

Novel Human IgG2b/Murine Chimeric Antitenascin Monoclonal Antibody Construct Radiolabeled with ^{131}I and Administered into the Surgically Created Resection Cavity of Patients with Malignant Glioma: Phase I Trial Results

David A. Reardon^{1,2}, Jennifer A. Quinn³, Gamal Akabani⁴, R. Edward Coleman⁴, Allan H. Friedman¹, Henry S. Friedman^{1,2}, James E. Herndon II⁵, Roger E. McLendon⁶, Charles N. Pegram⁶, James M. Provenzale⁴, Jeannette M. Dowell⁵, Jeremy N. Rich³, James J. Vredenburgh³, Annick Desjardins³, John H. Sampson¹, Sridharan Gururangan², Terence Z. Wong⁴, Michael A. Badrudoja³, Xiao-Guang Zhao⁶, Darell D. Bigner⁶, and Michael R. Zalutsky⁴

¹Department of Surgery, The Preston Robert Tisch Brain Tumor Center, Duke University Medical Center, Durham, North Carolina;

²Department of Pediatrics, The Preston Robert Tisch Brain Tumor Center, Duke University Medical Center, Durham, North Carolina;

³Department of Medicine, The Preston Robert Tisch Brain Tumor Center, Duke University Medical Center, Durham, North Carolina;

⁴Department of Radiology, The Preston Robert Tisch Brain Tumor Center, Duke University Medical Center, Durham, North Carolina;

⁵Cancer Center Biostatistics, The Preston Robert Tisch Brain Tumor Center, Duke University Medical Center, Durham, North Carolina;

and ⁶Department of Pathology, The Preston Robert Tisch Brain Tumor Center, Duke University Medical Center, Durham, North Carolina

Results from animal experiments have shown that human IgG2/ mouse chimeric antitenascin 81C6 (ch81C6) monoclonal antibody exhibited higher tumor accumulation and enhanced stability compared with its murine parent. Our objective was to determine the effect of these differences on the maximum tolerated dose (MTD), pharmacokinetics, dosimetry, and antitumor activity of ^{131}I -ch81C6 administered into the surgically created resection cavity (SCRC) of malignant glioma patients. **Methods:** In this phase I trial, eligible patients received a single injection of ^{131}I -ch81C6 administered through a Rickham catheter into the SCRC. Patients were stratified as newly diagnosed and untreated (stratum A), newly diagnosed after external beam radiotherapy (XRT) (stratum B), and recurrent (stratum C). ^{131}I -ch81C6 was administered either before (stratum A) or after (stratum B) conventional XRT for newly diagnosed patients. In addition, chemotherapy was prescribed for all patients after ^{131}I -ch81C6 administration. Dose escalation was performed independently for each stratum. Patients were observed for toxicity and response until death or progressive disease. **Results:** We treated 47 patients with ^{131}I -ch81C6 doses up to 4.44 GBq (120 mCi), including 35 with newly diagnosed tumors (strata A and B) and 12 with recurrent disease (stratum C). Dose-limiting hematologic toxicity defined the MTD to be 2.96 GBq (80 mCi) for all patients, regardless of treatment strata. Neurologic dose-limiting toxicity developed in 3 patients; however, none required further surgery to debulk radiation necrosis. Median survival was 88.6 wk and 65.0

wk for newly diagnosed and recurrent patients, respectively. **Conclusion:** The MTD of ^{131}I -ch81C6 is 2.96 GBq (80 mCi) because of dose-limiting hematologic toxicity. Although encouraging survival was observed, ^{131}I -ch81C6 was associated with greater hematologic toxicity, probably due to the enhanced stability of the IgG2 construct, than previously observed with ^{131}I -murine 81C6.

Key Words: radioimmunotherapy; monoclonal antibody; malignant glioma; glioblastoma multiforme; brain neoplasms

J Nucl Med 2006; 47:912–918

Although a recent phase III study demonstrated that temozolomide combined with radiotherapy improves outcome of patients with newly diagnosed glioblastoma multiforme (GBM), the vast majority of patients still die from progressive disease (PD) within 1–2 y of diagnosis (1). Salvage therapies after progression have also proven ineffective (2). Thus, there is no established therapy for patients with PD after standard external beam radiotherapy (XRT) and temozolomide chemotherapy. In addition, current treatments contribute to a limited quality of life (3).

Most patients progress at the primary tumor site (4), indicating that better local control represents a critical first step to improve overall outcome. Prior studies using regionally administered radiolabeled monoclonal antibodies (mAbs) directed against molecular targets on malignant glioma (MG) tumors have noted an encouraging rate of overall survival (5–8).

Received Dec. 6, 2005; revision accepted Feb. 11, 2006.

For correspondence or reprints contact: David A. Reardon, MD, Division of Neurosurgery, Department of Surgery, Duke University Medical Center, Box 3624, Durham, NC 27710.

E-mail: reard003@mc.duke.edu

mAb 81C6 (m81C6), a murine IgG2b, binds an isoform of tenascin C abundantly expressed in the extracellular matrix of gliomas but does not react with normal adult brain (9,10). Preclinical studies confirmed the specificity and efficacy of ^{131}I -m81C6 therapy (11–13), whereas initial clinical studies confirmed its specificity and selectivity in human subjects (14,15).

Subsequent phase I studies defined the MTD of ^{131}I -m81C6 to be 4.44 and 3.7 GBq (120 and 100 mCi) for newly diagnosed and recurrent patients, respectively, with the dose-limiting toxicity (DLT) being delayed neurologic and hematologic toxicities (6,7). A recent phase II study demonstrated a median survival of 79.4 wk among newly diagnosed GBM patients treated with ^{131}I -m81C6 (8).

