

PET Imaging of Tumor Hypoxia Using ^{18}F -Fluoroazomycin Arabinoside in Stage III–IV Non–Small Cell Lung Cancer Patients

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Tumor hypoxia hampers the efficacy of radiotherapy because of its increased resistance to ionizing radiation. The aim of the present study was to estimate the potential added clinical value of the specific hypoxia tracer ^{18}F -fluoroazomycin arabinoside (^{18}F -FAZA) over commonly used ^{18}F -FDG in the treatment of advanced-stage non–small cell lung cancer (NSCLC). **Methods:** Eleven patients with stage III or stage IV NSCLC underwent ^{18}F -FDG and ^{18}F -FAZA PET before chemoradiotherapy. The maximum standardized uptake value (SUV_{max}) was used to depict ^{18}F -FDG uptake, and the tumor-to-background (T/B) ratio and tumor fractional hypoxic volume (FHV) were used to quantify hypoxia. The spatial correlation between ^{18}F -FDG and ^{18}F -FAZA uptake values was investigated using voxel-based analysis. Partial-volume correction was applied. **Results:** All 11 patients showed clear uptake of ^{18}F -FAZA in the primary tumor. However, different patterns of ^{18}F -FDG and ^{18}F -FAZA uptake distributions were observed and varied widely among different tumors. No significant correlation was observed between ^{18}F -FDG SUV_{max} and ^{18}F -FAZA T/B ratio ($P = 0.055$). The median FHV of 1.4 was 48.4% (range, 5.0–91.5). A significant positive correlation was found between the ^{18}F -FAZA T/B ratio and FHV of 1.4 ($P < 0.001$). There was no correlation between the lesion size and FHV or between the ^{18}F -FDG SUV_{max} and FHV. The pattern of tumoral ^{18}F -FDG uptake was rather homogeneous, whereas ^{18}F -FAZA uptake was more heterogeneous, suggesting that ^{18}F -FAZA identifies hypoxic areas within metabolically active areas of tumor. A significant correlation between ^{18}F -FDG SUV_{max} and lesion size ($P = 0.002$) was observed. **Conclusion:** ^{18}F -FAZA PET imaging is able to detect heterogeneous distributions of hypoxic subvolumes out of homogeneous ^{18}F -FDG background in a clinical setting. Therefore, ^{18}F -FAZA might be considered a tool for guiding dose escalation to the hypoxic fraction of the tumor.

Key Words: oncology; PET/CT; respiratory; ^{18}F -FAZA; ^{18}F -FDG; PET; tumor hypoxia

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Lung cancer is the leading cause of worldwide cancer mortality, and non–small cell lung cancer (NSCLC) accounts for 85% of all cases. At presentation, approximately 30% of patients with NSCLC have locally advanced stage III disease (1). The median overall survival for (pathologically) stage IIIA and IIIB disease is 22 and 13 mo, respectively, with corresponding 5-y overall survival rates of 24% and 9%, respectively (2). For patients with stage IV clinical disease, median overall survival is only 6 mo, and the estimated 5-y overall survival a poor 2% (2).

The treatment of stage III and IV patients currently consists of either chemotherapy or combined chemoradiotherapy. Unfortunately, the efficacy of these therapies is limited because of chemoradioresistance, which can be attributed to a large extent to the presence of hypoxia (3). The hallmark of tumor hypoxia is upregulation of hypoxia-inducible factor, leading to an increased expression of hypoxia-responsive elements such as the vascular endothelial growth factor, which induces angiogenesis for growth and differentiation of cancer cells; glycolytic enzymes, which stimulate anaerobic metabolism to meet demands at the cost of an acidic tumor environment and that are prone to metastases; erythropoietin (EPO) release-stimulating erythropoiesis, proliferation, and survival; and BNIP3 (Bcl-2 and 19-kDa interacting protein-3), which leads to genomic instability by evading apoptosis. All these factors contribute to multifactorial treatment resistance (4–7).

The gold standard to measure tumor hypoxia is Eppendorf polarographic electrodes. However, the oxygen electrode method has only been applied in a research setting, because the methodology is technically demanding and invasive and therefore not attractive in a clinical setting. Moreover, the method can only be applied in well-accessible superficial tumors. These limitations have made researchers concentrate on noninvasive techniques such as PET using hypoxia tracers.

