

Imaging CXCR4 Expression in Human Cancer Xenografts: Evaluation of Monocyclam ^{64}Cu -AMD3465

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The chemokine receptor 4 (CXCR4) is overexpressed in several cancers and metastases and as such presents an enticing target for molecular imaging of metastases and metastatic potential of the primary tumor. CXCR4-based imaging agents could also be useful for diagnosis, staging, and therapeutic monitoring. Here we evaluated a positron-emitting monocyclam analog, ^{64}Cu -{N-[1,4,8,11-tetraazacyclotetradecanyl-1,4-phenylenebis(methylene)]-2-(aminomethyl)pyridine} (^{64}Cu -AMD3465), in subcutaneous U87 brain tumors and U87 tumors stably expressing CXCR4 (U87-stb-CXCR4) and in colon tumors (HT-29) using dynamic and whole-body PET supported by ex vivo biodistribution studies. Both dynamic and whole-body PET/CT studies show specific accumulation of radioactivity in U87-stb-CXCR4 tumors, with the percentage injected dose per gram reaching a maximum of 102.70 ± 20.80 at 60 min and tumor-to-muscle ratios reaching a maximum of 362.56 ± 153.51 at 90 min after injection of the radiotracer. Similar specificity was also observed in the HT-29 colon tumor model. Treatment with AMD3465 inhibited uptake of radioactivity by the tumors in both models. Our results show that ^{64}Cu -AMD3465 is capable of detecting lesions in a CXCR4-dependent fashion, with high target selectivity, and may offer a scaffold for the synthesis of clinically translatable agents.

Key Words: PET; tumor microenvironment; chemokine; stem cells; molecular imaging; colon cancer; brain cancer; metastasis

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Compelling evidence demonstrates that chemokine receptor 4 (CXCR4) expression plays an important role in several diseases including HIV, cancer, and lupus (1–3). Although CXCR4 was discovered as a coreceptor for HIV-1 cell entry in the 1990s (4), recently much attention has been paid to investigating its role in cancer progression and metastasis. CXCR4 is overexpressed in more than 20

different human cancers (4,5); is required for tumor development, growth, vascularization, and metastases (5,6); and is considered a therapeutic target (7).

CXCR4 has been shown to be a prognostic factor in several cancers, including colorectal carcinoma (CRC). CXCR4 is differentially expressed in CRC and is required for the outgrowth of colon cancer metastasis (8,9). CXCR4 expression in primary CRC is associated with recurrence, poor survival, and liver metastasis (8). Most recently, in patients treated with the folinic acid, fluorouracil, and oxaliplatin regimen, enhanced CXCR4 expression was reported in resected liver metastases, suggesting that CXCR4 may be contributing to drug resistance and indicating an expanding role of CXCR4 in cancer biology (10).

In addition to the aforementioned biologic roles, several comprehensive studies have demonstrated that CXCR4 is necessary for cancer metastasis (11,12). Homing of cancer cells to the common destinations of cancer metastasis is mediated through the binding of CXCR4 to its cognate ligand, CXCL12, which is secreted in tissues that eventually harbor metastases (6,13). Extensive immunohistochemical studies have revealed that tumors with elevated CXCR4 expression often have an aggressive phenotype and that metastases often have elevated expression, compared with the primary tumor (14–16). Inhibition of CXCR4–CXCL12 signaling by antibodies, peptide analogs, small molecules, or small interfering RNA knockdown has demonstrated reduced metastatic burden in orthotopic and metastatic models of various cancers (11,12,17–22). Taken together, those preclinical and clinical observations demonstrate that CXCR4 plays a critical role in metastasis. In addition, the pleiotropic activity of CXCR4 and CXCL12 is critical for neural, vascular, and hematopoietic organogenesis, underscoring the need to stratify the patient population with high tumor CXCR4 levels. Accordingly, development of imaging agents that can noninvasively and repeatedly detect CXCR4 expression levels may prove useful in managing cancer patients. CXCR4-targeted imaging agents can be used to evaluate primary tumors for elevated CXCR4 expression and therapeutic intervention, to screen for secondary metastatic spread to both local and distant sites, and for therapeutic monitoring.

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To image CXCR4 expression in tumor models, macro-molecular agents such as ^{111}In - and ^{18}F -labeled peptides and ^{125}I -labeled monoclonal antibodies have been investigated using either SPECT/CT or PET (23–25). Others and our own group reported the imaging of CXCR4 expression using a low-molecular-weight agent, ^{64}Cu -AMD3100, in control and tumor-bearing mice (26,27). Our studies with ^{64}Cu -AMD3100 in human cancer xenografts showed clear detection of CXCR4 expression in an orthotopic breast cancer model and in experimental lung metastases. Though promising, the bicyclam AMD3100 has a relatively low affinity ($\sim 651 \pm 37$ nM) (28) and a structurally restricted scaffold. In search of agents that are amenable to structural modification, we pursued the monocyclam analog $\{N$ -[1,4,8,11-tetraazacyclotetradecanyl-1,4-phenylenebis(methylene)]-2-(aminomethyl)pyridine} (AMD3465) to image CXCR4 expression. Compared with AMD3100, AMD3465 has higher affinity, reduced size, and charge (29). The properties of AMD3465 as a CXCR4 inhibitor have been well characterized (30). AMD3465 is reported to have a 50% inhibitory concentration (IC₅₀) of 41.7 ± 1.2 nM and 12.07 ± 2.02 nM for ^{125}I -CXCL12 binding and calcium flux stimulated by stromal-derived factor-1 α (29,30), respectively. Also, AMD3465 was shown to be 10-fold more effective as a CXCR4 antagonist than the bicyclam AMD3100 (31). Because of those favorable qualities, we examined the potential of AMD3465 as an imaging agent for CXCR4 expression. We used the ability of the cyclam to form strong complexes with copper to develop ^{64}Cu -AMD3465 as an imaging agent for PET. Here, we report a detailed evaluation of ^{64}Cu -AMD3465 in human subcutaneous brain tumor xenografts stably expressing CXCR4 by whole-body pseudodynamic imaging, whole-body static imaging, and ex vivo biodistribution studies. We also show the potential of ^{64}Cu -AMD3465 to image graded levels of CXCR4 expression in a colon cancer model that has an intermediate level of CXCR4, compared with that found in the brain tumor model.