Human/mouse chimeric mAbs join murine antigen-binding domains to the human immunoglobulin constant domains (16). Because of difficulties producing murine 81C6 in sufficient quantity to support a multiinstitutional randomized trial, we developed a human/mouse chimeric 81C6 mAb (ch81C6) that was capable of bulk production and that incorporated a human IgG2 constant region because of its low affinity for Fc receptors. Although the specificity and binding affinity of ch81C6 is virtually identical to m81C6, ch81C6 unexpectedly also demonstrated increased tumor uptake in human glioma xenografts and enhanced *in vivo* stability compared with m81C6 (17). Similar results in humans would suggest that ^{131}I -ch81C6 might be a better reagent than m81C6 for brain tumor–targeted radiotherapy. To investigate this hypothesis, the current phase I study was conducted to determine the MTD, dosimetry, and evidence of therapeutic benefit of ^{131}I -ch81C6.

MATERIALS AND METHODS

Patient Eligibility and Treatment

Eligibility criteria for the current trial were the same as those used in our published phase I and II studies with ^{131}I -m81C6. Briefly, eligible patients had resectable, unifocal, supratentorial, recurrent MG tumors that expressed tenascin. Additional inclusion criteria included age over 18 y, Karnofsky performance status (KPS) over 60, and adequate bone marrow, hepatic, and renal function (5). Pregnant patients and those with an iodine allergy were ineligible.

After resection and Rickham catheter placement into the surgically created resection cavity (SCRC), patency of the Rickham catheter and intactness of the SCRC were confirmed by injecting $^{99\text{m}}\text{Tc}$ -labeled albumin or diethylenetriaminepentaacetic acid into the reservoir followed by analysis of γ -camera images immediately, 4 h, and 24 h after injection. Patients with leakage of $^{99\text{m}}\text{Tc}$ -labeled tracer from the SCRC were not eligible. All patients underwent peripheral blood leukopheresis of CD34+ stem cells (PBSC) before treatment. Circulating antibodies to ch81C6 were measured before and 30–120 d after treatment (5).

Patients were stratified as newly diagnosed with no prior therapy (stratum A), newly diagnosed after XRT (stratum B), or recurrent (stratum C). The initial dose for all strata was 2.96 GBq (80 mCi) because this dose was below the MTD previously established for ^{131}I -m81C6 (6,7). Dose escalation, defined in consultation with the Food and Drug Administration, was empirically set in 0.74-GBq (20 mCi) increments and was performed independently for each stratum. To maintain optimal immunoreactivity, 20 mg of ch81C6 were used with doses of 3.7 GBq (100 mCi) or greater. A minimum of 3 patients was treated per dose level. Patients with a SCRC volume of $<5\text{ cm}^3$ were eligible for treatment; however, they were not included in the dose-escalation schema because of concern for increased toxicity risk. The absorbed doses to the cavity interface and 2-cm margin were estimated using S values as a function of cavity volume. These S values were calculated by means of β - and photon Monte Carlo transport using previously described methods (18).

Suppression of thyroid radioiodine accumulation and administration of ^{131}I -ch81C6 were performed as described (8). Patients on stratum A received XRT after ^{131}I -ch81C6. Patients on all strata also received chemotherapy for approximately 1 y using a “best clinical management” regimen determined individually, which included conventional dosing schedules of temozolomide, lomustine, irinotecan, and etoposide (Fig. 1).

The Duke Investigational Review Board approved this trial and each patient provided informed consent.

Antibody Production and Labeling

Genomic cloning was used to combine the murine 81C6 variable region genes with those for the human IgG2 constant regions (19). ch81C6 was grown in a Mini-Max hollow fiber bioreactor (Biovest Inc.) with CD Hybridoma media (Invitrogen) without serum or protein additives. Purification was by affinity chromatography over a Sepharose–staphylococcal protein-A column, followed by polyethyleneimine ion-exchange chromatography. Food and Drug Administration Manufacture and Testing

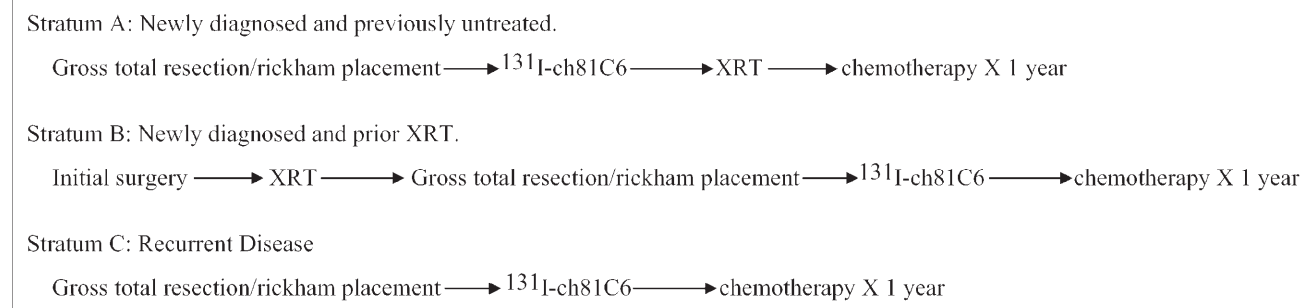


FIGURE 1. Treatment schema for each stratum.

Guidelines were followed for each clinical batch (20). ^{131}I -ch81C6, prepared using the IODO-GEN method (19), had an immunoreactivity of >75%, with >95% of the label eluting as IgG on size-exclusion high-performance liquid chromatography and precipitating with trichloroacetic acid. The dose of ^{131}I -ch81C6 was prepared the day of administration and was administered to patients within 1–3 wk of surgical resection.