Among the hypoxia PET tracers available, nitroimidazole derivatives are most frequently used. These compounds undergo reductive metabolism under hypoxic cell conditions, resulting in reactive tumor metabolite markers that selectively bind to macromolecular cell components and thus are trapped in the cell but wash out from normoxic cells (8). Among nitroimidazole derivatives, ^{18}F -fluoromisonidazole was the first-generation

TABLE 1
Patient Characteristics and Imaging Data

Patient no.	Sex	Age (y)	Weight (kg)	World Health Organization performance status	Stage	Primary tumor		Lesion size (mm)	¹⁸ F-FDG SUV _{max}	¹⁸ F-FAZA T/B ratio	FHV	
						Histology	Site				≥1.2	≥1.4
1	M	44	77	0	IV	Adenocarcinoma	RUL	22	8.6	1.9	66.4	36
2	F	56	73	1	IV	Adenocarcinoma	LUL	63	29	4.6	95.9	89.8
3	F	49	75	0	IV	Adenocarcinoma	RLL	15	4.9	1.7	34.0	20.0
4	F	50	53	2	III	SCC	LUL	58	25	3.5	95.7	90.4
5	F	52	79	1	IV	Adenocarcinoma	LLL	21	4.9	2.7	64.7	48.4
6	M	61	68	1	III	SCC	LUL	26	7.9	1.7	69.3	54.3
7	F	62	67	2	IV	SCC	Clavicle*	13	4.4	2.6	85.0	30.0
8	M	71	70	0	III	SCC	RUL	28	6.1	2.9	81.8	65.0
9	F	46	53	1	IV	Adenocarcinoma	RUL	30	6.7	1.8	17.8	5.0
10	M	72	89	0	III	SCC	RUL	61	25	2.8	58.2	40.6
11	M	72	62	1	IV	Adenocarcinoma	RUL	24	14.7	2.9	97.4	91.5

*Patient had lobectomy and presented with metastasis in clavicle; ¹⁸F-FDG SUV_{max} and ¹⁸F-FAZA T/B values represent uptake of clavicular metastasis. RUL = right upper lobe; LUL = left upper lobe; SCC = squamous cell carcinoma.

compound that has been studied extensively in humans and animals (9,10). More recently, ¹⁸F-fluoroazomycin arabinoside (¹⁸F-FAZA) has been developed as a hypoxia tracer, because it exhibits more favorable tumor-to-background (T/B) ratios in most anatomic regions than ¹⁸F-fluoromisonidazole (11). In addition, ¹⁸F-FAZA exhibits in vivo stability against enzymatic activity, rendering it a promising tracer for tumor hypoxia in clinical studies (12).

It has been suggested that ¹⁸F-FDG uptake in tumors also reflects hypoxia, because the uptake depends on the presence of glycolytic enzymes and glucose transporters (GLUT-1) (13). The expression of GLUT-1 is stimulated by hypoxia-inducible factor 1α activity, which is upregulated in a hypoxic environment (14). Hence, it has been assumed that ¹⁸F-FDG uptake of tumors is affected by tumor hypoxia (15). Previous studies have reported significant discrepancies between ¹⁸F-fluoromisonidazole and ¹⁸F-FDG uptake, indicating that tumor hypoxia and glucose metabolism do not always correlate (16,17). However, to our knowledge, the relationship between tumor hypoxia determined by ¹⁸F-FAZA PET and glucose metabolism determined by ¹⁸F-FDG PET has not been established yet in a clinical setting.

The aim of the present study was, therefore, to estimate the potential added clinical value of the specific hypoxia tracer ¹⁸F-FAZA over the commonly used ¹⁸F-FDG in the treatment of advanced-stage NSCLC. Voxel-by-voxel analysis was performed to determine the spatial correlation between ¹⁸F-FDG and ¹⁸F-FAZA uptake assessed in the same patients at close time points.

MATERIALS AND METHODS

Patients

Patients with stage III or IV NSCLC treated with primary chemotherapy or concurrent chemoradiotherapy according to our institutional protocol were selected for inclusion. The patient work-up minimally included bronchoscopy with biopsy, contrast-enhanced staging CT and ¹⁸F-FDG PET/CT of the thorax and abdomen, and lung-function testing including a flow-volume curve at baseline. Eleven patients (5 men, 6 women) with a median age of 57 y (age range, 44–72 y) were included into the study. The patient characteristics and imaging data are shown in Table 1. The study was approved by the Institutional Ethics Review Board, and written informed consent was obtained from all patients.

¹⁸F-FDG PET/CT

¹⁸F-FDG PET scans were obtained at the Department of Nuclear Medicine and Molecular imaging of the University Medical Center Groningen on a Siemens mCT machine according to guidelines of the European Association of Nuclear Medicine (18). Blood samples were taken before tracer injection to confirm an acceptable blood sugar level (<11 mmol/L) after an overnight fast minimum of 5–6 h. Patients were injected with 3 MBq/kg of bodyweight intravenously. After a waiting period of 60 min, a scan was obtained from the mid thigh to the brain. The maximum standardized uptake value (SUV_{max}) was obtained by delineating the volume of interest (VOI) comprising the entire tumor volume using the Imalytics system (Philips), a computer-based workstation for visualization, quantification, and analysis of PET images. The data were reconstructed with time-of-flight, high-definition, ordered-subsets expectation maximization using 3 iterations, 21 subsets, and a 8-mm gaussian postprocessing filter (NEDPASS Protocol) and had a spatial resolution of 2.04 × 2.04 × 2 mm³.

