

Clinical Relevance of Targeting the Gastrin-Releasing Peptide Receptor, Somatostatin Receptor 2, or Chemokine C-X-C Motif Receptor 4 in Breast Cancer for Imaging and Therapy

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Imaging and therapy using radioligands targeting receptors overexpressed on tumor cells is successfully applied in neuroendocrine tumor patients. Because expression of the gastrin-releasing peptide receptor (GRPR), somatostatin receptor 2 (SSTR2), and chemokine C-X-C motif receptor 4 (CXCR4) has been demonstrated in breast cancer, targeting these receptors using radioligands might offer new imaging and therapeutic opportunities for breast cancer patients. The aim of this study was to correlate messenger RNA (mRNA) expression of *GRPR*, *SSTR2*, and *CXCR4* with clinicopathologic and biologic factors, and with prognosis and prediction to therapy response, in order to identify specific breast cancer patient groups suited for the application of radioligands targeting these receptors. **Methods:** First, we studied GRPR and SSTR2 expression in 13 clinical breast cancer specimens by in vitro autoradiography and correlated this with corresponding mRNA levels to investigate whether mRNA levels reliably represent cell surface expression. Next, *GRPR*, *SSTR2*, and *CXCR4* mRNA levels were measured by quantitative reverse transcriptase polymerase chain reaction in 915 primary breast cancer tissues and correlated with known clinicopathologic and biologic factors, disease-free survival, distant metastasis-free survival, and overall survival (DFS, MFS, and OS, respectively). In 224 adjuvant hormonal treatment-naïve estrogen receptor (ER, *ESR1*)-positive patients who received tamoxifen as first-line therapy for recurrent or metastatic disease, the expression levels of the receptors were correlated with progression-free survival. **Results:** Our results showed a significant positive correlation between GRPR and SSTR2 expression analyzed by in vitro autoradiography and by quantitative reverse transcriptase polymerase chain reaction (Spearman's rank correlation coefficient [R_s] = 0.94, $P < 0.001$, and $R_s = 0.73$, $P = 0.0042$, respectively). Furthermore, high *GRPR* and *SSTR2* mRNA levels were observed more frequently in *ESR1*-positive specimens, whereas high *CXCR4* expression was associated with *ESR1*-negative specimens. Also, high mRNA expression of *CXCR4* was associated with a prolonged DFS, MFS, and OS (multivariate hazard ratio MFS = 0.76 [95% confidence interval, 0.64–0.90], $P = 0.001$), whereas high mRNA levels of *GRPR* were associated with a prolonged progression-free survival after the start of first-line tamoxifen treatment (multivariate hazard ratio = 0.68 [95% confidence interval, 0.48–0.97], $P = 0.031$). **Conclusion:** Our data indicate that imaging and therapy

using GRPR or SSTR2 radioligands might especially be beneficial for *ESR1*-positive breast cancer and CXCR4 radioligands for *ESR1*-negative breast cancer.

Key Words: breast cancer; GRPR; SSTR2; CXCR4; PRS/PRRT

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Breast cancer is the most common cancer found in women worldwide. An estimated 1.7 million new cases were diagnosed in 2012 worldwide, and 522,000 people died as a consequence of the disease, making it the fifth cause of death by cancer overall (1).

Multiple subtypes of breast cancer exist, with different molecular characteristics such as the absence or presence of estrogen receptor (ER, *ESR1*), progesterone receptor (PR, *PGR*), and human epidermal growth factor 2 (HER2, *ERBB2*) (2). In the case of ER and HER2, these receptors also serve as therapeutic targets. ER-positive patients are treated with either aromatase inhibitors or ER antagonists, most commonly tamoxifen, whereas HER2-positive patients are often treated with the HER2-specific monoclonal antibody trastuzumab (2). However, in the recurrent or metastatic setting nearly all patients acquire resistance against tamoxifen and trastuzumab after an initial response (3,4).

Mammography is the standard method used for breast cancer screening, in some cases supplemented with MR imaging or ultrasound (5). Unfortunately these methods may lead to false-positive and false-negative results (6,7). Because current imaging and the above-mentioned therapy options, in particular, have limitations and are not always successful, new imaging and therapeutic options are urgently needed.

