

Comparative Assessment of Methods for Estimating Tumor Volume and Standardized Uptake Value in ^{18}F -FDG PET

Perrine Tyłski¹, Simon Stute¹, Nicolas Grotus¹, Kaya Doyeux², Sébastien Hapdey², Isabelle Gardin², Bruno Vanderlinden³, and Irène Buvat¹

¹IMNC UMR 8165 CNRS–Paris 7 and Paris 11 Universities, Orsay, France; ²LITIS EA 4108 Laboratory, University of Rouen, Rouen, France; and ³Nuclear Medicine Department, Bordet Institute, Université Libre de Bruxelles, Brussels, Belgium

In ^{18}F -FDG PET, tumors are often characterized by their metabolically active volume and standardized uptake value (SUV). However, many approaches have been proposed to estimate tumor volume and SUV from ^{18}F -FDG PET images, none of them being widely agreed upon. We assessed the accuracy and robustness of 5 methods for tumor volume estimates and of 10 methods for SUV estimates in a large variety of configurations.

Methods: PET acquisitions of an anthropomorphic phantom containing 17 spheres (volumes between 0.43 and 97 mL, sphere-to-surrounding-activity concentration ratios between 2 and 68) were used. Forty-one nonspheric tumors (volumes between 0.6 and 92 mL, SUV of 2, 4, and 8) were also simulated and inserted in a real patient ^{18}F -FDG PET scan. Four threshold-based methods (including one, T_{bgd} , accounting for background activity) and a model-based method (Fit) described in the literature were used for tumor volume measurements. The mean SUV in the resulting volumes were calculated, without and with partial-volume effect (PVE) correction, as well as the maximum SUV (SUV_{max}). The parameters involved in the tumor segmentation and SUV estimation methods were optimized using 3 approaches, corresponding to getting the best of each method or testing each method in more realistic situations in which the parameters cannot be perfectly optimized. **Results:** In the phantom and simulated data, the T_{bgd} and Fit methods yielded the most accurate volume estimates, with mean errors of $2\% \pm 11\%$ and $-8\% \pm 21\%$ in the most realistic situations. Considering the simulated data, all SUV not corrected for PVE had a mean bias between -31% and -46% , much larger than the bias observed with SUV_{max} ($-11\% \pm 23\%$) or with the PVE-corrected SUV based on T_{bgd} and Fit ($-2\% \pm 10\%$ and $3\% \pm 24\%$). **Conclusion:** The method used to estimate tumor volume and SUV greatly affects the reliability of the estimates. The T_{bgd} and Fit methods yielded low errors in volume estimates in a broad range of situations. The PVE-corrected SUV based on T_{bgd} and Fit were more accurate and reproducible than SUV_{max} .

Key Words: PET; standardized uptake value; tumor volume; partial volume effect correction; tumor segmentation

J Nucl Med 2010; 51:268–276

DOI: 10.2967/jnumed.109.066241

Received May 18, 2009; revision accepted Jul. 13, 2009.

For correspondence or reprints contact: Perrine Tyłski, IMNC, Campus d'Orsay, Bâtiment 104, 91406 Orsay Cedex, France.

E-mail: tyłski@imnc.in2p3.fr

COPYRIGHT © 2010 by the Society of Nuclear Medicine, Inc.

Automatic tumor delineation in ^{18}F -FDG PET images is highly desirable for improved quantification, objective patient monitoring, and refinement of CT-based treatment planning in radiotherapy. However, the tumor segmentation task is challenging given the modest spatial resolution and the relatively high noise level in PET images. A large number of approaches have been proposed to segment tumors in PET images. Many assume that voxels belonging to the tumor have an uptake greater than a certain threshold. This threshold can be set as a percentage of the maximum voxel value in the tumor (1), possibly accounting for surrounding activity (2). Alternatively, the threshold can be calibrated as a function of the mean activity in a growing region around the tumor (3), adjusted using iterative approaches (4–7), or even applied to images of the glucose metabolic rate derived from dynamic PET (8). Apart from threshold-based approaches, gradient-based segmentation relying on morphologic information or on active contours has been proposed (9–11). Methods including various statistical models have also been described (12,13).

To date, there is no consensus on which methods should be preferred for tumor segmentation, because of the difficulty in assessing tumor volumes in vivo (14). Although the performance of the different segmentation methods has been studied in specific configurations, a comprehensive comparison of various segmentation approaches for a broad range of cases has not been reported. Comparative studies considering patient tumors in the context of radiotherapy planning have underlined the great variability of the volumes defined from the PET images as a function of the segmentation method (2,15,16) but have not investigated accuracy in tumor volume estimates. It has also been shown that the standardized uptake values (SUV in g/cm^3 , units will not be specified hereafter) strongly depended on the methods used to define the tumor volume (17,18).

The purpose of this study was to assess the accuracy, precision, and robustness of 5 volume estimation methods (1–3,6,19). Using these segmentation methods, 10 SUV estimates were also compared. The comparisons were

performed using phantom data and simulations of patient PET scans. A detailed analysis of the performance of the methods depending on whether the parameters they involved were perfectly optimized was also included.

MATERIALS AND METHODS

Phantom Data

Seventeen spheres (Table 1) were inserted in the Data Spectrum model ECT/TOR/P torso phantom (volume of 10.3 L), containing a liver insert (1.2 L) and 2 lung inserts (0.9 and 1.1 L) (Fig. 1). Five phantom configurations were considered, with activity concentration ratios between the spheres and the background compartment of 10.1, 8.3, 6.5, 4.8, and 2.9. This yielded a 1.2–67.9 range of sphere-to-surrounding-activity concentration ratios, depending on the location of the sphere (Table 2). For each phantom configuration, a 3-min acquisition was performed on a Siemens Biograph PET/CT scanner. Images were reconstructed using ordered-subsets expectation maximization (6 iterations, 8 subsets), corrected for attenuation using a CT-derived map, for random coincidences using delayed coincidences, and for scatter using a model-based correction (20). The voxel size was $2 \times 2 \times 2$ mm. The reconstructed images were postfiltered with a 3-dimensional (3D) gaussian function of 5 mm in full width at half maximum (FWHM). Among the 85 spheres (17 spheres \times 5 acquisitions), 7 (the 5 smallest spheres and 2 spheres located in the liver insert in acquisition 5; Table 2) could not be visually detected and were excluded from further analysis, resulting in 78 spheres in this dataset.