MATERIALS AND METHODS

Cell Lines

All cell culture reagents were purchased from Invitrogen unless otherwise specified. The human glioblastoma cell line U87 and colorectal adenocarcinoma cell line HT-29 were purchased from American Type Culture Collection and cultured in our laboratory in minimum essential medium and McCoy 5A medium supplemented with 10% fetal bovine serum, 100 units of penicillin per milliliter, and 100 mg of streptomycin per milliliter, respectively. A U87 cell line stably transfected with human CD4 and CXCR4 (U87-stb-CXCR4) was obtained from the National Institutes of Health AIDS Research Reference Reagent Program (32) and cultured in Dulbecco's modified Eagle's medium supplemented with 15% fetal bovine serum, 1 μg of puromycin per milliliter, 300 μg of G418 per milliliter, 100 units of penicillin per milliliter, and 100 mg of streptomycin per milliliter. Cell lines were maintained in a humidified incubator with 5% CO_2 .

Radiopharmaceutical Preparation

AMD3465 was a kind gift from Genzyme Corp., and ^{64}Cu - CuCl_2 was obtained from the University of Wisconsin. AMD3465 was radiolabeled with ^{64}Cu - CuCl_2 using a standard procedure, as described previously (26). Briefly, 200 μg of AMD3465·6HCl was added to 370–740 MBq (10–20 mCi) of ^{64}Cu - CuCl_2 concentrated in vacuo and adjusted to an approximate pH of 5–5.5 with 0.1 M sodium acetate, and the pH-adjusted reaction mixture was heated at 55–60°C for 45 min. ^{64}Cu -AMD3465 was purified on a reversed-phase high-performance liquid chromatography system (Varian) using a C-18 (Luna, 5 μm , 10 \times 250 mm; Phenomenex) semipreparative column. A mobile phase with 15% MeOH (with 0.1% trifluoroacetic acid) and 85% H_2O (with 0.1% trifluoroacetic acid) at a flow rate of 5 mL/min was used. ^{64}Cu -AMD3465 was collected between 16 and 18 min, dried, and diluted in saline for cell and animal studies. Radioactive signal was detected using a single-channel radiation detector (model 105S; Bioscan) with ultraviolet absorbance monitored at 266 nm.

Determination of Partition Coefficient

The partition coefficient (log P) of ^{64}Cu -AMD3465 complex was determined by adding 148–185 kBq (4–5 μCi) of the complex to a solution containing 1 mL of water and 1 mL of octanol ($n = 3$). The resulting solution was then shaken well for 1 h at room temperature. From each phase, 100- μL aliquots were ($n = 3$) counted separately. The partition coefficient was calculated as a ratio between counts in the octanol phase to counts in the water phase. An average log P value was obtained from those ratios.

Flow Cytometry

Surface CXCR4 expression levels were analyzed using a CXCR4 monoclonal antibody (clone 44716) conjugated to phycoerythrin (R&D Systems) according to procedures described previously (26).

Receptor Binding Assays

U87, U87-stb-CXCR4, and HT-29 cells seeded in 6-well plates at 60%–80% confluence were used for receptor binding assays. Cells were incubated with 37 kBq/mL (1 $\mu\text{Ci/mL}$) of ^{64}Cu -AMD3465 in phosphate-buffered saline binding buffer (containing 5 mM MgCl_2 , 1 mM CaCl_2 , 0.25% bovine serum albumin, pH 7.4) for 30 min at 4°C. After incubation, cells were washed quickly 4 times with 4°C binding buffer, trypsinized using nonenzymatic buffer, and cell-associated activity was determined in a γ -spectrometer (1282 Compugamma CS; Pharmacia/LKB Nuclear, Inc.).

For internalization assays, cells were detached using nonenzymatic buffer, and aliquots of 1 million cells per tube were incubated with 37 kBq (1 μCi) of ^{64}Cu -AMD3465 per milliliter for various times up to 4 h at 4°C or 37°C in the phosphate-buffered saline binding buffer. Assuming minimal receptor endocytosis at 4°C, the internalization assay was performed only with cells incubated at 37°C. At 15-, 30-, 60-, and 240-min intervals, the medium was removed and cells were washed once with binding buffer followed by a mild acidic buffer (50 mM glycine, 150 mM NaCl [pH 3.0]) at 4°C for 5 min. Then the acidic buffer was collected, and cells were washed twice with binding buffer. Pooled washes (containing cell surface-bound ^{64}Cu -AMD3465) and cell pellets (containing internalized ^{64}Cu -AMD3465) were counted in an automated γ -counter along with the standards. All of the radioactivity values were converted into percentage of incubated dose (%ID) per million cells. Experiments were performed in triplicate.

and repeated 3 times. Data were fitted according to linear regression analysis using PRIZM software.

