

A Pretargeted PET Imaging Strategy Based on Bioorthogonal Diels–Alder Click Chemistry

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The specificity of antibodies have made immunoconjugates promising vectors for the delivery of radioisotopes to cancer cells; however, their long pharmacologic half-lives necessitate the use of radioisotopes with long physical half-lives, a combination that leads to high radiation doses to patients. Therefore, the development of targeting modalities that harness the advantages of antibodies without their pharmacokinetic limitations is desirable. To this end, we report the development of a methodology for pretargeted PET imaging based on the bioorthogonal Diels–Alder click reaction between tetrazine and transcyclooctene. **Methods:** A proof-of-concept system based on the A33 antibody, SW1222 colorectal cancer cells, and ⁶⁴Cu was used. The huA33 antibody was covalently modified with transcyclooctene, and a NOTA-modified tetrazine was synthesized and radiolabeled with ⁶⁴Cu. Pretargeted in vivo biodistribution and PET imaging experiments were performed with athymic nude mice bearing A33 antigen-expressing, SW1222 colorectal cancer xenografts. **Results:** The huA33 antibody was modified with transcyclooctene to produce a conjugate with high immunoreactivity, and the ⁶⁴Cu-NOTA-labeled tetrazine ligand was synthesized with greater than 99% purity and a specific activity of 9–10 MBq/μg. For in vivo experiments, mice bearing SW1222 xenografts were injected with transcyclooctene-modified A33; after allowing 24 h for accumulation of the antibody in the tumor, the mice were injected with ⁶⁴Cu-NOTA-labeled tetrazine for PET imaging and biodistribution experiments. At 12 h after injection, the retention of uptake in the tumor (4.1 ± 0.3 percent injected dose per gram), coupled with the fecal excretion of excess radioligand, produced images with high tumor-to-background ratios. PET imaging and biodistribution experiments performed using A33 directly labeled with either ⁶⁴Cu or ⁸⁹Zr revealed that although absolute tumor uptake was higher with the directly radiolabeled antibodies, the pretargeted system yielded comparable images and tumor-to-muscle ratios at 12 and 24 h after injection. Further, dosimetry calculations revealed that the ⁶⁴Cu pretargeting system resulted in only a fraction of the absorbed background dose of A33 directly labeled with ⁸⁹Zr (0.0124 mSv/MBq vs. 0.4162 mSv/MBq, respectively). **Conclusion:** The high quality of the images produced by this pretargeting approach, combined with the ability of the methodology to dramatically reduce nontarget radiation doses to patients, marks this system as a strong candidate for clinical translation.

Key Words: positron emission tomography (PET); pretargeting; click chemistry; antibody; colorectal cancer

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The remarkable specificity and affinity of antibodies make them extremely attractive vectors for the delivery of diagnostic and therapeutic radioisotopes to cancer cells (1). Over the last 2 decades, a wide variety of antibody-based radiopharmaceuticals has been developed using isotopes ranging from ⁶⁴Cu for imaging to ²²⁵Ac for therapy. Importantly, the relatively slow pharmacokinetics of antibodies require that the radioactive half-life of the isotope be compatible with the biologic half-life of the target-tissue localization of the vector (2). In practice, this means that antibodies are often labeled with isotopes with multiday physical half-lives. Such long physical half-lives allow time for the antibodies to accumulate in the tumor while simultaneously preventing the premature decay of their radioactive payload. However, this combination of long biologic and physical half-lives gives rise to a critical limiting complication: high activity concentrations in and radiation doses to nontarget organs.

To circumvent this problem, considerable attention has been dedicated to the development of targeting methodologies that combine the advantages of antibodies with the pharmacokinetics of smaller molecules. One particularly appealing method of achieving this balance while still using intact antibodies is termed pretargeting (3–5). Generally, pretargeted methodologies involve 4 steps: the injection into the bloodstream of a bivalent antibody with the ability to bind both an antigen and a radioligand; the slow accumulation of the antibody in the tumor and concomitant clearance of the antibody from the blood; the injection into the bloodstream of the small-molecule radioligand; and the binding of the radioligand to the antibody, followed by the rapid clearance of excess radioactivity. The pharmacokinetics of the small-molecule ligands not only reduces background radiation dose to nontarget organs but also facilitates the use of radioisotopes with short half-lives that would normally be incompatible with antibody-based imaging.

Two strategies for pretargeted systems predominate in the literature. In the first, bispecific antibodies capable of binding both an antigen and a radiolabeled hapten—such as a diethylenetriaminepentaacetic acid-chelated radiometal or a chelator-modified biotin—are used (6,7). In the second strategy, antibodies covalently modified with oligomeric nucleic acids are used as the targeting vectors while radiolabeled, complementary oligonucleotide sequences are

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used as the radioligands (8). Both systems are creative solutions and have proven successful in vivo (9,10). Each, however, possesses significant limitations that threaten their clinical applicability. For the former, these include the complexity and expense associated with the generation of bispecific antibodies and the immunogenicity of streptavidin-based systems; for the latter, both the in vivo stability of the oligonucleotides and the expense of using nonnatural nucleic acid analogs are concerns.

In recent years, a chemical methodology has been developed that has the potential to aid in the creation of novel, antibody-based pretargeting systems without these limitations: bioorthogonal click chemistry (11). Although the [3 + 2] Huisgen cycloaddition between an azide and alkyne remains the preeminent form of click chemistry (12), another promising click ligation variant has recently garnered considerable interest: the rapid, selective, and bioorthogonal inverse electron-demand [4 + 2] Diels–Alder cycloaddition between a tetrazine and a strained alkene dienophile (Fig. 1) (13,14). To date, the ligation has been used in a variety of settings, including fluorescence imaging with antibodies and nanoparticles (13) and PET imaging with both ^{18}F - and radiometal-based probes (15–17). Recently, and most applicable here, 2 reports have been made of pretargeting systems based on the tetrazine–dienophile ligation: one using a small molecule tetrazine moiety equipped with a SPECT isotope and the second using a larger, tetrazine-modified, ^{18}F -labeled dextran-based radioligand (18,19).

Herein, we report the development of a pretargeted PET imaging methodology based on the [4 + 2] cycloaddition reaction between tetrazine and transcyclooctene (Fig. 2). For proof of concept, we have used a colorectal cancer cell line, the A33 antibody, and the positron-emitting radiometal ^{64}Cu as a model system. Ultimately, we have found that the in vivo click methodology is highly effective at delineating tumor from normal tissue, producing PET images with contrast comparable to that obtained with directly radiolabeled antibodies.