Simulated Data

The GATE Monte Carlo simulation software (21) was used to simulate PET data as acquired from the Philips Gemini GXL PET scanner. A cylindrical water phantom (22 cm in diameter and 19 cm in height) including 6 spheres of 1.1, 2.1, 3.6, 8.6, 16.8, and 28.7 mL was first simulated. The spheres were in the central transaxial plane of the cylinder, the center of each sphere being equidistant and 5.5 cm from the axis. The activity in the phantom background was 3.4 kBq/mL, and 4 acquisitions were simulated with sphere-to-background-activity ratios of 2, 4, 8, and 12. These simulations

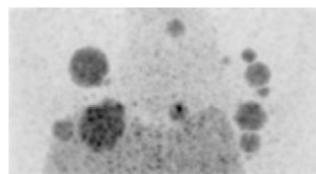


FIGURE 1. Maximum intensity projection of phantom containing 17 spheres.

were used for calibration of the segmentation methods applied to the patient simulations.

GATE was also used to simulate realistic tumors within the PET scan of a patient (66 y old; 70 kg) with no tumor in the lungs (Fig. 2) (22). The PET “tumor-free” sinogram of the patient was first simulated based on his actual Philips Gemini GXL PET/CT scan by estimating the activity distribution from the reconstructed PET images and the propagation medium from the CT scan. Tumors were then placed in the healthy lungs, and a PET sinogram of the tumors only was simulated using the patient CT as the propagation medium, in which the attenuation of the tumor (considered as soft tissues) had been added. The 3D contours of the simulated tumors were derived from a nuclear physician’s manual delineation of 41 lung tumors in fourteen ^{18}F -FDG PET scans of patients with non-small cell lung cancer. The tumor volumes ranged from 0.6 to 91.8 mL (mean = 13.01 ± 19.5 mL). Each of the 41 tumors was simulated with SUV of 2, 4, and 8 to yield 123 simulated tumors with various activities and volumes. The simulated “tumor-free” sinogram and the “tumor-only” sinogram were corrected for attenuation using the appropriate attenuation maps and were added after all counts originating from the tumor locations had been removed from the “tumor-free” sinogram. The summed sinograms were finally reconstructed.

All simulated sinograms corresponded to 2-min scans and covered an 18-cm axial field of view. Because sinograms containing only true coincidences were considered, images did not require random and scatter corrections. Images were reconstructed using 3D ordered-subsets expectation maximization (5 iterations and 5 subsets) and postfiltered with a 5-mm FWHM 3D gaussian function. The voxel size was $4 \times 4 \times 4$ mm. The whole simulated activity in the patients with tumors was between 39.8 and 43.0 MBq.

Volume Estimates

Five methods for tumor delineation were considered. All were applied to large manually defined volumes of interest (VOI) containing the spheres or simulated tumors and including at least 80% of background voxels.

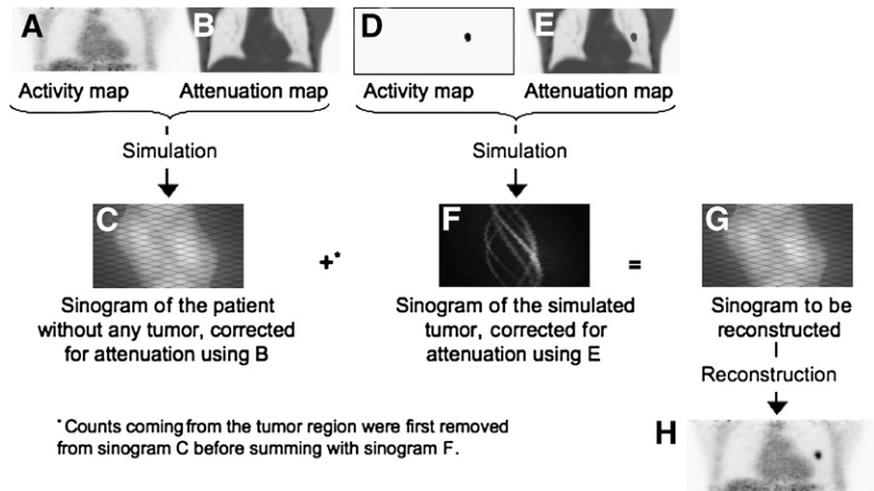
TABLE 1. Volumes and Locations of the 17 Spheres in the Phantom

Sphere	Location	Volume (mL)
1	Lung insert	0.4
2	Background	0.6
3	Background	1.0
4	Background	1.2
5	Background	1.6
6	Liver insert	2.1
7	Background	2.7
8	Lung insert	3.8
9	Background	5.6
10	Background	11.6
11	Background	11.7
12	Background	13.6
13	Lung insert	19.3
14	Background	26.6
15	Background	27.9
16	Lung insert	58.1
17	Liver insert	97.3

TABLE 2. Activity Concentrations in Lung and Liver Inserts and in Background of Phantom for Each Acquisition and Corresponding Sphere-to-Surrounding-Activity Concentration Ratios

Acquisition no.	Activity concentration (kBq/mL)			
	Sphere	Lung	Liver	Body
1	28.2	0.4 (67.9)	6.8 (4.1)	2.8 (10.1)
2	18.4	0.3 (55.6)	5.4 (3.4)	2.2 (8.3)
3	15.1	0.4 (41.2)	5.7 (2.6)	2.3 (6.5)
4	11.3	0.4 (29.9)	5.9 (1.9)	2.4 (4.8)
5	8.2	0.4 (19.6)	6.8 (1.2)	2.8 (2.9)

Data in parentheses are corresponding sphere-to-surrounding-activity concentration ratios.



Four methods, denoted T_{\max} , T_{reg} , T_{mean} , and T_{bgd} , considered that all connected voxels with an intensity greater than a given threshold belonged to the tumor.

In T_{\max} , the threshold was defined as a percentage of the SUV_{\max} in the VOI.

In T_{reg} (6), the threshold was estimated iteratively using

$$T_{\text{reg}} = \beta \times \text{SUV}_{\text{mean}} + \gamma, \quad \text{Eq. 1}$$

where β and γ were calibration factors.

In this approach, the tumor volume was first delineated using T_{\max} with $\alpha = 0.4$ and the mean SUV (SUV_{mean}) in this volume was deduced. The corresponding threshold T_{reg} was derived from Equation 1 and a new mean SUV in the corresponding tumor region was calculated. This procedure was repeated until the segmented region differed by less than 1 voxel between 2 iterations.

In T_{mean} , the threshold was defined as a percentage δ of the mean SUV in a growing region R_{grow} (3). The algorithm was initialized with R_{grow} corresponding to a single voxel in the tumor. If voxels connected to R_{grow} had an intensity of at least $\delta \times \text{mean SUV}$, they were included in R_{grow} . Mean SUV was updated and the process was repeated until no additional voxel could be included in R_{grow} .

In T_{bgd} , the threshold depended on the activity I_{bgd} surrounding the tumor and on the mean activity $I_{0.7}$ in the volume defined by the voxels with an intensity higher than $0.7 \times I_{\max}$ (2):

$$T_{\text{bgd}} = \varepsilon \times I_{0.7} + I_{\text{bgd}}. \quad \text{Eq. 2}$$

In our implementation, I_{bgd} was determined automatically. As the original VOIs included about 80% of background voxels, the histogram in this VOI had a gaussian shape roughly centered on the mean background value. The mean background value was deduced by fitting the histogram with a gaussian.