Production of ^{18}F -FAZA

Procedures for good-manufacturing-practice production of the hypoxia tracer ^{18}F -FAZA have been developed previously. The synthesis of ^{18}F -FAZA was optimized using a Micro Fluid Chemistry Module (Advion). The routine production was performed using a robot system (Zymark). Briefly, the precursor (2 nitro imidazole) for labeling ^{18}F -FAZA was reacted with dried ^{18}F /K222 complex in dimethyl sulfoxide and thereafter deprotected with 0.1 M NaOH. After high-performance liquid chromatography purification of the reaction mixture, ^{18}F -FAZA was formulated using an Oasis HLB plus cartridge. The final sterile solution was analyzed using high-performance liquid chromatography and released for administration to the patient.

^{18}F -FAZA PET/CT

^{18}F -FAZA PET scans were acquired on the same mCT machine as the ^{18}F -FDG images according to local standard operating procedures for ^{18}F -FAZA PET scans. Patients were injected with 370 MBq intravenously. After a waiting period of 120 min, a scan was obtained from the mid thigh to the brain and analyzed using the above-mentioned research workstation. ^{18}F -FAZA SUV_{max} was estimated in the same way as ^{18}F -FDG SUV_{max} , including correction for the partial-volume effect. The median time interval between ^{18}F -FDG PET and ^{18}F -FAZA PET was 7 d. The data were reconstructed with time-of-flight, high-definition, ordered-subsets expectation maximization using 3 iterations, 21 subsets, and a 5-mm gaussian postprocessing filter and had a spatial resolution of $2.04 \times 2.04 \times 2 \text{ mm}^3$.

Calculation of Voxel-Based Spatial Correlation Between ^{18}F -FDG and ^{18}F -FAZA PET/CT

^{18}F -FDG whole-body PET and ^{18}F -FAZA thorax PET were registered in a 2-step procedure using the (low-dose) CT part of both PET/CT examinations. In a first step, the whole-body CT of the ^{18}F -FDG PET/CT was registered to the CT of the thorax ^{18}F -FAZA PET/CT using rigid registration. In a second step, CT-CT registration was locally adjusted for motion with a constrained, intensity-based, free-form deformable image registration (DIR) algorithm using an Advanced Work station MIM Vista (version 5.6; MIM Vista Corp.). The quality of the deformation results was assessed visually based on the CT data. The algorithm used for this study has been previously validated (19,20). Next, the rigid transformation and nonrigid deformation field obtained from aligning the CT data were used to align the ^{18}F -FDG and ^{18}F -FAZA PET data. A 3-dimensional VOI was defined on the ^{18}F -FDG data comprising the metabolically most active part of the tumor by thresholding the lesion using 34% of maximum ^{18}F -FDG uptake (21). This VOI was then projected onto the aligned ^{18}F -FAZA PET data, yielding a spatial correspondence between ^{18}F -FAZA and ^{18}F -FDG uptake for all tumor voxels (Fig. 1). In this way, a 2-dimensional histogram was generated representing the joint distribution of corresponding ^{18}F -FDG and ^{18}F -FAZA standardized uptake value (SUV) for the tumor lesion.

Calculation of Fractional Hypoxic Volume (FHV)

The FHV was defined as the volume within the tumor exhibiting a T/B ratio ≥ 1.2 or ≥ 1.4 on the ^{18}F -FAZA scans (10,22). The FHV of the tumor was determined along the following steps. First, the VOI representing the gross tumor volume on CT was created, and the SUVs within the VOI were expressed per voxel. Next, a tumor-free area in the mediastinum of at least 30-mm diameter was chosen as a reference background. The mean SUV of this background area was calculated. Finally, the ^{18}F -FAZA T/B ratio was assessed by calculating the ratio between SUV_{max} within the tumor and mean SUV background.

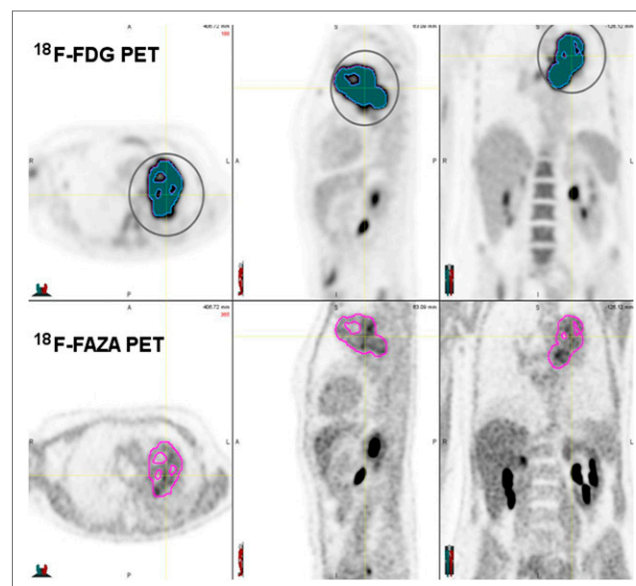


FIGURE 1. Thirty-four percent of ^{18}F -FDG SUV_{max} VOI defined on ^{18}F -FDG PET image and then projected onto aligned ^{18}F -FAZA PET image.

