility Evaluation.
45. Darley-Usmar V, Wiseman H, Halliwell B. Nitric oxide and oxygen radicals: a question of balance. *FEBS Lett* 1995; 369: 131-135.
46. Talebi AR. Sperm Nuclear Maturation: a basic and clinical approach: Nova Science Publishers; 2011.
47. Aitken RJ. Molecular mechanisms regulating human sperm function. *Mol Hum Reprod* 1997; 3: 169-173.
48. Tremellen K, Miari G, Froiland D, Thompson J. A randomised control trial examining the effect of an antioxidant (Menevit) on pregnancy outcome during IVF/ICSI treatment. *Aust N Z J Obstet Gynaecol* 2007; 47: 216-221.
49. Kessopoulou E, Tomlinson M, Barratt C, Bolton A, Cooke I. Origin of reactive oxygen species in human semen: spermatozoa or leucocytes? *J Reprod Fertil* 1992; 94: 463-470.
50. Agarwal A, Saleh RA, Bedaiwy MA. Role of reactive oxygen species in the pathophysiology of human reproduction. *Fertil Steril* 2003; 79: 829-843.
51. Gil-Guzman E, Ollerio M, Lopez M, Sharma R, Alvarez J, Thomas A, et al. Differential production of reactive oxygen species by subsets of human spermatozoa at different stages of maturation. *Hum Reprod* 2001; 16: 1922-1930.
52. Lamirande ED, Gagnon C. A positive role for the superoxide anion in triggering hyperactivation and capacitation of human spermatozoa. *Int J Androl* 1993; 16: 21-25.
53. Olugbenga OM, Olukole SG, Adeoye AT, Adejoke AD. Semen characteristics and sperm morphological studies of the West African Dwarf Buck treated with Aloe vera gel extract. *Iran J Reprod Med* 2011; 9: 83-88.
54. Yamagishi SI, Edelstein D, Du XI, Kaneda Y, Guzman M, Brownlee M. Leptin induces mitochondrial superoxide production and monocyte chemoattractant protein-1 expression in aortic endothelial cells by increasing fatty acid oxidation via protein kinase A. *J Biol Chem* 2001; 276: 25096-25100.
55. Ford W. Regulation of sperm function by reactive oxygen species. *Hum Reprod Update* 2004; 10: 387-399.
56. Visconti PE, Moore GD, Bailey JL, Leclerc P, Connors SA, Pan D, et al. Capacitation of mouse spermatozoa. II. Protein tyrosine phosphorylation and capacitation are regulated by a cAMP-dependent pathway. *Development* 1995; 121: 1139-1150.
57. Aitken R, Vernet P. Maturation of redox regulatory mechanisms in the epididymis. *J Reprod Fertil* 1997; 53: 109-118.
58. Aitken R. Possible redox regulation of sperm motility activation. *J Androl* 2000; 21: 491-496.
59. Sharma R, Said T, Agarwal A. Sperm DNA damage and its clinical relevance in assessing reproductive outcome. *Asian J Androl* 2004; 6: 139-148.
60. Alexander NJ. Male evaluation and semen analysis. *Clin Obstet Gynecol* 1982; 25: 463-482.
61. Talebi AR, Sarcheshmeh AA, Khalili MA, Tabibnejad N. Effects of ethanol consumption on chromatin condensation and DNA integrity of epididymal spermatozoa in rat. *Alcohol* 2011; 45: 403-409.
62. Goverde H, Dekker HS, Janssen H, Bastiaans BA, Rolland R, Zielhuis GA. Semen quality and frequency of smoking and alcohol consumption--an explorative study. *Int J Fertil Menopaus Stud* 1994; 40: 135-138.
63. Agarwal A, Prabakaran SA. Mechanism, measurement and prevention of oxidative stress in male reproductive physiology. *Indian J Exp Biol* 2005; 43: 963.
64. Saleh RA, Agarwal A, Sharma RK, Nelson DR, Thomas AJ. Effect of cigarette smoking on levels of seminal oxidative stress in infertile men: a prospective study. *Fertil Steril* 2002; 78: 491-499.
65. Künzle R, Mueller MD, Hänggi W, Birkhäuser MH, Drescher H, Bersinger NA. Semen quality of male smokers and nonsmokers in infertile couples. *Fertil Steril* 2003; 79: 287-291.