The fifth method, Fit, was derived from a previously published method (23). It assumed that the tumor image could be modeled as the convolution of the actual tumor volume of uniform activity with a 3D gaussian function describing the local spatial resolution ζ in the reconstructed image. In our implementation (19), the tumor volume was initialized using the T_{bgd} method with $\varepsilon = 0.25$. Assuming that this volume was always larger than the true tumor volume, this volume was eroded using a 1-voxel structuring element. The 3 model parameters (number of erosions, activities

in the tumor, and activities outside the tumor) that best fit the observed tumor image in the least-square sense were identified. The tumor image modeling and erosion were performed after resampling the tumor model images to a $1 \times 1 \times 1$ mm voxel size with a piecewise cubic Hermite interpolating polynomial interpolation (24), whereas the comparison of the model image with the original PET images was performed in the original PET sampling.

SUV Estimates

For each tumor segmented using T_{\max} , T_{reg} , T_{mean} , and T_{bgd} , the mean SUV in the segmented volumes was calculated. Each of these 4 mean SUV was also corrected for partial-volume effect (PVE) using a recovery coefficient (RC). The RC was deduced from the segmented volume by convolving the binary mask corresponding to this volume with the 3D gaussian function of FWHM θ modeling the spatial resolution effect in the reconstructed images (25). The PVE correction also accounted for surrounding activity (25), which was estimated as I_{bgd} in the T_{bgd} method previously described.

The mean SUV in the tumor volume estimated using the Fit method was intrinsically corrected for PVE as the sampling and resolution effects were modeled.

The maximum SUV (SUV_{\max}) in the tumor VOI was also systematically calculated.

In summary, for each tumor, 5 volume estimates (from T_{\max} , T_{reg} , T_{mean} , T_{bgd} , and Fit) and 10 SUV estimates (from T_{\max} , T_{reg} , T_{mean} , T_{bgd} without and with PVE correction, Fit, and SUV_{\max}) were available. The SUV estimates were denoted $\text{SUV}_{T_{\text{reg}}}$, $\text{SUV}_{T_{\text{regRC}}}$, $\text{SUV}_{T_{\text{mean}}}$, $\text{SUV}_{T_{\text{meanRC}}}$, $\text{SUV}_{T_{\max}}$, $\text{SUV}_{T_{\maxRC}}$, $\text{SUV}_{T_{\text{bgd}}}$, $\text{SUV}_{T_{\text{bgdRC}}}$, SUV_{Fit} , and SUV_{\max} .

Comparison Protocols

Optimization of the Segmentation Methods. All segmentation methods involved 1 or 2 parameters. To ensure a fair comparison of the methods, these parameters have to be optimized for each type of scanner, acquisition, and processing protocol.

Using the phantom acquisitions and the cylindrical phantom simulation, for each sphere we calculated the optimal parameters minimizing the absolute value of the error in sphere volume estimate for each method. We also determined the corresponding FWHM θ giving the smallest absolute value of the error in SUV corrected for PVE with the RC.

Given these optimal parameters (1 or 2 per sphere and per segmentation method), the segmentation methods were first assessed on the real phantom data using a leave-one-out procedure (26): for each segmentation method, the parameters used to segment a given sphere were the average of the 77 optimal parameter values obtained for all other spheres. Similarly, the θ value used to calculate the RC used for PVE correction was taken as the average of the 77 optimal θ obtained for all other spheres.

As this leave-one-out procedure assessed the optimal performance of the methods that cannot be achieved in real situations, we also used a hold-out procedure. The phantom dataset of 78 spheres was randomly split into 2 groups of 39 spheres. The first group was used to optimize the segmentation and resolution parameters for each sphere. The means over all spheres of these optimized parameters were then used for segmenting the spheres of the other group. The validation and test groups were identical for all segmentation methods.

For the simulated patient data, a realistic approach was used for optimizing the segmentation parameters. It consisted of optimizing the parameters for each of the 24 spheres (6 spheres \times 4 acquisitions) of the simulated cylindrical phantom. The averaged parameter values were considered when applying the segmentation methods to the simulated patient data.

Figures of Merit and Statistical Analysis. For each segmentation method and each sphere or simulated tumor, the percentage error in volume estimate was calculated:

$$E_{\text{volume}}(\%) = \frac{\text{volume of the segmented region} - \text{true volume}}{\text{true volume}} \times 100. \quad \text{Eq. 3}$$

The absolute value $|E_{\text{volume}}|$ was also used to compare the segmentation methods.

Similarly, the percentage error in SUV estimates was calculated:

$$E_{\text{SUV}}(\%) = \frac{\text{SUV} - \text{true SUV}}{\text{true SUV}} \times 100. \quad \text{Eq. 4}$$

The mean error and associated SD were calculated for each estimation method and optimization procedure.

Sign tests were performed to test if the median of the E_{volume} or E_{SUV} distribution was significantly different from zero, using a Bonferroni adjustment to account for multiple comparisons (27). A Friedman test based on ranks and appropriate for multiple comparisons on paired data was used to test whether several distributions of $|E_{\text{volume}}|$ or $|E_{\text{SUV}}|$ were identical. A Tukey procedure appropriate for the multiple comparisons of dispersions (28) was used to rank the estimation methods as a function of the variability of the errors. All tests were performed with $\alpha = 0.05$.

RESULTS

Volume Estimates

We distinguished the results obtained for all the spheres or simulated tumors and the results obtained for spheres or tumors with a volume of 2 mL or more, as it has been suggested that no accurate volume estimate could be achieved for tumor volumes less than 2 mL (29). In the phantom data, T_{mean} and T_{reg} did not converge in 2 of 78 spheres (0.43- and 0.99-mL spheres with a contrast of 20

and 3, respectively) and in 11 of 123 simulated tumors (volumes from 0.64 to 2.4 mL with an SUV of 2). These spheres and tumors were excluded from further analysis.

Figure 3 shows the tradeoff between the mean percentage errors in volume estimate and the variability of the error as measured by the SD of E_{volume} , for the 5 segmentation methods and the 3 datasets (phantom data and leave-one-out optimization, phantom data and hold-out optimization, simulated patient data). These plots suggest that for all datasets, the 2 segmentation methods performing the best (lowest bias and smallest variability) are T_{bgd} and Fit. T_{mean} suffers from a large variability for all datasets. T_{max} and T_{reg} had variable performance depending on the dataset.