Peptide receptor scintigraphy and peptide receptor radionuclide therapy are methods based on targeting receptors overexpressed on tumor cells using radioligands for diagnostic and therapeutic purposes. Within nuclear medicine, radiolabeled somatostatin (SST) analogs are most widely and successfully used for the localization, treatment, and evaluation of neuroendocrine tumors (8). These SST analogs bind to SST receptors (SSTR, especially SSTR2) overexpressed on tumor cells, enabling imaging when labeled with γ or positron emitters and therapy when labeled with β^- or α -particle emitters. Currently, multiple radiolabeled SST analogs targeting SSTR2 are available and used in the clinic (9).

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In the past decade, imaging of breast cancer patients using SSTR2 radioligands has been studied with varying results (10,11). Currently, considerably improved SSTR2-directed radiotracers and imaging equipment are available.

Other promising targeting radioligands for breast cancer comprise radiolabeled gastrin-releasing peptide (GRP) analogs, earlier applied for the visualization and therapy of prostate cancer lesions, because significant GRP receptor (GRPR) levels are present in most primary prostate cancer tissues (12–14). Previous studies by Reubi et al. (15) showed a high expression of both SSTR2 and GRPR in breast cancer. SSTR2 and high-density GRPR expression was found in 75% and 74% of breast cancer cases, respectively.

Moreover, chemokine C-X-C motif receptor 4 (CXCR4) expression has been reported in most breast cancers. In a study by Salvucci et al. (16), in which 2,022 breast cancer specimens were analyzed for CXCR4 expression using immunohistochemistry, 67% of invasive tumors showed high nuclear staining and 41% of tumors showed cytoplasmic staining (12). Promising radiolabeled peptide derivatives binding to CXCR4 have been synthesized to target these receptors (17,18). So ⁶⁸Ga-pentaxifor, a CXCR4 radioligand, has successfully been used in a clinical study for the imaging of multiple myeloma patients (19). Thus, these 3 promising categories of radiolabeled compounds could be of promise in breast cancer patients.

Until now, little was known about the correlation between GRPR, SSTR2, and CXCR4 expression levels in breast cancer lesions and important molecular and prognostic characteristics, such as hormone receptor expression, as well as the association of GRPR, SSTR2, and CXCR4 expression with disease-free survival, distant metastasis-free survival, or overall survival (DFS, MFS, and OS, respectively) and with progression-free survival (PFS) after endocrine treatment.

In this study, we first analyzed the correlation between messenger RNA (mRNA) levels and protein expression of GRPR and SSTR2. Subsequently, we analyzed the mRNA expression of *GRPR*, *SSTR2*, and *CXCR4* in human breast cancer specimens. The aims of this study were to correlate *GRPR*, *SSTR2*, and *CXCR4* mRNA expression levels with clinicopathologic and biologic factors as well as with prognosis and outcome on tamoxifen therapy, to assess the potential impact of radioligands targeting these receptors for imaging and therapeutic purposes in breast cancer, and to thereby identify patient subgroups that potentially would benefit from application of these radiopharmaceuticals.

MATERIALS AND METHODS

Human Breast Cancer Cases

The study (MEC02-953) was approved by the Erasmus MC Medical Ethical Committee and adhered to the Code of Conduct of the Federation of Medical Scientific Societies in The Netherlands.

The primary breast cancer tissue of 915 female patients (mean age ± SD, 58 ± 13 y) (684 M0 [no metastasis at diagnosis] lymph-node-negative [LNN], 194 M0 lymph-node-positive [LNP], 24 M1 LNP, and 13 patients with unknown nodal status at time of primary treatment) who visited the clinic between 1979 and 2000 were selected from the Erasmus MC fresh-frozen tissue bank as described before (20). The inclusion criteria and the determination of clinicopathologic and biologic factors are described in the supplemental data (supplemental materials are available at <http://jnm.snmjournals.org>). *GRPR*, *SSTR2*, and *CXCR4* expression was initially correlated with clinicopathologic and biologic factors in the LNN M0 patient group ($n = 194$). A representative group of LNP tumors ($n = 194$) was added to study the influence of positive nodal status on the correlation analyses.

For prognosis, we focused our analyses on the cohort of 684 systemic treatment-naïve patients with LNN disease; for prediction of therapy response, a cohort of 224 hormonal treatment-naïve ER-positive patients who received tamoxifen as first-line therapy for recurrent or metastatic disease was analyzed. The clinicopathologic and biologic factors of the LNN M0 tumors are shown in Table 1, and clinicopathologic and biologic factors for the LNN and LNP M0 patient group and the ER-positive first-line tamoxifen-treated subcohort are shown in Supplemental Tables 1A and 1B, respectively. Patients were censored at 120-mo follow-up after surgical removal of the primary tumor in the regression analysis for DFS (283 events), MFS (241 events), and OS (223 events) and at 36 mo after the start of tamoxifen treatment for analysis of PFS (24 events). The study design is depicted in Figure 1.