66. Waris G, Ahsan H. Reactive oxygen species: role in the development of cancer and various chronic conditions. *J Carcinogen* 2006; 5: 14.
67. Jalili C, Salahshoor MR, Naseri A. Protective effect of *Urtica dioica* L against nicotine-induced damage on sperm parameters, testosterone and testis tissue in mice. *Iran J Reprod Med* 2014; 12: 401-408.
68. Mohammadi S, Jalali M, Nikravesheh MR, Fazel A, Ebrahimzadeh A, Gholamin M, et al. Effects of Vitamin-E treatment on CatSper genes expression and sperm quality in the testis of the aging mouse. *Iran J Reprod Med* 2013; 11: 989-998.
69. Mostafa T, Tawadrous G, Roaia M, Amer M, Kader R, Aziz A. Effect of smoking on seminal plasma ascorbic acid in infertile and fertile males. *Andrologia* 2006; 38: 221-224.
70. Lee BM, Lee SK, Kim HS. Inhibition of oxidative DNA damage, 8-OHdG, and carbonyl contents in smokers treated with antioxidants (vitamin E, vitamin C, β -carotene and red ginseng). *Cancer Lett* 1998; 132: 219-227.
71. Dietrich M, Block G, Norkus EP, Hudes M, Traber MG, Cross CE, et al. Smoking and exposure to environmental tobacco smoke decrease some plasma antioxidants and increase γ -tocopherol in vivo after adjustment for dietary antioxidant intakes. *Am J Clin Nutr* 2003; 77: 160-166.
72. Majo J, Ghezzi H, Cosio M. Lymphocyte population and apoptosis in the lungs of smokers and their relation to emphysema. *Eur Resp J* 2001; 17: 946-953.
73. Jorsaraei SGA, Shibahara H. The in-vitro effects of nicotine, cotinine and leptin on sperm parameters analyzed by CASA system. *Iran J Reprod Med* 2008; 6: 157-165.
74. Rubes J, Selevan SG, Evenson DP, Zudova D, Vozdova M, Zudova Z, et al. Episodic air pollution is associated with increased DNA fragmentation in human sperm without other changes in semen quality. *Hum Reprod* 2005; 20: 2776-2783.
75. Sun JG, Jurisicova A, Casper RF. Detection of

- deoxyribonucleic acid fragmentation in human sperm: correlation with fertilization in vitro. *Biol Reprod* 1997; 56: 602-607.
76. Zavos PM, Correa JR, Karagounis CS, Ahparaki A, Phoroglou C, Hicks CL, et al. An electron microscope study of the axonemal ultrastructure in human spermatozoa from male smokers and nonsmokers. *Fertil Steril* 1998; 69: 430-434.
 77. Vine M, Tse C-K, Hu P, Truong KY. Cigarette smoking and semen quality. *Fertil Steril* 1996; 65: 835-842.
 78. Barbieri ER, Hidalgo ME, Venégas A, Smith R, Lissi EA. Varicocele Associated Decrease in Antioxidant Defenses. *J Androl* 1999; 20: 713-717.
 79. Agarwal A, Said TM. Role of sperm chromatin abnormalities and DNA damage in male infertility. *Hum Reprod Update* 2003; 9: 331-345.
 80. Meucci E, Milardi D, Mordente A, Martorana GE, Giacchi E, De Marinis L, et al. Total antioxidant capacity in patients with varicoceles. *Fertil Steril* 2003; 79: 1577-1583.
 81. Agarwal A, Prabakaran S, Allamaneni SS. Relationship between oxidative stress, varicocele and infertility: a meta-analysis. *Reprod BioMed Online* 2006; 12: 630-633.
 82. Talebi A, Moein M, Tabibnejad N, Ghasemzadeh J. Effect of varicocele on chromatin condensation and DNA integrity of ejaculated spermatozoa using cytochemical tests. *Andrologia* 2008; 40: 245-251.
 83. Ha HK, Park HJ, Park NC. Expression of E-cadherin and α -catenin in a varicocele-induced infertility rat model. *Asian J Androl* 2011; 13: 470-475.
 84. Santoro G, Romeo G, Impellizzeri P, Ientile R, Cutroneo G, Trimarchi F, et al. Nitric oxide synthase patterns in normal and varicocele testis in adolescents. *BJU Int* 2001; 88: 967-973.