Statistical Analysis

The data were analyzed with SPSS software (version 18.0; SPSS Inc.) for Windows (Microsoft). The ^{18}F -FDG PET SUV_{max} , ^{18}F -FAZA PET T/B ratio, FHV ≥ 1.2 , FHV ≥ 1.4 , and lesion size were considered as variables and reported as median values. Correlations between variables were calculated with the Spearman correlation coefficient. Because this analysis involved multiple comparisons, Bonferroni adjustment was applied, and the significance level was set at $\alpha = 0.05/9 = 0.0055$.

RESULTS

Relationship Between ^{18}F -FDG SUV_{max} and ^{18}F -FAZA T/B Ratio in Tumor

All 11 patients showed clear uptake of ^{18}F -FAZA in the primary tumor, although the accompanying CT scan was indispensable to establish the uptake of ^{18}F -FAZA more accurately with respect to anatomic reference. The ^{18}F -FDG uptake was also clearly visible in all tumors studied with reduced or absent uptake in necrotic areas of the tumor. The median ^{18}F -FAZA T/B ratio was 2.8 (range, 1.8–4.6), and the median ^{18}F -FDG SUV_{max} of the tumors was 7.9 (range, 4.4–29.0). The correlation between tumor hypoxia and glucose metabolism was not significant ($r = 0.59$; $P = 0.055$). Visual analysis showed that the pattern of tumoral ^{18}F -FDG uptake was rather homogeneous, whereas ^{18}F -FAZA uptake was more heterogeneous (Fig. 2).

Spatial Correlation Between ^{18}F -FDG SUV_{max} and FAZA T/B Ratio in Tumor

A detailed voxel-by-voxel analysis revealed different patterns of ^{18}F -FDG and ^{18}F -FAZA uptake and varied widely among different tumors. In Figure 3, patients 1, 7, 8, 9, and 11 showed a moderate to good spatial correlation between ^{18}F -FDG and ^{18}F -FAZA uptake. A good correlation between the 2 tracers suggests well-demarcated hypoxic areas colocalized in metabolically active tumor volume. In contrast, the scatter pattern of ^{18}F -FDG and ^{18}F -FAZA voxels in the other patients was irregular, and no clear correlation was observed between the uptake of either tracer in identical voxels. These findings support the notion that

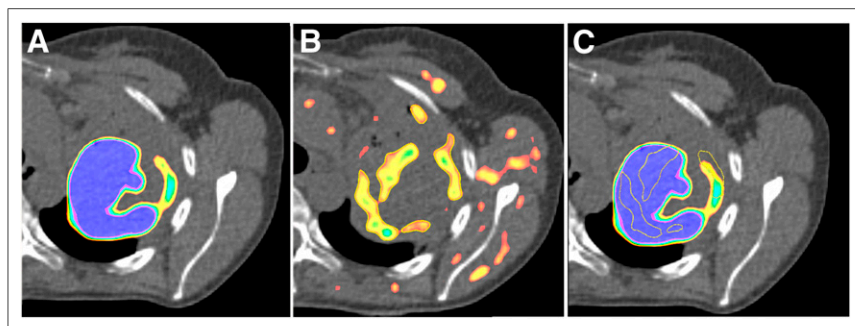


FIGURE 2. Representative transaxial ^{18}F -FDG and ^{18}F -FAZA PET/CT images of patient 4. (A) Thirty-four percent of maximum ^{18}F -FDG accumulation is shown on corresponding CT image. (B) ^{18}F -FAZA accumulation is shown on corresponding CT image. (C) Transposition of areas with ^{18}F -FAZA accumulation onto areas with ^{18}F -FDG accumulation.

^{18}F -FDG and ^{18}F -FAZA signals indeed reflect different biologic functions if measured in the same tumor. Consequently, a poor correlation between these 2 tracers suggests a highly heterogeneous tumor microenvironment.

Relationship Between ^{18}F -FDG SUV_{max} or ^{18}F -FAZA T/B Ratio and Lesion Size

The median lesion size was 26 mm (range, 13–63 mm). Lesion size significantly correlated with the ^{18}F -FDG SUV_{max} ($r = 0.82$; $P = 0.002$), but the correlation between lesion size and ^{18}F -FAZA T/B ratio was not significant ($r = 0.55$; $P = 0.07$). This suggests that larger tumors are not necessarily more hypoxic than smaller tumors but do exhibit higher metabolic activity.