E_{volume} was significantly different from zero only for T_{max} and T_{mean} and spheres larger than 2 mL for the leave-

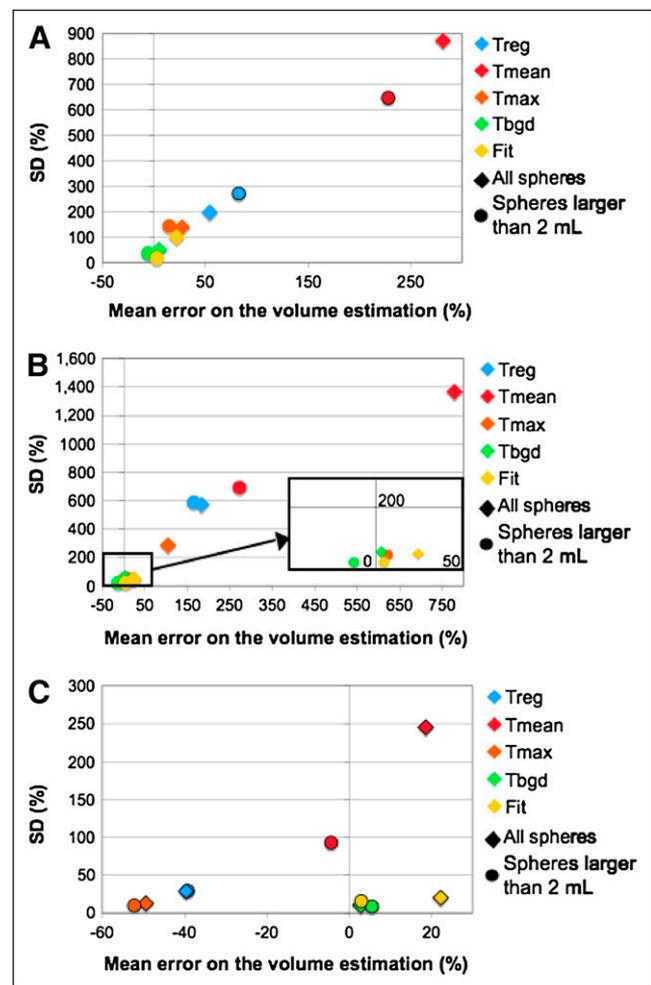


FIGURE 3. SD of volume percentage error as function of mean of volume percentage error E_{volume} for all spheres and tumors (diamonds) and for spheres and tumors > 2 mL (circles) using 5 segmentation methods: leave-one-out phantom data (A), hold-out phantom data (B), and simulated patient data (C). Black-edged symbols show cases in which E_{volume} was significantly different from zero.

one-out dataset. For the simulated data, all the error distributions had a significant bias.

Figure 4 shows the mean rank of $|E_{\text{volume}}|$ for the 5 volume estimation methods (Friedman test), when one is considering all spheres and tumors of the leave-one-out phantom data, hold-out phantom data, and simulated data. The smallest rank corresponds to the smallest $|E_{\text{volume}}|$. Fit and T_{bgd} were significantly less biased than T_{max} for the leave-one-out phantom data (Fig. 4A, red line). These 2 methods had the smallest ranks for the 3 datasets, consistent with their lowest bias in tumor volume estimates seen in Figure 3. Fit and T_{bgd} had nonsignificantly different accuracy, except in the simulated data, where T_{bgd} was significantly less biased than Fit.

For the 2 phantom datasets, T_{mean} led, on average, to the largest error in volume estimate, and the differences with the 4 other methods were systematically significant (green and brown lines in Figs. 4A and 4B). However in the simulated data, the mean rank of T_{mean} was significantly smaller than the mean rank of T_{max} (Fig. 4C, purple line).

When only the spheres and tumors with volumes larger than 2 mL were considered (results not shown), identical trends were found.

When the variability of the errors in volume estimates were compared for all datasets (y-axis in Fig. 3), the 2 methods yielding the most variable errors were T_{mean} and T_{reg} , with T_{mean} being systematically more variable than T_{reg} .

For the phantom data, the smallest variability of the error was systematically observed for T_{bgd} and Fit, with no consistent difference between them in terms of variability. For the simulated data, T_{bgd} had the smallest variability, but this variability was not significantly lower than that of T_{max} . Fit had a significantly larger variability than T_{bgd} and T_{max} for the simulated tumors.

SUV Estimates

The performance of the 10 SUV estimation methods was assessed for the 76 spheres and 112 simulated tumors for which the segmentation methods converged.

Figure 5 shows the SD of the SUV percentage errors as a function of the mean percentage errors in SUV estimates for the 3 datasets and for the 10 SUV estimation methods. The mean SUV not corrected for PVE clearly showed a negative bias. Focusing on the 6 methods including a PVE correction (y-axis in Fig. 5), the SDs of the error were consistent between datasets, between 19% and 31% for SUV_{Fit} , SUV_{max} , and $\text{SUV}_{\text{TmaxRC}}$. The variability of the errors as a function of the dataset was greater for $\text{SUV}_{\text{TmeanRC}}$, $\text{SUV}_{\text{TregRC}}$, and $\text{SUV}_{\text{TbgdRC}}$. However, the variability of the errors in SUV was less different between methods than the variability of the errors in tumor volume (compare y-axes of Figs. 3 and 5), except for $\text{SUV}_{\text{TregRC}}$, which had the significantly largest variability for the simulated data.

The hypothesis that the median of the SUV error distribution was equal to zero was always rejected except for $\text{SUV}_{\text{TregRC}}$, $\text{SUV}_{\text{TmeanRC}}$, and SUV_{max} for the