Animal Models

All experimental procedures using animals were conducted according to protocols approved by the Johns Hopkins Animal Care and Use Committee. Female nonobese diabetic (NOD)/severe combined immune deficient (SCID) mice (age, 6–8 wk; weight, 25–30 g) were purchased from the Johns Hopkins Immune Compromised Animal Core. Mice were implanted subcutaneously with U87 and U87-stb-CXCR4 cells (4×10^6 cells/100 μ L) in the upper left and right flanks, respectively. Similarly, HT-29 colon adenocarcinoma cells (4×10^6 cells/100 μ L) were inoculated in the upper right flanks of the mice. Animals were used for biodistribution and PET/CT experiments when the tumor size reached 300–400 mm³.

PET/CT and Analysis

Dynamic and whole-body PET and CT images were acquired on an eXplore VISTA small-animal PET (GE Healthcare) and an X-SPECT small SPECT/CT system (Gamma Medica Ideas), respectively. For imaging studies, mice were induced with 3% and maintained under 1.5% isoflurane (v/v). Pseudodynamic imaging studies were performed on NOD/SCID mice bearing U87 and U87-stb-CXCR4 brain tumors. After intravenous injection of ⁶⁴Cu-AMD3465 (range, 9.0–9.6 MBq; mean, 9.4 MBq), changes in radiotracer accumulation were recorded over the whole body using an imaging sequence consisting of 16 frames for a total of 70 min with variable dwell times (2×60 s, 6×120 s, 4×240 s, and 4×600 s), as described previously (33). After dynamic imaging, whole-body PET images (2 bed positions, 15-min emission per bed position) were acquired at 1.5, 4, 8, and 24 h after injection of radiotracer. For binding specificity studies, a separate group of mice ($n = 2$) was subcutaneously administered with a blocking dose of AMD3465 (25 mg/kg) at 1 h before the injection of ⁶⁴Cu-AMD3465, and another set of 2 mice was injected with ⁶⁴Cu-CuCl₂ alone. In colon cancer xenografts, whole-body images were acquired at 90 min after injection of ⁶⁴Cu-AMD3465 alone or with a blocking dose. After each PET scan, a CT scan was acquired in 512 projections for anatomic coregistration. PET emission data were corrected for decay and dead time and reconstructed using the

3-dimensional ordered-subsets expectation maximization algorithm. Data were analyzed based on regions of interest drawn within the tumors or tissue. The percentage of injected dose per gram (%ID/g) was calculated based on a calibration factor that was determined using a known quantity of radioactivity. Time-activity curves were calculated as an average from the region-of-interest analyses of 4 mice. Data were analyzed using AMIDE software (SourceForge), and volume-rendered images were generated using Amira 5.2.0 software (Visage Imaging Inc.).

Ex Vivo Biodistribution

NOD/SCID mice harboring either brain or colon xenografts were injected intravenously with 740 kBq (20 μ Ci) of ⁶⁴Cu-AMD3465 in 200 μ L of saline. At 30, 60, 90, 240, and 600 min after injection, animals were sacrificed; blood, tumors, and selected tissues were harvested and weighed; and the radioactivity in the tissues was measured in an automated γ -spectrometer. In the case of colon cancer xenografts, biodistribution studies were performed at 90 min after ⁶⁴Cu-AMD3465 injection.

To demonstrate the in vivo specificity of ⁶⁴Cu-AMD3465, a set of mice received a blocking dose of AMD3465 (25 mg/kg) subcutaneously at 1 h before the injection of radiotracer, and another set of mice received ⁶⁴Cu-CuCl₂ alone. At 90 min after injection of radiotracers, ex vivo biodistribution studies were performed. Aliquots of the injected dose were counted as reference standards for the calculation of %ID/g values. A minimum of 4 animals per time point was used.

Immunohistochemistry

HT-29 tumors from biodistribution experiments were used for histologic examination. Sections of tumors were stained with hematoxylin and eosin, and immunohistochemistry was performed on tumor tissues as previously described using rabbit polyclonal antibodies that recognize amino acids 328–338 of human CXCR4 (34), with minimum cross-reactivity to mouse CXCR4 (Imgenex) (26).

Data Analysis

Statistical analysis was performed using PRIZM software. An unpaired 2 tailed *t* test was used, and *P* values less than 0.05 for the comparison between tumors expressing high and tumors expressing low CXCR4 uptake were considered to be significant.