Relationship Between FHV of Tumor and ^{18}F -FAZA T/B Ratio, ^{18}F -FDG SUV_{max} , or Lesion Size

The median FHV was 69% (range, 17.0%–97.0%) and 48% (range, 5.0%–91.0%) when a cutoff of ≥ 1.2 and ≥ 1.4 , respectively, was applied. We observed a strong and significant positive correlation between the ^{18}F -FAZA T/B ratio and FHV cutoff of 1.4 ($r = 0.88$; $P < 0.001$); however, no significant correlation was found between the ^{18}F -FAZA T/B ratio and FHV cutoff of 1.2 ($r = 0.74$; $P = 0.009$), suggesting that an increase of the ^{18}F -FAZA T/B ratio is due to heterogeneously distributed high-uptake regions rather than to

homogeneous uptake. Also, no correlation was found between lesion size and an FHV cutoff of ≥ 1.2 ($r = 0.15$; $P = 0.65$) or ≥ 1.4 ($r = 0.40$; $P = 0.21$) or between an ^{18}F -FDG SUV_{max} and an FHV cutoff of ≥ 1.2 ($r = 0.38$; $P = 0.24$) or ≥ 1.4 ($r = 0.56$; $P = 0.07$).

DISCUSSION

The results of our study show that the pattern of tumoral uptake of ^{18}F -FAZA varies among tumors and is more heterogeneous than ^{18}F -FDG, signifying the importance of spatial distribution of tumor hypoxia or FHV. This distribution may arise as the result of transitory functional changes in a tumor microenvironment such as variation in tumor physiology and adaptation of blood supply

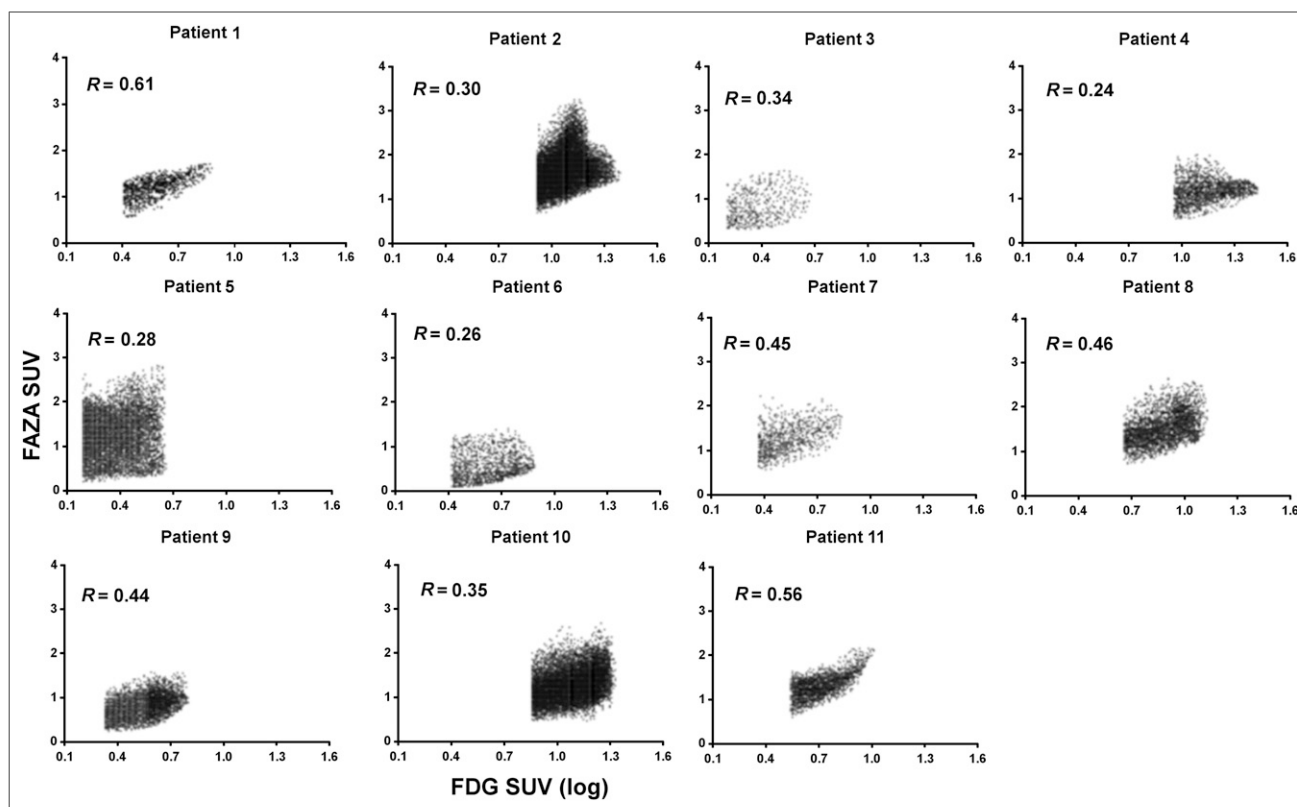


FIGURE 3. ^{18}F -FDG and ^{18}F -FAZA scatterplots for all patients.