 85. Mitropoulos D, Deliconstantinos G, Zervas A, Villiotou V, Dimopoulos C, Stavrides J. Nitric oxide synthase and xanthine oxidase activities in the spermatic vein of patients with varicocele: a potential role for nitric oxide and peroxynitrite in sperm dysfunction. *J Urol* 1996; 156: 1952-1958.
 86. Fischer MA, Willis J, Zini A. Human sperm DNA integrity: correlation with sperm cytoplasmic droplets. *Urology* 2003; 61: 207-211.
 87. Kashou AH, du Plessis SS, Agarwal A. The role of obesity in ROS generation and male infertility. *Studies on Men's Health and Fertility*: Springer; 2012: 571-590.
 88. Furukawa S, Fujita T, Shimabukuro M, Iwaki M, Yamada Y, Nakajima Y, et al. Increased oxidative stress in obesity and its impact on metabolic syndrome. *J Clin Invest* 2004; 114: 1752-1761.
 89. Hjollund NHI, Bonde JPE, Jensen TK, Olsen J. Diurnal scrotal skin temperature and semen quality. *Int J Androl* 2000; 23: 309-318.
 90. Magnúsdóttir EV, Thorsteinsson T, Thorsteinsdóttir S, Heimisdóttir M, Olafsdóttir K. Persistent organochlorines, sedentary occupation, obesity and human male subfertility. *Hum Reprod* 2005; 20: 208-215.
 91. Baynes JW. Role of oxidative stress in development of complications in diabetes. *Diabetes* 1991; 40: 405-412.
 92. Bloch-Damti A, Bashan N. Proposed mechanisms for the induction of insulin resistance by oxidative stress. *Antioxidant Redox Signal* 2005; 7: 1553-1567.
 93. Valko M, Leibfritz D, Moncol J, Cronin MT, Mazur M, Telser J. Free radicals and antioxidants in normal physiological functions and human disease. *Int J Biochem Cell Biol* 2007; 39: 44-84.
 94. Du XL, Edelstein D, Dimmeler S, Ju Q, Sui C, Brownlee M. Hyperglycemia inhibits endothelial nitric oxide synthase activity by posttranslational modification at the Akt site. *J Clin Invest* 2001; 108: 1341-1348.
 95. Agbaje I, Rogers D, McVicar C, McClure N, Atkinson A, Mallidis C, et al. Insulin dependant diabetes mellitus: implications for male reproductive function. *Hum Reprod* 2007; 22: 1871-1877.
 96. Vignera S, Condorelli R, Vicari E, D'Agata R, Calogero AE. Diabetes mellitus and sperm parameters. *J Androl* 2012; 33: 145-153.
 97. Alves MG, Oliveira PF, Oliveira P. Diabetes Mellitus and male re-productive function: where we stand. *Int J Diabetes Vasc Dis Res* 2013; 1: 1-2.
 98. Mangoli E, Talebi AR, Anvari M, Pourntezari M. Effects of experimentally-induced diabetes on sperm parameters and chromatin quality in mice. *Iran J Reprod Med* 2013; 11: 53-60.
 99. Talebi AR, Mangoli E, Nahangi H, Anvari M, Pourntezari M, Halvaei I. Vitamin C attenuates detrimental effects of diabetes mellitus on sperm parameters, chromatin quality and rate of apoptosis in mice. *Eur J Obstet Gynecol Reprod Biol* 2014; 181: 32-36.
 100. Peake JM, Suzuki K, Coombes JS. The influence of antioxidant supplementation on markers of inflammation and the relationship to oxidative stress after exercise. *J Nutr Biochem* 2007; 18: 357-371.
 101. Manna I, Jana K, Samanta P. Effect of different intensities of swimming exercise on testicular oxidative stress and reproductive dysfunction in mature male albino Wistar rats. *Indian J Exp Biol* 2004; 42: 816-822.
 102. Fenster L, Katz DF, Wyrobek AJ, Pieper C, Rempel DM, Oman D, et al. Effects of psychological stress on human semen quality. *J Androl* 1997; 18: 194-202.
 103. Eskiocak S, Gozen A, Taskiran A, Kilic A, Eskiocak M, Gulen S. Effect of psychological stress on the L-arginine-nitric oxide pathway and semen quality. *Braz J Med Biol Res* 2006; 39: 581-588.
 104. Eskiocak S, Gozen AS, Kilic AS, Molla S. Association between mental stress & some antioxidant enzymes of seminal plasma. *Indian J Med Res* 2005; 122: 491-496.