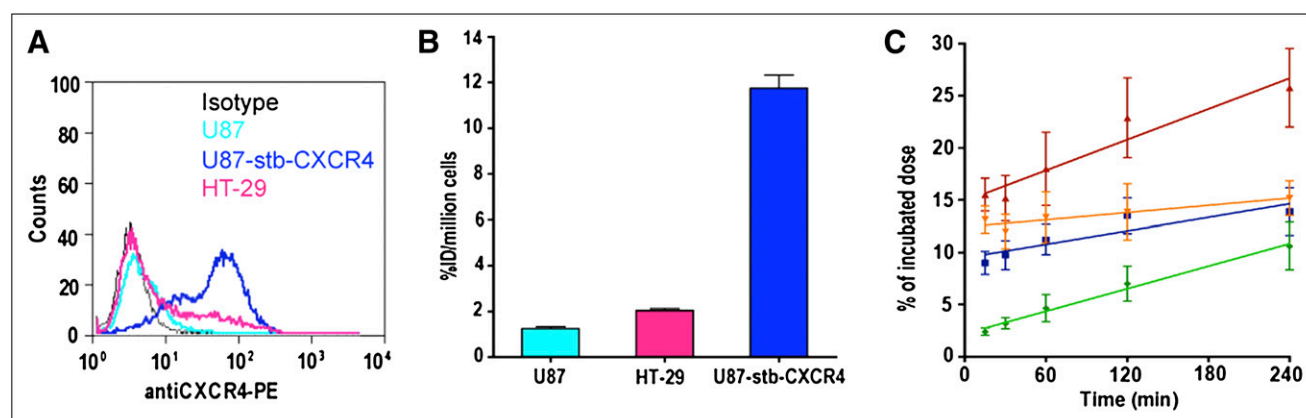


FIGURE 1. In vitro characterization of CXCR4 expression. (A) Evaluation of surface CXCR4 expression in cancer cell lines used by flow cytometry. (B) Uptake analysis of ⁶⁴Cu-AMD3465 in cancer cell lines. (C) Internalization of CXCR4 in U87-stb-CXCR4 cells: 4°C (green), 37°C total (red), 37°C cell surface (orange), and 37°C internalized (blue).

RESULTS

Radiolabeling and Partition Coefficient

Radio-high-performance liquid chromatography assessment showed the radiochemical purity of ^{64}Cu -AMD3465 to be greater than 98% (Supplemental Fig. 1; supplemental materials are available online only at <http://jnm.snmjournals.org>), with specific radioactivity in the range of $6.0 \pm 3.1 \text{ GBq}/\mu\text{mol}$ ($162 \pm 84 \text{ mCi}/\mu\text{mol}$). The log P value for the complex was -2.71 ± 0.37 .

Receptor Expression, In Vitro Radioligand Binding, and Internalization

Flow cytometric analysis using CXCR4 antibody (clone 44716) revealed that 2%, 95%, and 30% of U87, U87-stb-CXCR4, and HT-29 cells, respectively, were positive for CXCR4 (Fig. 1A). The radioligand binding studies, supported by these data, show a gradual increase in the accumulation of radioactivity on the order of U87-stb-CXCR4 > HT-29 > U87 (Fig. 1B). The internalization assays were done retrospectively to understand the retained radioactivity uptake observed in the U87-stb-CXCR4 tumors. Over the 4-h incubation period, a continuous accumulation of radioactivity was observed in cells incubated at 37°C , with slopes of 0.0360 ± 0.0022 and 0.0492 ± 0.0081 for internalized and total fractions, respectively. For cells incubated at 4°C , a lower slope of 0.0215 ± 0.0060 was observed (Fig. 1C).

PET

The time-activity curves acquired in mice harboring U87 and U87-stb-CXCR4 subcutaneous brain tumor xenografts over 70 min showed continuous ^{64}Cu -AMD3465 uptake in U87-stb-CXCR4 tumors (Fig. 2). The %ID/g for U87-stb-CXCR4 tumors reached up to 110 at 65 min (Fig. 2). The %ID/g values for U87-stb-CXCR4 tumors at 65 min were nearly 3-fold greater than those of kidneys and liver and nearly 18-fold greater than for the U87 tumors. The whole-body PET/CT images acquired on U87 and U87-stb-CXCR4 brain xenografts at different times are shown

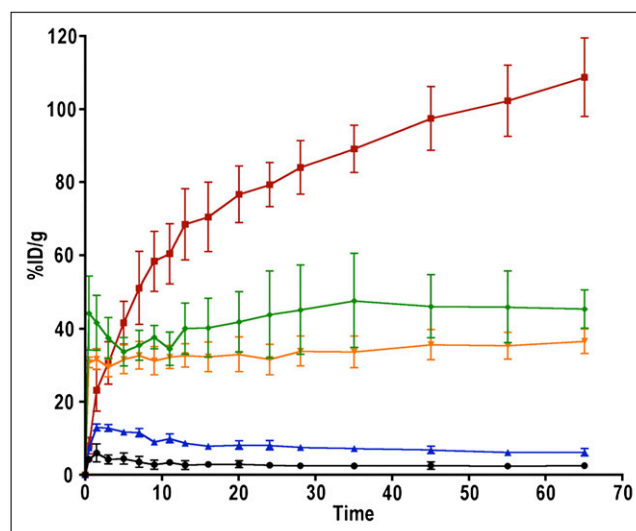


FIGURE 2. Dynamic imaging of CXCR4 expression in subcutaneous U87 xenografts with ^{64}Cu -AMD3465. NOD/SCID mice bearing U87 and U87-stb-CXCR4 glioblastoma xenografts on left and right flanks, respectively, were given approximately 9.25 MBq (250 μCi) of ^{64}Cu -AMD3465 via tail vein injection, and whole-body pseudodynamic imaging was performed for 70 min. Dynamic time-activity curves are for various tissues: muscle (black), U87 (blue), kidney (orange), liver (green), and U87-stb-CXCR4 (red). Data are mean \pm SD of 4 animals. Specific accumulation of radioactivity in U87-stb-CXCR4 (red line) over U87 (blue line) is apparent.