MATERIALS AND METHODS

Reagents and General Procedures

All chemicals, unless otherwise noted, were acquired from Sigma-Aldrich and used as received without further purification. All water used was ultra-pure ($>18.2\text{ M}\Omega\text{cm}^{-1}$), and all dimethyl sulfoxide was of molecular biology grade ($>99.9\%$). Both 2,2',2''-(2-(4-aminobenzyl)-1,4,7-triazonane-1,4,7-triyl)triacetic acid ($\text{NH}_2\text{-Bn-NOTA}$) and *p*-isothiocyanatobenzyl-desferrioxamine (SCN-DFO) were purchased from Macrocyclics, Inc. Amine-reactive transcyclooctene (TCO-NHS) and amine-reactive tetrazine (Tz-NHS) were synthesized as described previously (13). Humanized A33 antibody was generously provided by the Ludwig Institute for Cancer Immunotherapy.

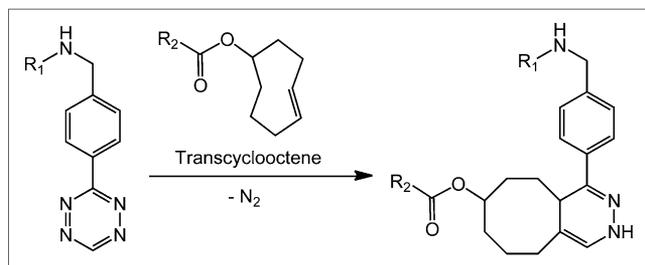


FIGURE 1. Inverse electron-demand Diels–Alder [4 + 2] cycloaddition click ligation between tetrazine and transcyclooctene.

^{64}Cu was purchased from Washington University, St. Louis, where it was produced on the Washington University School of Medicine Cyclotron (model CS-15; Cyclotron Corp.) by the $^{64}\text{Ni}(p,n)^{64}\text{Cu}$ reaction and purified to yield $^{64}\text{Cu-CuCl}_2$ with a specific activity of 7.4–14.8 GBq/ μg . ^{89}Zr was produced at Memorial Sloan-Kettering Cancer Center on a TR19/9 cyclotron (Ebc Industries Inc.) via the $^{89}\text{Y}(p,n)^{89}\text{Zr}$ reaction and purified to yield ^{89}Zr with a specific activity of 196–496 MBq/ μg . Activity measurements were made using a CRC-15R Dose Calibrator (Capintec). For the quantification of activities, experimental samples were counted on an Automatic Wizard (2) γ -Counter (Perkin Elmer). The labeling of antibodies with both ^{64}Cu and ^{89}Zr was monitored using silica gel-impregnated instant thin-layer chromatography (TLC) paper (Pall Corp.) and analyzed on an AR-2000 radio-TLC plate reader (Bioscan Inc.). The human colorectal cancer cell line SW1222 was obtained from the Ludwig Institute for Cancer Immunotherapy and grown by serial passage. All in vivo experiments were performed according to protocols approved by the Memorial Sloan-Kettering Institutional Animal Care and Use Committee.

Modification of A33 with TCO-NHS

A33 (2 mg) was dissolved in 500 μL of phosphate-buffered saline (PBS, pH 7.4), and the pH of the solution was adjusted to 8.8–9.0 with NaHCO_3 (0.1 M). To this solution was added an appropriate volume of TCO-NHS in *N,N*-dimethylformamide (10 mg/mL) to yield a TCO:mAb reaction stoichiometry of 10:1. The resulting solution was incubated with gentle shaking for 30 min at room temperature. After 30 min, the modified antibody was purified using centrifugal filter units with a 50,000-Dalton molecular weight cutoff (Amicon Ultra 4; Millipore Corp.) and PBS.

Radiolabeling of Tz-Bn-NOTA with ^{64}Cu

A solution of Tz-Bn-NOTA (5–6 μg) in NH_4OAc buffer (0.2 M, pH 5.5) was prepared. $^{64}\text{CuCl}_2$ in 0.1 M HCl (74–81 MBq) was added to this solution, and the reaction mixture was heated to 80°C for 10 min. After heating, the solution was allowed to cool to room temperature and purified via high-performance liquid chromatography (HPLC) (retention time, 9.6 min). Solvent was removed via rotary evaporation to yield purified product in an $87\% \pm 3\%$ decay-corrected yield with a radiochemical purity of more than 99% and a specific activity of $8.9 \pm 1.2\text{ MBq}/\mu\text{g}$.

Pretargeted PET Imaging

Pretargeted PET imaging experiments were conducted on a micro-PET Focus 120 scanner (Concorde Microsystems). Mice bearing subcutaneous SW1222 xenografts (100–150 mm^3 , 9–12 d after inoculation) were administered A33-TCO (100 μg in 200 μL of 0.9% sterile saline) via tail vein injection. After 24 h, mice were administered ^{64}Cu -Tz-Bn-NOTA (10.2–12.0 MBq [275–325 μCi] in 200 μL of 0.9% sterile saline) via tail vein injection ($t = 0$). Approximately 5 min before PET imaging, mice were anesthetized by inhalation of a 2% isoflurane (Baxter Healthcare):oxygen gas mixture and placed on the scanner bed; anesthesia was maintained using a 1% isoflurane:gas mixture. PET data for each mouse were recorded in list mode at various time points between 2 and 18 h (further technical specifications are given in the supplemental materials, “Methods” section, available online only at <http://jnm.snmjournals.org>).

RESULTS

System Design

The first step in the development of the pretargeting methodology was the design of the model system. Five components needed to be chosen: antibody, tetrazine, dienophile, radionuclide, and chelator. The antibody selected, A33, is a humanized antibody that targets the A33 antigen, a transmembrane glycoprotein

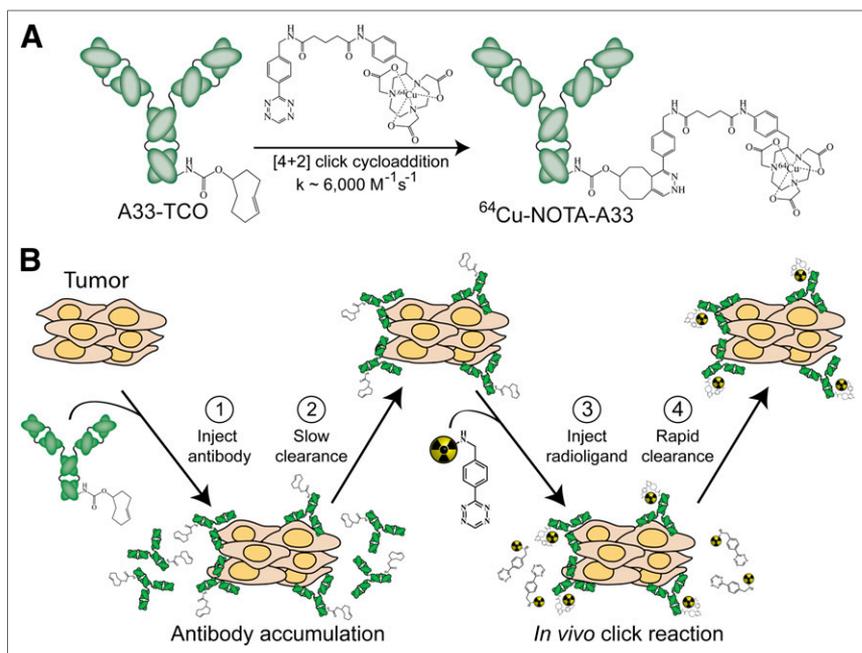


FIGURE 2. Click chemistry-based pretargeting system. (A) Inverse electron-demand Diels-Alder click reaction between A33-TCO and ^{64}Cu -Tz-Bn-NOTA. (B) Illustration of the 4 steps of the pretargeting methodology.