 105. Desai N, Sabanegh E, Kim T, Agarwal A. Free radical theory of aging: implications in male infertility. *Urology* 2010; 75: 14-19.
 106. Wyrobek AJ, Eskenazi B, Young S, Arnheim N, Tiemann-Boege I, Jabs E, et al. Advancing age has differential effects on DNA damage, chromatin integrity, gene mutations, and aneuploidies in sperm. *Proc Nat Acad Sci* 2006; 103: 9601-9606.
 107. Moskovtsev SI, Willis J, Mullen JBM. Age-related decline in sperm deoxyribonucleic acid integrity in patients evaluated for male infertility. *Fertil Steril* 2006; 85: 496-499.
 108. Youssry M, Ozmen B, Orief Y, Zohni K, Al-Hasani S. Human sperm DNA damage in the context of assisted reproductive techniques. *Iran J Reprod Med* 2007; 5: 137-150.
 109. Junqueira VB, Barros SB, Chan SS, Rodrigues L,

- Giavarotti L, Abud RL, et al. Aging and oxidative stress. *Mol Aspects Med* 2004; 25: 5-16.
110. Weir CP, Robaire B. Spermatozoa have decreased antioxidant enzymatic capacity and increased reactive oxygen species production during aging in the Brown Norway rat. *J Androl* 2007; 28: 229-240.
111. Said TM, Gokul SR, Agarwal A. Clinical Consequences of Oxidative Stress in Male Infertility. *Studies on Men's Health and Fertility*: Springer; 2012: 535-549.
112. Nadapdap T, Lutan D, Arsyad K, Ilyas S. Influence of Chitosan from Shrimp Skin to Quality and Quantity of Sperm of Albino Rats after Administration of Lead. *Andrology* 2014; 3: 114.
113. Agarwal D, Maronpot R, Lamb J, Kluwe W. Adverse effects of butyl benzyl phthalate on the reproductive and hematopoietic systems of male rats. *Toxicology* 1985; 35: 189-206.
114. Nagao T, Ohta R, Marumo H, Shindo T, Yoshimura S, Ono H. Effect of butyl benzyl phthalate in Sprague-Dawley rats after gavage administration: a two-generation reproductive study. *Reprod Toxicol* 2000; 14: 513-532.
115. Lee E, Ahn MY, Kim HJ, Kim IY, Han SY, Kang TS, et al. Effect of di (n-butyl) phthalate on testicular oxidative damage and antioxidant enzymes in hyperthyroid rats. *Environment Toxicol* 2007; 22: 245-255.
116. Hamada AJ, Singh A, Agarwal A. Cell phones and their impact on male fertility: fact or fiction. *Open Reprod Sci J* 2011; 5: 125-137.
117. Agarwal A, Deepinder F, Sharma RK, Ranga G, Li J. Effect of cell phone usage on semen analysis in men attending infertility clinic: an observational study. *Fertil Steril* 2008; 89: 124-128.
118. Aitken RJ, Baker MA. Oxidative stress, sperm survival and fertility control. *Mol Cell Endocrinol* 2006; 250: 66-69.
119. Gonzales G, Munoz G, Sanchez R, Henkel R, Gallegos-Avila G, Díaz-Gutierrez O, et al. Update on the impact of Chlamydia trachomatis infection on male fertility. *Andrologia* 2004; 36: 1-23.
120. Mazzilli F, Rossi T, Sabatini L, Pulcinelli F, Rapone S, Dondero F, et al. Human sperm cryopreservation and reactive oxygen species (ROS) production. *Acta Eur Fertil* 1994; 26: 145-148.
121. D'agata R, Vicari E, Moncada M, Sidoti G, Calogero A, Fornito M, et al. Generation of reactive oxygen species in subgroups of infertile men. *Int J Androl* 1990; 13: 344-351.
122. Wang A, Fanning L, Anderson D, Loughlin K. Generation of reactive oxygen species by leukocytes and sperm following exposure to urogenital tract infection. *Syst Biol Reprod Med* 1997; 39: 11-17.
123. Zhang ZH, Zhu HB, Li LL, Yu Y, Zhang HG, Liu RZ. Decline of semen quality and increase of leukocytes with cigarette smoking in infertile men. *Iran J Reprod Med* 2013; 11: 589-596.