Pharmacokinetics and Dosimetry

Absorbed-dose calculations for the SCRC, whole body, and bone marrow were performed as previously described using a serial, 2-compartment system in which the SCRC and body (excluding the SCRC) were assumed to be the first and second compartments, respectively. Specifically, depth-dose calculations and corresponding isodose curves for the SCRC, SCRC interface, and normal brain were performed using the 3-dimensional discrete Fourier transform convolution method based on a dose kernel for ^{131}I (21).

Toxicity and Response Determinations

Patients were monitored for toxicity (Common Toxicity Criteria database, version 2.0; National Cancer Institute; <http://ctep.cancer.gov/reporting/ctc.html>) until PD or death. Complete blood counts were performed weekly for the first 8 wk. Initial follow-up occurred 1 mo after ^{131}I -ch81C6 and then regularly thereafter as described (8). ^{18}F -FDG PET scans were obtained as needed to help determine whether progressive enhancement on follow-up MRI represented either progressive tumor or treatment-related inflammatory changes. A thyroid panel was obtained 1–2 mo after ^{131}I -ch81C6 and every 4–6 mo thereafter.

DLT was defined as either (a) attributable grade ≥ 3 nonhematologic toxicity or (b) hematologic toxicity within 6 wk of ^{131}I -ch81C6 administration consisting of >14 d of either an absolute neutrophil count of <500 cells per milliliter or a platelet count of <20,000 platelets per milliliter.

Seizures were recorded but were not considered neurologic toxicity attributable to ^{131}I -ch81C6 because of their expected occurrence in this disease setting. The precise etiology of non-seizure neurologic toxicity after ^{131}I -ch81C6 was difficult to define. Neither clinical features nor radiographic findings observed on either MRI or ^{18}F -FDG PET reliably distinguished between recurrent tumor and treatment-induced radiation necrosis. Although stereotactic needle biopsy is limited with regard to volume sampling, it remains the state of the art for diagnosis of focal brain lesions. Therefore, the etiology of observed neurologic toxicity was determined based on stereotactic needle biopsy whenever possible.

Statistical Analysis

A single-center phase I study with a classical “3 + 3” design was performed to determine the MTD of ^{131}I -ch81C6 (6). Kaplan–Meier survival distributions were estimated from initial treatment to either death or last contact (22). Logistic regression was used to examine the effect of cavity size as well as ^{131}I -ch81C6 absorbed and cumulative (^{131}I -ch81C6 plus XRT) radiation doses to the 2-cm-thick SCRC interface on toxicity.

RESULTS

Patient Characteristics

Forty-seven patients accrued between October 1999 and February 2002. Patient characteristics were comparable among the 3 strata (Table 1). All stratum C patients received prior radiation therapy and 9 of these patients received prior

TABLE 1
Patient Characteristics on Phase I ^{131}I -ch81C6

Characteristic	Stratum A (n = 19)	Stratum B (n = 16)	Stratum C (n = 12)	Total (n = 47)
Age median/ range (y)	55/30–70	52/36–66	52/28–69	52/28–70
Sex				
Male	11 (58)	8 (50)	7 (58)	26 (55)
Female	8 (42)	8 (50)	5 (42)	21 (45)
KPS				
100	17 (89)	12 (75)	5 (42)	34 (72)
90	1 (5)	3 (19)	4 (33)	8 (17)
80	1 (5)	0	2 (17)	3 (6)
70	0	0	1 (8)	1 (2)
60	0	1 (6)	0	1 (2)
Histology				
GBM	16 (84)	14 (88)	8 (67)	38 (81)
AA	3 (16)	1 (6)	3 (25)	7 (15)
AO	0	1 (6)	1 (8)	2 (4)

AA = anaplastic astrocytoma; AO = anaplastic oligodendroglioma.

Values in parentheses are percentages.

chemotherapy. The median number of episodes of prior PD for patients enrolled on stratum C was 1 (range, 1–2). The median number of prior chemotherapy agents administered to these patients was 1 (range, 0–4).

Dosimetry Results

Dosimetry findings for patients on the 3 strata were comparable and displayed significant interpatient variability. The average SCRC volumes were 21 cm³ (range, 2–81 cm³), 17 cm³ (range, 2–65 cm³), and 18 cm³ (range, 2–57 cm³), whereas the average (range) ^{131}I -ch81C6 SCRC residence times were 79 h (10–113 h), 85 (24–177 h), and 86 h (54–161 h) for patients on strata A, B, and C, respectively. The average absorbed dose to the 2-cm SCRC margin was 32 Gy (3–59 Gy), 45 Gy (9–97 Gy), and 40 Gy (18–110 Gy) among patients enrolled on strata A, B, and C, respectively. The average (range) residence time for the whole body, excluding the SCRC, for all patients was 69 h (48–106 h).

Toxicity

The MTD established for all 3 strata was 2.96 GBq (80 mCi), due primarily to hematologic toxicity (Table 2). Reversible, grade 4 hematologic toxicity developed in 17 patients (36%) within 6 wk of ^{131}I -ch81C6 administration, including 9 patients on stratum A, 5 patients on stratum B, and 3 patients on stratum C. Criteria for hematologic DLT were met in 7 patients (15%), all of whom received an ^{131}I -ch81C6 dose that exceeded the MTD. The median duration of grade 4 hematologic toxicity among patients who did not meet criteria for DLT was 6 d (range, 3–14 d),

TABLE 2
Treatment Levels and Toxicity

Stratum	<2.96 GBq (<80 mCi)			2.96 GBq (80 mCi)			3.7 GBq (100 mCi)		
	No. Treated	No. with DLT*	Type DLT*	No. Treated	No. with DLT*	Type DLT*	No. Treated	No. with DLT*	Type DLT*
A (n = 19)	2	2 (100)	Hematologic; MDS [†]	8	0	None	9	3 (33)	Hematologic; MDS [†]
B (n = 16)	3	2 (67)	Neuromotor; aphasia	10	0	None	3	2 (67)	Hematologic
C (n = 12)	0	0	None	6	0	None	6 [‡]	3 (50)	Hematologic; aphasia
Total	5	4 (80)	Hematologic; MDS [†] ; neuromotor	24	0	None	18	7 (39)	Hematologic; MDS [†] ; aphasia

*DLT: (a) grade 3 or greater nonhematologic toxicity attributable to ¹³¹I-ch81C6; or (b) hematologic toxicity occurring within 6 wk of ¹³¹I-ch81C6 administration and consisting of >14 d of either an absolute neutrophil count of <500 cells per milliliter or a platelet count of <20,000 platelets per milliliter.