RNA Isolation, Complementary DNA Synthesis, and Quantitative Reverse Transcriptase Polymerase Chain Reaction (RT-qPCR)

Tissue processing, RNA isolation, complementary DNA synthesis, and RT-qPCR were performed and normalized using the δ Cq method on the average of 3 reference genes (*HMBS*, *HPRT1*, and *TBP*) as described (21). All RNA samples that required more than 25 rounds of real-time PCR for detectable products of our 3 reference genes at a fixed input of 10 ng of total RNA and at a threshold of 0.1 were considered of insufficient quality and were excluded from further analysis. Target genes were quantified using the following intron-spanning Taqman probe-based gene expression assays (Applied BioSystems/Life Technologies): *GRPR*, Hs01055872 m1; *SSTR2*, Hs0099356 m1; and *CXCR4*, Hs00237052 m1, according to the manufacturer's instructions in a MX3000P Real-Time PCR System (Agilent). Genomic grade index (GGI), a gene expression pattern of histologic tumor grade, and *ESR1*, *PGR*, and *ERBB2* levels and status of the samples were already known based on quantification as previously described (22–24).

Radioligands and In Vitro Autoradiography

Peptide analogs targeting the SSTR2 and GRPR, DOTA-Tyr³-octreotate (Mallinckrodt) and AMBA (BioSynthema), respectively, were radiolabeled with ¹¹¹In (Covidien), as previously described (25). Quenchers (10 mM methionine, 3.5 mM ascorbic acid, and 3.5 mM gentisic acid) were used to prevent radiolysis (26). Specific activity of both radiotracers was 50 MBq/nmol. Radiometal incorporation (>99%) and radiochemical purity (>90%) were measured by instant thin-layer chromatography on silica gel and high-pressure liquid chromatography as previously described (26).

The CXCR4 radioligand, pentaxifor, available to us showed reduced receptor affinity when radiolabeled with ¹¹¹In, and thus satisfying in vitro autoradiography studies using this compound could not be performed.

In the in vitro autoradiography assay, tissue sections of 13 fresh-frozen breast cancer specimens (10 μ m) were incubated with 10⁻⁹M ¹¹¹In-AMBA and ¹¹¹In-DOTA-Tyr³-octreotate for 1 h, without and with 10⁻⁶ M unlabeled tracer as control for nonspecific binding. H69 (SSTR2-positive, GRPR-negative) and PC3 xenografts (GRPR-positive, SSTR2-negative) were used as controls. Results were quantified using OptiQuant software (Perkin Elmer), and the net percentage binding of added dose was calculated. The in vitro autoradiography assay and quantification of the results are described in more detail in the supplemental data.

Statistics

Statistical analyses are described in the supplemental data.

RESULTS

In Vitro Autoradiography and Correlation with mRNA Expression

Specific binding to tumor cells of the GRPR- and SSTR2-mediated radiotracers, ¹¹¹In-AMBA and ¹¹¹In-DOTA-Tyr³-octreotate,

TABLE 1
Associations of *GRPR*, *SSTR2*, and *CXCR4* mRNA Levels in LNN M0 Patients