in response to tumor growth. Koh et al. (23) and Rasey et al. (10) were first to publish data about the use of bioreductive molecules to visualize hypoxic subvolumes. On the basis of experimental and clinical data (10,23), they proposed the following definition for hypoxic regions: voxels exhibiting an ^{18}F -fluoromisonidazole T/B ratio ≥ 1.4 . Alternatively, Busk et al. proposed a ^{18}F -FAZA T/B ratio of ≥ 1.2 to distinguish hypoxia (22). In our study, we decided to test both of these T/B ratio cutoff levels to determine the FHV and its relationship to lesion size, ^{18}F -FDG SUV_{max} , and ^{18}F -FAZA SUV_{max} . We observed no correlation between the FHV and tumor size or ^{18}F -FDG SUV_{max} . Our data are similar to those reported by Rasey et al. (10) and Koh et al. (23), who also did not find a correlation between FHV measured by ^{18}F -fluoromisonidazole and lesion size in NSCLC. Similarly, Rajendran et al. (24) and Stadler et al. (25) did not observe a correlation between FHV and tumor size in head and neck cancer patients. The fact that the hypoxic fraction of the tumor holds no relation to tumor size and ^{18}F -FDG SUV_{max} is in agreement with the visual appearance of ^{18}F -FDG and ^{18}F -FAZA images of single tumors, where greater heterogeneity is seen on ^{18}F -FAZA scans than on ^{18}F -FDG scans (26,27).

It has been suggested that tumor ^{18}F -FDG uptake is dependent on hypoxic environment (15,27,28). Hence, ^{18}F -FDG uptake might serve as surrogate marker for hypoxia, rendering ^{18}F -FAZA scanning redundant. The degree of ^{18}F -FDG accumulation primarily depends on GLUT-1 expression. Upregulation of GLUTs, due to hyperglycolysis driven by hypoxia-inducible factor 1α , is observed in hypoxic regions (14). In our study, a detailed voxel-by-voxel analysis of ^{18}F -FDG and ^{18}F -FAZA scatterplots manifested large inter- and intratumor variability distribution patterns. In short, all possible combinations of hypoxia and glucose metabolism parameters were found: active metabolic areas, which are severely hypoxic; inactive metabolic areas, which are also hypoxic; metabolically active tumor areas, which are nonhypoxic; and finally, tumor areas that were metabolically inactive and nonhypoxic. As a result of our observations, it appears that hypoxia is heterogeneously distributed within a tumor and can occur in both metabolically active and inactive zones. Similar results were reported by Thorwarth et al. (29), underlying the need for a specific PET hypoxia tracer. The question arises as to whether these different patterns of uptake reflect different biologic mechanisms and, in addition, different clinical behavior and outcome. High ^{18}F -FDG and ^{18}F -FAZA uptake levels may correspond to active metabolic cells with high expression of GLUT, characterizing diffusion-limited hypoxia, whereas low ^{18}F -FDG and ^{18}F -FAZA uptake may characterize perfusion-limited hypoxia. High ^{18}F -FDG uptake and no trapping of ^{18}F -FAZA may indicate well-perfused nonhypoxic areas. Low ^{18}F -FDG uptake and ^{18}F -FAZA trapping might signal tumor cells that are deprived of oxygen and glucose due to perfusion insufficiency (e.g., remote blood vessel).

Recent studies have shown that hypoxia imaging during treatment could be a promising way to stratify patients who may benefit from hypoxia modification or dose escalation strategies (30,31). Future studies should evaluate the degree of spatial heterogeneity of hypoxia, retention of ^{18}F -FAZA uptake using blood flow measurements, and spatial correlation between ^{18}F -FDG and ^{18}F -FAZA uptake within a single tumor during curative radiotherapy or chemoradiotherapy. In patients with a diffuse distribution of hypoxia, this pattern indicates a higher degree of spatial heterogeneity with poor prognosis; these patients might benefit from a more systemic approach, such as chemotherapy with an addition of a hypoxia-targeting drug (e.g., tirapazamine)

(32). On the other hand, a well-demarcated hypoxic area with lesser degree of spatial heterogeneity can be treated with focal radiation dose escalation (33). In the past few years, highly sophisticated radiotherapy technologies have been introduced in the field of radiation oncology, such as intensity-modulated radiotherapy, which allows for dose painting—that is, the delivery of a higher dose to specific tumor areas and subvolumes. This approach is only effective in the case of static hypoxia because the boost dose is delivered to the same region every day. In the case of dynamic hypoxia, a single high-dose fraction using stereotactic body radiotherapy delivered to the hypoxic region on the same day of the ^{18}F -FAZA PET/CT may be more appropriate.

There are some limitations in this study. First, our study lacks an innovative scanning technique such as dynamic PET scanning capable of quantifying differences between tumor oxygen perfusion and hypoxia tracer permeability within the same tumor to understand tumor heterogeneity. Future studies should focus on the spatiotemporal distribution of tumor hypoxia over time to select the most optimal treatment technique. Second, although we tried to keep the time interval between ^{18}F -FDG and ^{18}F -FAZA PET/CT scans as short as possible, this was not always feasible because of patient logistics and the fact that 2 tracers were used. It should be stressed that in our study patients did not receive any treatment between the 2 scans. Although we do not expect significant changes in tumor oxygenation levels, it cannot be ruled out that this may have occurred.