present in more than 95% of human colorectal cancers (20). In addition, because of its association with tight junction proteins, the A33 antigen has been shown to exhibit surface persistence, even when bound to the targeting antibody (21). This is an extremely important trait in the context of pretargeting: the internalization and consequent sequestration of the antibody before the administration of the radioligand would severely reduce the likelihood of *in vivo* click ligations. 3-(4-benzylamino)-1,2,4,5-tetrazine and transcyclooct-4-en-1-yl hydrogen carbonate were chosen as the tetrazine-dienophile click pair because of their convenient conjugation handles and their balance of relative stability with rapid reaction kinetics ($k = 6,000 \text{ M}^{-1}\text{s}^{-1}$ at 37°C) (13,14). Finally, ^{64}Cu was selected as the radioisotope. Despite the utility and ubiquity of ^{18}F , a radio-metal-based system offers greater versatility and modularity. In particular, ^{64}Cu displays favorable imaging characteristics and exhibits a half-life (12.7 h) normally considered suboptimal for clinical antibody imaging. The choice of ^{64}Cu , in turn, narrowed the choice of chelator, and NOTA was selected because of its relatively rapid chelation kinetics and high *in vivo* stability with ^{64}Cu (22).

Synthesis, Characterization, and Reactivity of Components

The NOTA-modified tetrazine (Tz-Bn-NOTA) was synthesized in high yield (>95%) via peptide coupling from NH_2 -Bn-NOTA and 3-(4-benzylamino)-1,2,4,5-tetrazine bearing an amine-reactive

linker (Fig. 3). The compound was purified using reversed-phase C_{18} HPLC and characterized by ultraviolet-visible spectroscopy, proton nuclear magnetic resonance spectroscopy, and electrospray ionization mass spectrometry. Tz-Bn-NOTA was, in turn, labeled with ^{64}Cu via incubation with ^{64}Cu - CuCl_2 at 90°C for 10 min in NH_4OAc buffer. The radioligand for 10 min in NH_4OAc buffer. The radioligand was obtained in $87\% \pm 3\%$ decay-corrected yield and purified via reversed-phase HPLC to more than 98% radiochemical purity, and its identity was confirmed against a cold Cu-Tz-Bn-NOTA standard. The specific activity of the purified ^{64}Cu -Tz-Bn-NOTA was $8.9 \pm 1.2 \text{ MBq}/\mu\text{g}$ ($n = 10$).

The stability of the radioligand was assayed *in vitro* by incubation in human serum at 37°C and subsequent HPLC analysis. The ^{64}Cu -Tz-Bn-NOTA remained relatively stable after 1 h, with $93.0\% \pm 2.5\%$ intact. With time, however, decomposition of the ^{64}Cu -Tz-Bn-NOTA becomes apparent: at 12 h, only $31.6\% \pm 6.1\%$ of the radioligand remained intact. In addition, *in vivo* stability experiments were performed, and these produced similar results, although some acceleration in decomposition can be noted: $77\% \pm 3\%$ of the ^{64}Cu -Tz-Bn-NOTA is intact in the blood after 60 min, followed by $47\% \pm 2\%$ after 2 h and $22\% \pm 3\%$ after 6 h. Although greater stability may be preferable, the speed of the cycloaddition reaction and the rapid pharmacokinetics of the radioligand led us to believe it unlikely that poor serum stability at later time points would limit the system (Supplemental Figs. 1 and 2; Supplemental Table 1).

The TCO-modified A33 (A33-TCO) was constructed via reaction of the antibody with 10 molar equivalents of the *N*-hydroxysuccinimidyl ester of TCO (TCO-NHS) for 30 min at room temperature in PBS adjusted to pH 8.8–9.0 with NaHCO_3 (0.1 M), and the bioconjugate was purified using centrifugal filtration. To determine the number of TCO per monoclonal antibody, the antibody was ligated with a 50-fold molar excess of Tz-Bn-NOTA in PBS (pH 7.4) and purified via size-exclusion chromatography. An isotopic dilution was performed on this NOTA-modified product to determine a degree of labeling of 5.0 ± 0.9 NOTA/mAb, in turn suggesting a degree of labeling of 5 TCO/mAb, given the quantitative nature of the click ligation (18).

With the 2 system components in hand, the reaction between A33-TCO and ^{64}Cu -Tz-Bn-NOTA was next investigated. A33-TCO was incubated with ^{64}Cu -Tz-Bn-NOTA in 1:5, 1:1, or 5:1 Tz-to-mAb molar ratios in either PBS or serum at 37°C for 30 min. As analyzed by radio-TLC, the reaction proceeded to

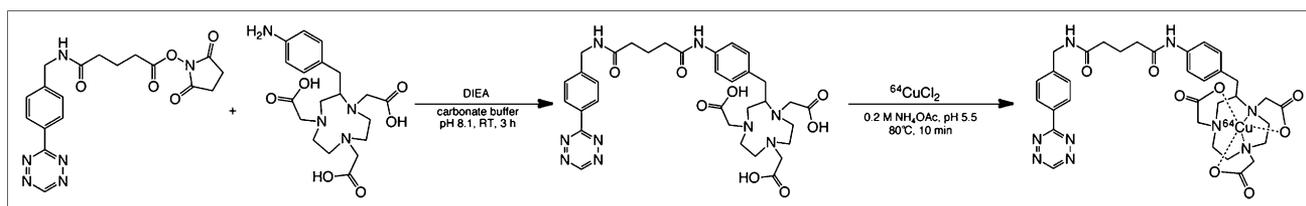


FIGURE 3. Synthesis of ^{64}Cu -Tz-Bn-NOTA.

near-complete conversion, with the 1:5 Tz-to-mAb reaction providing yields of $91.2\% \pm 0.9\%$ and $89.6\% \pm 1.8\%$ in PBS and serum, respectively, with only slightly diminished yields with the other ratios (18). As negative controls, the reactions of free ^{64}Cu with A33-TCO and ^{64}Cu -Tz-Bn-NOTA with unmodified A33 were investigated, and in each case less than 1% association of the activity with the antibody was observed (Supplemental Tables 2 and 3).