Characteristic	No. of patients*	Percentage of patients	<i>GRPR</i> mRNA ($\times 10^{-2}$)		<i>SSTR2</i> mRNA ($\times 10^{-2}$)		<i>CXCR4</i> mRNA ($\times 10^{-2}$)	
			Median	Interquartile range	Median	Interquartile range	Median	Interquartile range
All patients in this cohort	684	100%	0.72	7.07	0.58	1.75	11.78	13.13
Age at surgery (y)								
≤40	60	9%	1.17	12.72	0.90	2.99	14.06	13.75
41–55	252	37%	0.97	9.20	0.61	1.64	11.58	13.18
56–70	218	32%	0.52	5.38	0.52	1.68	12.19	11.34
>70	154	23%	0.72	4.44	0.62	1.61	9.99	12.98
<i>P</i> [†]			0.52		0.68		0.0403	
Menopausal status								
Premenopausal	273	40%	1.26	10.95	0.62	1.82	11.81	13.73
Postmenopausal	411	60%	0.60	4.87	0.55	1.57	11.76	12.12
<i>P</i> [†]			0.13		0.53		0.39	
Surgery								
Lumpectomy	378	55%	0.61	7.69	0.57	1.82	11.67	13.15
Ablation	306	45%	0.90	6.79	0.60	1.56	11.90	13.00
<i>P</i> [†]			0.69		0.59		0.65	
Pathologic tumor size								
pT1	307	45%	1.25	8.54	0.69	1.87	12.03	13.40
pT2 + unknown	351	51%	0.41	5.25	0.51	1.65	11.53	12.84
pT3 + pT4	26	4%	0.58	3.05	0.50	1.38	12.19	13.53
<i>P</i> [†]			0.0014		0.24		0.92	
<i>ESR1</i> mRNA status [‡]								
Negative < 0.2	184	27%	0.09	0.13	0.28	0.42	14.74	13.83
Positive ≥ 0.2	500	73%	2.46	10.98	0.81	2.59	10.98	12.16
<i>P</i> [†]			<0.001		<0.001		<0.001	
<i>PGR</i> mRNA status [‡]								
Negative < 0.1	285	42%	0.12	0.32	0.32	0.56	14.36	14.31
Positive ≥ 0.1	399	58%	3.67	12.68	1.02	2.98	10.45	11.09
<i>P</i> [†]			< 0.001		<0.001		<0.001	
<i>ERBB2</i> mRNA status [‡]								
Negative < 18	574	84%	0.99	8.28	0.61	1.92	11.64	12.96
Positive ≥ 18	107	16%	0.30	1.51	0.49	1.00	13.88	13.32
<i>P</i> [¶]			<0.001		0.0344		0.22	
Grade (GGI)								
1	227	33%	2.42	10.46	0.75	2.11	10.83	11.03
2	229	33%	0.89	6.92	0.63	2.41	11.44	14.59
3	224	33%	0.13	1.42	0.34	0.98	13.83	13.58
<i>P</i> [¶]			<0.001		<0.001		<0.001	
Percentage invasive tumor cells								
≤70%	470	69%	0.81	6.84	0.63	1.88	12.57	13.92
>70%	214	31%	0.64	8.28	0.43	1.28	9.13	10.70
<i>P</i> [†]			0.87		0.002		<0.001	

*Because of missing numbers, not all categories add up to 684.

[†]*P* for Mann–Whitney U or Kruskal–Wallis test when appropriate.

[‡]*ESR1*, *PGR*, and *ERBB2* were determined by real-time PCR; cut points were as follows: *ESR1* = 0.2, *PGR* = 0.1, and *ERBB2* = 18.0 (mRNA level relative to reference gene set).

[¶]*P* for Spearman rank-correlation test.