Another technical issue with lung cancer treatment is tumor movement during respiration. The same problem pertains to the voxel-based analysis (Fig. 3). Hence, the accurate delineation of tumor volume, taking into account respiratory motion, is essential for radiotherapy planning. To overcome this problem, optimal gating techniques or 4-dimensional PET would be an attractive option (34). However, further research needs to be done to determine the practical feasibility. Lung lesions that were included in this study, compared with the locations close to the diaphragm, were situated mostly in the upper lobes of the lungs, where respiratory movement is less pronounced (35).

DIR is a field of extreme complexity, and it is hard to evaluate its results. In this study, we expected that the influence of possible DIR inaccuracies on the PET registration was limited, because the registered volumes did not show dramatic deformations. The latter was explained by little changes in tumor size and by the short time interval between the scans (1 wk). Furthermore, no treatment had taken place in the time interval, so no dramatic deformations were expected. We considered visual inspection of the results of the DIR adequate for the application of DIR in this study.

CONCLUSION

^{18}F -FAZA PET imaging is able to detect heterogeneous distributions of hypoxic subvolumes even within homogeneous ^{18}F -FDG background. Therefore, ^{18}F -FAZA when added to ^{18}F -FDG provides information on tumor hypoxia and might be developed into a tool for guiding individualization of treatment of advanced NSCLC.

DISCLOSURE

The costs of publication of this article were defrayed in part by the payment of page charges. Therefore, and solely to indicate this fact, this article is hereby marked “advertisement” in accordance with 18 USC section 1734. This project was performed within the

framework of CTMM, the Center for Translational Molecular Medicine (www.ctmm.nl), project AIRFORCE no. 030-103. No other potential conflict of interest relevant to this article was reported.