Finally, to probe the effect of the TCO conjugation on the immunoreactivity of the antibody, A33-TCO was reacted with a 5-fold excess of ^{64}Cu -Tz-Bn-NOTA in PBS and purified via size-exclusion chromatography to form ^{64}Cu -NOTA-A33 in more than 99% radiochemical purity and a specific activity of 125.8 ± 11.1 MBq/mg (Supplemental Figs. 3–5). Immunoreactivity assays with A33 antigen-expressing SW1222 human colorectal cancer cells yielded an immunoreactivity for the bioconjugate of $95.6\% \pm 4.3\%$.

Pretargeted PET Imaging and Biodistribution

After the synthesis and characterization of the components of the system, the next step was to perform in vivo pretargeted biodistribution and PET imaging experiments. Not surprisingly, one of the most important variables in the development of a pretargeted system is the length of the interval between the injection of the antibody and the administration of the radioligand. In this case, biodistribution studies with clicked and preassembled ^{64}Cu -NOTA-A33 (see the “Discussion” section) indicate that the maximum loading of ^{64}Cu -NOTA-A33, and thus by proxy A33-TCO, in the tumor occurs at 24 h after injection, and relatively little antibody remains in the blood pool at the same time point. Thus, it becomes clear that 24–48 h represents a promising interval between the administration of antibody and the subsequent injection of radioligand.

For the pretargeted biodistribution experiments, mice bearing subcutaneous SW1222 xenografts were administered A33-TCO (100 μg) via tail vein injection. After a 24-h interval for the accumulation of the antibody in the tumor and its simultaneous clearance from the blood, the mice were administered ^{64}Cu -Tz-Bn-NOTA (0.55–0.75 MBq, 0.06–0.09 μg , a 0.14–0.18 Tz-to-A33 ratio) via tail vein injection. The data reveal rapid accumulation and retention of radioactivity in the tumor, with 4.1 ± 0.3 percent injected dose per gram (%ID/g) at 1 h after injection, 4.2 ± 0.8 %ID/g at 12 h, and 4.0 ± 0.9 %ID/g at 24 h (Table 1; Supplemental Tables 4 and 5). The compound is excreted through the feces, with high levels of activity at 1 h after injection dropping to 2.5 ± 0.1 %ID/g at 12 h. The amount of uptake in all other tissues remains low, generally less than 1 %ID/g. The rapid and steady uptake in the tumor indicates that ^{64}Cu -Tz-Bn-NOTA is clicking with A33-TCO localized at the tumor, rather than clicking with A33-TCO in the blood pool that subsequently accumulates in the tumor.

PET imaging experiments were performed in a similar manner; the only change was the higher amount of injected activity (10.2–12.0 MBq, 1.2–1.4 μg , a 2.5–2.8 Tz-to-A33 ratio) (Fig. 4; Supplemental Figs. 6 and 7). Tumor uptake is apparent at 2 h after injection, although consistent with the biodistribution, it is much less than the uptake in the gut (feces). However, over the course of the experiment, the fecal radioactivity clears dramatically, resulting in excellent delineation of tumor from background by 12 h after injection, in agreement with the 26.6 ± 6.6 tumor-to-muscle activity concentration ratio measured at the same time point in the biodistribution study.

Imaging experiments in which 300 μg of A33-TCO were administered showed similar results; however, these experiments revealed higher activity levels in the blood, suggesting a greater

TABLE 1
Biodistribution Data for In Vivo ^{64}Cu
Pretargeting Experiment

Organ	1 h	4 h	12 h	24 h
Blood	3.5 ± 0.6	2.6 ± 0.8	2.3 ± 0.4	2.1 ± 0.5
Tumor	4.1 ± 0.3	4.1 ± 0.6	4.2 ± 0.8	4.0 ± 0.9
Heart	1.1 ± 0.2	0.9 ± 0.3	0.9 ± 0.1	0.8 ± 0.2
Lung	1.6 ± 0.5	1.6 ± 0.4	1.1 ± 0.4	1.0 ± 0.3
Liver	2.2 ± 0.3	1.3 ± 0.3	0.9 ± 0.2	1.1 ± 0.2
Spleen	0.6 ± 0.1	0.5 ± 0.2	0.6 ± 0.3	0.4 ± 0.1
Stomach	0.5 ± 0.1	0.3 ± 0.1	0.5 ± 0.6	0.2 ± 0.0
Large intestine	0.5 ± 0.0	3.1 ± 1.1	2.3 ± 1.0	1.1 ± 0.4
Feces	11.9 ± 4.4	8.8 ± 3.4	2.5 ± 0.1	1.4 ± 0.5
Small intestine	0.0 ± 0.0	0.4 ± 0.1	0.8 ± 0.5	0.3 ± 0.0
Kidney	1.3 ± 0.1	0.9 ± 0.3	0.9 ± 0.3	0.7 ± 0.2
Muscle	0.2 ± 0.0	0.1 ± 0.0	0.2 ± 0.0	0.1 ± 0.0
Bone	0.3 ± 0.2	0.3 ± 0.2	0.4 ± 0.1	0.3 ± 0.1
Skin	0.3 ± 0.1	0.3 ± 0.1	0.4 ± 0.1	0.5 ± 0.2

Values are %ID/g \pm SD. Mice ($n = 4$) bearing subcutaneous SW1222 xenografts were administered A33-TCO (100 μg) via tail vein injection. After 24 h, the same mice were administered ^{64}Cu -Tz-Bn-NOTA (0.55–0.75 MBq [15–20 μCi]) via tail vein injection ($t = 0$).

incidence of in vivo click reactions in the blood in addition to ligations at the tumor (Supplemental Figs. 8 and 9; Supplemental Tables 6 and 7). Similarly, imaging experiments performed with a pretargeting interval of 12 h resulted in a higher degree of activity in the blood, likely for the same reason. Control imaging experiments using ^{64}Cu -Tz-Bn-NOTA alone, unmodified A33 rather than A33-TCO, and a vast excess of unlabeled tetrazine resulted in minimal uptake of activity in the tumor, in all cases less than 0.25 %ID/g. Further, pretargeting with a nonspecific IgG-TCO construct resulted in apparent yet slight tumor uptake (<1.5 %ID/g), not surprising given the tendency of nontargeted antibodies to accumulate in tumors because of the enhanced permeability and retention effect (Supplemental Figs. 10–14) (23). Finally, although preliminary pilot imaging experiments with longer pretargeting intervals (e.g., 72 h) strongly suggest that the 24-h interval used strikes the optimal balance between tumor uptake and background clearance, further interval optimization studies are currently under way in our laboratory.