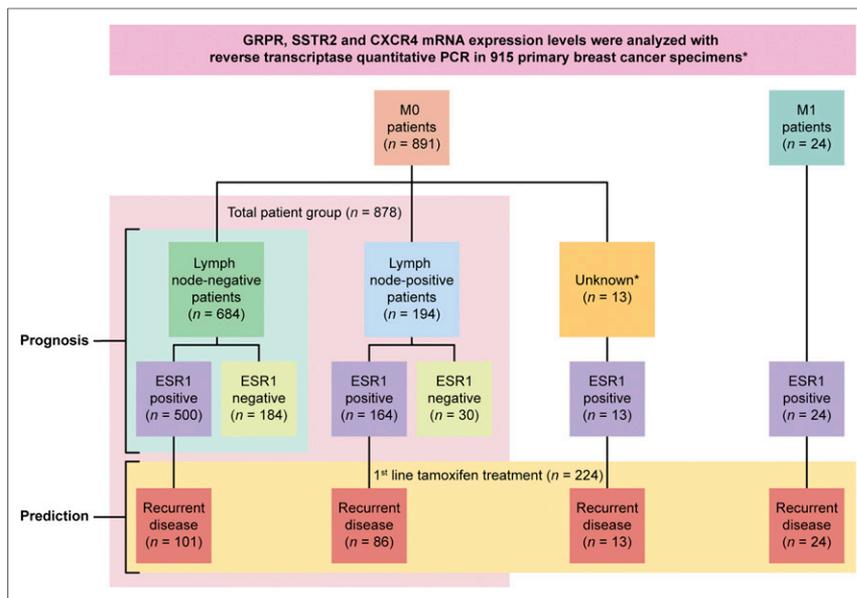


FIGURE 1. Study design. mRNA expression levels of *GRPR*, *SSTR2*, and *CXCR4* of 915 primary breast cancer specimens (684 M0 LNN, 194 M0 LNP, 13 with unknown nodal status, and 24 M1) were analyzed using RT-qPCR. LNN and LNP M0 patient groups were used to study association of *GRPR*, *SSTR2*, and *CXCR4* expression and clinicopathologic and biologic factors, with focus on M0 LNN patient group. Association of *GRPR*, *SSTR2*, and *CXCR4* with prognostic factors was studied in M0 LNN patients. mRNA levels of ER-positive primary tumors of patients with recurrent breast cancer who received first-line tamoxifen treatment were used to study association of *GRPR*, *SSTR2*, and *CXCR4* mRNA expression and PFS.

respectively, was demonstrated using in vitro autoradiography on 13 selected human breast cancer specimens with varying levels of mRNA receptor expression. Two mouse xenografts served as positive and negative control (Fig. 2A). Autoradiography results were quantified and correlated with the level of mRNA expression of the respective receptors, resulting in a significant positive correlation for both *GRPR* (Spearman's rank correlation coefficient [R_s] = R_s = 0.94, $P < 0.0001$) and *SSTR2* (R_s = 0.73, $P = 0.0042$) (Fig. 2B). Furthermore, binding of the tracers was observed only on tumor cells and not on the surrounding stromal cells. We thus concluded that mRNA expression for *GRPR* and *SSTR2* can be used as a predictor for binding of the radiotracers to tumor tissue.

Correlation of *GRPR*, *SSTR2*, and *CXCR4* mRNA Expression with Clinicopathologic and Biologic Factors

We focused on the 684 LNN M0 patients to study the correlation between *GRPR*, *SSTR2*, and *CXCR4* mRNA levels and known clinicopathologic and biologic factors. The results of the correlation analyses are shown in Table 1. To study the influence of positive nodal status on the correlation analyses, a representative group of 194 LNP M0 tumors were added to the study. Results of the LNN and LNP M0 patient group are described in Supplemental Table 1A.

A significant correlation was observed between *GRPR* mRNA levels and a smaller pathologic tumor size ($P = 0.0014$), a positive *ESR1* ($P < 0.001$) and *PGR* status ($P < 0.001$), a negative *ERBB2* ($P < 0.001$) status, and a favorable GGI ($P < 0.001$).

SSTR2 mRNA expression showed a significant correlation with a positive *ESR1* ($P < 0.001$) and *PGR* mRNA status ($P < 0.001$), a negative *ERBB2* status ($P = 0.0344$), favorable GGI ($P < 0.001$), and 70% or less invasive tumor cells ($P = 0.002$).

CXCR4 mRNA expression showed a significant negative correlation with *ESR1* ($P < 0.001$) and *PGR* mRNA status ($P < 0.001$) and was associated with an unfavorable GGI ($P < 0.001$). Furthermore, *CXCR4* mRNA levels were higher in tumors with 70% or less invasive tumor cells ($P < 0.001$).

Association of *GRPR*, *SSTR2*, and *CXCR4* mRNA Expression with Prognosis and Efficacy of Tamoxifen Treatment

To exclude the possible confounding effect of adjuvant therapy on prognosis, the association of *GRPR*, *SSTR2*, and *CXCR4* expression with prognosis was evaluated in the LNN patient group, which did not receive adjuvant systemic therapy. The results of the evaluation of *GRPR*, *SSTR2*, and *CXCR4* mRNA expression with DFS, MFS, and OS are shown in Supplemental Table 2.

No significant associations were observed between *GRPR* and *SSTR2* mRNA expression and DFS, MFS, or OS. For *CXCR4*, however, there was a significant association of its expression with a favorable DFS, MFS, and OS, both when analyzed as a continuous variable and when dichotomized at the median level. For the primary endpoint MFS, the results of the multivariate analysis were hazard ratio (HR) = 0.76 (95% confidence interval [CI], 0.64–0.90), $P = 0.001$, when analyzed as a continuous variable, and HR = 0.71 (95% CI, 0.55–0.91), $P = 0.011$, when dichotomized at the median level.

To visualize the association of the levels of *CXCR4* mRNA with MFS, Kaplan–Meier analysis was performed as a function of the quartile levels of *CXCR4* mRNA (Fig. 3). The results show a clear trend of quartiles, with lower expression having a worse MFS time.