in Figure 3 and Supplemental Figure 2. The highest signal intensity was observed in the U87-stb-CXCR4 tumors at all time points. The best image contrast for U87-stb-CXCR4 tumors was noticed at 90 min after injection. Even after two ^{64}Cu half-lives and 24 h after injection of the radiotracer, U87-stb-CXCR4 tumors were clearly visible. Other organs with noticeable uptake were the liver and kidneys. On the basis of these image contrast data, in vivo specificity studies and imaging studies in the colon tumor model were performed at 90 min after injection of tracers. A 25 mg/kg blocking dose of AMD3465 resulted

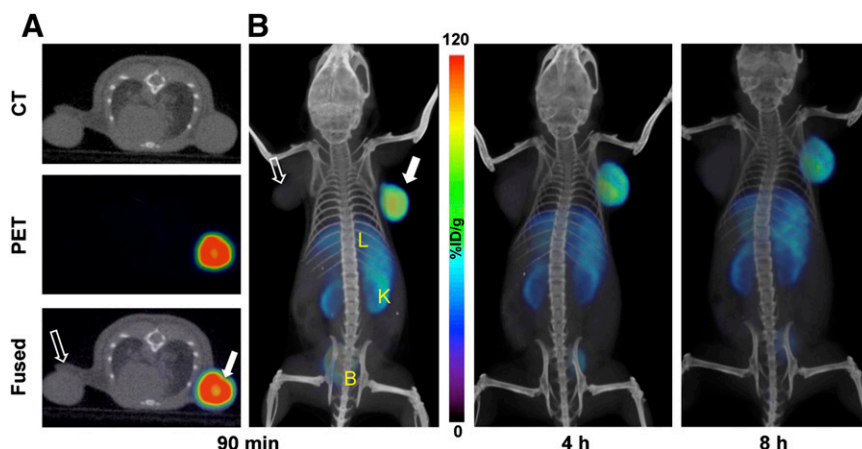


FIGURE 3. PET/CT images of CXCR4 expression in subcutaneous brain tumor xenografts with ^{64}Cu -AMD3465. NOD/SCID mice bearing U87 and U87-stb-CXCR4 glioblastoma xenografts on left and right flanks, respectively, were given approximately 9.25 MBq (250 μCi) of ^{64}Cu -labeled radiotracers via tail vein injection, and PET/CT images were acquired. (A) Representative transaxial PET, CT, and fused sections of both tumors from a mouse injected with ^{64}Cu -AMD3465 at 90 min after injection. (B) Representative volume-rendered whole-body images of a ^{64}Cu -AMD3465 injected mouse at 90 min (left), 4 h (middle), and 8 h (right) after injection. All images were decay-corrected and scaled to same maximum threshold value. B = bladder; K = kidney; L = liver; solid arrow = U87-stb-CXCR4 tumor; unfilled arrow = U87 tumor.

corrected and scaled to same maximum threshold value. B = bladder; K = kidney; L = liver; solid arrow = U87-stb-CXCR4 tumor; unfilled arrow = U87 tumor.

in a significant reduction of ^{64}Cu -AMD3465 uptake in U87 and U87-stb-CXCR4 tumors, demonstrating the specificity of the radiotracer (Fig. 4A). Further evidence of specificity was established by uniform uptake observed in mice injected with only ^{64}Cu - CuCl_2 (Fig. 4A). PET studies of mice with HT-29 tumors showed specific accumulation of activity in the tumors (Fig. 5A). Also, the blocking dose inhibited the radioactivity uptake in tumors, further demonstrating the ^{64}Cu -AMD3465 specificity in these tumor models (Fig. 5A). Supporting the CXCR4 expression levels observed *in vitro*, the uptake of ^{64}Cu -AMD3465 in HT-29 tumors was intermediate between that of U87 and U87-stb-CXCR4 tumors.