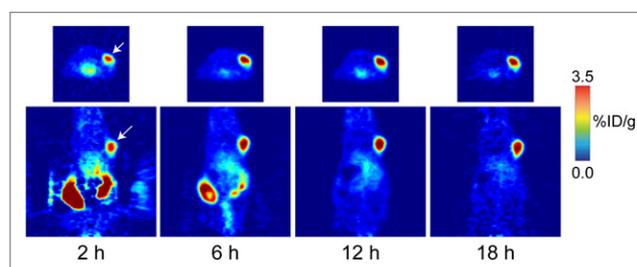


FIGURE 4. PET images of ^{64}Cu -Tz-Bn-NOTA/A33-TCO pretargeting strategy. Mice bearing subcutaneous SW1222 xenografts (100–150 mm^3 , arrow) were administered A33-TCO (100 μg) via tail vein injection. After 24 h, the same mice were administered ^{64}Cu -Tz-Bn-NOTA (10.2–12.0 MBq [275–325 μCi], 1.2–1.4 μg , for 2.5–2.8 Tz-to-A33 ratio) via tail vein injection. Transverse (top) and coronal (bottom) planar images intersect the center of the tumors.

Multimodality Imaging, Autoradiography, and Fluorescence Microscopy

Although the PET imaging experiments form the core of this investigation, they only allow us to monitor the fate of 1 of the 2 components in the system: the radioligand. To circumvent this issue, A33-TCO was modified with the near-infrared fluorophore AlexaFluor680 (AF680) to yield A33-TCO-AF680 with a degree of labeling of 1.6 AF680/mAb. Tumor-bearing mice were then injected with A33-TCO-AF680 (100 μ g), imaged with fluorescence at 24 h after antibody injection, injected immediately thereafter with ^{64}Cu -Tz-Bn-NOTA (10.2–12.0 MBq [275–325 μCi]), and then imaged with both fluorescence and PET 12 h after radiotracer injection. Not surprisingly, the PET imaging results from these experiments clearly mirror those reported above for the non-fluorescent antibody (Supplemental Figs. 15–17).

More important than the *in vivo* imaging experiments, this multimodality approach facilitated the tracking of the 2 system components at the microscopic level via autoradiography and fluorescence microscopy (Fig. 5). To this end, *ex vivo* analysis of resected tumors was performed to determine the relative distributions of A33-TCO-AF680 and ^{64}Cu -Tz-Bn-NOTA. A close correspondence of the 2 components was observed, with regions of high A33-TCO-AF680 uptake spatially matching regions of high ^{64}Cu -Tz-Bn-NOTA uptake. Further, A33-TCO-AF680 was associated exclusively with regions containing tumor cells and did not appear to associate with stromal or muscle tissue. More than any other piece of data in this investigation, the clear and striking microscopic colocalization of the fluorescently labeled A33-TCO and the ^{64}Cu -Tz-Bn-NOTA illustrates the occurrence of the *in vivo* click ligation. Finally, A33-TCO-AF680 shows a distinct cell surface distribution, which is easily visualized when compared with the nuclear counterstain 4',6-diamidino-2-phenylindole and is consistent with the cell surface expression of the A33 antigen.

DISCUSSION

Ultimately, an *in vivo* comparison with the preassembled and purified ^{64}Cu -NOTA-A33 construct is essential for the evaluation of the efficacy of this pretargeted system. To this end, the ^{64}Cu -labeled antibody was synthesized as described above and purified to more than 99% radiochemical purity, with a specific activity of 125.8 ± 11.1 MBq/mg and an immunoreactivity of more than 95%. In the biodistribution experiment, nude mice bearing SW1222 xenografts in the right shoulder were injected with ^{64}Cu -NOTA-A33 (0.55–0.75 MBq, 4–6 μ g) and were euthanized at 4, 12, 24, and 48 h, followed by the collection and weighing of tissues and assay of ^{64}Cu activity in each tissue.

High specific uptake of the radiotracer was observed in the SW1222 xenografts, with the %ID/g increasing from 18.2 ± 3.0 at 4 h to 35.0 ± 3.8 at 48 h. As is typical of antibody-based imaging, a concomitant decrease in the %ID/g in the blood also occurred over the course of the experiment. The organs with the highest background uptake were the lungs, liver, and spleen, though the uptake in these organs was at its highest point at 4 h; by 48 h, the tumor-to-organ ratios for each of these organs were 20.8 ± 13.1 , 10.0 ± 4.5 , and 19.8 ± 5.4 , respectively (Supplemental Tables 8–11).

These biodistribution data were corroborated by small-animal PET imaging. In these experiments, tumor-bearing mice were injected with ^{64}Cu -NOTA-A33, and PET images were acquired at various time points between 4 and 48 h. The results clearly indicate that the ^{64}Cu -NOTA-A33 constructs are taken up in the

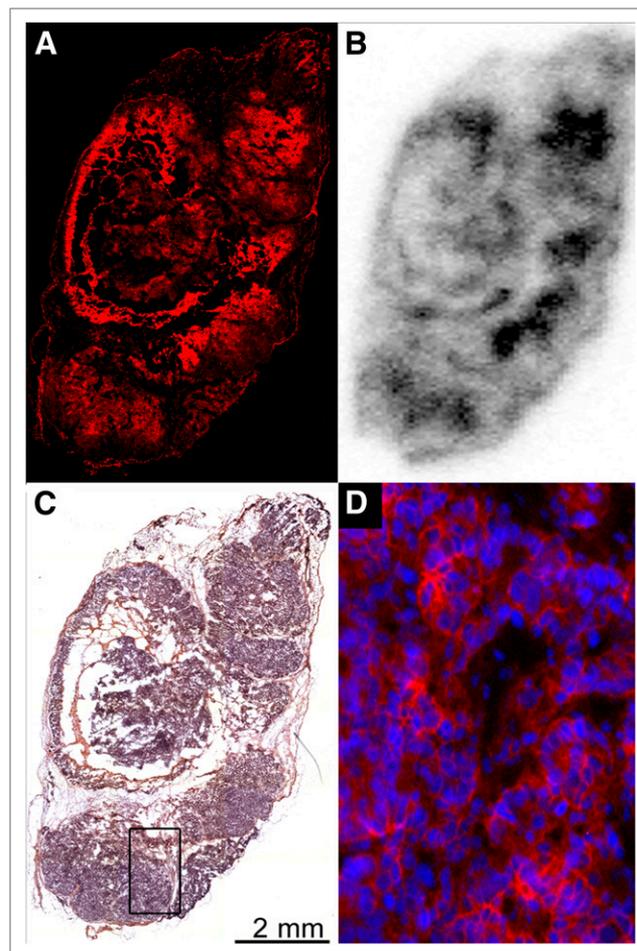


FIGURE 5. Autoradiography, histology, and fluorescence microscopy of resected SW1222 xenografts from multimodality pretargeting experiment: fluorescence microscopy indicating A33-TCO-AF680 localization (red) (A), autoradiography indicating ^{64}Cu -Tz-Bn-NOTA localization (B), hematoxylin and eosin staining (C), and fluorescence microscopy images with A33-TCO-AF680 (red) and 4',6-diamidino-2-phenylindole nuclear counterstain (blue) corresponding to box in C (D).

antigen-expressing SW1222 tumor (Fig. 6; Supplemental Fig. 18). High blood-pool activity and background uptake are evident at early time points, but over the course of the experiment, the tumor signal increases significantly to a point at which it is by far the most prominent feature in the image.