In addition, *GRPR*, *SSTR2*, and *CXCR4* mRNA expression levels were correlated with the efficacy of tamoxifen treatment in *ESR1*-positive patients with recurrent disease (Supplemental Table 1B). There was a significant correlation between high *GRPR* mRNA levels and prolonged PFS after the start of first-line tamoxifen treatment, indicating that *GRPR* expression has predictive value for the efficacy of tamoxifen therapy (Fig. 4; Supplemental Table 3) (25% high vs. 75% low, univariate HR = 0.65 [95% CI, 0.47–0.91], $P = 0.011$, and multivariate HR = 0.68 [95% CI, 0.48–0.97], $P = 0.031$).

DISCUSSION

We have analyzed *GRPR*, *SSTR2*, and *CXCR4* mRNA expression in 915 primary breast cancer tissues and correlated mRNA expression of these receptors with clinicopathologic and biologic factors and with prognosis and prediction to therapy response, to study the relevance of the application of radioligands targeting these receptors for imaging and therapy in breast cancer patients. For this, we first successfully demonstrated in vitro binding of radiotracers for *GRPR* and *SSTR2* to tissue sections and showed a significant positive correlation between radiotracer binding and mRNA expression, demonstrating that mRNA levels of these

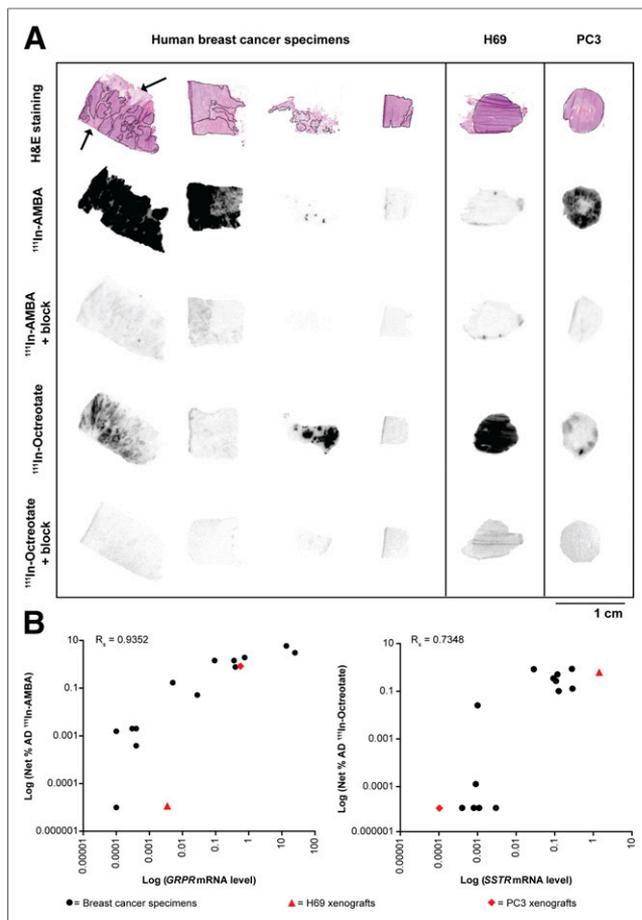


FIGURE 2. (A) In vitro autoradiography of human breast cancer specimens using ^{111}In -AMBA (GRP analog) and ^{111}In -DOTA-Tyr³-octreotate (SST analog) with and without block, demonstrating specific binding of radiotracers to receptor-positive tumor tissue. H69 (SSTR-positive, GRPR-negative) and PC3 xenografts (SSTR-negative, GRPR-positive) were used as controls. Tumor-containing areas are encircled in hematoxylin and eosin (H&E) stainings. As example, arrows indicate non-tumor-containing tissue in first H&E staining. (B) Significant correlation between *GRPR* and *SSTR2* mRNA levels and quantification of in vitro autoradiography results analyzed in 13 breast cancer specimens with variable receptor expression, demonstrating that mRNA levels of receptors can be used as predictor for radiotracer binding. AD = added dose.

receptors can be used as a predictor for specific radiotracer binding. The CXCR4 radioligand pentaxifor, available to us, showed reduced receptor affinity when radiolabeled with ^{111}In for in vitro autoradiography purposes, hampering reliable in vitro autoradiography studies for CXCR4. Thus, studies correlating CXCR4 radiotracer binding and *CXCR4* mRNA expression could not be performed. However, because Philip-Abbrederis et al. (19) reported on detecting *CXCR4* mRNA expression in cell lines and successful in vivo imaging of corresponding xenograft models using ^{68}Ga -pentaxifor, we concluded that *CXCR4* mRNA expression can also be used as a predictor for CXCR4 radioligand binding.