124. Chew BP, Park JS. Carotenoid action on the immune response. *J Nutr* 2004; 134: 257S-261S.
125. Nabel G, Baltimore D. An inducible transcription factor activates expression of human immunodeficiency virus in T cells. *Nature* 1987; 326: 711-713.
126. Tollon C, Soulié M, Pontonnier F, Plante P. [Prevention of sterility in spinal cord injured men]. *Progres en urologie*; 1997; 7: 203-208. (In French)
127. Keck C, Gerber-Schäfer C, Clad A, Wilhelm C, Breckwoldt M. Seminal tract infections: impact on male fertility and treatment options. *Hum Reprod Update* 1998; 4: 891-903.
128. Ochsendorf F. Infections in the male genital tract and reactive oxygen species. *Hum Reprod Update* 1999; 5: 399-420.
129. Nicopoullos JD, Almeida PA, Ramsay JW, Gilling-Smith C. The effect of human immunodeficiency virus on sperm parameters and the outcome of intrauterine insemination following sperm washing. *Hum Reprod* 2004; 19: 2289-2297.
130. Frodsham LC, Boag F, Barton S, Gilling-Smith C. Human immunodeficiency virus infection and fertility care in the United Kingdom: demand and supply. *Fertil Steril* 2006; 85: 285-289.
131. Safarnavadeh T, Rastegarpanah M. Antioxidants and infertility treatment, the role of Satureja Khuzestanica: A mini-systematic review. *Iran J Reprod Med* 2011; 9: 61-70.
132. Khosrowbeygi A, Zarghami N, Deldar Y. Correlation between sperm quality parameters and seminal plasma antioxidants status. *Iran J Reprod Med* 2004; 2: 58-64.
133. Halliwell B. Free radicals and antioxidants: a personal view. *Nutr Rev* 1994; 52: 253-265.
134. Venkatesh S, Gurdeep Singh M, Prasad Gupta N, Kumar R, Deecaraman M, Dada R. Correlation of sperm morphology and oxidative stress in infertile men. *Iran J Reprod Med* 2009; 7: 29-34.
135. Halliwell B. Antioxidant defence mechanisms: from the beginning to the end (of the beginning). *Free Rad Res* 1999; 31: 261-272.
136. Halliwell B. Free radicals and other reactive species in disease. John Wiley & Sons, Ltd, eLS; 2005.
137. Dröge W. Free radicals in the physiological control of cell function. *Physiol Rev* 2002; 82: 47-95.
138. Fraga CG, Motchnik PA, Shigenaga MK, Helbock HJ, Jacob RA, Ames BN. Ascorbic acid protects against endogenous oxidative DNA damage in human sperm. *Proc Nat Acad Sci* 1991; 88: 11003-11006.
139. Sheikh N, Amiri I, Farimani M, Najafi R, Hadeie J. Correlation between sperm parameters and sperm DNA fragmentation in fertile and infertile men. *Iran J Reprod Med* 2008; 6: 13-18.
140. Griveau J, Dumont E, Renard P, Callegari J, Le Lannou D. Reactive oxygen species, lipid peroxidation and enzymatic defence systems in human spermatozoa. *J Reprod Fertil* 1995; 103: 17-26.
141. German JB, Traber MG. Nutrients and oxidation: Actions, transport, and metabolism of dietary antioxidants. Rucker RB, Suttie JW, McCormick DB, Machlin LJ. Handbook of vitamins. 3rd Ed. New York, Marcel Dekker; 2001: 569-88.
142. Jube S, Borthakur D. Recent advances in food biotechnology research. Blackwell Publishing, Oxford, UK 2006: 35-70.
143. Ling Coh GM. Potential for Ginkgo biloba as a functional food [Ph.D. thesis]. Potential for Ginkgo biloba as a functional food; 2004.
144. Lenzi A, Gandini L, Picardo M, Tramer F, Sandri G, Panfili E. Lipoperoxidation damage of spermatozoa polyunsaturated fatty acids (PUFA): scavenger mechanisms and possible scavenger therapies. *Front Biosci* 2000; 5: 1-15.
145. Lanzafame FM, La Vignera S, Vicari E, Calogero AE.

- Oxidative stress and medical antioxidant treatment in male infertility. *Reprod BioMed Online* 2009; 19: 638-659.
146. Saraswat S, Kharche S, Jindal S. Impact of Reactive Oxygen Species on Spermatozoa: A Balancing Act between Beneficial and Detrimental Effects. *Iran J Appl Anim Sci* 2014; 4: 247-255.