REFERENCES

- Mountain CF. Revisions in the international system for staging lung cancer. *Chest*. 1997;111:1710–1717.
- Goldstraw P, Crowley J, Chansky K, et al. The IASLC lung cancer staging project: proposals for the revision of the TNM stage groupings in the forthcoming (seventh) ed. of the TNM classification of malignant tumours. *J Thorac Oncol*. 2007;2:706–714.
- Wilson WR, Hay MP. Targeting hypoxia in cancer therapy. *Nat Rev Cancer*. 2011;11:393–410.
- Ivan M, Kondo K, Yang H, et al. HIF α targeted for VHL-mediated destruction by proline hydroxylation: implications for O₂ sensing. *Science*. 2001;292:464–468.
- Jaakkola P, Mole DR, Tian YM, et al. Targeting of HIF- α to the von Hippel-Lindau ubiquitylation complex by O₂-regulated prolyl hydroxylation. *Science*. 2001;292:468–472.
- Ikeda Y, Taveira-DaSilva AM, Pacheco-Rodriguez G, et al. Erythropoietin-driven proliferation of cells with mutations in the tumor suppressor gene TSC2. *Am J Physiol Lung Cell Mol Physiol*. 2011;300:L64–L72.
- Walenta S, Salameh A, Lyng H, et al. Correlation of high lactate levels in head and neck tumors with incidence of metastasis. *Am J Pathol*. 1997;150:409–415.
- Foo SS, Abbott DF, Lawrentschuk N, Scott AM. Functional imaging of intratumoral hypoxia. *Mol Imaging Biol*. 2004;6:291–305.
- Piert M, Machulla HJ, Becker G, Aldinger P, Winter E, Bares R. Dependency of the [¹⁸F]fluoromisonidazole uptake on oxygen delivery and tissue oxygenation in the porcine liver. *Nucl Med Biol*. 2000;27:693–700.
- Rasey JS, Koh WJ, Evans ML, et al. Quantifying regional hypoxia in human tumors with positron emission tomography of [¹⁸F]fluoromisonidazole: a pretherapy study of 37 patients. *Int J Radiat Oncol Biol Phys*. 1996;36:417–428.
- Sorger D, Patt M, Kumar P, et al. [¹⁸F]fluoroazomycinabinofuranoside (¹⁸FAZA) and [¹⁸F]fluoromisonidazole (¹⁸FMISO): a comparative study of their selective uptake in hypoxic cells and PET imaging in experimental rat tumors. *Nucl Med Biol*. 2003;30:317–326.
- Souvatoglou M, Grosu AL, Roper B, et al. Tumour hypoxia imaging with [¹⁸F]FAZA PET in head and neck cancer patients: a pilot study. *Eur J Nucl Med Mol Imaging*. 2007;34:1566–1575.
- Higashi K, Ueda Y, Sakurai A, et al. Correlation of GLUT-1 glucose transporter expression with. *Eur J Nucl Med*. 2000;27:1778–1785.
- Semenza GL. Hypoxia, clonal selection, and the role of HIF-1 in tumor progression. *Crit Rev Biochem Mol Biol*. 2000;35:71–103.
- Burgman P, Odonoghue JA, Humm JL, Ling CC. Hypoxia-induced increase in FDG uptake in MCF7 cells. *J Nucl Med*. 2001;42:170–175.
- Cherk MH, Foo SS, Poon AM, et al. Lack of correlation of hypoxic cell fraction and angiogenesis with glucose metabolic rate in non-small cell lung cancer assessed by ¹⁸F-fluoromisonidazole and ¹⁸F-FDG PET. *J Nucl Med*. 2006;47:1921–1926.
- Gagel B, Reinartz P, Demirel C, et al. [¹⁸F] fluoromisonidazole and [¹⁸F] fluorodeoxyglucose positron emission tomography in response evaluation after chemo-radiotherapy of non-small-cell lung cancer: a feasibility study. *BMC Cancer*. 2006;6:51.
- Boellaard R, O'Doherty MJ, Weber WA, et al. FDG PET and PET/CT: EANM procedure guidelines for tumour PET imaging: version 1.0. *Eur J Nucl Med Mol Imaging*. 2010;37:181–200.
- Piper J. SU-FF-I-68: Evaluation of an intensity-based free-form deformable = registration algorithm. *Med Phys*. 2007;34:2353–2354.
- La Macchia M, Fellin F, Amichetti M, et al. Systematic evaluation of three different commercial software solutions for automatic segmentation for adaptive therapy in head-and-neck, prostate and pleural cancer. *Radiat Oncol*. 2012;7:160-717X-7-160.
- Daisne JF, Duprez T, Weynand B, et al. Tumor volume in pharyngolaryngeal squamous cell carcinoma: comparison at CT, MR imaging, and FDG PET and validation with surgical specimen. *Radiology*. 2004;233:93–100.
- Busk M, Horsman MR, Overgaard J. Resolution in PET hypoxia imaging: voxel size matters. *Acta Oncol*. 2008;47:1201–1210.
- Koh WJ, Bergman KS, Rasey JS, et al. Evaluation of oxygenation status during fractionated radiotherapy in human nonsmall cell lung cancers using [¹⁸F]fluoromisonidazole positron emission tomography. *Int J Radiat Oncol Biol Phys*. 1995;33:391–398.
- Rajendran JG, Schwartz DL, O'Sullivan J, et al. Tumor hypoxia imaging with [¹⁸F] fluoromisonidazole positron emission tomography in head and neck cancer. *Clin Cancer Res*. 2006;12:5435–5441.
- Stadler P, Becker A, Feldmann HJ, et al. Influence of the hypoxic subvolume on the survival of patients with head and neck cancer. *Int J Radiat Oncol Biol Phys*. 1999;44:749–754.
- Nordmark M, Bentzen SM, Overgaard J. Measurement of human tumour oxygenation status by a polarographic needle electrode: an analysis of inter- and intratumour heterogeneity. *Acta Oncol*. 1994;33:383–389.
- Pugachev A, Ruan S, Carlin S, et al. Dependence of FDG uptake on tumor microenvironment. *Int J Radiat Oncol Biol Phys*. 2005;62:545–553.
- Kaira K, Endo M, Abe M, et al. Biologic correlation of 2-[¹⁸F]-fluoro-2-deoxy-D-glucose uptake on positron emission tomography in thymic epithelial tumors. *J Clin Oncol*. 2010;28:3746–3753.
- Thorwarth D, Eschmann SM, Holzner F, Paulsen F, Alber M. Combined uptake of [¹⁸F]FDG and [¹⁸F]FMISO correlates with radiation therapy outcome in head-and-neck cancer patients. *Radiother Oncol*. 2006;80:151–156.
- Zips D, Zophel K, Abolmaali N, et al. Exploratory prospective trial of hypoxia-specific PET imaging during radiochemotherapy in patients with locally advanced head-and-neck cancer. *Radiother Oncol*. 2012;105:21–28.
- Mortensen LS, Johansen J, Kallehauge J, et al. FAZA PET/CT hypoxia imaging in patients with squamous cell carcinoma of the head and neck treated with radiotherapy: results from the DAHANCA 24 trial. *Radiother Oncol*. 2012;105:14–20.
- von Pawel J, von Roemeling R, Gatzemeier U, et al. Tirapazamine plus cisplatin versus cisplatin in advanced non-small-cell lung cancer: a report of the international CATAPULT I study group—cisplatin and tirapazamine in subjects with advanced previously untreated non-small-cell lung tumors. *J Clin Oncol*. 2000;18:1351–1359.
- Kong FM, Ten Haken RK, Schipper MJ, et al. High-dose radiation improved local tumor control and overall survival in patients with inoperable/unresectable non-small-cell lung cancer: long-term results of a radiation dose escalation study. *Int J Radiat Oncol Biol Phys*. 2005;63:324–333.
- van Elmpt W, Hamill J, Jones J, De Ruyscher D, Lambin P, Ollers M. Optimal gating compared to 3D and 4D PET reconstruction for characterization of lung tumours. *Eur J Nucl Med Mol Imaging*. 2011;38:843–855.
- Cohade C, Osman M, Marshall LN, Wahl RN. PET-CT: Accuracy of PET and CT spatial registration of lung lesions. *Eur J Nucl Med Mol Imaging*. 2003;30:721–726.



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