As expected for an intact antibody, the ^{64}Cu -NOTA-A33 displays higher absolute uptake in the tumor than the pretargeted system: 33.1 ± 7.0 %ID/g at 24 h, compared with 4.0 ± 0.9 %ID/g for the click chemistry approach at the same time point. However, for imaging purposes, the far more important question is target-to-background ratio, and indeed, when speaking in terms of relative uptake, the methodologies become far more similar. At 24 h after injection, the pretargeting methodology provides a tumor-to-muscle activity ratio that is statistically identical to that of ^{64}Cu -NOTA-A33: 27.0 ± 7.4 versus 33.8 ± 6.7 , respectively. In addition, at all time points assayed, the uptake in nontarget tissues is significantly higher for the intact antibody than for the pretargeting system in all organs save the large intestine.

Not surprisingly given the pharmacokinetics of intact antibodies, at later time points, the tumor-to-muscle activity ratio for

the intact ^{64}Cu -NOTA-A33 surpasses the maximum values measured for the pretargeting system, reaching 52.2 ± 14.7 at 48 h. Likewise, whereas the tumor-to-blood ratios of ^{64}Cu -NOTA-A33 and the pretargeting system are comparable at 24 h (2.9 ± 0.4 and 1.9 ± 0.6 , respectively), at 48 h, the tumor-to-blood ratio of ^{64}Cu -NOTA-A33 (24.5 ± 11.6) exceeds the maximum ratio observed with the pretargeting system. Yet despite the high absolute and relative uptake of ^{64}Cu -NOTA-A33 at these later time points, the data clearly illustrate that the pretargeted system represents a qualitatively and quantitatively comparable alternative to ^{64}Cu -based antibody imaging.

The preceding comparison to ^{64}Cu -NOTA-A33 undoubtedly provides the most direct measure of the comparative efficacy of the pretargeting system. However, the recent and rapid advent of clinical ^{89}Zr -immunoPET likely means that many of the antibody-based imaging agents in the future will be labeled with ^{89}Zr . Thus, we believe that a comparison to directly labeled ^{89}Zr -DFO-A33 could also be valuable by providing insight into the merits of ^{64}Cu -based pretargeting as an alternative to antibodies labeled with ^{89}Zr . To this end, ^{89}Zr -DFO-A33 was synthesized by modifying A33 with the ^{89}Zr chelator desferrioxamine via isothiocyanate coupling (3.5 ± 1.1 DFO/A33) and radiolabeling this construct with ^{89}Zr using standard procedures (2). The purified radiopharmaceutical exhibited more than 99% radiochemical purity, an immunoreactivity of $92\% \pm 5\%$, more than 95% stability over 7 d at 37°C , and a specific activity of 159.1 ± 22.2 MBq/mg (Supplemental Figs. 19–21).

For both PET imaging and biodistribution experiments, SW1222 tumor-bearing animals were injected with ^{89}Zr -DFO-A33 (10.2–12.0 MBq for PET, 0.55–0.75 MBq for biodistribution) and subsequently imaged or euthanized for biodistribution at time points between 4 and 120 h after injection. Both the PET and the biodistribution experiments display all the hallmarks of ^{89}Zr -based antibody imaging (Fig. 7). High early levels of activity in the blood (58.4 ± 8.5 %ID/g at 4 h) decrease over the first 48 h as the activity levels in the tumor increase dramatically to a maximum of 44.7 ± 10.5 %ID/g at 48 h. All other tissues exhibit much lower activities after 48 h with the exception of bone, which displays uptake values of approximately 10 %ID/g, a consequence of the mineralization of liberated $^{89}\text{Zr}^{4+}$ in bone (Supplemental Fig. 22; Supplemental Tables 12 and 13).

In comparative terms, the directly labeled ^{89}Zr -DFO-A33, like ^{64}Cu -NOTA-A33, is characterized by higher absolute tumor uptake, with 43.3 ± 9.0 %ID/g in the xenograft at 24 h compared with 4.0 ± 0.9 %ID/g for the pretargeting system. Again, however, the pretargeting methodology excels with regard to the more important metric: tumor-to-background ratio. Here, the pretargeting approach produces a higher tumor-to-muscle activity ratio than ^{89}Zr -DFO-A33 at 24 h after injection (27.0 ± 7.4 vs. 12.7 ± 3.3 , respectively) as well as a comparable tumor-to-blood activity ratio at the same time point (1.9 ± 0.6 vs. 2.4 ± 0.7 , respectively). Further, the background associated with the intact antibody is significantly higher than in the pretargeted system.

Not surprisingly, though, both the absolute uptake and the tumor-to-background ratios for ^{89}Zr -DFO-A33 increase further at later time points. For example, despite the higher tumor-to-muscle ratio produced by the pretargeting approach at 24 h, the tumor-to-muscle activity ratios for ^{89}Zr -DFO-A33 surpass this value at later time points, ultimately reaching a maximum of 57.7 ± 7.0 at 72 h.

Critically, for both ^{64}Cu -NOTA-A33 and ^{89}Zr -DFO-A33, the high absolute uptake values and tumor-to-background ratios ob-

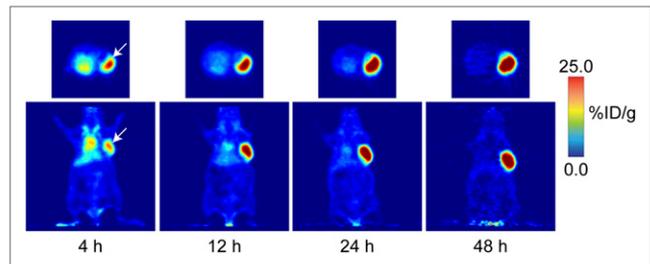


FIGURE 6. PET imaging with ^{64}Cu -NOTA-A33. Mice bearing subcutaneous SW1222 xenografts ($100\text{--}150\text{ mm}^3$, arrow) were administered ^{64}Cu -NOTA-A33 (10.2–12.0 MBq [$275\text{--}325\text{ }\mu\text{Ci}$]) via tail vein injection and imaged between 4 and 48 h after injection. Transverse and coronal planar images intersect the center of the tumors.

served at later time points do not come without cost. In both cases— ^{89}Zr , in particular—the pharmacokinetics of the antibodies mean that high levels of radioactivity persist in the patient for extended periods of time; although these levels of activity are often highest in the tumor, the levels in background organs are relatively high as well. It is in this regard that the pretargeted strategy offers an exciting and innovative alternative, for the data clearly indicate that the pretargeted approach produces images comparable to those created with directly radiolabeled antibodies but with both accelerated pharmacokinetics and dramatically lower background uptake in nontarget tissues.