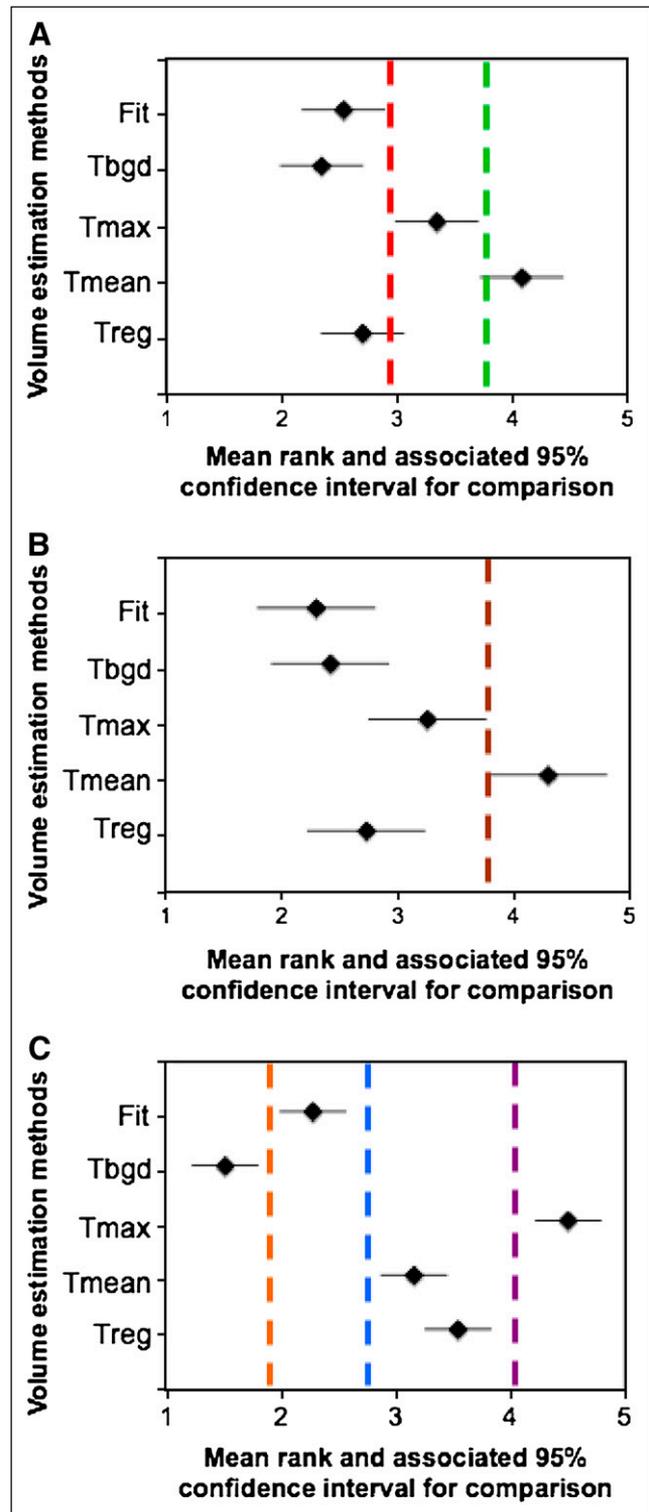


FIGURE 4. Comparison of mean rank of $|E_{\text{volume}}|$ for the 5 volume estimates: leave-one-out phantom data (A), hold-out phantom data (B), and simulated patient data (C). Colored lines highlight significant differences between methods.

leave-one-out sphere data. It was never rejected for the methods corrected for PVE, except for Fit in the hold-out sphere data.

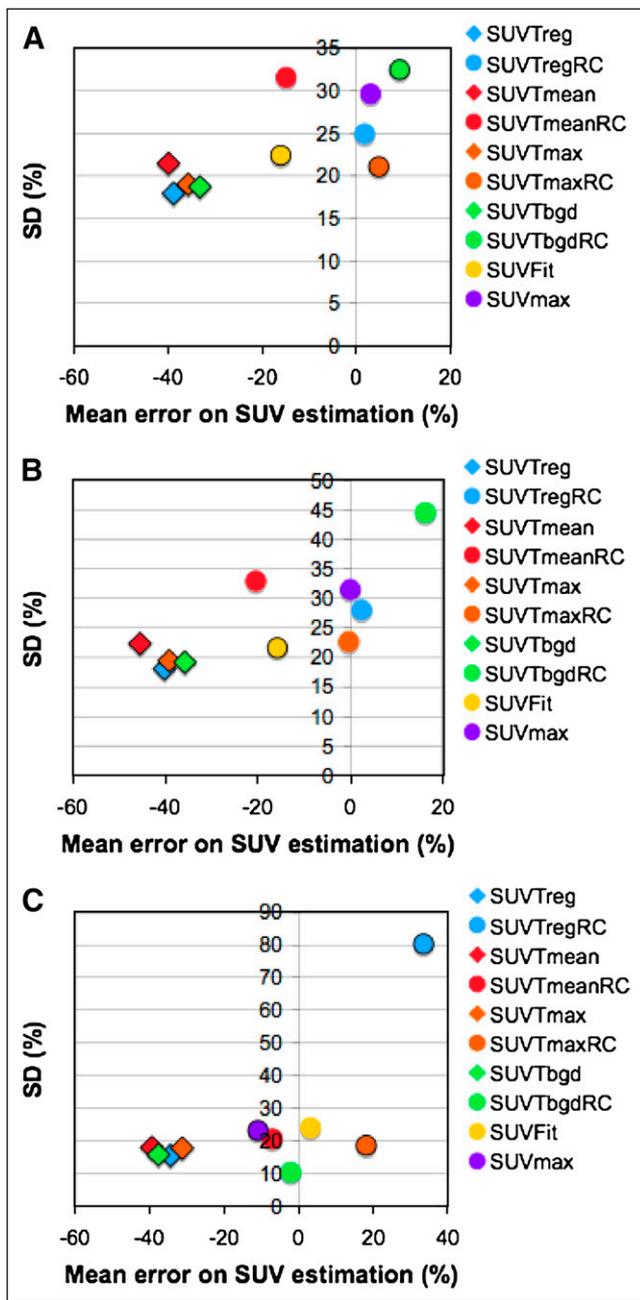


FIGURE 5. SD of SUV percentage error as function of mean of SUV percentage errors: leave-one-out phantom data (A), hold-out phantom data (B), and simulated patient data (C). SUV corrected for PVE and maximum value are shown using circles, whereas SUV not corrected for PVE are shown using diamonds. Black-edged symbols show cases in which mean error was significantly different from zero.

For noncorrected mean SUV, this hypothesis was rejected for all datasets. For the simulated tumors, the median bias was always significantly different from zero except for $SUV_{TmeanRC}$, SUV_{TbgdRC} , and SUV_{Fit} .

Figure 6 shows the mean rank of $|E_{SUV}|$ for the 10 SUV estimates when all spheres and tumors of the leave-one-out phantom data, hold-out phantom data, and simulated data

are considered. The smaller the rank, the more accurate the SUV estimate. The mean SUV not corrected for PVE was significantly different from the SUV corrected for PVE and from SUV_{max} for the leave-one-out and hold-out data (blue line in Fig. 6A and orange line in Fig. 6B). Comparing SUV_{max} with the SUV corrected for PVE, the only significant differences were between SUV_{max} and