[†]MDS = myelodysplasia.

[‡]Includes 1 patient inadvertently treated with 4.44 GBq (120 mCi) who did not experience DLT.

Values in parentheses are percentages.

whereas the median duration for those who met DLT criteria was 20 d (range, 18–31 d). Six patients received PBSC reinfusion because of acute hematologic toxicity associated with ¹³¹I-ch81C6 administration.

Neurologic DLT occurred in 3 patients (6%) and seemed to be related to a small SCRC volume (<5 cm³) or proximity to critical functional centers in the brain. Reversible grade 3 aphasia developed in 1 patient on stratum B the day after receiving 2.22 GBq (60 mCi) of ¹³¹I-ch81C6. This patient remains alive with a good quality of life 6.3 y from diagnosis of an anaplastic astrocytoma (AA). Irreversible neurologic toxicity developed in 2 patients (4%). Irreversible grade 3 left-sided weakness developed in 1 stratum B patient approximately 4 mo after administration of 1.85 GBq (50 mCi) of ¹³¹I-ch81C6. A stereotactic biopsy performed on this patient revealed gliosis and necrosis. Unfortunately, this patient died acutely of a presumed pulmonary embolism approximately 9 mo after ¹³¹I-ch81C6 administration. Grade 3 aphasia developed in 1 stratum C patient with recurrent AA 8 mo after receiving 3.7 GBq (100 mCi) of ¹³¹I-ch81C6, which gradually resolved over the next year. He remains alive approximately 5.4 y from original diagnosis.

Grade 2 hypothyroidism developed in 4 patients (9%) 4–33 mo after ¹³¹I-ch81C6, including 2 treated at the 3.7-GBq (100 mCi) level and 2 treated with 2.96 GBq (80 mCi). All of these patients also received prior XRT. One patient experienced an initial seizure after ¹³¹I-ch81C6 therapy; however, the overall frequency and severity of seizures experienced by study patients were within the expected range for this patient population. No significant hepatic, renal, or other organ toxicities attributable to therapy with ¹³¹I-ch81C6 were observed.

Of note, myelodysplasia (MDS) developed in 2 patients with newly diagnosed GBM. Grade 2 leukopenia, neutropenia, and thrombocytopenia developed in 1 patient treated with 3.7 GBq (100 mCi) of ¹³¹I-ch81C6 followed by XRT approximately 4 mo after ¹³¹I-ch81C6 therapy. A bone marrow examination 10 mo after ¹³¹I-ch81C6 revealed moder-

ate hypocellularity (20%) but no dyspoiesis and <3% blasts. Flow cytometry was unremarkable, but cytogenetic examination revealed 46,XX,t(3;21)(q26;q22) and fluorescence in situ hybridization analysis showed rearrangement of the *AML1* locus. A histocompatibility locus antigen (HLA)-matched, sibling, nonmyeloablative, T-cell depleted, allogeneic stem cell transplantation was performed and 99% donor engraftment with a normal karyotype was noted 2 mo after transplantation. However, biopsy-proven recurrent GBM developed and she died 4 mo after transplantation. Persistent pancytopenia developed in the second patient approximately 26 mo after administration of 2.22 GBq (60 mCi) of ¹³¹I-ch81C6, followed by conventional XRT and 10 cycles of systemic chemotherapy, including temozolomide (4 cycles), lomustine (2 cycles), and irinotecan (4 cycles). Bone marrow examination revealed clonal rearrangements, including monosomy 7 and deletion of 11q23. Fatal pneumonia developed in this patient 32 mo from original diagnosis.

Human Antimouse Antibody (HAMA)

The serum of 19 of 41 evaluable patients (46%) reacted with ch81C6. However, all 25 available patient serum samples tested against murine single-fragment-chain region were negative. No observed toxicity was related to HAMA reactivity.

Biopsies and Further Surgery

Twenty-seven patients (57%) underwent 30 surgical procedures for progressive clinical or radiographic changes, including 25 stereotactic biopsies and 5 resections. Seventeen biopsies revealed gliosis, whereas 8 confirmed recurrent tumor. Four patients had recurrent tumor on resection, whereas 1 asymptomatic patient had only gliosis at the time of resection. Five patients who had an initial biopsy demonstrating gliosis underwent a second surgical procedure. Repeated biopsy in 2 patients confirmed gliosis, whereas the remaining 3 patients had recurrent tumor.

Pattern of Recurrence

PD was local in most cases. Distant recurrence occurred in 14 patients (30%), including 9 patients with simultaneous local and distant progression and 5 patients with solely distant recurrence.

Response/Survival Data

Survival was the most important criterion for efficacy because all patients underwent total or near-total resection leaving little or no residual tumor. With a median follow-up of 185 wk, the median survival for all newly diagnosed patients, those with GBM, and those with World Health Organization (WHO) grade III tumors was 88.6 wk (95% confidence interval [CI], 72.9; 128.7 wk), 86.1 wk (95% CI, 70.1; 99.1 wk), and 230.3 wk (95% CI, 88.6 to not estimable), respectively (Fig. 2A). Of note, there was no significant difference in outcome between stratum A and stratum B patients.