Concerning prognosis, we found no association between *GRPR* and *SSTR2* expression and DFS, MFS, and OS in the M0 LNN patients. Surprisingly, we found that high *CXCR4* levels correlated with better prognosis despite its negative correlation with ER, PR, and unfavorable GGI, indicating that a component of CXCR4 expression that is independent of these factors determines good outcome.

Other studies on CXCR4 expression in breast cancer have associated CXCR4 expression with poor patient survival (16). The discrepancy in study outcome might be explained by the fact that in our study we analyzed mRNA expression of the receptors (independent of receptor localization), whereas in the study by Salvucci et al. (16) tissue microarrays were analyzed by immunohistochemistry and nuclear and cytoplasmic CXCR4 staining were analyzed separately. In agreement with our study, Salvucci et al. (16) reported more cytoplasmic CXCR4 staining in ER-negative (54%) than ER-positive tumors (38%).

Furthermore, we found that high *GRPR* expression was of modest predictive value for increased time to progression on tamoxifen treatment, suggesting GRPR radioligands to be useful in monitoring tumor response to treatment with tamoxifen. Recently, preclinical ^{68}Ga -AMBA PET imaging in a mouse model also demonstrated the feasibility for monitoring tumor response after treatment with tamoxifen (27).

For the association with clinicopathologic and biologic characteristics analyzed in the M0 LNN patients, we observed a significant positive correlation between *GRPR* and *SSTR2* expression and *ESR1*- and *PGR*-positive tumors. In line with our findings, significant positive correlation between *SSTR2* and ER expression was reported previously (28), whereas van den Bossche et al. (29) reported estrogen-mediated regulation of *SSTR2* expression in breast cancer cell lines. Because *ESR1* and *PGR* positivity correlates with breast cancer of the luminal subtype (2), tumors of this subtype could benefit most from GRPR- or *SSTR2*-mediated imaging or therapy. Moreover, *ESR1*-negative tumors showed low to no *GRPR* expression, and thus patients with *ESR1*-negative primary tumors are likely not suited for the application of GRPR radioligands. Because *ESR1*- or *PGR*-positive tumors account for 75% of the breast cancer tumors (2), GRPR- and *SSTR2*-mediated imaging and therapy might be of benefit for the larger part of the breast cancer patient population.

Concerning therapy, GRPR or *SSTR2* radioligands can especially be of benefit for patients with *ESR1*-positive tumors who have progressed on various lines of endocrine treatment, because nearly all patients with recurrent disease become resistant against current antiestrogen treatments (4).

Previous studies we performed on GRPR and *SSTR2* expression in human breast cancer specimens showed GRPR expression in 48

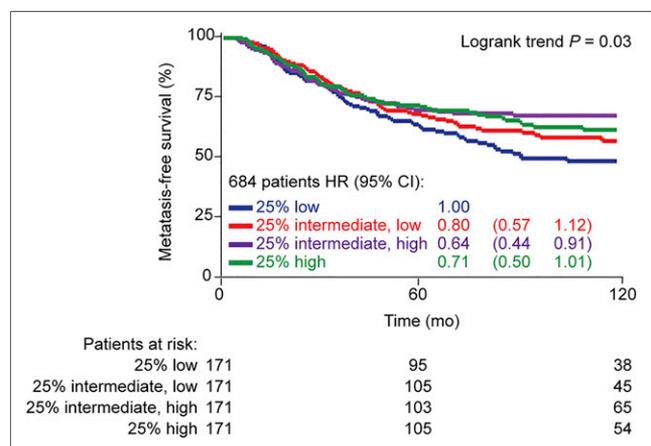


FIGURE 3. Distant MFS in 684 LNN patients as function of levels of CXCR4. CI = confidence interval; HR = hazard ratio.