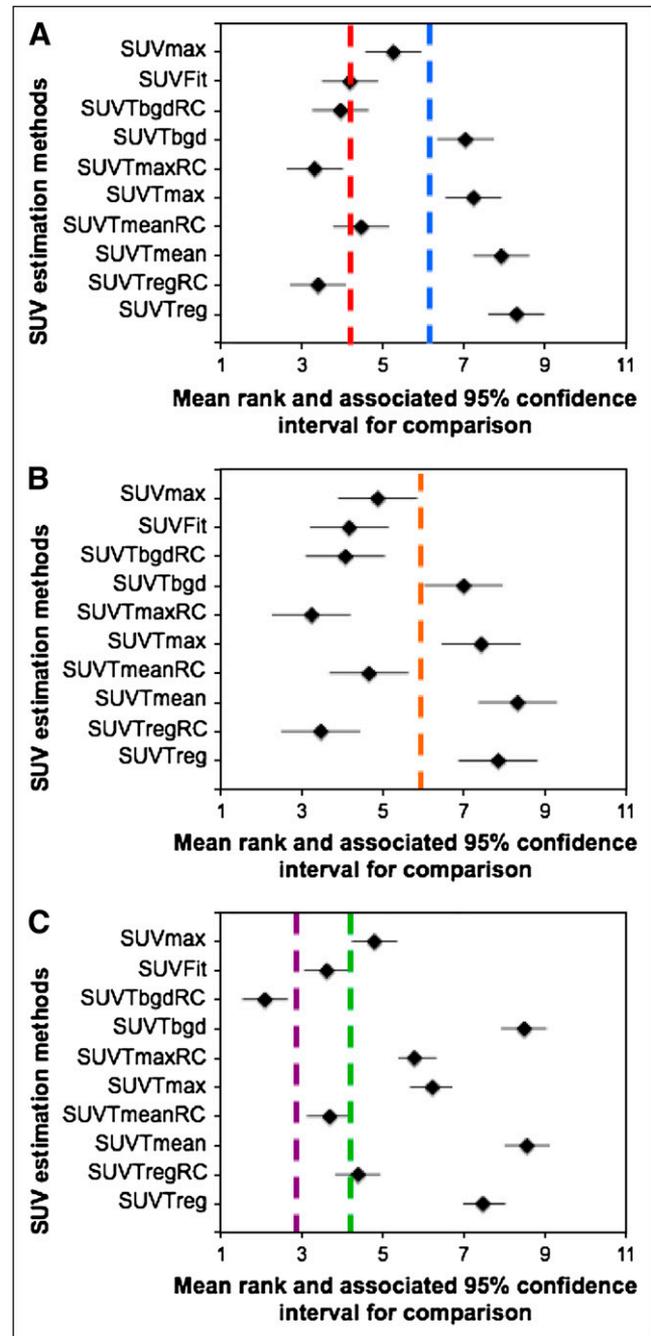


FIGURE 6. Comparison of mean rank of $|E_{SUV}|$ for 10 SUV estimation methods: leave-one-out phantom data (A), hold-out phantom data (B), and simulated patient data (C). Colored lines highlight significant differences between methods.

$SUV_{T_{max}RC}$ and between SUV_{max} and $SUV_{T_{reg}RC}$ for the leave-one-out data (red line in Fig. 6A).

For the phantom data and simulated data, the mean SUV not corrected for PVE had the largest ranks. However, $SUV_{T_{max}RC}$ was not significantly different from $SUV_{T_{max}}$ for simulated data. $SUV_{T_{bgd}RC}$ had a significantly smaller rank, compared with all other methods (purple line in Fig. 6C). SUV_{Fit} and $SUV_{T_{mean}RC}$ were significantly less biased than SUV_{max} (green line in Fig. 6C).

DISCUSSION

Although many methods have been proposed for tumor delineation in ^{18}F -FDG PET images (1–13), it is not clear yet which method should be preferred. Comparing the performance of these methods from the data published in the literature is almost impossible given the variety of situations in which evaluation studies have been conducted. In addition, the performance of each method depends on the proper optimization of its parameters. It is thus extremely important to consider the robustness of any method with respect to the setting of its parameters, given that the optimal parameters can never be identified in clinical configurations. The same observations are true for SUV estimates.

Our study compared several methods for estimating the tumor volumes or SUV , using 2 datasets and different optimization strategies for setting their parameters. We first considered a phantom including spheres as often used to characterize the performance of tumor segmentation methods (1,5,7,29) or SUV estimation methods (17,30). However, tumors are rarely spheric, and the activity distribution in tissues is far more complex in patients than in phantoms. We thus considered highly realistic Monte Carlo simulations of patient PET scans, based on real patient PET/CT scans. Such simulations were more representative of clinical situations than the phantom datasets in 2 respects: first, the background activity distribution reflected heterogeneities observed in a real patient (Fig. 2). Second, the tumors had shapes observed in real patients.

Three strategies were considered for setting the parameters of each method. The leave-one-out strategy ensures the best operating conditions for each method. Although the associated performance could not be achieved in practical situations, this strategy allowed us to characterize the potential of each method. The hold-out procedure relying on training and test dataset is conventionally used to assess the performance of estimation methods but remains unrealistic for patient data. The realistic optimization strategy when dealing with patient data consists of optimizing the parameters using phantom data acquired and reconstructed under the same conditions as the patient data. We thus also considered this third optimization strategy.

Volume Estimates

We first compared the accuracy in volume estimates for the 5 tumor segmentation methods (Figs. 3 and 4).

Whatever the dataset and optimization strategy, the T_{bgd} and Fit methods offered the best trade-off between bias and variability in volume estimates. As expected, the errors and SD were systematically larger for the hold-out optimization than for the leave-one-out strategy when the phantom data were considered (Fig. 3). However, the ranking of the 5 estimation methods was consistent (Fig. 4), although 2 differences (T_{bgd} and Fit vs. T_{max}) found significant with the leave-one-out optimization were no more significant with the hold-out data. The ranking of the methods with the simulated data was different from that with the phantom data (Fig. 4). T_{bgd} and Fit still yielded the most accurate volume estimates. Unlike in the phantom data, T_{bgd} was significantly better than Fit, and T_{max} performed the worst. These differences might be due to the shapes of the “tumors,” the different background patterns, or the different optimization strategies. To determine whether the optimization strategy used for the simulated data explained the significant difference between T_{bgd} and Fit, we also optimized the T_{bgd} and Fit parameters using a leave-one-out procedure for the simulated data (results not shown), but the resulting parameters were not significantly different from the parameters derived from the simulated phantom. The poorest performance of Fit, compared with T_{bgd} , for the simulated data was actually due to the inability of Fit to properly recover the shape of the simulated tumors using the erosion strategy. T_{bgd} might thus be more accurate than Fit for highly nonspheric tumors.

The better performance of T_{mean} for the simulated data, compared with the phantom data, could be due to the smaller range of “tumor”-to-background activity ratio in the simulated tumors (from 8 to 32) than in the phantom spheres (from 1.9 to 67.9). In particular, for spheres with a sphere-to-background ratio smaller than 8 (34/76), T_{mean} often led to severe volume overestimates, with a mean percentage error of 800% on these spheres, much poorer than previously reported (3). T_{mean} thus does not appear to be a good option when the processed images can include a large variety of tumor-to-background activity ratios.

T_{max} accuracy also depends on the tumor-to-background activity ratio (31). For the simulated data, T_{max} parameters were optimized using data with sphere-to-background-activity ratios between 1 and 12, which did not match those in the simulated tumors (8–32), yielding a systematic underestimation of the volumes of the simulated tumors. The optimization of the T_{max} parameter using a leave-one-out procedure for the simulated data (results not shown) actually led to parameters significantly different from those derived from the simulated phantom ($P < 0.001$). Similar to T_{mean} , the performance of T_{max} is thus highly dependent on whether its parameter has been optimized considering tumor-to-background-activity ratios similar to those observed in the images subsequently processed.