With a median follow-up of 228 wk, the median survival for all patients with recurrent disease, those with GBM, and those with WHO grade III tumors was 65 wk (95% CI, 45.0; 142.4 wk), 48.9 wk (95% CI, 30.4; 83.3 wk), and not

estimable, respectively (Fig. 2B). Figure 3 depicts representative radiographic findings after ^{131}I -ch81C6.

DISCUSSION

Recurrence immediately adjacent to or at the primary tumor site occurs predictably in nearly all MG patients following current standard therapies, indicating that local control remains elusive and represents the critical first step required to improve overall outcome (4,23).

Radioimmunotherapy, consisting of radiolabeled mAbs targeting tumor antigens, is associated with encouraging survival and acceptable toxicity for patients with both newly diagnosed and recurrent MG (24–29). Our group has extensively evaluated the clinical activity of radioimmunotherapy using ^{131}I -m81C6 administered directly into the SCRC for MG patients. In previously published phase I studies, dose-limiting neurotoxicity defined the MTD of ^{131}I -m81C6 to be 3.7 and 4.44 GBq (100 and 120 mCi) for patients with recurrent and newly diagnosed MG, respectively. Of note, only 1 of the 76 patients treated on these 2 protocols (1.3%) required further surgery to debulk radionecrosis. In addition, patients with recurrent and newly diagnosed GBM treated on our phase I studies achieved median survivals of 56 and 69 wk, respectively (6,7), whereas patients with newly diagnosed GBM treated with 4.44 GBq (120 mCi) of ^{131}I -m81C6 in a recently completed phase II study achieved a median survival of 79.4 wk (8).

However, in the past, sufficient m81C6 for clinical use could be produced only in ascites of athymic mice because of instability of the murine 81C6 hybridoma. Although that difficulty has been overcome recently, human/mouse chimeric 81C6 mAb (ch81C6) was developed by combining the variable region genes of m81C6 with human IgG2 constant region domains, to have a stable hybridoma capable of producing 81C6 in cell culture (17). The human IgG2 constant region was selected because IgG2 exhibits the lowest Fc receptor affinity of human IgG, thereby minimizing potential reactivity with normal tissues. There are few clinical studies of human IgG2 constructs and, to our knowledge, the current study may be the first involving use of this type of construct as a carrier for therapeutic levels of radionuclide.

Although ch81C6 has the same reactivity with tenascin as m81C6 and exhibits an identical affinity constant for recombinant tenascin CD fragment, paired-label experiments demonstrated a significantly higher tumor uptake of ch81C6 in both subcutaneous and intracranial xenografts compared with m81C6. This behavior is likely attributed to enhanced *in vivo* stability of ^{131}I -ch81C6 (30).

Because of its greater stability, we hypothesized that ch81C6 might remain in the SCRC longer than its murine counterpart, thereby increasing delivered radiation dose. The average residence time for ^{131}I -ch81C6 for all patients in the current study was 83 h, which is longer than that seen previously for m81C6 (7,8). Furthermore, comparable clinical

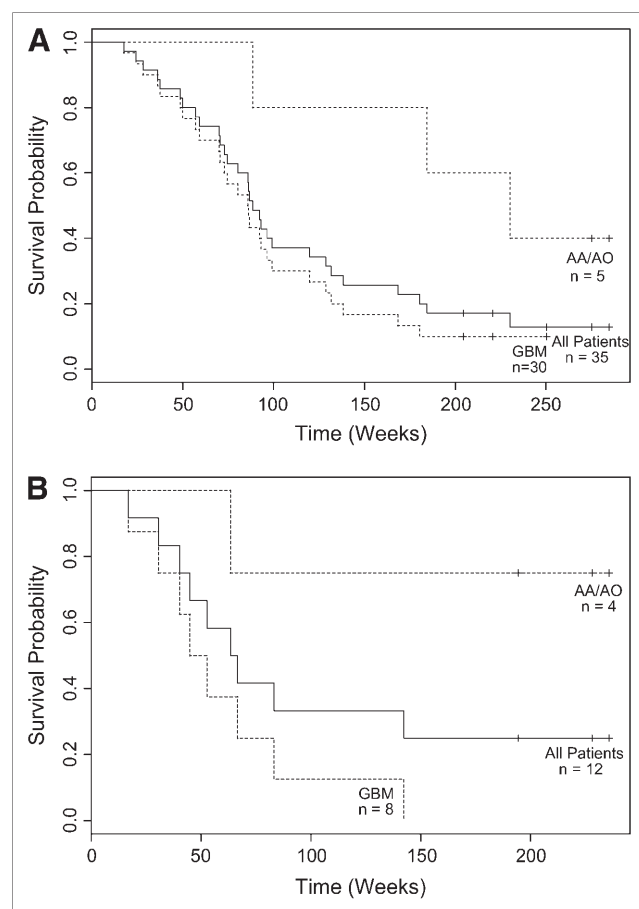


FIGURE 2. Kaplan-Meier overall survival estimates for newly diagnosed patients (A) and recurrent patients (B) after stratification by histology. AO = anaplastic oligodendroglioma.