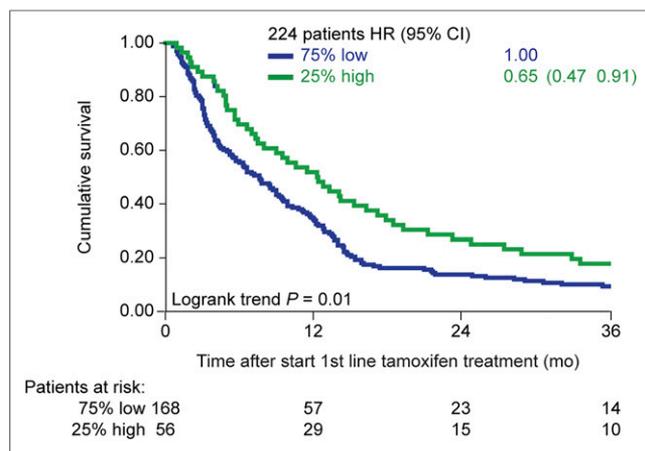


FIGURE 4. Association of GRPR expression with PFS on first-line tamoxifen treatment. CI = confidence interval; HR = hazard ratio.

of 50 (30) and SSTR2 expression in 26 of 53 (SU Dalm, CHM van Deurzen, M Melis, M de Jong, unpublished data, 2014) of the specimens analyzed by in vitro autoradiography, emphasizing that GRPR- and SSTR2-mediated imaging and therapy could be applied in a large group of breast cancer patients.

Contrary to *GRPR* and *SSTR2*, high *CXCR4* mRNA expression was correlated with *ESR1*- and *PGR*-negative tumors, associated with breast cancer of the basal like subtype (2), indicating that these tumors, in particular, might be suitable for *CXCR4*-mediated imaging or therapy. Patients with triple-negative tumors, especially, might benefit from *CXCR4*-mediated therapy, because effective therapy options for this aggressive subtype of breast cancer are scarce. Differences in *CXCR4* expression between *ESR1*- and *PGR*-negative and *ESR1*- and *PGR*-positive patients were less pronounced than for *GRPR* and *SSTR2*. *ESR1*- and *PGR*-positive patients should therefore not be ruled out for *CXCR4*-mediated imaging or therapy.

Except for the presence of the receptors, for the selection of patients for imaging or treatment with radioligands, also the density of GRPR, SSTR2, and CXCR4 might determine the target of choice. In a study by Reubi et al. (15), among other receptors, GRPR and SSTR2 expression in 77 breast cancer tissues was analyzed using in vitro autoradiography. Results showed that high-density GRPR expression was observed in 50 of 77 tumors, compared with 14 of 77 tumors with high-density SSTR2 expression. Similarly, in our previous work we found homogeneous GRPR expression in 56% of the breast cancer specimens analyzed (30), whereas homogeneous SSTR2 expression was seen in 29% only (SU Dalm, CHM van Deurzen, M Melis, M de Jong, unpublished data, 2014).

One of the benefits of targeted imaging and therapy using GRPR, SSTR2, and CXCR4 radioligands is the possibility to upfront select patients who could benefit from these methods using one of the radioligands. For this, either frozen material from breast cancer biopsies can be used to perform in vitro autoradiography with radioligands or formalin-fixed paraffin-embedded material can be used for immunohistochemistry, or both can be used to perform RT-qPCR, to identify patients suited for imaging or therapy.

There are, however, also limitations to our study. First, mRNA expression was used as a surrogate for radiotracer binding and ER, PGR, and HER2 protein expression, which may, despite our current and previously published data (22,23), turn out not to be entirely

equivalent with protein expression. Second, for the prognostic part only, even though our study was relatively large no independent validation was performed. In addition, this is a retrospective study and might not completely represent the current situation in patients.

CONCLUSION

We successfully identified potential breast cancer patient groups for the application of radioligands targeting GRPR, SSTR2, or CXCR4 by analyzing associations between receptor expression and clinicopathologic, biologic, and prognostic factors. Our data show compelling evidence that sensitive and specific nuclear medicine-based imaging and therapy using radioligands might be of great benefit for selected breast cancer patients in a personalized setting. GRPR and SSTR2 radioligands in ER-positive and PR-positive tumors and CXCR4 radioligands in ER-negative patients might offer new, promising tools for imaging and therapy of breast cancer.

DISCLOSURE

The costs of publication of this article were defrayed in part by the payment of page charges. Therefore, and solely to indicate this fact, this article is hereby marked “advertisement” in accordance with 18 USC section 1734. This study was funded by the Erasmus MC grant “The application of radiolabeled peptides in Breast Cancer” and the Cancer Genomics Netherlands funded by the Netherlands Organization for Scientific Research. No other potential conflict of interest relevant to this article was reported.

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