This discussion leads us to another potential advantage to consider for the pretargeting system: reduced radiation dose to nontarget tissues. Using biodistribution data, we performed dosimetry calculations for the pretargeting system, ^{64}Cu -NOTA-A33, and ^{89}Zr -DFO-A33 using the OLINDA computer program to determine mean organ absorbed doses and effective dose in mGy/MBq and mSv/MBq respectively (Table 2) (24). The data indicate that the pretargeting system provides a significant dosimetric advantage, compared with the ^{89}Zr -labeled antibody: the effective dose with ^{64}Cu pretargeting is 0.0124 mSv/MBq whereas that due to ^{89}Zr -DFO-A33 is 0.416 mSv/MBq. A more pronounced disparity between the two exists when considering the mean absorbed dose to the bone, specifically red marrow and osteogenic cells; with ^{89}Zr -DFO-A33, the dose to these tissues is 0.843 and 1.646 mGy/MBq, respectively, compared with 0.0413 and 0.0230 with ^{64}Cu pretargeting. Not surprisingly given the shorter half-life of ^{64}Cu , these advantages are not as pronounced when comparing the pretargeting system to ^{64}Cu -NOTA-A33. The effective dose of ^{64}Cu -NOTA-A33 is 0.0359 mSv/MBq, approximately 3 times larger than the pretargeting value of 0.0124 mSv/MBq. Thus, whereas the dosimetric differences are somewhat attenuated when making comparisons to the

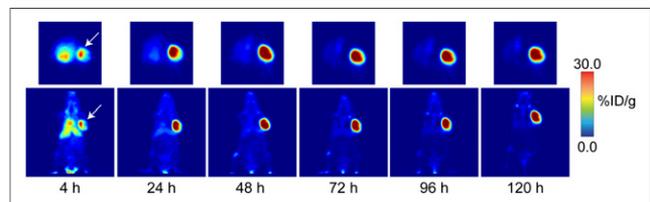


FIGURE 7. PET imaging with ^{89}Zr -DFO-A33. Mice bearing subcutaneous SW1222 xenografts ($100\text{--}150\text{ mm}^3$, arrow) were administered ^{89}Zr -DFO-A33 (10.2–12.0 MBq [$275\text{--}325\text{ }\mu\text{Ci}$]) via tail vein injection and imaged between 4 and 120 h after injection. Transverse and coronal planar images intersect the center of the tumors.

^{64}Cu -labeled antibody, it becomes clear that the pretargeting system provides qualitatively and quantitatively comparable imaging results at only a fraction of the background radiation dose of the directly labeled antibodies, particularly the ^{89}Zr -labeled construct.

CONCLUSION

Herein, we have reported the development of a modular pretargeting system for PET imaging based on the inverse electron-demand Diels–Alder cycloaddition between tetrazine and transcyclooctene. In a model system, we have found that this *in vivo* click methodology produces high-quality images with tumor-to-background contrast comparable to that achieved with directly radiolabeled antibodies. Further, the system facilitates the production of these images at a dramatically reduced absorbed dose to nontarget organs compared with imaging with a ^{89}Zr -labeled antibody.

Despite these successes, the system is not without slight limitations. First, the system requires that the antibody either be non-internalizing or have a slow rate of internalization; although this applies to both A33 and several other clinically interesting antibodies (e.g., CC49), there are certainly other clinically relevant antibodies that are known to internalize upon antigen binding and thus would be much more challenging to use in a pretargeted strategy. And second, the relatively slow fecal excretion of the excess radioligand limits both the ability to image abdominal tumors rapidly after the administration of the radio-

ligand and the clinical use of radionuclides with short half-lives, such as ^{18}F or ^{68}Ga . Currently, experiments are under way to develop novel tetrazine constructs that have altered pharmacokinetics and pharmacodynamics and thus should enable more rapid imaging. However, these changes may not be strictly necessary in the clinic. If other methods for clearing radioactive feces from the patient are adopted, images can be recorded at earlier time points, and radionuclides with short half-lives such as ^{18}F or ^{68}Ga may be used, further augmenting the dosimetric benefits of the approach.

Ultimately, we believe that this pretargeted PET imaging methodology could be an extremely valuable and versatile clinical tool. Specifically, the success of the A33-based proof-of-concept system described here—both in terms of tumor delineation and in terms of dosimetry—has identified the system as a prime candidate for translation to the clinic for the imaging of colorectal cancer. Translation of this approach is an enticing prospect for two principal reasons. First, from a dosimetry perspective, the $^{64}\text{Cu}/\text{A33}$ pretargeting system would almost certainly prove a safer alternative for patients than the $^{124}\text{I}\text{-A33}$ PET radiotracer currently used, without undue sacrifice in image quality. Second, the targeting of A33 to normal bowel has proved a major concern in clinical trials using A33 (25). Kinetic models, however, have suggested that the half-life of the antibody in normal colon is far less than that of the antibody in the tumor. A pretargeting strategy would be a near

TABLE 2
Dosimetry Calculations for Imaging Constructs Studied

Target organ*	^{64}Cu pretargeting [†]	^{64}Cu -NOTA-A33 [‡]	^{89}Zr -DFO-A33 [§]
Adrenals	0.0068	0.0196	0.4432
Brain	0.0064	0.0150	0.2065
Breasts	0.0056	0.0138	0.1678
Gallbladder wall	0.0074	0.0200	0.3892
Lower large intestine wall	0.0449	0.0522	0.3622
Small intestine	0.0089	0.0225	0.3000
Stomach wall	0.0072	0.0239	0.2565
Upper large intestine wall	0.0308	0.0392	0.3243
Heart wall	0.0079	0.0292	0.4189
Kidneys	0.0085	0.0503	0.6838
Liver	0.0084	0.0524	0.7676
Lungs	0.0078	0.0484	0.6108
Muscle	0.0037	0.0148	0.3432
Ovaries	0.0081	0.0184	0.2946
Pancreas	0.0070	0.0191	0.3703
Red marrow	0.0143	0.0832	0.8432
Osteogenic cells	0.0230	0.1186	1.6459
Skin	0.0052	0.0125	0.1830
Spleen	0.0049	0.0324	0.6811
Testes	0.0061	0.0141	0.1846
Thymus	0.0061	0.0158	0.2670
Thyroid	0.0062	0.0152	0.2559
Urinary bladder wall	0.0071	0.0165	0.2232
Uterus	0.0075	0.0176	0.2543
Total body	0.0074	0.0231	0.3757
Effective dose	0.0124	0.0359	0.4162

*Mean organ absorbed doses and effective dose are expressed in mGy/MBq and mSv/MBq, respectively.

[†]100 μg of A33-TCO injected 24 h before injection of ^{64}Cu -Tz-Bn-NOTA.