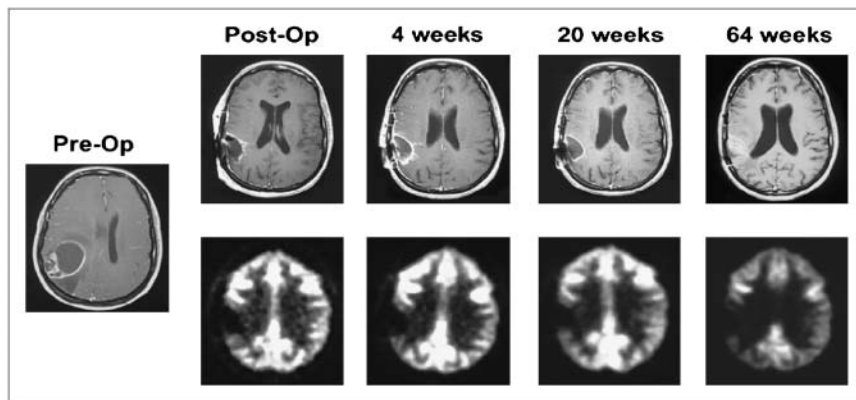


FIGURE 3. Serial MRI (top and middle) and ^{18}F -FDG PET scan results of representative patient after ^{131}I ch81C6 therapy. Corresponding ^{18}F -FDG PET scan images (bottom) demonstrate a lack of increased metabolic activity in region of SCRC.

benefit was observed with ^{131}I -ch81C6 at lower administered activity levels than those used previously with m81C6 (8).

In the current study, we determined the MTD of ^{131}I -ch81C6 to be 2.96 GBq (80 mCi) for patients with either newly diagnosed or recurrent MG. The DLT was hematologic in most cases. The frequency and severity of hematologic toxicity associated with ^{131}I -ch81C6 increased significantly at doses above the MTD, regardless of prior treatment status, whereas hematologic DLT was not observed among patients treated at the MTD. The lower MTD and higher hematologic toxicity for ^{131}I -ch81C6 compared with its murine parent is consistent with the prolonged retention of the chimeric construct in the whole body. Of note, the average whole-body residence time determined for ^{131}I -ch81C6, 69 h, was nearly twice that determined previously for ^{131}I -m81C6 (7,8,21). Neurotoxicity developed in 3 patients, including 2 with a SCRC volume of $<5\text{ cm}^3$, suggesting that additional modifications are required to safely administer ^{131}I -ch81C6 to patients with small SCRC volumes. However, none of the patients on this study required further surgery for symptomatic radionecrosis.

It is possible that ^{131}I exposure contributed to the cases of MDS observed in our study. Secondary cancers, including leukemia and MDS, have been noted after radiotherapy with ^{131}I for thyroid disorders (31,32). In addition, cases of secondary MDS have been recently reported among lymphoma patients treated with ^{131}I -labeled mAbs (33). The cytogenetic abnormalities observed in our patients who developed secondary MDS are associated with therapy-induced hematologic malignancies (34). Of note, we have not observed secondary leukemia or MDS among >300 patients with MG treated with intracavitary ^{131}I -m81C6. It is likely that the hematologic toxicities observed with ^{131}I -ch81C6 result from the long plasma half-life of the human IgG2 construct, which, in turn, yields greater radiation dose to the bone marrow than ^{131}I -m81C6.

The median survival achieved among newly diagnosed and recurrent patients treated in the current study with ^{131}I -ch81C6 therapy is encouraging and compares favorably with “standard of care” treatments (1,2,35,36). However, this result, as well as those of our previously published trials incorporating ^{131}I -m81C6, must be interpreted with caution.

Our current and prior studies are limited by enrollment of patients with favorable prognostic features and by a single-institutional study design. Definitive conclusions regarding the efficacy of radioimmunotherapy approaches such as ours will only be possible after the completion of randomized, multiinstitutional trials.

A question that often arises is the extent to which radioimmunotherapy provides a tangible advantage in efficacy compared with brachytherapy. An important factor is the extent to which the labeled mAb penetrates beyond the cavity margin, potentially providing a means of irradiating tumor cells infiltrating proximal normal brain. Unfortunately, this important question cannot be addressed adequately using existent animal brain tumor models and it is unethical to subject human patients to repeated, invasive procedures to address this question. Thus, current information on this subject is limited. Using multiple biopsies obtained at autopsy, Hopkins et al. demonstrated that mAb penetration is much greater than expected, with a mean R_0 of 0.6 cm (37). Furthermore, in a recent dosimetry and SPECT study, we have shown that ^{131}I -m81C6 can penetrate into areas of edema associated with the resection cavity of patients with malignant tumors of the central nervous system (38). Penetration of radiolabeled, tumor-antigen-specific, mAbs into areas of adjacent edema may provide a potential advantage of radioimmunotherapy over other approaches to boost radiation delivered to the resection cavity perimeter such as stereotactic radiosurgery, which has failed to demonstrate a survival advantage for newly diagnosed MG patients in a randomized, phase III setting (39), and interstitial brachytherapy with ^{125}I -labeled interstitial beads (40,41).

CONCLUSION

The encouraging median survival observed on the current study compares favorably with that achieved with ^{131}I -m81C6; however, the clinical benefit of ^{131}I -ch81C6 was partially offset by increased hematologic toxicity. The efficacy, as well as the toxicity of ^{131}I -ch81C6, most likely reflects the enhanced stability of the human IgG₂ construct. Because we now have stabilized the m81C6 hybridoma and can produce

amounts of m81C6 needed for phase III multiinstitutional trials, no further studies with ^{131}I -ch81C6 are planned.

ACKNOWLEDGMENTS

This research was supported by National Institutes of Health grants 1-P50-CA108786-01, NS20023, and CA11898 and by grant MO1 RR 30 through the General Clinical Research Centers Program, National Center for Research Resources, National Institutes of Health.