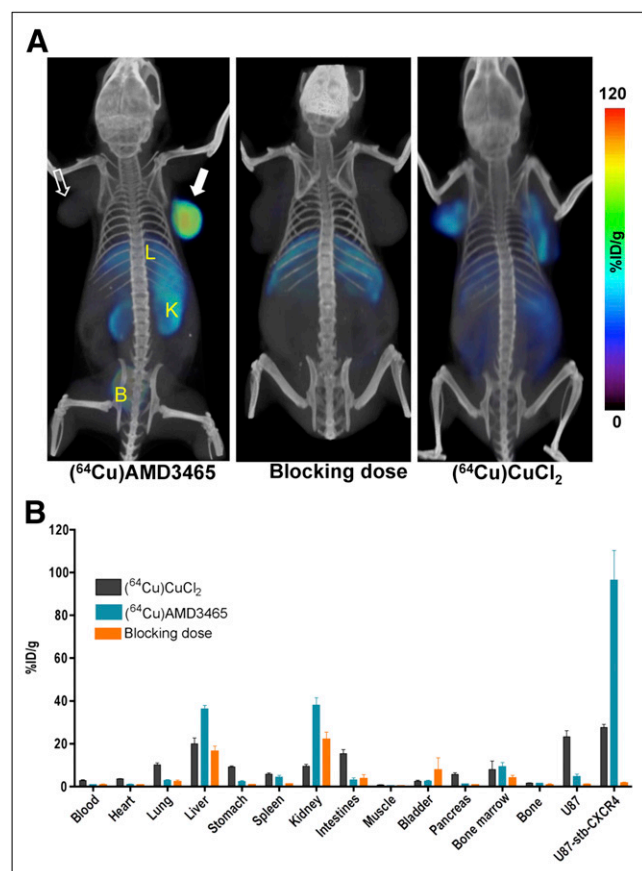


FIGURE 4. In vivo specificity of ^{64}Cu -AMD3465 in subcutaneous brain tumor xenografts. NOD/SCID mice bearing U87 and U87-stb-CXCR4 glioblastoma xenografts on left and right flanks, respectively, were given approximately 9.25 MBq (250 μCi) of ^{64}Cu -labeled radiotracers via tail vein injection, and PET/CT images were acquired. (A) Representative volume-rendered whole-body images of ^{64}Cu -AMD3465, AMD3465 blocking dose (25 mg/kg) followed by ^{64}Cu -AMD3465, and ^{64}Cu - CuCl_2 alone. All images were scaled to same maximum threshold value. (B) Biodistribution analysis of selected tissues from mice injected with 740 kBq (20 μCi) of radiotracers and sacrificed at 90 min after injection. All radioactivity values were converted into %ID/g. Biodistribution data are mean \pm SD of 4–5 animals. B = bladder; K = kidney; L = liver; solid arrow = U87-stb-CXCR4 tumor; unfilled arrow = U87 tumor.

Ex Vivo Biodistribution

To validate the imaging studies and further quantify the ^{64}Cu -AMD3465 uptake, biodistribution studies were performed at 30, 60, 90, 240, and 600 min in the brain tumor model (Table 1). Those biodistribution studies showed the highest uptake in the U87-stb-CXCR4 tumor at all time points. Other tissues with noticeable uptake were the liver, kidneys, and bone marrow. All other tissues had a low %ID/g, in agreement with PET data. The tumor-to-muscle and tumor-to-blood ratios for both tumors are shown in Table 2 and support the best contrast observed in PET/CT at 90 min after injection. The blocking dose of AMD3465 resulted in a greater than 95% reduction in the tumor uptake of radioactivity (Fig. 4B). The %ID/g for the kidneys, liver, spleen, and bone marrow reduced roughly by half, suggesting that a significant portion of that uptake was CXCR4-mediated. In mice injected with 740 kBq (20 μCi) of ^{64}Cu - CuCl_2 , relatively low but uniform distribution of radioactivity was observed in several tissues and both tumors.

Biodistribution studies in mice bearing HT-29 colon tumors were performed at 90 min after injection (Fig. 5B). The %ID/g for the HT-29 tumors was 5.62 ± 0.90 , and the tumor-to-muscle and tumor-to-blood ratios were 24.78 ± 2.38 and 8.05 ± 0.60 , respectively. No significant differences in radioactivity distribution were observed in other tissues, compared with the brain tumor model. Representative images of sections stained with hematoxylin and eosin or CXCR4 are shown in Figure 5C.

DISCUSSION

We investigated a radiolabeled version of a known high-affinity monocyclam inhibitor of CXCR4, AMD3465, as an imaging agent for CXCR4 expression. ^{64}Cu -AMD3465 evaluation in U87 glioblastoma cells, U87 cells stably expressing CXCR4, and a colon tumor expressing intermediate levels of CXCR4 showed specific accumulation of radioactivity. The specificity, target selectivity, and tumor-to-muscle ratios observed suggest that ^{64}Cu -AMD3465, compared with other known agents such as AMD3100, is a superior agent for PET of CXCR4 expression and can delineate graded levels of CXCR4 expression in tumors.

To determine the pharmacokinetics and establish the specificity, we initially investigated ^{64}Cu -AMD3465 in subcutaneous brain tumor models stably expressing CXCR4. In vitro binding studies showed specific binding of radioactivity to U87-stb-CXCR4 cells, compared with the parental U87 cells. Also, the *in vivo* dynamic and whole-body PET/CT studies clearly demonstrated selective accumulation of radioactivity in the U87-stb-CXCR4 tumors, starting as early as 5 min after injection of the radiotracer. The tumor-to-muscle ratios reached a maximum at 90 min after injection of the radiotracer. The radioactivity uptake in liver and kidneys reached a maximum within few minutes after injection and remained constant up to 70 min. The radiotracer accumulation in the U87-stb-CXCR4 tumors was evident even at 24 h. Retrospective *in vitro* analysis of