SUV Estimates

Overall, the comparison of SUV estimation methods showed a systematic underestimation of SUV with the methods that did not include any PVE correction, except for SUV_{\max} (Fig. 5), which often minimizes PVE (25).

Similar to what was observed for the volume estimates, the ranking of the SUV estimation methods was identical whatever the optimization strategy when the phantom data were considered (Figs. 6A and 6B), but differences ($SUV_{T_{\text{regRC}}}$ and $SUV_{T_{\text{maxRC}}}$ vs. SUV_{\max}) that were found significant with the leave-one-out optimization were no more significant with the hold-out optimization.

$SUV_{T_{\text{bgdRC}}}$ was not significantly different from the other PVE-corrected SUV in the phantom data and significantly less biased than all other SUV estimates in the simulated data. This latter result is consistent with the ranking of T_{bgd} for volume estimates.

SUV_{Fit} is corrected for PVE but has a negative bias in the phantom data, compared with SUV_{\max} , $SUV_{T_{\text{maxRC}}}$, and $SUV_{T_{\text{bgdRC}}}$. This is because the Fit method uses only 1 parameter for both volume and SUV estimations. This parameter has been optimized for volume recovery rather than for activity recovery on phantoms, making the method more accurate in estimating volumes than activity. In the simulations, SUV_{Fit} has the second smallest mean bias (3% against -2% for $SUV_{T_{\text{bgdRC}}}$), which suggests that the optimization of a single parameter for Fit still makes it robust enough to properly assess volume and SUV in a variety of situations. The observed systematic negative bias of T_{\max} in the simulated data leads to a systematic positive bias in $SUV_{T_{\text{maxRC}}}$.

The variability in the SUV estimation error is related to the variability in the volume estimates: for instance, T_{mean} had variable errors in volume and also in SUV estimates. Moreover, the relationship between the error in volume estimate and in SUV corrected for PVE is nonlinear (32). Small underestimation in small-volume estimates can lead to high overestimation of PVE corrected activity. Most methods overestimated the small volumes, but T_{bgd} underestimated volumes less than 2 mL in 41% of the cases (14% for T_{reg} , 4% for T_{mean} , and 14% for T_{max}) in the leave-one-out phantom data. This yielded outliers in $SUV_{T_{\text{bgdRC}}}$ error distribution and explained its positive bias and higher dispersion in the phantom data, compared with other methods.

Robustness of the Estimation Methods

Some methods were more sensitive than others to the proper setting of the parameters they involved. For the volume estimates, T_{\max} , T_{mean} , and T_{reg} had very different biases for the simulated data, compared with the phantom data, whereas T_{bgd} and Fit had a more constant bias across the datasets and optimization strategies. This finding suggests that T_{bgd} and Fit are more robust than the other methods with respect to the setting of the parameters.

For the SUV estimates, $SUV_{T_{\text{bgdRC}}}$ and SUV_{Fit} did not have the smallest rank for the phantom data (Fig. 6) but

were not significantly less accurate than $SUV_{T_{\text{regRC}}}$ and $SUV_{T_{\text{maxRC}}}$. For the simulated data, they yielded the most accurate SUV estimates ($SUV_{T_{\text{bgdRC}}}$ rank being significantly smaller than SUV_{Fit} rank), although training and evaluation datasets had different characteristics. $SUV_{T_{\text{bgdRC}}}$ and SUV_{Fit} thus seemed to yield the most robust SUV estimates. They were also significantly more accurate than SUV_{\max} for all the datasets.

Limitations to the Current Study

The evaluation based on the simulated data was supposed to be closer to real clinical situations than is the phantom. However, our simulations remained too simple in at least 2 regards: neither respiratory motion nor heterogeneity in tumor uptake was modeled. Our results might still be representative of the performance to be expected for images compensated for respiratory motion, such as respiratory-gated images with appropriate signal-to-noise ratios (33). Respiratory motion compensation (e.g., based on gated PET/CT) before tumor volume or SUV estimates is certainly more appropriate than optimizing tumor segmentation methods for data corrupted by motion, given the large variability of the motion blur in patients, as a function of the respiratory amplitude or tumor location, for instance. Further studies regarding the reliability of tumor volume estimates and SUV estimates for heterogeneous tumors are still needed. For heterogeneous tumors, however, the very concept of tumor volume and tumor SUV might have to be reconsidered, and approaches such as activity-volume histograms might appear more relevant for describing tumors (34) than is a single volume or SUV per tumor.

CONCLUSION

A comprehensive evaluation of 5 volume and 10 SUV estimation methods demonstrated that 2 segmentation methods (T_{bgd} and Fit) and 2 SUV indices corrected for PVE ($SUV_{T_{\text{bgdRC}}}$ and SUV_{Fit}) yielded the most accurate tumor volume and SUV estimates.

ACKNOWLEDGMENTS

We thank Dr. Michelle Dusart from the Citadelle Hospital of Liège (Belgium) for the delineation of PET tumors in patients and the GDR Stic Santé for supporting the collaboration between the IMNC and LITIS laboratories.

REFERENCES

1. Erdi YE, Mawlawi O, Larson SM, et al. Segmentation of lung lesion volume by adaptive positron emission tomography image thresholding. *Cancer*. 1997;80(12, suppl):2505–2509.
2. Nestle U, Kremp S, Schaefer-Schuler A, et al. Comparison of different methods for delineation of ^{18}F -FDG PET-positive tissue for target volume definition in radiotherapy of patients with non-small cell lung cancer. *J Nucl Med*. 2005;46:1342–1348.
3. Green AJ, Francis RJ, Baig S, Begent RH. Semiautomatic volume of interest drawing for ^{18}F -FDG image analysis—method and preliminary results. *Eur J Nucl Med Mol Imaging*. 2008;35:393–406.