[‡]5 μg of ^{64}Cu -NOTA-A33 per injection.

[§]5 μg of ^{89}Zr -DFO-A33 per injection.

ideal way to circumvent this complication, because the pretargeted radioligand could be injected after the shedding of the antibody from the normal colon but before the departure of the antibody from the cancerous tissue.

Equally important, this methodology could also have a significant impact beyond the system described here. Along these lines, the modularity of the system is an incredibly important asset: the transcyclooctene moiety can be appended to any antibody of interest, and the simplicity of the tetrazine-based radioligand facilitates the creation of a library of constructs bearing various radioisotopes. Indeed, preliminary experiments are under way in the creation of pretargeting systems for both ^{89}Zr -based PET imaging and ^{177}Lu - and ^{90}Y -based radiotherapy. However, the paramount value of the system plainly lies in its ability to harness the advantages of antibody-based radiopharmaceuticals while skirting their pharmacokinetic limitations. The methodology produces high tumor-to-background contrast at relatively early time points while limiting uptake in, and thus dose to, normal organs, especially compared with antibodies bearing radionuclides with long physical half-lives. Consequently, this approach may ultimately offer an effective and safer alternative to both immunoPET and immunoSPECT imaging with long-lived isotopes such as ^{89}Zr , ^{111}In , or ^{124}I and radioimmunotherapy with isotopes such as ^{177}Lu , ^{90}Y , or ^{131}I .

DISCLOSURE

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REFERENCES

1. Wu AM. Antibodies and antimatter: the resurgence of immuno-PET. *J Nucl Med.* 2009;50:2–5.
2. Zeglis B, Lewis JS. A practical guide to the construction of radiometallation bioconjugates for positron emission tomography. *Dalton Trans.* 2011;40:6168–6195.
3. Hollander N. Bispecific antibodies for cancer therapy. *Immunotherapy.* 2009;1:211–222.
4. Sharkey RM, Goldenberg DM. Advances in radioimmunotherapy in the age of molecular engineering and pretargeting. *Cancer Invest.* 2006;24:82–97.
5. Boerman OC, van Schaijk FG, Oyen WJG, Corstens FHM. Pretargeted radioimmunotherapy of cancer: progress step by step. *J Nucl Med.* 2003;44:400–411.
6. Press OW, Corcoran M, Subbiah K, et al. A comparative evaluation of conventional and pretargeted radioimmunotherapy of CD20-expressing lymphoma xenografts. *Blood.* 2001;98:2535–2543.
7. Goldenberg DM, Chang CH, Rossi EA, McBride WJ, Sharkey RM. Pretargeted molecular imaging and radioimmunotherapy. *Theranostics.* 2012;2:523–540.
8. Liu G, Dou S, Liu Y, Wang Y, Ruszkowski M, Hnatowich DJ. ^{90}Y -labeled phosphorodiamidate morpholino oligomer for pretargeting radiotherapy. *Bioconjug Chem.* 2011;22:2539–2545.
9. Lewis MR, Wang M, Axworthy DB, et al. In vivo evaluation of pretargeted ^{64}Cu for tumor imaging and therapy. *J Nucl Med.* 2003;44:1284–1292.
10. Mohsin H, Jia F, Bryan JN, et al. Comparison of pretargeted and conventional CC49 radioimmunotherapy using ^{149}Pm , ^{166}Ho , and ^{177}Lu . *Bioconjug Chem.* 2011;22:2444–2452.
11. Kolb HC, Finn MG, Sharpless KB. Click chemistry: diverse chemical function from a few good reactions. *Angew Chem Int Ed Engl.* 2001;40:2004–2021.
12. Sletten EM, Bertozzi CR. Bioorthogonal chemistry: fishing for selectivity in a sea of functionality. *Angew Chem Int Ed Engl.* 2009;48:6973–6998.
13. Devaraj NK, Upadhyay R, Hatini JB, Hilderbrand SA, Weissleder R. Fast and sensitive pretargeted labeling of cancer cells through a tetrazine/trans-cyclooctene cycloaddition. *Angew Chem Int Ed Engl.* 2009;48:7013–7016.
14. Blackman ML, Royzen M, Fox JM. Tetrazine ligation: fast bioconjugation based on inverse electron demand Diels-Alder reactivity. *J Am Chem Soc.* 2008;130:13518–13519.
15. Zeglis BM, Mohindra P, Weissmann GI, et al. Modular strategy for the construction of radiometalated antibodies for positron emission tomography based on inverse electron demand Diels-Alder click chemistry. *Bioconjug Chem.* 2011;22:2048–2059.
16. Li Z, Cai H, Hassink M, et al. Tetrazine-trans-cyclooctene ligation for the rapid construction of 18-F labeled probes. *Chem Commun (Camb).* 2010;46:8043–8045.
17. Reiner T, Keliher EJ, Earley S, Marinelli B, Weissleder R. Synthesis and in vivo imaging of a ^{18}F -labeled PARP1 inhibitor using a chemically orthogonal scavenger-assisted high-performance method. *Angew Chem Int Ed Engl.* 2011;50:1922–1925.
18. Rossin R, Verkerk PR, van den Bosch SM, et al. In vivo chemistry for pretargeted tumor imaging in live mice. *Angew Chem Int Ed Engl.* 2010;49:3375–3378.
19. Devaraj NK, Thurber GM, Keliher EJ, Marinelli B, Weissleder R. Reactive polymer enables efficient in vivo bioorthogonal chemistry. *Proc Natl Acad Sci USA.* 2012;109:4762–4767.
20. Carrasquillo JA, Pandi-Taskar N, O'Donoghue JA, et al. ^{124}I -huA33 antibody PET of colorectal cancer. *J Nucl Med.* 2011;52:1173–1180.
21. Ackerman ME, Chalouni C, Schmidt MM, et al. A33 antigen displays persistent surface expression. *Cancer Immunol Immunother.* 2008;57:1017–1027.
22. Wadas TJ, Wong EH, Weisman GR, Anderson CJ. Coordinating radiometals of copper, gallium, indium, yttrium, and zirconium for PET and SPECT imaging of disease. *Chem Rev.* 2010;110:2858–2902.
23. Heneweer C, Holland JP, Divilov V, Carlin S, Lewis JS. Magnitude of enhanced permeability and retention effect in tumors with different phenotypes: Zr-89-albumin as a model system. *J Nucl Med.* 2011;52:625–633.
24. Stabin MG, Sarks RB, Crowe E. OLINDA/EXM: the second-generation personal computer software for internal dose assessment in nuclear medicine. *J Nucl Med.* 2005;46:1023–1027.
25. Welt S, Scott AM, Divgi CR, et al. Phase I/II study of iodine 125-labeled monoclonal antibody A33 in patients with advanced colon cancer. *J Clin Oncol.* 1996;14:1787–1797.



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