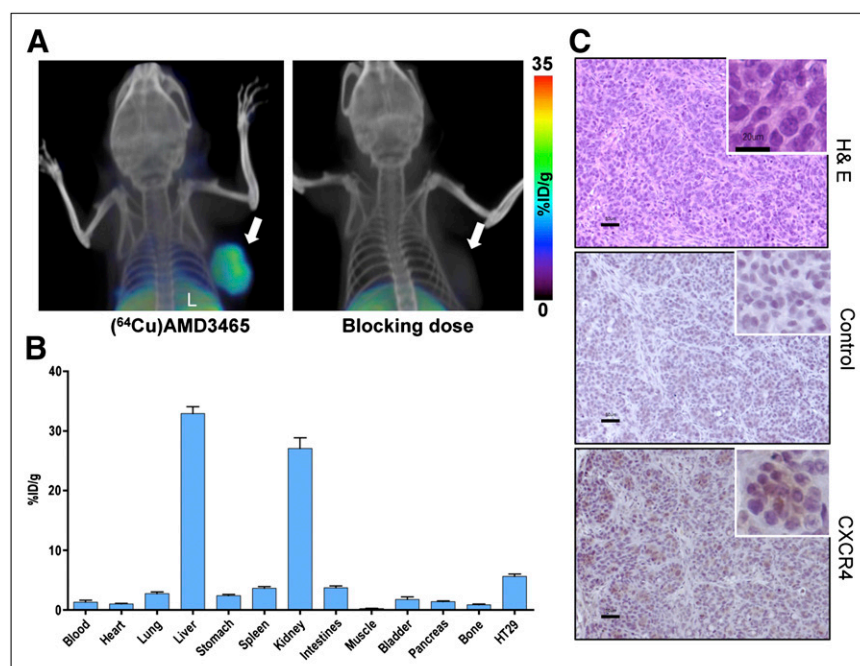


FIGURE 5. CXCR4 expression imaging in colon cancer xenografts with ^{64}Cu -AMD3465. NOD/SCID mice harboring HT-29 colon cancer xenografts in upper right flank received either approximately 9.25 MBq (250 µCi) of ^{64}Cu -AMD3465 or AMD3465 (25 mg/kg) blocking dose followed by ^{64}Cu -AMD3465 via tail vein injection, and whole-body images were acquired at 90 min after injection. (A) Representative volume-rendered whole-body images showing clear and specific accumulation of radioactivity in HT-29 tumors. (B) Biodistribution analysis of selected tissues from mice injected with 740 kBq (20 µCi) of ^{64}Cu -AMD3465 and sacrificed at 90 min after injection. All radioactivity values were converted into %ID/g. Biodistribution data are mean \pm SD of 6 animals. (C) Representative microscopy images of 10-µm-thick sections stained with hematoxylin and eosin and CXCR4 obtained at $\times 10$ magnification from HT-29 tumors. A $\times 20$ magnification is also shown in boxed region. H & E = hematoxylin and eosin; L = liver; solid arrow = HT-29 tumor.

^{64}Cu -AMD3465 uptake at 37°C in U87-stb-CXCR4 cells revealed that accumulation and retention of radioactivity was due to receptor-mediated endocytosis and perhaps binding of this radioactivity to cytosolic proteins. CXCR4-mediated endocytosis has been previously described using fluorescently labeled CXCL12 (35).

To validate further the utility of ^{64}Cu -AMD3465, we pursued imaging and biodistribution studies in a colorectal adenocarcinoma cell line, HT-29, that has 30% of its cells expressing CXCR4 as determined by flow cytometry. Both PET and biodistribution studies showed clear and specific

accumulation of radioactivity in tumors derived from those cells. This radioactivity uptake in tumors was significantly reduced in blocking studies, further validating the specificity of ^{64}Cu -AMD3465. Furthermore, the %ID/g values of HT-29 tumors were intermediate to U87 and U87-stb-CXCR4 tumors, suggesting that variable levels of CXCR4 expression could be delineated using this imaging agent.

Previously, we investigated the bicyclam AMD3100 as a PET agent for CXCR4 expression in the same glioblastoma models, which allows for comparison between these 2 agents (26). The tumor-to-muscle and tumor-to-blood ratios

TABLE 1
 ^{64}Cu -AMD3465 Ex Vivo Biodistribution Studies for Subcutaneous Brain Tumor Xenografts

Organ/tissue	Time (min)				
	30	60	90	240	600
Blood	1.93 \pm 0.30	0.90 \pm 0.08	0.73 \pm 0.23	0.61 \pm 0.09	1.20 \pm 0.36
Heart	1.19 \pm 0.16	1.00 \pm 0.20	0.90 \pm 0.25	0.93 \pm 0.06	0.72 \pm 0.07
Lung	4.48 \pm 0.56	3.70 \pm 0.99	2.78 \pm 0.47	3.39 \pm 0.85	2.10 \pm 0.13
Liver	32.08 \pm 2.03	35.06 \pm 7.77	36.15 \pm 1.79	33.65 \pm 3.13	27.27 \pm 2.57
Stomach	1.91 \pm 0.31	2.02 \pm 0.76	2.17 \pm 0.65	1.98 \pm 0.102	1.21 \pm 0.03
Spleen	6.23 \pm 1.17	5.68 \pm 1.75	4.28 \pm 1.00	4.88 \pm 0.42	2.64 \pm 0.12
Kidney	32.45 \pm 2.33	32.05 \pm 4.03	37.93 \pm 3.57	33.57 \pm 6.39	25.15 \pm 3.29
Small intestines	3.12 \pm 0.53	3.33 \pm 0.87	2.99 \pm 1.01	2.77 \pm 0.69	1.79 \pm 0.19
Muscle	0.75 \pm 0.52	0.37 \pm 0.12	0.30 \pm 0.13	0.23 \pm 0.03	0.19 \pm 0.02
Bladder	2.65 \pm 0.32	4.75 \pm 3.43	2.41 \pm 0.51	2.17 \pm 0.71	0.96 \pm 0.21
Pancreas	1.19 \pm 0.18	1.53 \pm 0.32	1.13 \pm 0.22	1.47 \pm 0.62	0.68 \pm 0.05
Bone	2.79 \pm 0.74	1.53 \pm 0.44	1.38 \pm 0.19	1.12 \pm 0.19	0.80 \pm 0.36
Bone marrow	6.39 \pm 0.00	10.22 \pm 4.97	9.18 \pm 2.04	8.10 \pm 3.38	9.42 \pm 3.88
U87	3.34 \pm 0.59	2.56 \pm 0.20	4.15 \pm 0.94	4.23 \pm 1.30	4.03 \pm 0.94
U87-stb-CXCR4	106.58 \pm 5.99*	102.70 \pm 20.80*	96.29 \pm 13.98*	50.95 \pm 6.38*	28.92 \pm 2.65*

* $P < 0.0001$, and comparative reference is U87 tumor uptake.