REFERENCES

- Stupp R, Mason WP, van den Bent MJ, et al. Radiotherapy plus concomitant and adjuvant temozolomide for glioblastoma. *N Engl J Med*. 2005;352:987–996.
- Wong ET, Hess KR, Gleason MJ, et al. Outcomes and prognostic factors in recurrent glioma patients enrolled onto phase II clinical trials. *J Clin Oncol*. 1999;17:2572–2578.
- Imperato JP, Paleologos NA, Vick NA. Effects of treatment on long-term survivors with malignant astrocytomas. *Ann Neurol*. 1990;28:818–822.
- Gaspar LE, Fisher BJ, Macdonald DR, et al. Supratentorial malignant glioma: patterns of recurrence and implications for external beam local treatment. *Int J Radiat Oncol Biol Phys*. 1992;24:55–57.
- Bigner DD, Brown M, Coleman RE, et al. Phase I studies of treatment of malignant gliomas and neoplastic meningitis with ^{131}I -radiolabeled monoclonal antibodies anti-tenascin 81C6 and anti-chondroitin proteoglycan sulfate Me1-14 F(ab')₂: a preliminary report. *J Neurooncol*. 1995;24:109–122.
- Bigner DD, Brown MT, Friedman AH, et al. Iodine-131-labeled antitenascin monoclonal antibody 81C6 treatment of patients with recurrent malignant gliomas: phase I trial results. *J Clin Oncol*. 1998;16:2202–2212.
- Cokgor I, Akabani G, Kuan CT, et al. Phase I trial results of iodine-131-labeled antitenascin monoclonal antibody 81C6 treatment of patients with newly diagnosed malignant gliomas. *J Clin Oncol*. 2000;18:3862–3872.
- Reardon DA, Akabani G, Coleman RE, et al. Phase II trial of murine ^{131}I -labeled antitenascin monoclonal antibody 81C6 administered into surgically created resection cavities of patients with newly diagnosed malignant gliomas. *J Clin Oncol*. 2002;20:1389–1397.
- Bourdon MA, Wikstrand CJ, Furthmayr H, Matthews TJ, Bigner DD. Human glioma-mesenchymal extracellular matrix antigen defined by monoclonal antibody. *Cancer Res*. 1983;43:2796–2805.
- Bourdon MA, Matthews TJ, Pizzo SV, Bigner DD. Immunochemical and biochemical characterization of a glioma-associated extracellular matrix glycoprotein. *J Cell Biochem*. 1985;28:183–195.
- Bourdon MA, Coleman RE, Blasberg RG, Groothuis DR, Bigner DD. Monoclonal antibody localization in subcutaneous and intracranial human glioma xenografts: paired-label and imaging analysis. *Anticancer Res*. 1984;4:133–140.
- Lee YS, Bullard DE, Wikstrand CJ, Zalutsky MR, Muhlbaier LH, Bigner DD. Comparison of monoclonal antibody delivery to intracranial glioma xenografts by intravenous and intracarotid administration. *Cancer Res*. 1987;47:1941–1946.
- Lee YS, Bullard DE, Zalutsky MR, et al. Therapeutic efficacy of anti-glioma mesenchymal extracellular matrix ^{131}I -radiolabeled murine monoclonal antibody in a human glioma xenograft model. *Cancer Res*. 1988;48:559–566.
- Zalutsky MR, Moseley RP, Coakham HB, Coleman RE, Bigner DD. Pharmacokinetics and tumor localization of ^{131}I -labeled anti-tenascin monoclonal antibody 81C6 in patients with gliomas and other intracranial malignancies. *Cancer Res*. 1989;49:2807–2813.
- Schold SC Jr, Zalutsky MR, Coleman RE, et al. Distribution and dosimetry of I-123-labeled monoclonal antibody 81C6 in patients with anaplastic glioma. *Invest Radiol*. 1993;28:488–496.
- Bouliaume GL, Hozumi N, Shulman MJ. Production of functional chimaeric mouse/human antibody. *Nature*. 1984;312:643–646.
- He X, Archer GE, Wikstrand CJ, et al. Generation and characterization of a mouse/human chimeric antibody directed against extracellular matrix protein tenascin. *J Neuroimmunol*. 1994;52:127–137.
- Akabani G, Poston JW Sr, Bolch WE. Estimates of beta absorbed fractions in small tissue volumes for selected radionuclides. *J Nucl Med*. 1991;32:835–839.
- Zalutsky MR, Archer GE, Garg PK, Batra SK, Bigner DD. Chimeric antitenascin antibody 81C6: increased tumor localization compared with its murine parent. *Nucl Med Biol*. 1996;23:449–458.
- U.S. Department of Health and Human Services, Food and Drug Administration, Center for Biologics Evaluation and Research: Points to Consider in the Manufacture and Testing of Monoclonal Antibody Products for Human Use. Rockville, MD. February 28, 1997.
- Akabani G, Reist CJ, Cokgor I, et al. Dosimetry of ^{131}I -labeled 81C6 monoclonal antibody administered into surgically created resection cavities in patients with malignant brain tumors. *J Nucl Med*. 1999;40:631–638.
- Kaplan E, Meier P. Nonparametric estimation from incomplete observations. *J Am Stat Assoc*. 1958;53:457–481.
- Hochberg FH, Pruitt A. Assumptions in the radiotherapy of glioblastoma. *Neurology*. 1980;30:907–911.
- Paganelli G, Bartolomei M, Ferrari M, et al. Pre-targeted locoregional radioimmunotherapy with ^{90}Y -biotin in glioma patients: phase I study and preliminary therapeutic results. *Cancer Biother Radiopharm*. 2001;16:227–235.
- Bartolomei M, Mazzetta C, Handkiewicz-Junak D, et al. Combined treatment of glioblastoma patients with locoregional pre-targeted ^{90}Y -biotin radioimmunotherapy and temozolomide. *Q J Nucl Med Mol Imaging*. 2004;48:220–228.
- Riva P, Franceschi G, Frattarelli M, et al. Loco-regional radioimmunotherapy of high-grade malignant gliomas using specific monoclonal antibodies labeled with ^{90}Y : a phase I study. *Clin Cancer Res*. 1999;5(10 suppl):3275s–3280s.
- Riva P, Franceschi G, Arista A, et al. Local application of radiolabeled monoclonal antibodies in the treatment of high grade malignant gliomas: a six-year clinical experience. *Cancer*. 1997;80(12 suppl):2733–2742.
- Brady LW, Miyamoto C, Woo DV, et al. Malignant astrocytomas treated with iodine-125 labeled monoclonal antibody 425 against epidermal growth factor receptor: a phase II trial. *Int J Radiat Oncol Biol Phys*. 1992;22:225–230.
- Paganelli G, Grana C, Chinol M, et al. Antibody-guided three-step therapy for high grade glioma with yttrium-90 biotin. *Eur J Nucl Med*. 1999;26:348–357.
- Reist CJ, Bigner DD, Zalutsky MR. Human IgG2 constant region enhances in vivo stability of anti-tenascin antibody 81C6 compared with its murine parent. *Clin Cancer Res*. 1998;4:2495–2502.
- Hall P, Boice JD Jr, Berg G, et al. Leukaemia incidence after iodine-131 exposure. *Lancet*. 1992;340:1–4.
- de Vathaire F, Schlumberger M, Delisle MJ, et al. Leukaemias and cancers following iodine-131 administration for thyroid cancer. *Br J Cancer*. 1997;75:734–739.
- Kaminski MS, Estes J, Zasadny KR, et al. Radioimmunotherapy with iodine ^{131}I tositumomab for relapsed or refractory B-cell non-Hodgkin lymphoma: updated results and long-term follow-up of the University of Michigan experience. *Blood*. 2000;96:1259–1266.
- Roulston D, Espinosa R 3rd, Nucifora G, Larson RA, Le Beau MM, Rowley JD. CBFA2(AML1) translocations with novel partner chromosomes in myeloid leukemias: association with prior therapy. *Blood*. 1998;92:2879–2885.
- Yung WK, Albright RE, Olson J, et al. A phase II study of temozolomide vs. procarbazine in patients with glioblastoma multiforme at first relapse. *Br J Cancer*. 2000;83:588–593.
- Yung WK, Prados MD, Yaya-Tur R, et al. Multicenter phase II trial of temozolomide in patients with anaplastic astrocytoma or anaplastic oligoastrocytoma at first relapse: Temodal Brain Tumor Group. *J Clin Oncol*. 1999;17:2762–2771.
- Hopkins K, Chandler C, Eatough J, Moss T, Kemshead JT. Direct injection of ^{90}Y MoAbs into glioma tumor resection cavities leads to limited diffusion of the radioimmunconjugates into normal brain parenchyma: a model to estimate absorbed radiation dose. *Int J Radiat Oncol Biol Phys*. 1998;40:835–844.
- Akabani G, Reardon DA, Coleman RE, et al. Dosimetry and radiographic analysis of ^{131}I -labeled anti-tenascin 81C6 murine monoclonal antibody in newly diagnosed patients with malignant gliomas: a phase II study. *J Nucl Med*. 2005;46:1042–1051.
- Souhami L, Seiferheld W, Brachman D, et al. Randomized comparison of stereotactic radiosurgery followed by conventional radiotherapy with carmustine to conventional radiotherapy with carmustine for patients with glioblastoma multiforme: report of Radiation Therapy Oncology Group 93-05 protocol. *Int J Radiat Oncol Biol Phys*. 2004;60:853–860.
- Loeffler JS, Alexander E 3rd, Wen PY, et al. Results of stereotactic brachytherapy used in the initial management of patients with glioblastoma. *J Natl Cancer Inst*. 1990;82:1918–1921.
- Shrieve DC, Alexander E 3rd, Wen PY, et al. Comparison of stereotactic radiosurgery and brachytherapy in the treatment of recurrent glioblastoma multiforme. *Neurosurgery*. 1995;36:275–282.