 147. Ko EY, Sabanegh ES, Agarwal A. Male infertility testing: reactive oxygen species and antioxidant capacity. *Fertil Steril* 2014; 102: 1518-1527.
 148. Keskes-Ammar L, Feki-Chakroun N, Rebai T, Sahnoun Z, Ghoszi H, Hammami S, et al. Sperm oxidative stress and the effect of an oral vitamin E and selenium supplement on semen quality in infertile men. *Syst Biol Reprod Med* 2003; 49: 83-94.
 149. Blount JD, Møller AP, Houston DC. Antioxidants, showy males and sperm quality. *Ecol Lett* 2001; 4: 393-396.
 150. Nouri M, Ghasemzadeh A, Farzadi L, Shahnazi V, Ghaffari Novin M. Vitamins C, E and lipid peroxidation levels in sperm and seminal plasma of asthenoteratozoospermic and normozoospermic men. *Iran J Reprod Med* 2008; 6: 1-5.
 151. Greco E, Iacobelli M, Rienzi L, Ubaldi F, Ferrero S, Tesarik J. Reduction of the incidence of sperm DNA fragmentation by oral antioxidant treatment. *J Androl* 2005; 26: 349-353.
 152. Oda SS, El-Maddawy ZK. Protective effect of vitamin E and selenium combination on deltamethrin-induced reproductive toxicity in male rats. *Exp Toxicol Pathol* 2012; 64: 813-819.
 153. Adesiyun AC, Oyejola TO, Abarikwu SO, Oyeyemi MO, Farombi EO. Selenium provides protection to the liver but not the reproductive organs in an atrazine-model of experimental toxicity. *Exp Toxicol Pathol* 2011; 63: 201-207.
 154. Omu A, Al-Azemi M, Kehinde E, Anim J, Oriowo M, Mathew T. Indications of the mechanisms involved in improved sperm parameters by zinc therapy. *Med Princ Pract* 2008; 17: 108-116.
 155. Zini A, Fischer MA, Nam RK, Jarvi K. Use of alternative and hormonal therapies in male infertility. *Urology* 2004; 63: 141-143.
 156. Dragsted LO. Biomarkers of exposure to vitamins A, C, and E and their relation to lipid and protein oxidation markers. *Eur J Nutr* 2008; 47: 3-18.
 157. Hwang K, Walters RC, Lipshultz LI. Contemporary concepts in the evaluation and management of male infertility. *Nat Rev Urol* 2011; 8: 86-94.
 158. Agarwal A, Sekhon LH. Oxidative stress and antioxidants for idiopathic oligoasthenoteratospermia: Is it justified? *Indian J Urol* 2011; 27: 74.
 159. Wang Y-N, Wang B, Liang M, Han C-Y, Zhang B, Cai J, et al. Down-regulation of CatSper1 channel in epididymal spermatozoa contributes to the pathogenesis of asthenozoospermia, whereas up-regulation of the channel by Sheng-Jing-San treatment improves the sperm motility of asthenozoospermia in rats. *Fertil Steril* 2013; 99: 579-587.
 160. Imhof M, Lackner J, Lipovac M, Chedraui P, Riedl C. Micronutrient supplementation increases sperm quality in the sub-fertile male. *Eur Urol Rev* 2011; 6: 45-49.
 161. Aliabadi E, Mehranjani MS, Borzoei Z, Talaei-Khozani T, Mirkhani H, Tabesh H. Effects of L-carnitine and L-acetyl-carnitine on testicular sperm motility and chromatin quality. *Iran J Reprod Med* 2012; 10: 77-82.
 162. Dehghani F, Hassanpour A, Poost-pasand A, Noorafshan A, Karbalay-Doust S. Protective effects of L-carnitine and homogenized testis tissue on the testis and sperm parameters of busulfan-induced infertile male rats. *Iran J Reprod Med* 2013; 11: 693-704.
 163. Arcaniolo D, Favilla V, Tiscione D, Pisano F, Bozzini G, Creta M, et al. Is there a place for nutritional supplements in the treatment of idiopathic male infertility? *Arch Ital Urol Androl* 2014; 86: 164-170.
 164. Babaei H, Abshenas J. Zinc therapy improves adverse effects of long term administration of copper on epididymal sperm quality of rats. *Iran J Reprod Med* 2013; 11: 577-582.