TABLE 2
Tumor-to-Tissue Ratios for ^{64}Cu -AMD3465 for Subcutaneous Brain Tumor Xenografts

Ratio	Time (min)				
	30	60	90	240	600
U87-stb-CXCR4 to U87	32.57 \pm 5.51	40.19 \pm 7.53	21.62 \pm 4.36	12.85 \pm 4.03	5.38 \pm 2.97
U87-stb-CXCR4 to muscle	193.82 \pm 108.55	297.78 \pm 93.83	362.56 \pm 153.51	222.95 \pm 27.62	151.00 \pm 29.91
U87 to muscle	5.88 \pm 2.93	7.31 \pm 1.59	17.53 \pm 7.64	18.91 \pm 7.25	42.04 \pm 35.98
U87-stb-CXCR4 to blood	56.10 \pm 7.93	114.76 \pm 23.09	138.78 \pm 30.87	83.47 \pm 4.63	27.80 \pm 9.93
U87 to blood	1.73 \pm 0.09	2.85 \pm 0.10	6.41 \pm 0.63	6.97 \pm 2.01	10.04 \pm 2.95

for ^{64}Cu -AMD3465 at 90 min after injection are 7- to 8-fold higher than those of ^{64}Cu -AMD3100, suggesting the superiority of ^{64}Cu -AMD3465 as an imaging agent. Even though ^{64}Cu -AMD3465 has improved affinity and kinetics, compared with ^{64}Cu -AMD3100, considerable uptake in the liver and kidneys was also observed. CXCR4 expression in the liver and kidneys has been reported (34,36), and reduced uptake has been observed in blocking studies in these tissues, suggesting that some of the uptake seen is receptor-mediated. Also, copper-cyclam complexes are reported to be thermodynamically stable; however, several in vitro studies have shown dissociation of copper from the complex (37,38). Therefore, some of this accumulation could be attributed to possible transchelation of ^{64}Cu from ^{64}Cu -AMD3465 to plasma proteins. Our previous studies with bicyclam ^{64}Cu -AMD3100 have shown that copper bound in the cyclam is stable for at least 4 h after injection of the radiotracer. These previous reports, combined with the loss of binding observed in our blocking studies and rather uniform uptake of radioactivity seen in ^{64}Cu - CuCl_2 -injected mice, suggest that the observed uptake is not related to transchelation but is receptor-mediated. Furthermore, the radioactivity in the blood of mice injected with ^{64}Cu -AMD3465 was 4-fold less than the %ID/g values in blood from the mice injected with ^{64}Cu - CuCl_2 . If there was significant transchelation, we would have observed higher %ID/g values in blood in mice injected with ^{64}Cu -AMD3465; however, higher values were not observed. The increase in %ID/g in blood at 600 min also suggests transchelation of copper at later time points. Cross-bridged cyclam-based inhibitors due to stable copper binding may lead to improved copper-based CXCR4 imaging agents (39). Also, possible metabolites such as ^{64}Cu -cyclam have been shown to have low affinity for CXCR4 (40,41), suggesting that the uptake observed in our studies is due to intact ^{64}Cu -AMD3465.

CONCLUSION

We have synthesized and performed a detailed evaluation of ^{64}Cu -AMD3465 as a PET agent to detect CXCR4 expression in tumor models with graded levels of CXCR4 expression. The results presented indicate that AMD3465 may serve as a suitable scaffold for the synthesis of clinically translatable CXCR4-targeted imaging agents.

DISCLOSURE STATEMENT

The costs of publication of this article were defrayed in part by the payment of page charges. Therefore, and solely to indicate this fact, this article is hereby marked "advertisement" in accordance with 18 USC section 1734.

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Erratum

The Invited Perspective “The Real Cost of Theoretic Risk Avoidance: The Need to Challenge Unsubstantiated Concerns About ¹³¹I Therapy,” by Goldsmith (*J Nucl Med*. 2011;52:681–682) contains an error in the first sentence of the sixth paragraph. According to title 10 of *Code of Federal Regulations* part 35.75(a), the originally stated value of 500 mSv in that sentence should have been 5 mSv. The corrected sentence appears below. The author regrets the error.

After the 1997 regulatory modification allowing release of patients receiving more than 1,110 MBq of ¹³¹I—provided the nuclear practitioner has demonstrated to the licensing authority that patients have been instructed on reasonable isolation and that conditions are such that no member of the public is likely to be exposed beyond 5 mSv—Grigsby et al. (2) distributed radiation-monitoring devices to family members (adults, children, and pets) of patients receiving 3.7–5.5 GBq of ¹³¹I.



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