The Journal of
NUCLEAR MEDICINE

Novel Human IgG2b/Murine Chimeric Antitenascin Monoclonal Antibody Construct Radiolabeled with ^{131}I and Administered into the Surgically Created Resection Cavity of Patients with Malignant Glioma: Phase I Trial Results

David A. Reardon, Jennifer A. Quinn, Gamal Akabani, R. Edward Coleman, Allan H. Friedman, Henry S. Friedman, James E. Herndon II, Roger E. McLendon, Charles N. Pegram, James M. Provenzale, Jeannette M. Dowell, Jeremy N. Rich, James J. Vredenburgh, Annick Desjardins, John H. Sampson, Sridharan Gururangan, Terence Z. Wong, Michael A. Badruddoja, Xiao-Guang Zhao, Darell D. Bigner and Michael R. Zalutsky

J Nucl Med. 2006;47:912-918.

This article and updated information are available at:
<http://jnm.snmjournals.org/content/47/6/912>

Information about reproducing figures, tables, or other portions of this article can be found online at:
<http://jnm.snmjournals.org/site/misc/permission.xhtml>

Information about subscriptions to JNM can be found at:
<http://jnm.snmjournals.org/site/subscriptions/online.xhtml>

The Journal of Nuclear Medicine is published monthly.
SNMMI | Society of Nuclear Medicine and Molecular Imaging
1850 Samuel Morse Drive, Reston, VA 20190.
(Print ISSN: 0161-5505, Online ISSN: 2159-662X)

© Copyright 2006 SNMMI; all rights reserved.

The logo for the Society of Nuclear Medicine and Molecular Imaging (SNMMI) consists of the letters 'S', 'N', 'M', and 'I' arranged in a 2x2 grid, each within its own red square. To the right of this graphic, the full name of the society is written in a sans-serif font.
SOCIETY OF
NUCLEAR MEDICINE
AND MOLECULAR IMAGING