4. van Dalen JA, Hoffmann AL, Dicken V, et al. A novel iterative method for lesion delineation and volumetric quantification with FDG PET. *Nucl Med Commun*. 2007;28:485–493.
5. Jentzen W, Freudenberg L, Eising EG, Heinze M, Brandau W, Bockisch A. Segmentation of PET volumes by iterative image thresholding. *J Nucl Med*. 2007;48:108–114.
6. Black QC, Grills IS, Kestin LL, et al. Defining a radiotherapy target with positron emission tomography. *Int J Radiat Oncol Biol Phys*. 2004;60:1272–1282.
7. Drever L, Roa W, McEwan A, Robinson D. Iterative threshold segmentation for PET target volume delineation. *Med Phys*. 2007;34:1253–1265.
8. Visser EP, Philippens MEP, Kienhorst L, et al. Comparison of tumor volumes derived from glucose metabolic rate maps and SUV maps in dynamic ¹⁸F-FDG PET. *J Nucl Med*. 2008;49:892–898.
9. Geets X, Lee JA, Bol A, Lonnew M, Gregoire V. A gradient-based method for segmenting FDG-PET images: methodology and validation. *Eur J Nucl Med Mol Imaging*. 2007;34:1427–1438.
10. Drever LA, Roa W, McEwan A, Robinson D. Comparison of three image segmentation techniques for target volume delineation in positron emission tomography. *J Appl Clin Med Phys*. 2007;8:93–109.
11. Li H, Thorstad WL, Biehl KJ, et al. A novel PET tumor delineation method based on adaptive region-growing and dual-front active contours. *Med Phys*. 2008;35:3711–3721.
12. Hatt M, Lamare F, Bousson N, et al. Fuzzy hidden Markov chains segmentation for volume determination and quantitation in PET. *Phys Med Biol*. 2007;52:3467–3491.
13. Montgomery DW, Amira A, Zaidi H. Fully automated segmentation of oncological PET volumes using a combined multiscale and statistical model. *Med Phys*. 2007;34:722–736.
14. Daisne JF, Duprez T, Weynand B, et al. Tumor volume in pharyngolaryngeal squamous cell carcinoma: comparison at CT, MR imaging, and FDG PET and validation with surgical specimen. *Radiology*. 2004;233:93–100.
15. Schinagl DA, Vogel WV, Hoffmann AL, van Dalen JA, Oyen WJ, Kaanders JH. Comparison of five segmentation tools for ¹⁸F-fluoro-deoxy-glucose-positron emission tomography-based target volume definition in head and neck cancer. *Int J Radiat Oncol Biol Phys*. 2007;69:1282–1289.
16. Jarritt PH, Carson KJ, Hounsell AR, Visvikis D. The role of PET/CT scanning in radiotherapy planning. *Br J Radiol*. 2006;79:S27–S35.
17. Boellaard R, Krak NC, Hoekstra OS, Lammertsma AA. Effects of noise, image resolution, and ROI definition on the accuracy of standard uptake values: a simulation study. *J Nucl Med*. 2004;45:1519–1527.
18. Krak NC, Boellaard R, Hoekstra OS, Twisk JW, Hoekstra CJ, Lammertsma AA. Effects of ROI definition and reconstruction method on quantitative outcome and applicability in a response monitoring trial. *Eur J Nucl Med Mol Imaging*. 2005;32:294–301.
19. Tyłski P, Grotus N, Giraud P, Rosenwald J, Buvat I. Experimental comparison of three methods for estimating tumor volume in FDG PET [abstract]. *J Nucl Med*. 2007;48(suppl):43P.
20. Ollinger JM. Model-based scatter correction for fully 3D PET. *Phys Med Biol*. 1996;41:153–176.
21. Jan S, Santin G, Strul D, et al. GATE: a simulation toolkit for PET and SPECT. *Phys Med Biol*. 2004;49:4543–4561.
22. Stute S, Tyłski P, Grotus N, Buvat I. LuCaS: Efficient Monte Carlo simulations of highly realistic PET tumor images. *IEEE Nucl Sci Symp Conf Rec*. 2008:4010–4012.
23. Chen CH, Myzick RF Jr, Nelson AD, Adler LP. Simultaneous recovery of size and radioactivity concentration of small spheroids with PET data. *J Nucl Med*. 1999;40:118–130.
24. Fritsch F, Carlson R. Monotone piecewise cubic interpolation. *SIAM J Numer Anal*. 1980;17:238–246.
25. Soret M, Bacharach SL, Buvat I. Partial-volume effect in PET tumor imaging. *J Nucl Med*. 2007;48:932–945.
26. Lunts A, Brailovskiy V. Evaluation of attributes obtained in statistical decision rules. *Eng Cybern*. 1967;3:982–1009.
27. Bland JM, Altman DG. Multiple significance tests: the Bonferroni method. *Br Med J*. 1995;310:170.
28. Zar JH. *Biostatistical Analysis*. 5th ed. Upper Saddle River, NJ: Pearson Education; 2009.
29. Daisne JF, Sibomana M, Bol A, Doumont T, Lonnew M, Gregoire V. Tridimensional automatic segmentation of PET volumes based on measured source-to-background ratios: influence of reconstruction algorithms. *Radiother Oncol*. 2003;69:247–250.
30. Feuardent J, Soret M, de Dreuille O, Foehrenbach H, Buvat I. Reliability of uptake estimates in FDG PET as a function of acquisition and processing protocols using the CPET. *IEEE Trans Nucl Sci*. 2005;52:1447–1452.
31. Brambilla M, Matheoud R, Secco C, Loi G, Krenqli M, Inglese E. Threshold segmentation for PET target volume delineation in radiation treatment planning: the role of target-to-background ratio and target size. *Med Phys*. 2008;35:1207–1213.
32. Geworski L, Knoop B, de Cabrejas M, Knapp W, Munz D. Recovery correction for quantitation in emission tomography: a feasibility study. *Eur J Nucl Med*. 2000;27:161–169.
33. Grotus N, Reader A, Stute S, Rosenwald J, Giraud P, Buvat I. Fully 4D list-mode reconstruction applied to respiratory-gated PET scans. *Phys Med Biol*. 2009;54:1705–1721.
34. El Naqa I, Grigsby P, Apte A, et al. Exploring feature-based approaches in PET images for predicting cancer treatment outcomes. *Pattern Recognit*. 2009;42:1162–1171.



The Journal of
NUCLEAR MEDICINE

Comparative Assessment of Methods for Estimating Tumor Volume and Standardized Uptake Value in ^{18}F -FDG PET

Perrine Tylski, Simon Stute, Nicolas Grotus, Kaya Doyeux, Sébastien Hapdey, Isabelle Gardin, Bruno Vanderlinden and Irène Buvat

J Nucl Med. 2010;51:268-276.

Published online: January 15, 2010.

Doi: 10.2967/jnumed.109.066241

This article and updated information are available at:

<http://jnm.snmjournals.org/content/51/2/268>

Information about reproducing figures, tables, or other portions of this article can be found online at:

<http://jnm.snmjournals.org/site/misc/permission.xhtml>

Information about subscriptions to JNM can be found at:

<http://jnm.snmjournals.org/site/subscriptions/online.xhtml>

The Journal of Nuclear Medicine is published monthly.
SNMMI | Society of Nuclear Medicine and Molecular Imaging
1850 Samuel Morse Drive, Reston, VA 20190.
(Print ISSN: 0161-5505, Online ISSN: 2159-662X)

© Copyright 2010 SNMMI; all rights reserved.

The logo for the Society of Nuclear Medicine and Molecular Imaging (SNMMI) consists of the letters 'S', 'N', 'M', and 'I' arranged in a 2x2 grid. Each letter is white and set within a red square. To the right of this grid, the full name of the society is written in a sans-serif font: 'SOCIETY OF NUCLEAR MEDICINE AND MOLECULAR IMAGING'.

SOCIETY OF
NUCLEAR MEDICINE
AND MOLECULAR IMAGING